

# Package ‘scCrossTalk’

April 14, 2023

**Title** Infer and visualize cell-cell communication for single-cell RNA-seq data

**Version** 1.0

**Depends** R (>= 4.0.0)

**Description**

This package can infer and visualize cell-cell communication for single-cell RNA-seq data.

**License** GPL (>= 3)

**Encoding** UTF-8

**LazyData** true

**Roxygen** list(markdown = TRUE)

**RoxygenNote** 7.1.2

**Imports** Seurat,

Matrix,  
graphics,  
ggplot2,  
ggrepel,  
methods,  
pheatmap,  
progress,  
reshape2,  
crayon,  
utils,  
scales,  
ggraph,  
igraph,  
circlize,  
heatmaply,  
networkD3,  
viridis

**Suggests** rmarkdown,  
knitr,  
prettydoc

**VignetteBuilder** knitr

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create_scCrossTalk	<i>Create scCrossTalk object</i>
--------------------	----------------------------------

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## Description

create scCrossTalk object using single-cell transcriptomics data

## Usage

```
create_scCrossTalk(sc_data, sc_celltype, species, if_normalize = TRUE)
```

## Arguments

sc_data	A data.frame or matrix or dgCMatrx containing raw counts of single-cell RNA-seq data. see <a href="#">demo_sc_data</a>
sc_celltype	A character containing the cell type of the single-cell RNA-seq data
species	A character meaning species of the single-cell transcriptomics data. 'Human' or 'Mouse'.
if_normalize	Normalize sc_data with Seurat LogNormalize. Set it FALSE when sc_data has been normalized.

## Value

scCrossTalk object

---

`demo_geneinfo`*Demo data of geneinfo*

---

**Description**

Demo data of geneinfo

**Usage**

```
demo_geneinfo()
```

**Details**

geneinfo must be a data.frame object with three columns, namely 'symbol', 'synonyms', 'species'.

**Examples**

```
geneinfo_demo <- demo_geneinfo()
```

---

`demo_lrpairs`*Demo data of lrpairs*

---

**Description**

Demo data of lrpairs

**Usage**

```
demo_lrpairs()
```

**Details**

lrpairs must be a data.frame object with three columns, namely 'ligand', 'receptor', 'species'.

**Value**

A data.frame.

**Examples**

```
lrpairs_demo <- demo_lrpairs()
```

---

demo_sc_data	<i>Demo data of sc_data</i>
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---

### Description

Demo data of sc\_data.

### Usage

```
demo_sc_data()
```

### Details

sc\_data can be a data.frame, matrix, or dgCMatrx object, each column representing a cell, each row representing a gene.

### Value

A dgCMatrx object.

### Examples

```
sc_data_demo <- demo_sc_data()
```

---

find_lrpairs	<i>Find highly expressed ligand-receptor pairs</i>
--------------	--

---

### Description

Find highly expressed ligands and receptors between pairwise cell types using Z score

### Usage

```
find_lrpairs(
  object,
  lrpairs,
  min_cell_num = 10,
  cell_min_pct = 0.1,
  p_value = 0.05
)
```

### Arguments

object	scCrossTalk object after <a href="#">create_scCrossTalk</a>
lrpairs	A data.frame of the system data containing ligand-receptor pairs of 'Human' and 'Mouse'. see <a href="#">demo_lrpairs</a>
min_cell_num	Min cell number for each cell type. Default is 10
cell_min_pct	Include the ligand and receptor gene detected in at least this many cells in each cluster. Default is 0.1
p_value	Include the significantly highly expressed ligands and receptors with this cutoff of p value from Z score. Default is 0.05

**Value**

scCrossTalk object containing the enriched ligand-receptor pairs and cell-cell communications

---

geneinfo	<i>geneinfo</i>
----------	-----------------

---

**Description**

A data.frame containing gene symbols of "Human" and "Mouse" updated on June 19, 2022 for revising gene symbols.

**Usage**

```
geneinfo
```

**Format**

An object of class data.frame with 288559 rows and 3 columns.

**Source**

<https://www.ncbi.nlm.nih.gov/gene>

---

get_top_lrpairs	<i>Get top n LR-pairs</i>
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---

**Description**

Get top n LR-pairs for each cell pairs

**Usage**

```
get_top_lrpairs(object, top_n = 10)
```

**Arguments**

object	scCrossTalk object after <a href="#">create_scCrossTalk</a>
top_n	Number of top LR-pairs. Default is 10

**Value**

A data.frame containing the top n LR-pairs for each cell pairs

---

lrpairs	<i>lrpairs</i>
---------	----------------

---

### Description

A data.frame containing ligand-receptor interactions for "Human" and "Mouse" from CellTalkDB.

### Usage

```
lrpairs
```

### Format

An object of class data.frame with 5427 rows and 3 columns.

### Source

<http://tcm.zju.edu.cn/celltalkdb/>

---

plot_cci_chord	<i>Chord plot of cell-cell communications</i>
----------------	---

---

### Description

Chord plot of cell-cell communications from senders to receivers with the sum of inferred number or score of ligand-receptor interactions

### Usage

```
plot_cci_chord(
  object,
  celltype = NULL,
  celltype_color = NULL,
  ligand = NULL,
  receptor = NULL,
  edge_color = NULL,
  edge_type = "big.arrow",
  show_type = "number",
  if_show_autocrine = FALSE,
  text_size = 1.5,
  y_scale = 0.1,
  ...
)
```

**Arguments**

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype	which cell types to plot by order. Default is to plot all cell types
celltype_color	Colors for the cell types, whose length must be equal to celltype
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
edge_color	Colors for the edges from the sender cell type, whose length must be equal to celltype
edge_type	Types for the edges from the sender cell type. Default is "big.arrow". "ellipse" for ellipse, "triangle" for triangle, "curved" for curved. Details see <a href="#">chordDiagram</a>
show_type	which type to show, "number" and "score" for sum of inferred LR number and score, respectively. Default is "number"
if_show_autocrine	Whether to show autocrine. Default is FALSE
text_size	Size of text labels. Default is 1.5
y_scale	y_scale to adjust the text. Default is 0.1
...	parameters pass to <a href="#">chordDiagram</a> , e.g., link.arr.width, link.arr.length, link.arr.col

**Value**

Chord plot of cell-cell communications mediated by ligand-receptor interactions

---

plot_cci_circle	<i>Circle plot of cell-cell communications</i>
-----------------	--

---

**Description**

Circle plot of cell-cell communications from senders to receivers with the sum of inferred number or score of ligand-receptor interactions

**Usage**

```
plot_cci_circle(
  object,
  celltype = NULL,
  ligand = NULL,
  receptor = NULL,
  celltype_color = NULL,
  edge_color = NULL,
  edge_type = "fan",
  show_type = "number",
  if_show_autocrine = FALSE,
  edge_alpha = 0.5,
  node_size = 10,
  text_size = 5
)
```

**Arguments**

object	scCrossTalk object after <code>find_lrpairs</code>
celltype	which cell types to plot. Default is to plot all cell types
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
celltype_color	Colors for the cell types, whose length must be equal to celltype
edge_color	Colors for the edges from the sender cell type, whose length must be equal to celltype
edge_type	Types for the edges. "fan" by default, "link", "hive"
show_type	which type to show, "number" and "score" for sum of inferred LR number and score, respectively. Default is "number"
if_show_autocrine	Whether to show autocrine. Default is FALSE
edge_alpha	Transparency of edge. Default is 0.5
node_size	Size of node. Default is 10
text_size	Size of text. Default is 5

**Value**

Circle plot of cell-cell communications mediated by ligand-receptor interactions

---

plot_cci_heatmap	<i>Heatmap plot of cell-cell communications</i>
------------------	---

---

**Description**

Heatmap plot of cell-cell communications from senders to receivers with the sum of inferred number or score of ligand-receptor interactions

**Usage**

```
plot_cci_heatmap(
  object,
  celltype = NULL,
  ligand = NULL,
  receptor = NULL,
  show_type = "number",
  text_size = 10,
  viridis_option = "D",
  ...
)
```



**Arguments**

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype	which cell types to plot by order. Default is to plot all cell types
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
show_type	which type to show, "number" and "score" for sum of inferred LR number and score, respectively. Default is "number"
text_size	Size of text labels. Default is 10
viridis_option	option in <a href="#">scale_color_viridis</a> , can be "A", "B", "C", "D", "E", "F", "G", "H". Default is "D".
...	parameters pass to <a href="#">heatmaply</a> , e.g., grid_color

**Value**

Heatmap plot of cell-cell communications mediated by ligand-receptor interactions

---

plot\_cci\_lrpairs\_bubble

*Bubble plot of inferred ligand-receptor pairs*

---

**Description**

Bubble plot of inferred ligand-receptor pairs from senders top receivers. Rows for cell pairs, and columns for LR pairs by default.

**Usage**

```
plot_cci_lrpairs_bubble(
  object,
  celltype = NULL,
  ligand = NULL,
  receptor = NULL,
  if_show_autocrine = FALSE,
  if_horizontal = TRUE,
  size = 2,
  viridis_option = "D"
)
```

**Arguments**

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype	which cell types to plot. Default is to plot all cell types
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
if_show_autocrine	Whether to show autocrine. Default is FALSE
if_horizontal	Whether to plot with the horizontal direction. Default is TRUE
size	Size of the bubble. Default is 2
viridis_option	option in <a href="#">scale_color_viridis</a> , can be "A", "B", "C", "D", "E", "F", "G", "H". Default is "D".

**Value**

Bubble plot of inferred ligand-receptor pairs

---

plot\_cci\_lrpairs\_heatmap

*Heatmap plot of all lrpairs among cell pairs*

---

**Description**

heatmap plot of inferred lrpairs. Rows for cell pairs, and columns for LR pairs by default.

**Usage**

```
plot_cci_lrpairs_heatmap(
  object,
  celltype = NULL,
  ligand = NULL,
  receptor = NULL,
  if_show_autocrine = FALSE,
  if_horizontal = TRUE,
  text_size = 10,
  viridis_option = "D",
  ...
)
```

**Arguments**

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype	which cell types to plot. Default is to plot all cell types
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
if_show_autocrine	Whether to show autocrine. Default is FALSE
if_horizontal	Whether to plot with the horizontal direction. Default is TRUE
text_size	Size of text labels. Default is 10
viridis_option	option in <a href="#">scale_color_viridis</a> , can be "A", "B", "C", "D", "E", "F", "G", "H". Default is "D".
...	parameters pass to <a href="#">heatmaply</a> , e.g., grid_color

**Value**

Heatmap plot of all lrpairs among cell pairs

---

plot_cci_sankey	<i>Sankey plot of cell-cell communications</i>
-----------------	--

---

## Description

Sankey plot of cell-cell communications from senders to receivers with the sum of inferred number or score of ligand-receptor interactions

## Usage

```
plot_cci_sankey(
  object,
  celltype = NULL,
  ligand = NULL,
  receptor = NULL,
  celltype_color = NULL,
  edge_color = NULL,
  show_type = "number",
  if_show_autocrine = FALSE,
  edge_alpha = 0.5,
  node_size = 40,
  text_size = 15,
  node_pad = 20,
  ...
)
```

## Arguments

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype	which cell types to plot. Default is to plot all cell types
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
celltype_color	Colors for the cell types, whose length must be equal to celltype
edge_color	Colors for the edges from the sender cell type, whose length must be equal to celltype, Or use "NO" to cancel it
show_type	which type to show, "number" and "score" for sum of inferred LR number and score, respectively. Default is "number"
if_show_autocrine	Whether to show autocrine. Default is FALSE
edge_alpha	Transparency of edge. Default is 0.5
node_size	Size of node. Default is 40
text_size	Size of text. Default is 15
node_pad	Size of node padding. Numeric essentially influences the width height. Default is 20
...	parameters pass to <a href="#">sankeyNetwork</a>

## Value

Sankey plot of cell-cell communications mediated by ligand-receptor interactions

---

plot_lrpairs_chord	<i>Chord plot of ligand-receptor pairs</i>
--------------------	--

---

### Description

Chord plot of ligand-receptor pairs from senders to receivers.

### Usage

```
plot_lrpairs_chord(
  object,
  celltype_sender,
  celltype_receiver,
  celltype_color = NULL,
  ligand = NULL,
  receptor = NULL,
  edge_color = NULL,
  edge_type = "circle",
  text_size = 0.5,
  y_scale = 1,
  ...
)
```

### Arguments

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype_sender	Name of celltype_sender. One or more cell types
celltype_receiver	Name of celltype_receiver. One or more cell types
celltype_color	Colors for the celltype_sender nodes and celltype_receiver nodes, or use "NO" to make it simple
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
edge_color	Colors for the edges from the sender cell type
edge_type	Types for the edges from the sender cell type. Default is "circle". "big.arrow" for big arrow, "triangle" for triangle, "ellipse" for ellipse, "curved" for curved. Details see <a href="#">chordDiagram</a>
text_size	Size of text labels. Default is 0.5
y_scale	y_scale to adjust the text. Default is 1
...	parameters pass to <a href="#">chordDiagram</a> , e.g., link.arr.width, link.arr.length, link.arr.col

### Value

Chord plot of ligand-receptor pairs from senders to receivers

---

plot\_lrpairs\_heatmap    *Heatmap plot of ligand-receptor pairs*

---

## Description

Heatmap plot of ligand-receptor pairs from senders to receivers.

## Usage

```
plot_lrpairs_heatmap(
  object,
  celltype_sender,
  celltype_receiver,
  ligand = NULL,
  receptor = NULL,
  text_size = 5,
  if_horizontal = TRUE,
  viridis_option = "D",
  ...
)
```

## Arguments

object	scCrossTalk object after <a href="#">find_lrpairs</a>
celltype_sender	Name of celltype_sender. One or more cell types
celltype_receiver	Name of celltype_receiver. One or more cell types
ligand	which ligand to use. Default is to plot all inferred ligands
receptor	which receptor to use. Default is to plot all inferred receptors
text_size	Size of text labels. Default is 3
if_horizontal	Whether to plot with the horizontal direction. Default is TRUE
viridis_option	option in <a href="#">scale_color_viridis</a> , can be "A", "B", "C", "D", "E", "F", "G", "H". Default is "D".
...	parameters pass to <a href="#">heatmaply</a> , e.g., grid_color

## Value

Heatmap plot of ligand-receptor pairs

---

rev_gene	<i>Pre-processing step: revising gene symbols</i>
----------	---

---

### Description

Revise genes according to NCBI Gene symbols updated in June 19, 2022 for count matrix and user-custom lrpairs data.frame

### Usage

```
rev_gene(data = NULL, data_type = NULL, species = NULL, geneinfo = NULL)
```

### Arguments

data	A data.frame or matrix or dgCMatrx
data_type	A character to define the type of data, select 'count' for the data matrix, 'lrpairs' for the lrpairs data.frame
species	Species of the data. 'Human' or 'Mouse'
geneinfo	A data.frame of the system data containing gene symbols of 'Human' or 'Mouse' updated on June 19, 2022 for revising gene symbols

### Value

A new data.frame, matrix, or dgCMatrx.

---

scCrossTalk	<i>Definition of 'scCrossTalk' class</i>
-------------	--

---

### Description

An S4 class containing the data, meta, and results of inferred cell-cell communications mediated by ligand-receptor interactions.

### Slots

data	A list containing the data.
meta	A data.frame containing the meta data.
species	A character containing the species.
lrdb	A data.frame containing expressed ligand and receptor genes.
cci	A data.frame containing enriched ligand-receptor interactions.

---

`show,scCrossTalk-method`

*Show scCrossTalk object*

---

### **Description**

Show scCrossTalk object

### **Usage**

```
## S4 method for signature 'scCrossTalk'  
show(object)
```

### **Arguments**

object                    scCrossTalk object after [create\\_scCrossTalk](#)

### **Value**

scCrossTalk object

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