

# Single-cell lineage tracing with a focus on cancer metastasis using macsGESTALT

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Pinello Lab Journal Club

Friday | June 25<sup>th</sup>, 2021



# Overview of today's presentation

Paper we are covering today:



First we will cover some history, background, motivation, and context:

The image is a collage of several academic publications and reviews, each with its own header, author list, and abstract. The publications include:

- RESEARCH ARTICLE**  
CELL LINEAGE TRACING  
**Whole-organism lineage tracing by combinatorial and cumulative genome editing**  
Aaron McKenna,<sup>1\*</sup> Gregory M. Findlay,<sup>1\*</sup> James A. Gagnon,<sup>2\*</sup> Marshall S. Horwitz,<sup>1</sup> Alexander F. Schier,<sup>2,4,5,6†</sup> Jay Shendure<sup>1,7†</sup>
- Developmental Cell**  
Review  
**Next-Generation Lineage Tracing and Fate Mapping to Interrogate Development**  
Cite as: Weinreb et al., *Science* 10.1126/science.aaw3381 (2020).
- Science**  
RESEARCH ARTICLES  
**Lineage tracing on transcriptional landscapes links state to fate during differentiation**  
Ríquez-Fraticelli<sup>2,3\*</sup>, Fernando D. Camargo<sup>2,3†</sup>, Allon M. Klein<sup>1†‡</sup>
- Cell**  
**ARTICLE**  
<https://doi.org/10.1038/s41586-018-0744-4>  
**Single-cell mapping of lineage and identity in direct reprogramming**  
Brent A. Biddy<sup>1,2,3</sup>, Wenjun Kong<sup>1,2,3</sup>, Kenji Kamimoto<sup>1,2,3</sup>, Chunru Guo<sup>1,2,3</sup>, Sarah E. Waye<sup>1,2,3</sup>, Tao Sun<sup>1,2,3,4</sup> & Samantha A. Morris<sup>1,2,3\*</sup>
- SINGLE-CELL OMICS**  
**Building a lineage from single cells: genetic techniques for cell lineage tracking**  
Mollie B. Woodworth<sup>1–3</sup>, Kelly M. Girsikis<sup>1–3</sup> and Christopher A. Walsh<sup>1–3</sup>
- Check for updates**
- Lineage tracing meets single-cell omics: opportunities and challenges**  
Daniel E. Wagner<sup>1,2</sup> and Allon M. Klein<sup>1</sup>
- age reconstruction from clonal correlations**  
inreb<sup>a</sup> and Allon M. Klein<sup>a,1</sup>

# Overview of today's presentation

Access all sides with links to papers and notes:



Screenshot of a GitHub repository page for [mvinyard/vintools](https://github.com/mvinyard/vintools).

The repository contains a single page titled "Pinello Lab Journal Club: June 25th, 2021".

**Featured paper:**

[Single-cell lineage tracing of metastatic cancer reveals selection of hybrid EMT states](#)

**Cancer Cell**

**Single-cell lineage tracing of metastatic cancer reveals selection of hybrid EMT states**

**Graphical abstract**

**Article**

**Authors**  
Kamen P. Simeonov, China N. Byrns, Megan L. Clark, ..., Jay Shendure, Aaron McKenna, Christopher J. Lengner

**Correspondence**  
kamen.simeonov@gmail.com (K.P.S.), shendure@uw.edu (J.S.), aaron.mckenna@dartmouth.edu (A.M.), lengner@vet.upenn.edu (C.J.L.)

**In brief**  
Simeonov et al. develop an inducible lineage recorder, enabling simultaneous

**Pages 3**

- Find a Page...
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- Pinello Lab Journal Club: June 25th, 2021

+ Add a custom sidebar

Clone this wiki locally  
<https://github.com/mvinyard/vintools>



**GitHub.com/mvinyard/vintools**



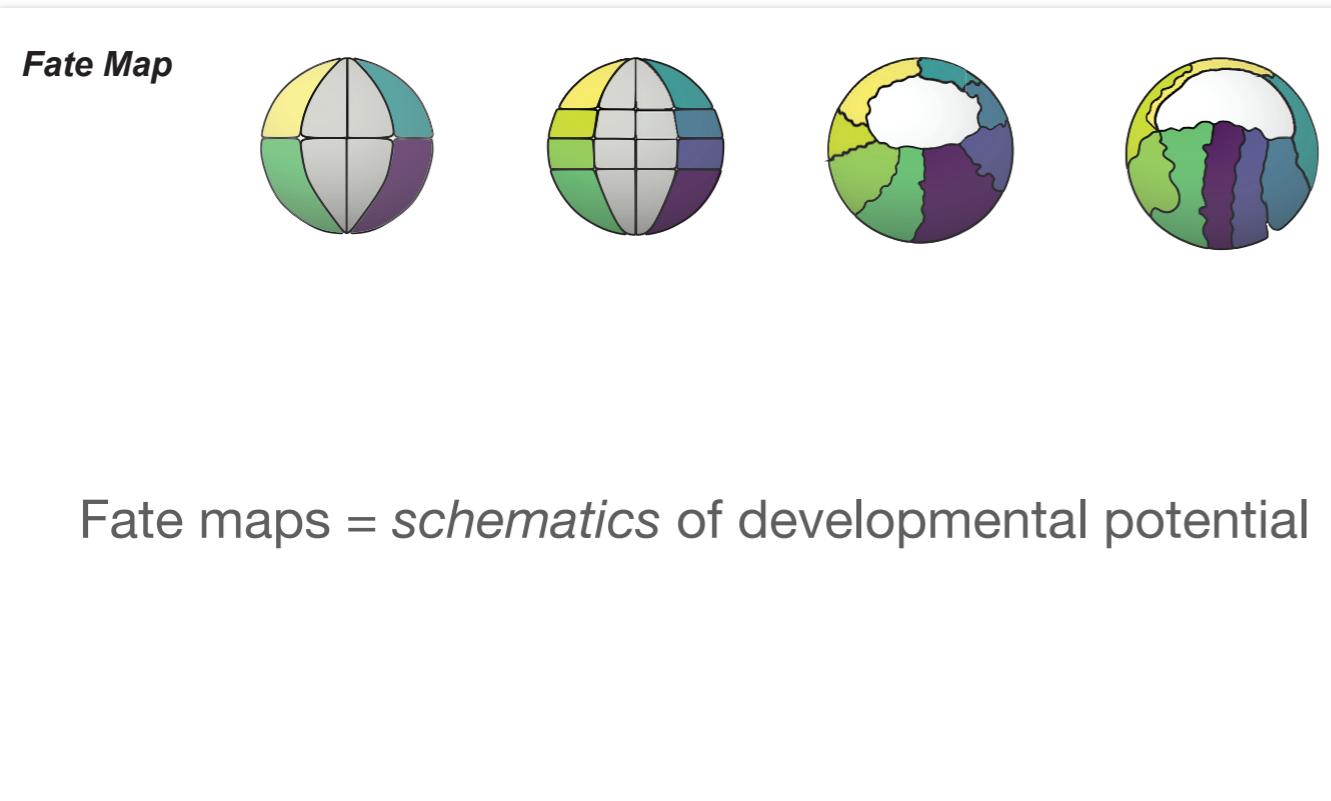
# Overview of today's presentation

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1. Brief history lesson and overview of tools / datasets / technologies available
2. Overview of GESTALT, the precursor technology to macsGESTALT
3. In-depth coverage of the macsGESTALT paper



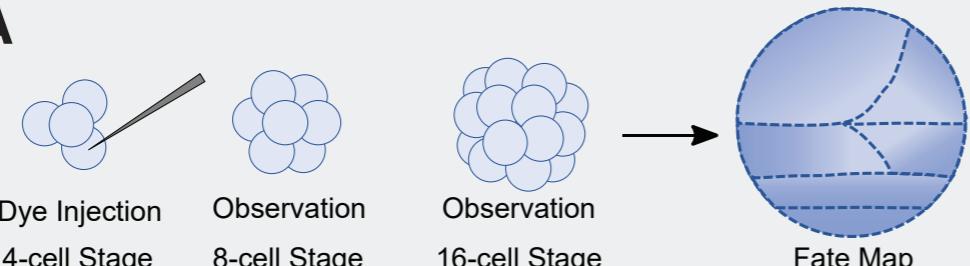
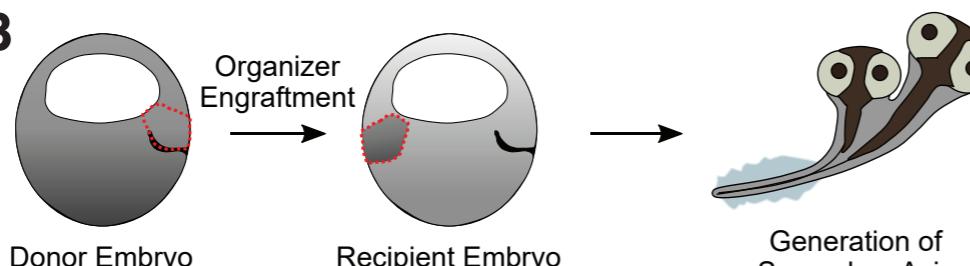
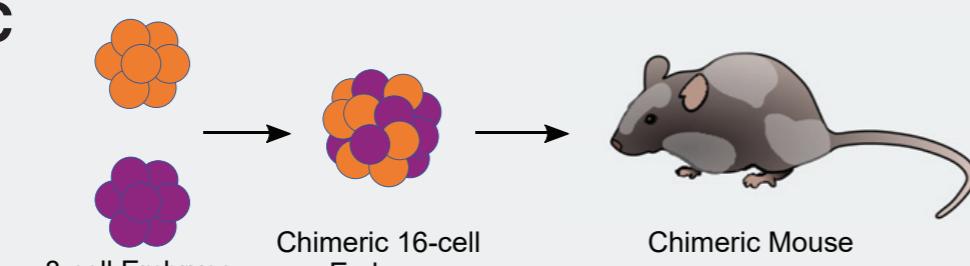
# Brief history of fate mapping and lineage tracing



- Fate mapping and lineage tracing are related but **distinct**
- Lineage tracing identifies progeny from a given ancestor cell

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Observational Biology	1890s	<b>A</b> Dye Injection 4-cell Stage → Observation 8-cell Stage → Observation 16-cell Stage → Fate Map	Single-cell limited by injection	10s of cells limited by observation	Observational data	Dye Injection and Time Lapse Conklin, 1905 Vogt, 1924
	1920-30s	<b>B</b> Donor Embryo → Organizer Engraftment → Recipient Embryo → Generation of Secondary Axis	N/A	Tissues	Observational data	Organizer Grafts Spemann and Mangold, 1924 Wetzel, 1929
Molecular Biology	1960s	<b>C</b> 8-cell Embryos → Chimeric 16-cell Embryo → Chimeric Mouse	N/A	Tissues	Only specific to embryo of origin	Chimera Generation Tarkowski, 1965 Mintz, 1965
	1980s	<b>D</b> Retroviral Transduction → Cellular Proliferation in vivo or in vitro → Marker Recovery (i.e. β-gal)	Theoretically single clones	10s of cells limited by observation	Observational data	Retroviral Labelling Cepko et al., 1987
	2000s	<b>E</b> Retroviral Transduction of Specialized Cre/lox Cassette → Cre Activation Creates Alternative XFP Readout → Lineage Determination	Theoretically single clones	100s of cells limited by observation	Observational data	Randomized Recombination Cassettes Livet et al., 2007 Snippert et al., 2010
	2010s	<b>F</b> Cas9 targeted editing of genomic DNA, transgene or endogenous → Accrual of Cas9-mediated scars on target sequences → Clonal tracking of regeneration in caudal fin	Theoretically single clones	100s of cells limited by observation	Dataset limitation based on collection method	Cas9 Targeted Scar Accrual McKenna et al., 2016 Junker et al., 2017
Single-cell Biology	2010s-	<b>G</b> Retroviral Transduction with CellTags → Rounds of CellTagging generate barcode combinations → scRNA-seq reads include transcriptomic and lineage barcode data	Single-cell	1000s - 10,000s of cells	Resolved to clonal and sub-clonal populations	Retroviral mRNA Barcode Accrual Yao et al., 2017 Biddy et al., 2018 Weinreb et al., 2020
	2010s-	<b>H</b> Cas9-targeted editing of genomic DNA sequence → Accrual of Cas9-mediated scars on target sequences → Generation of lineage trees based on mRNA barcode readout	Single-cell	1000s - 10,000s of cells	Information dropout due to Cas9 induced deletion of previous scars	Cas9 mRNA Scars Spanjaard et al., 2018 Raj et al., 2018 Chan et al., 2019 Bowling et al., 2020
	2010s-	<b>I</b> Tol2 transposase with barcoded GFP → Accrual of Tol2-mediated GFP-barcode Insertions → Lineage Tree Reconstruction	Single-cell	1000s - 10,000s of cells	None beyond usual scRNA-seq transgene dropout	Transposon mRNA Barcode Accrual Wagner et al., 2019

# Fate mapping and lineage tracing

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Observational Biology	1890s	<b>A</b> 	Single-cell limited by injection	10s of cells limited by observation	Observational data	<b>Dye Injection and Time Lapse</b> Conklin, 1905 Vogt, 1924
	1920-30s	<b>B</b> 	N/A	Tissues	Observational data	<b>Organizer Grafts</b> Spemann and Mangold, 1924 Wetzel, 1929
	1960s	<b>C</b> 	N/A	Tissues	Only specific to embryo of origin	<b>Chimera Generation</b> Tarkowski, 1965 Mintz, 1965

# Fate mapping and lineage tracing

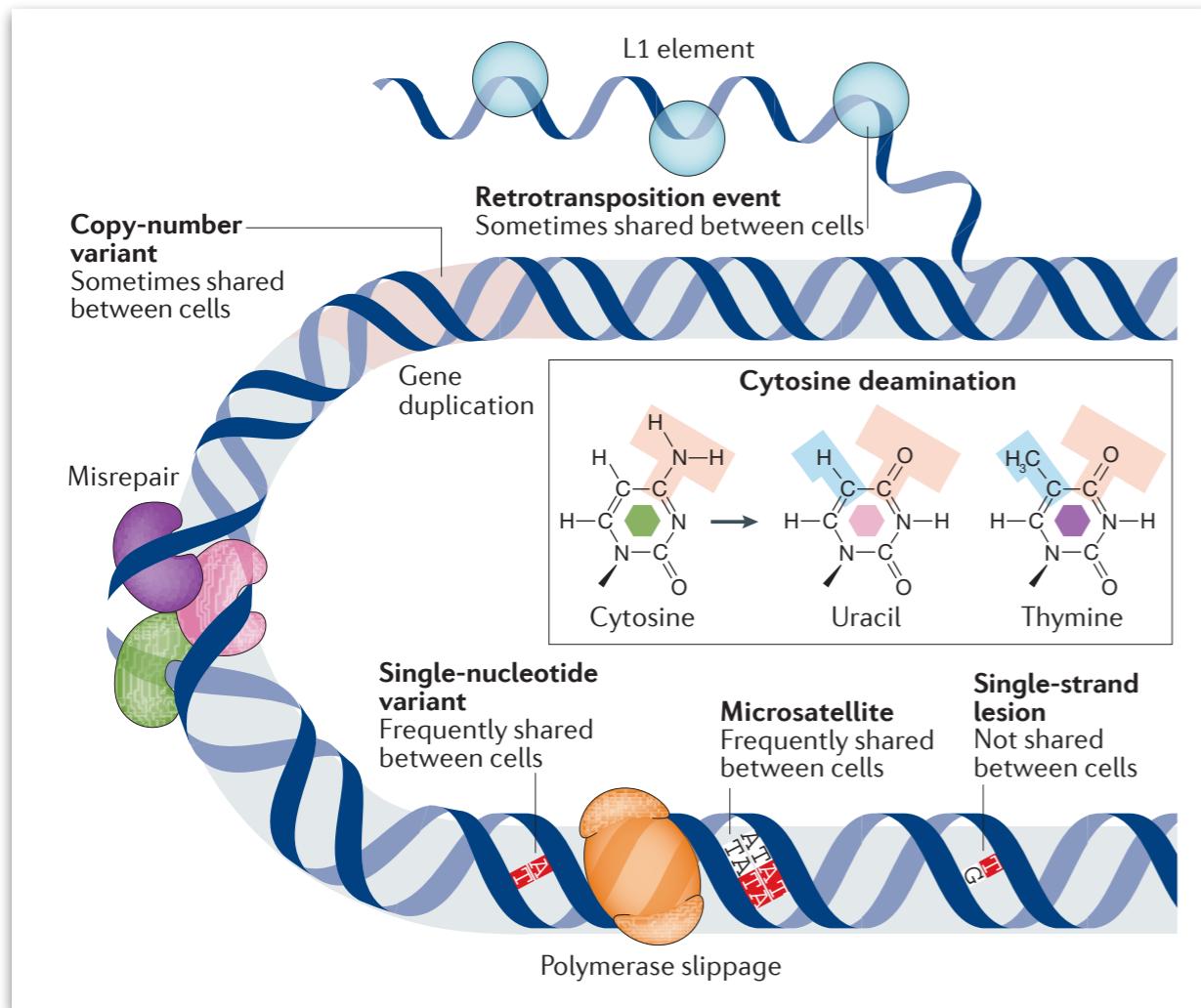
Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
1980s	D	Retroviral Transduction → Cellular Proliferation <i>in vivo</i> or <i>in vitro</i> → Marker Recovery (i.e. $\beta$ -gal)	Theoretically single clones	10s of cells limited by observation	Observational data	<b>Retroviral Labelling</b> Cepko <i>et al</i> , 1987
2000s	E	Retroviral Transduction of Specialized Cre/lox Cassette → Cre Activation Creates Alternative XFP Readout	Theoretically single clones	100s of cells limited by observation	Observational data	<b>Randomized Recombination Cassettes</b> Livet <i>et al</i> , 2007 Snippert <i>et al</i> , 2010
2010s	F	Cas9 targeted editing of genomic DNA, transgene or endogenous → Accrual of Cas9-mediated scars on target sequences	Theoretically single clones	100s of cells limited by observation	Dataset limitation based on collection method	<b>Cas9 Targeted Scar Accrual</b> McKenna <i>et al</i> , 2016 Junker <i>et al</i> , 2017

Still pre-“single-cell revolution” takeover

# Fate mapping and lineage tracing

Era	Year	Lineage Tracing Technique	Resolution	Scalability	Limitation	Technique & Citation
Single-cell Biology	2010s-	<p><b>G</b></p> <p>Retroviral Transduction with CellTags      Rounds of CellTagging generate barcode combinations      scRNA-seq reads include transcriptomic and lineage barcode data</p>	Single-cell	1000s - 10,000s of cells	Resolved to clonal and sub-clonal populations	<b>Retroviral mRNA Barcode Accrual</b> <i>Yao et al, 2017</i> <i>Biddy et al, 2018</i> <i>Weinreb et al, 2020</i>
	2010s-	<p><b>H</b></p> <p>Cas9-targeted editing of genomic DNA sequence      Accrual of Cas9-mediated scars on target sequences      Generation of lineage trees based on mRNA barcode readout</p>	Single-cell	1000s - 10,000s of cells	Information dropout due to Cas9 induced deletion of previous scars	<b>Cas9 mRNA Scars</b> <i>Spanjaard et al, 2018</i> <i>Raj et al, 2018</i> <i>Chan et al, 2019</i> <i>Bowling et al, 2020</i>
	2010s-	<p><b>I</b></p> <p>Tol2 transposase with barcoded GFP      Accrual of Tol2-mediated GFP-barcode Insertions      Lineage Tree Reconstruction</p>	Single-cell	1000s - 10,000s of cells	None beyond usual scRNA-seq transgene dropout	<b>Transposon mRNA Barcode Accrual</b> <i>Wagner et al, 2019</i>

# Using somatic mutations



From Figure 4 of Woodworth et al., Nat Rev Gen (2017)

**Advantage:** already “in the data” or “free”

Two main **limitations** of using somatic variation in lineage tracing:

1. **WGS** required (\$\$\$, unscalable)
2. Inherent read **sparsity** of scRNA-seq



# Using mtDNA mutations

## Article

### Lineage Tracing in Humans Enabled by Mitochondrial Mutations and Single-Cell Genomics

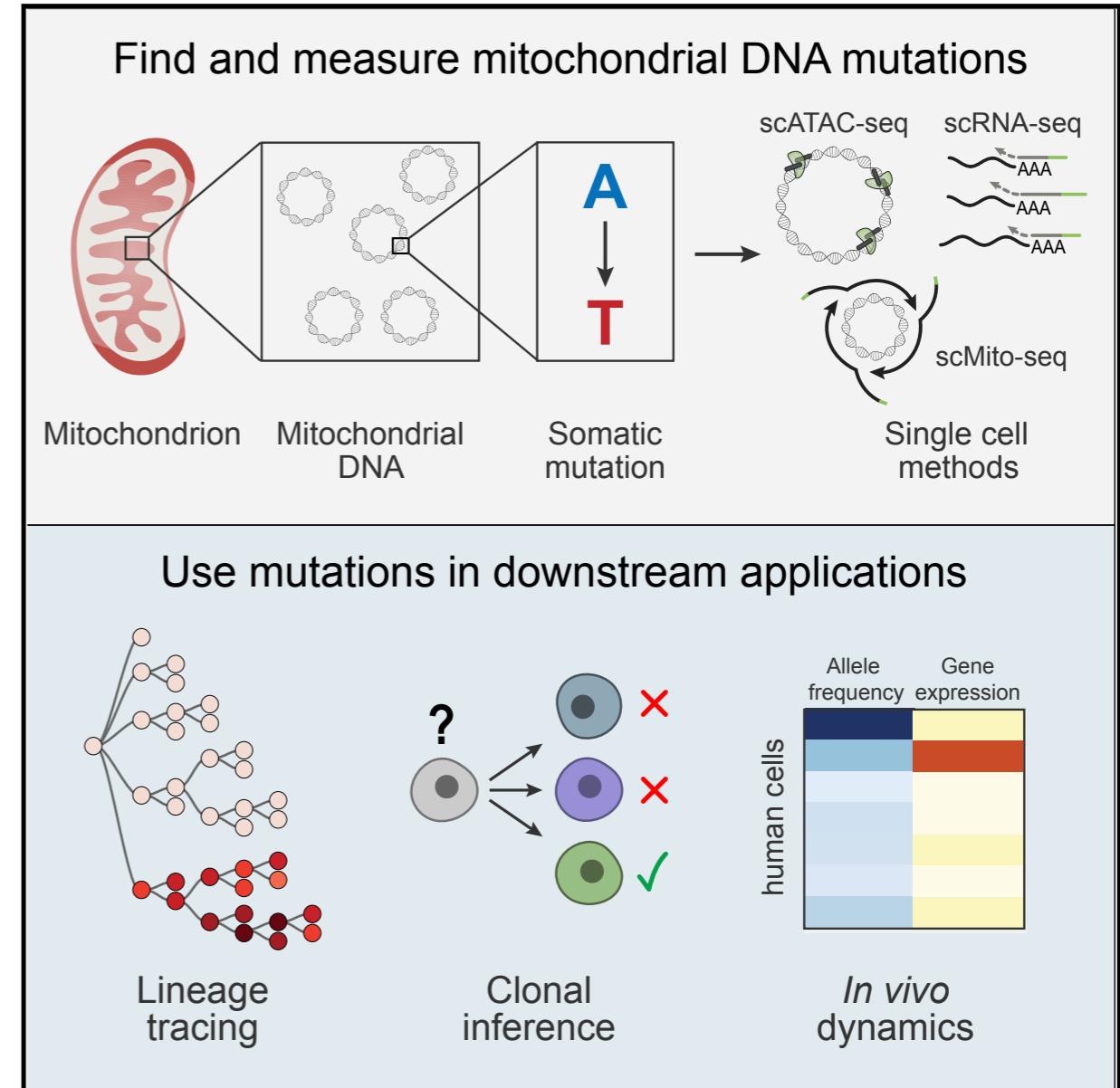
Leif S. Ludwig,<sup>1,2,15,\*</sup> Caleb A. Lareau,<sup>1,2,3,4,15</sup> Jacob C. Ulirsch,<sup>1,2,4,15</sup> Elena Christian,<sup>1</sup> Christoph Muus,<sup>1,5</sup> Lauren H. Li,<sup>1,2</sup> Karin Pelka,<sup>1,6,7</sup> Will Ge,<sup>1</sup> Yaara Oren,<sup>1,8</sup> Alison Brack,<sup>1</sup> Travis Law,<sup>1</sup> Christopher Rodman,<sup>1</sup> Jonathan H. Chen,<sup>1,9</sup> Genevieve M. Boland,<sup>6,10</sup> Nir Hacohen,<sup>1,6,7</sup> Orit Rozenblatt-Rosen,<sup>1</sup> Martin J. Aryee,<sup>1,3,11</sup> Jason D. Buenrostro,<sup>1,12</sup> Aviv Regev,<sup>1,13,\*</sup> and Vijay G. Sankaran<sup>1,2,14,16,\*</sup>

Cell

## Advantages:

1. Again, “free”
2. mtDNA mutation rates > gDNA mutation rates
3. Readily paired GEX / CA with clonal lineage information
4. No WGS or fancy barcoding required

## Graphical Abstract



From Graphical Abstract of Ludwig et al., Cell (2021)

## Limitations:

1. Current approaches offer limited coverage of mito genome
2. Potential for horizontal gene transfer in mito genome (unclear to what extent)



# GESTALT (g~~e~~nome e~~dit~~ing of s~~ynthetic~~ t~~arget~~ a~~rrays~~ for l~~ineage~~ t~~racing~~)

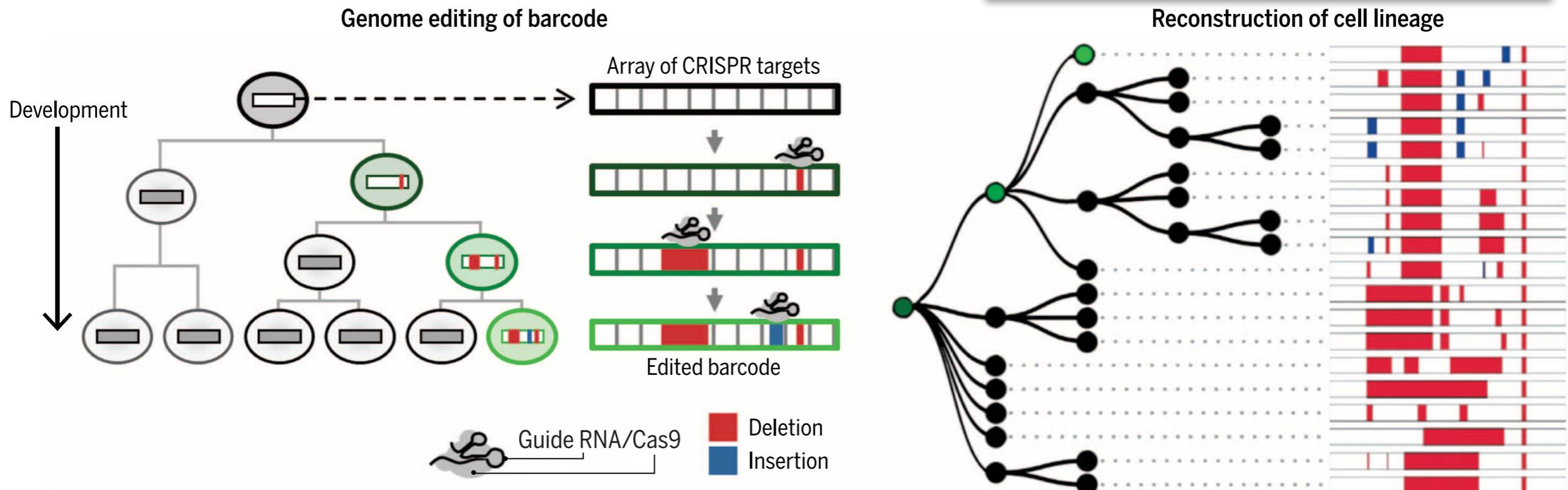
- This paper sets a precedent for large-scale lineage tracing - how? Tracing must...
  - Impart unique marks over division
  - Marks must accumulate over time
  - Easy single-cell readout

## RESEARCH ARTICLE

### CELL LINEAGE TRACING

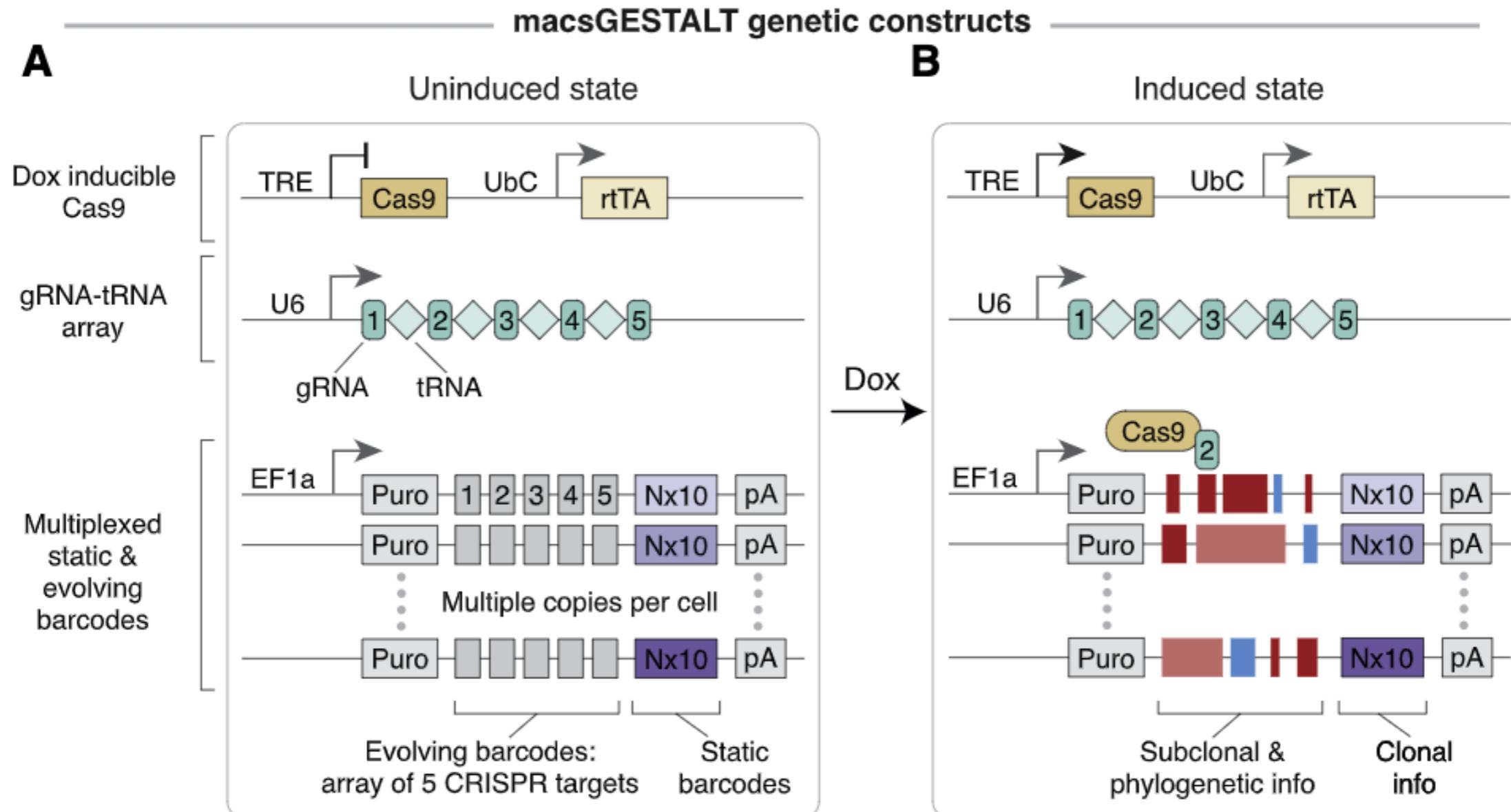
## Whole-organism lineage tracing by combinatorial and cumulative genome editing

Aaron McKenna,<sup>1,\*</sup> Gregory M. Findlay,<sup>1,\*</sup> James A. Gagnon,<sup>2,\*</sup> Marshall S. Horwitz,<sup>1,3</sup> Alexander F. Schier,<sup>2,4,5,6†</sup> Jay Shendure<sup>1,7†</sup>



- Applied to cell culture and zebrafish
- Found that most cells in adult organs derive from relatively few embryonic ancestor cells

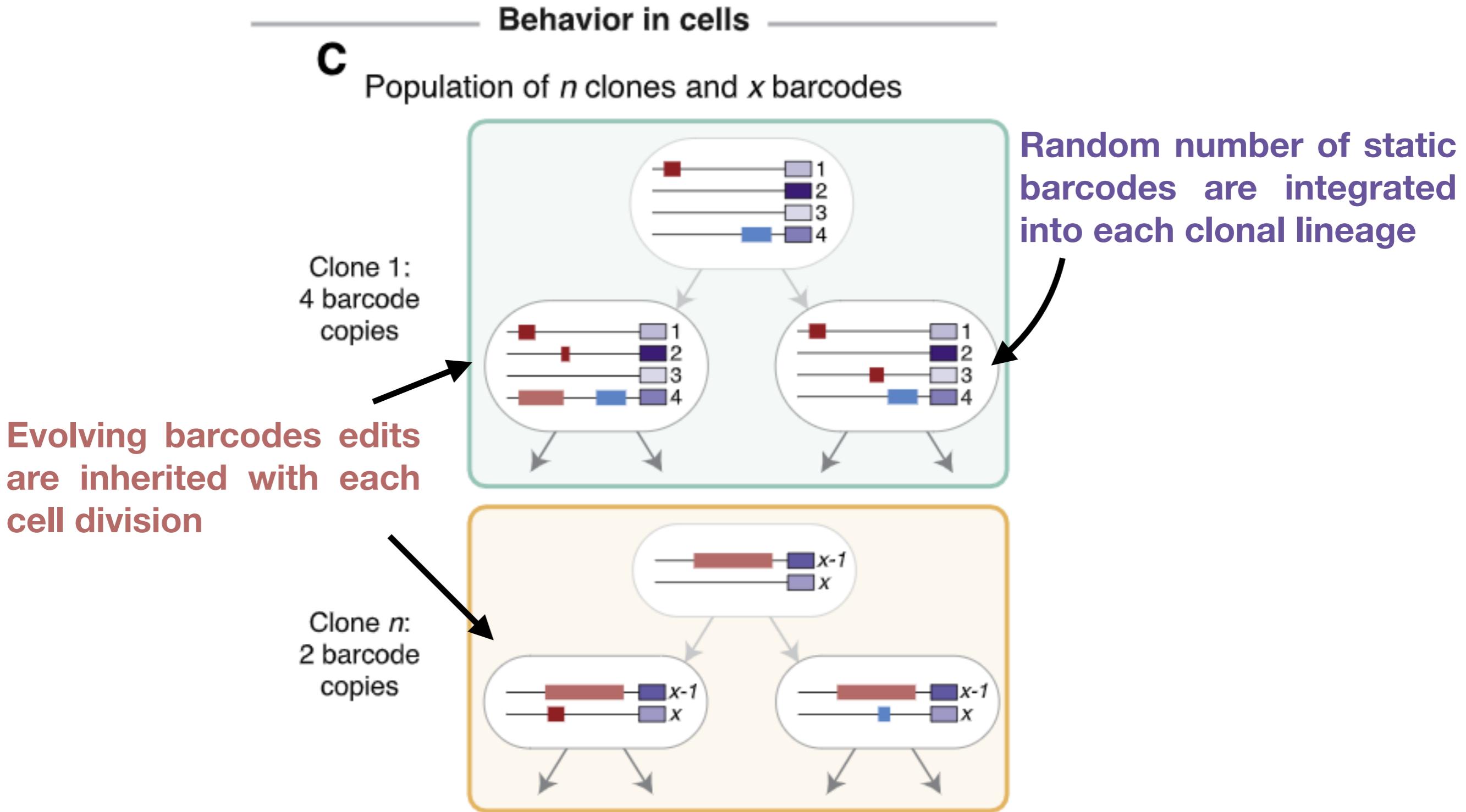
# macsGESTALT for high-res lineage tracing



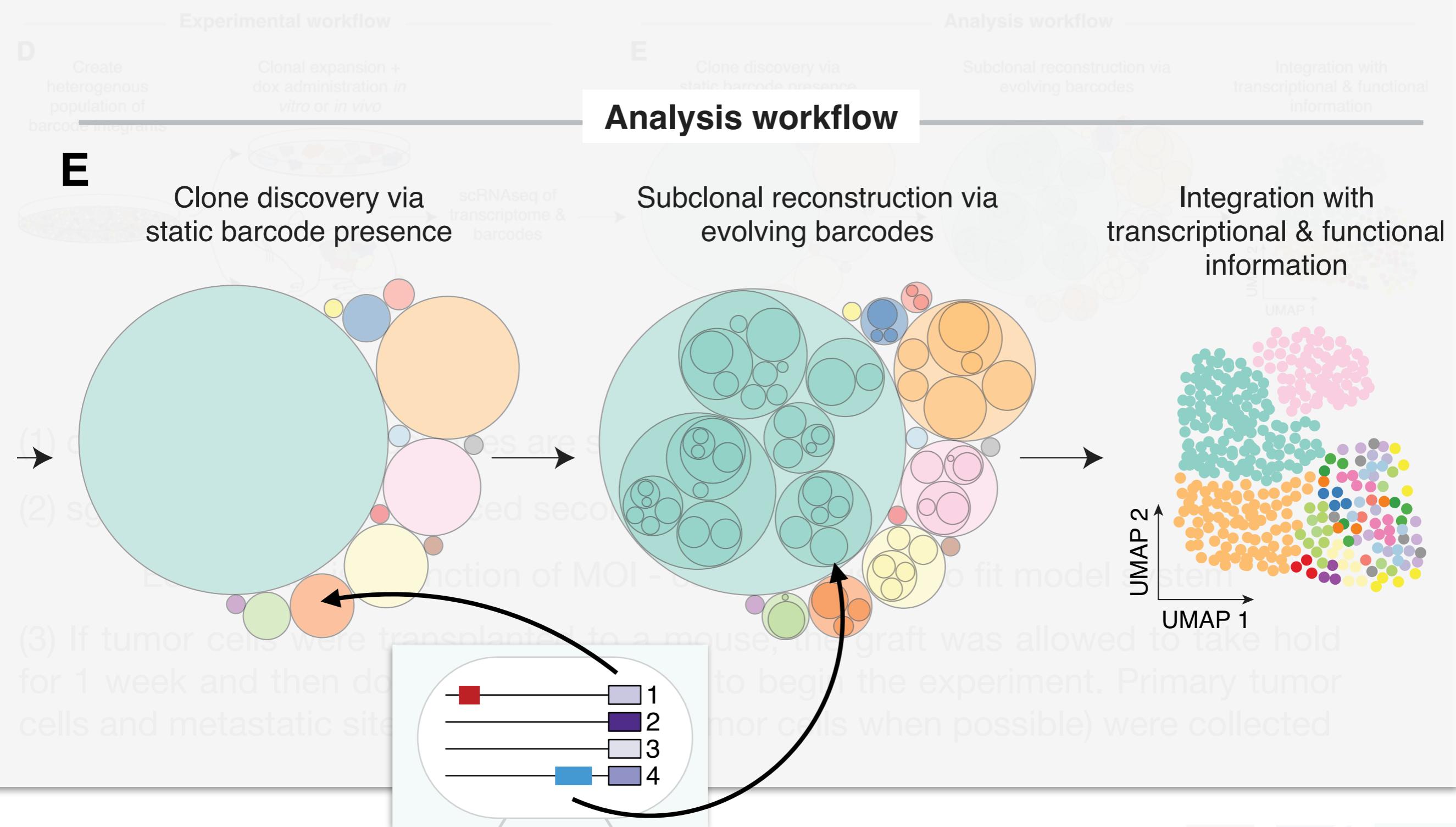
## Static barcodes track clonal information

## Evolving barcodes (via indel mutagenesis) track sub-clonal phylogenetic information

# macsGESTALT for high-res lineage tracing



# macsGESTALT for high-res lineage tracing

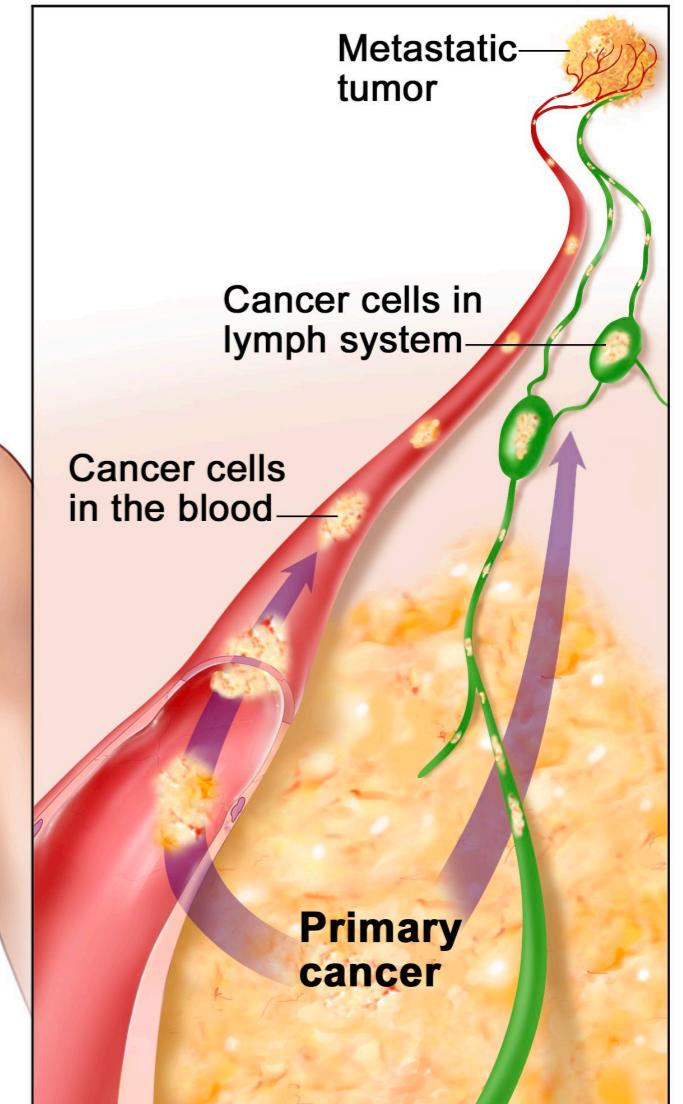
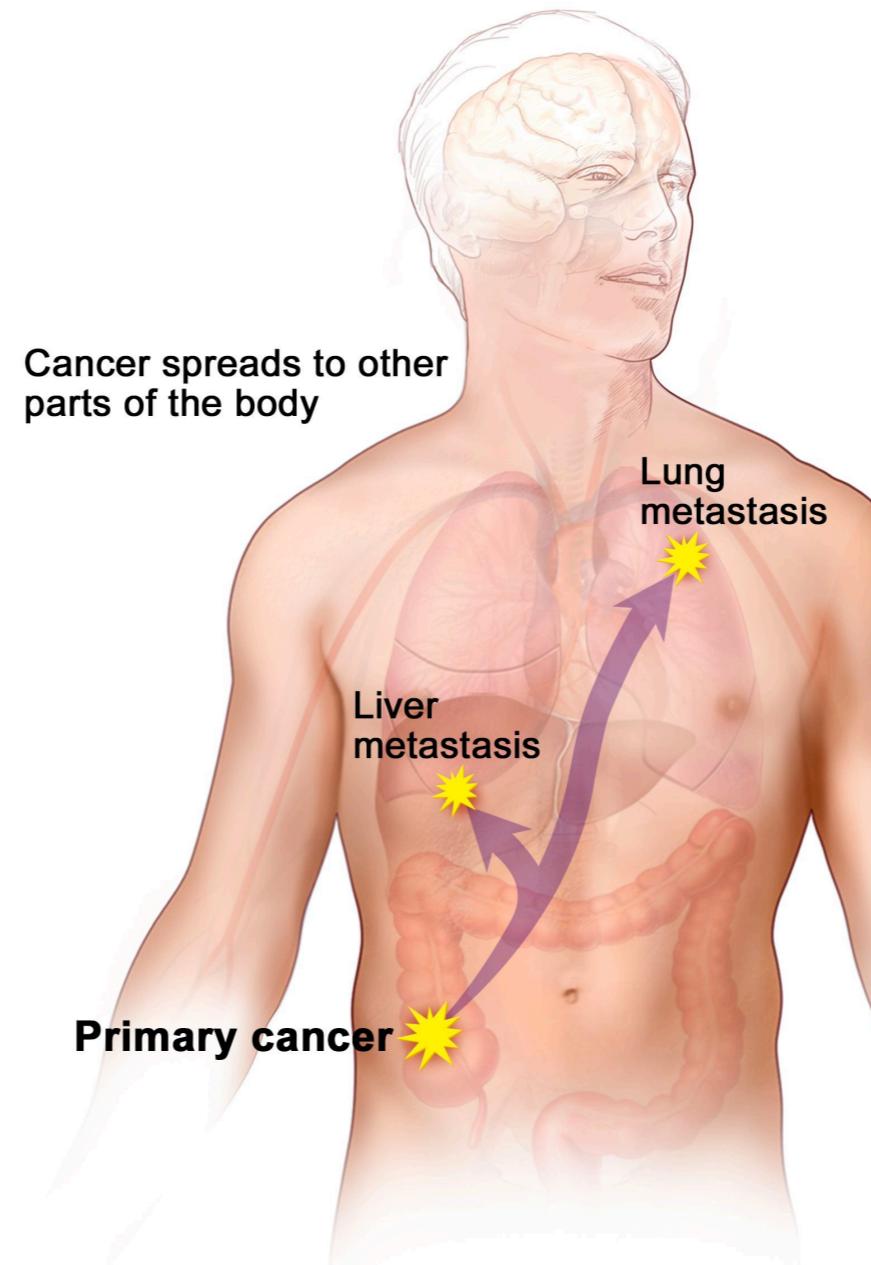


From Figure 1 of Simeonov et al., Cancer Cell (2021)



# What is metastasis and EMT?

- Metastasis = most cancer deaths
- EMT ~ metastasis
- EMT thought to play a role across many cancer types



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*Source: National Cancer Institute*



# Most metastases arise from rare clones

Experiment: combine macsGESTALT with scRNA-seq

A



From Figure 2 of Simeonov et al., Cancer Cell (2021)

PDAC is very deadly (5-year survival rate of 9%)

KPCY mouse tumor model cell line transplanted into non-tumor-bearing mice

Good model for two reasons:

- (1) This model exhibits consistent metastasis kinetics
- (2) Good model of human disease (Kras GoF and p53 LoF are the most common drivers of human PDAC)
- (3) minimal *in vitro* cell line culture time
- (4) pancreatic focal lesion disseminates to the same sites as in human PDAC (incl. liver and lung)



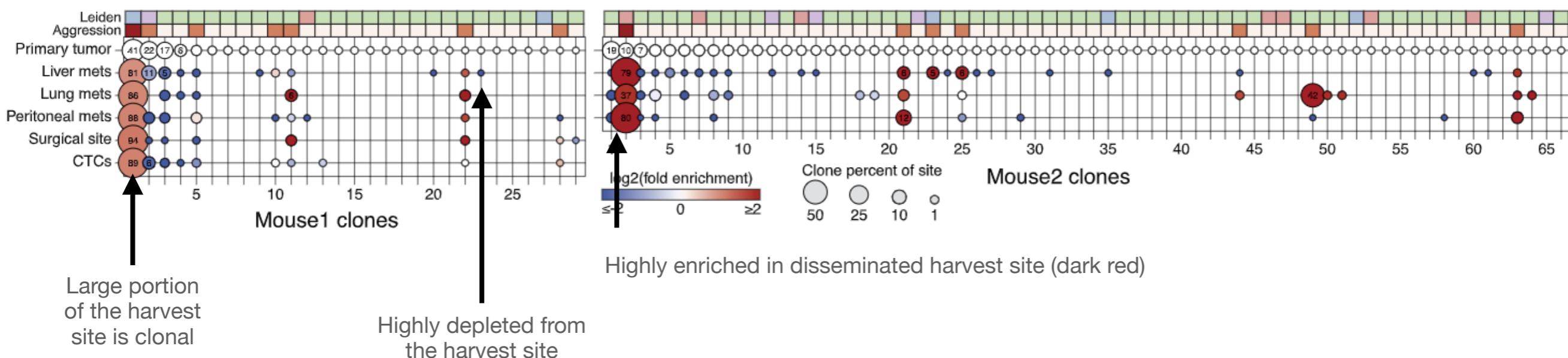
# Most metastases arise from rare clones

Clonal reconstruction via static (purple) barcodes:

Circle size = % contribution to harvest site

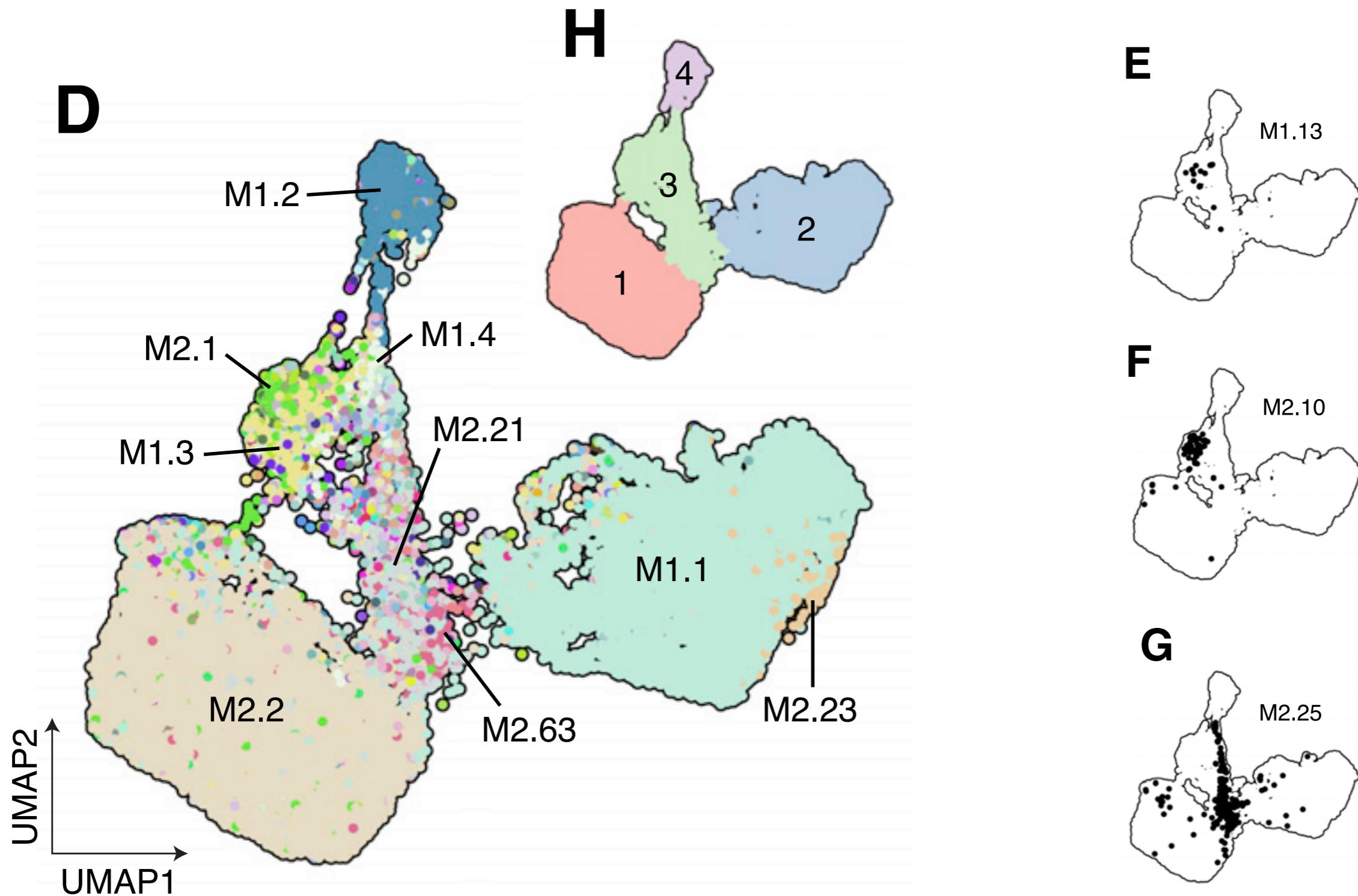
Circle color = enrichment compared to primary tumor

**B** Clones numbered by size in the primary tumor

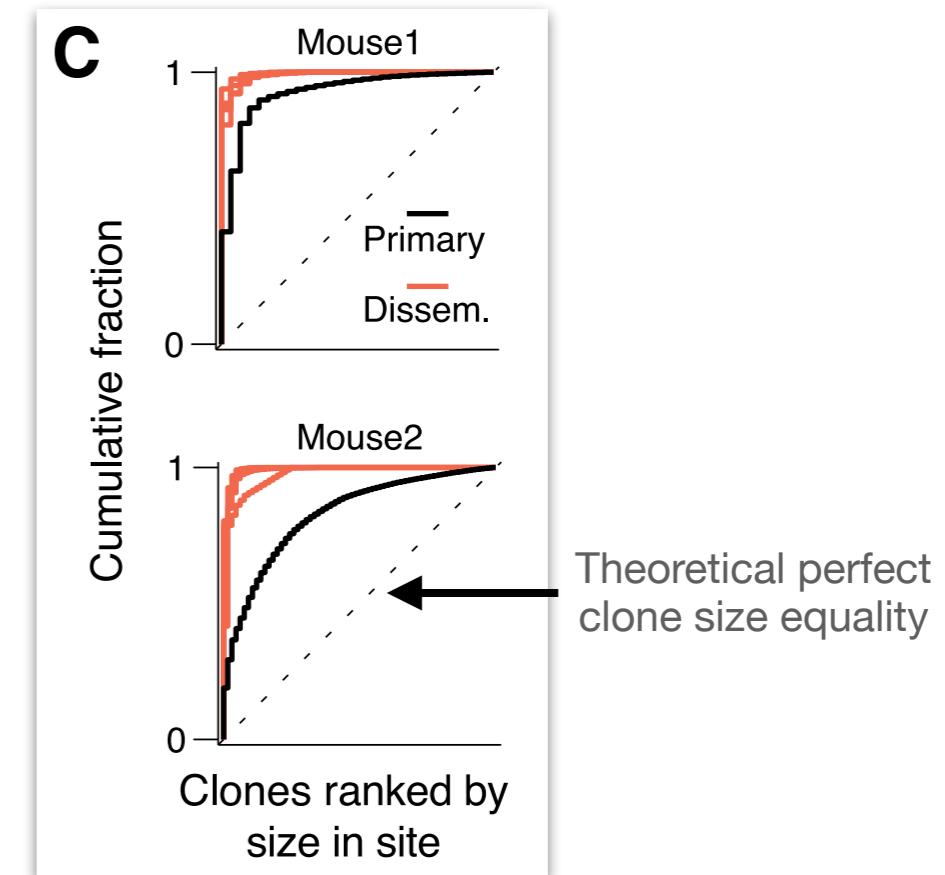
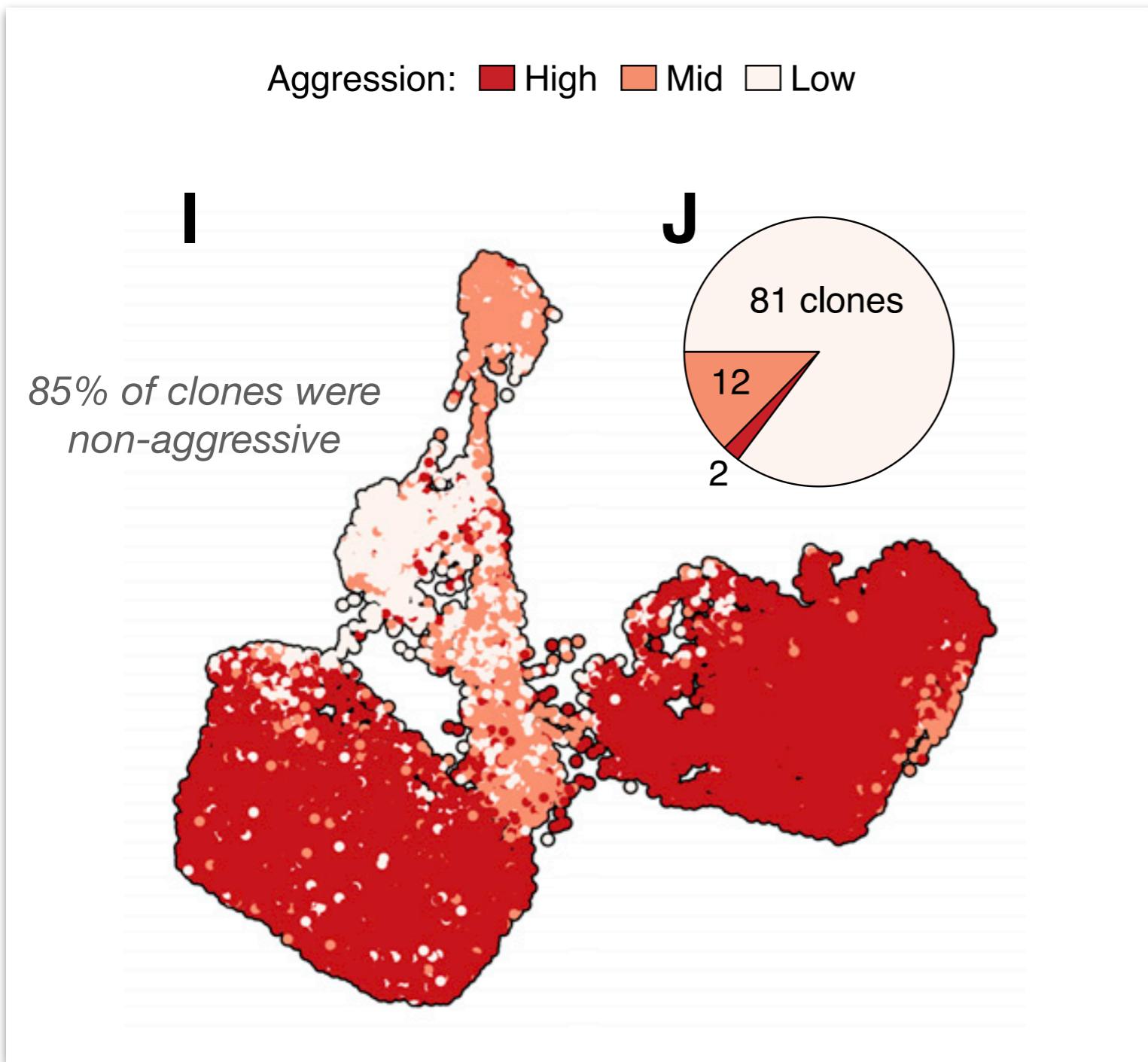


*“51% of clones (48/95) failed to metastasize at all, suggesting that mutations in Kras and p53 alone do not ensure metastatic success.”*

# Most metastases arise from rare clones

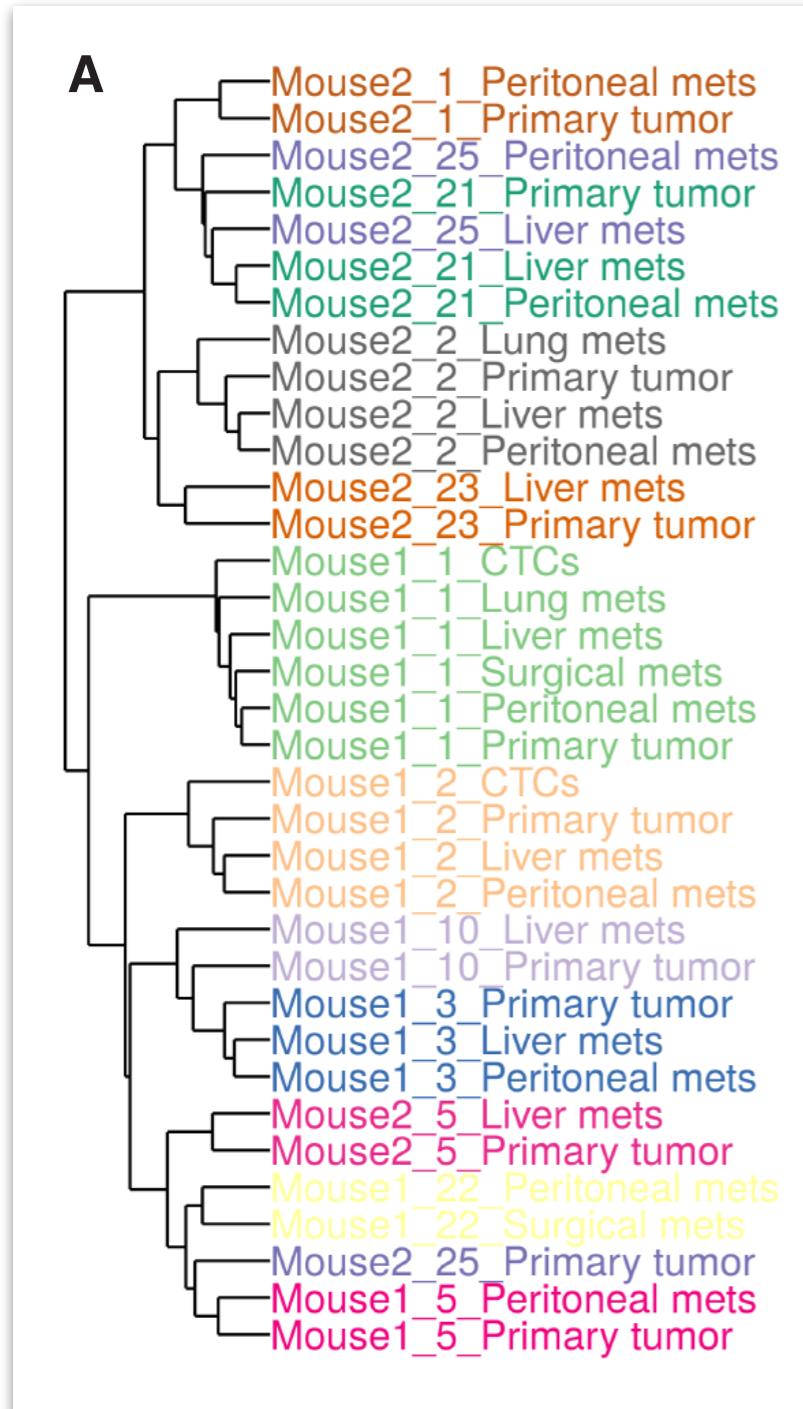


# Most metastases arise from rare clones

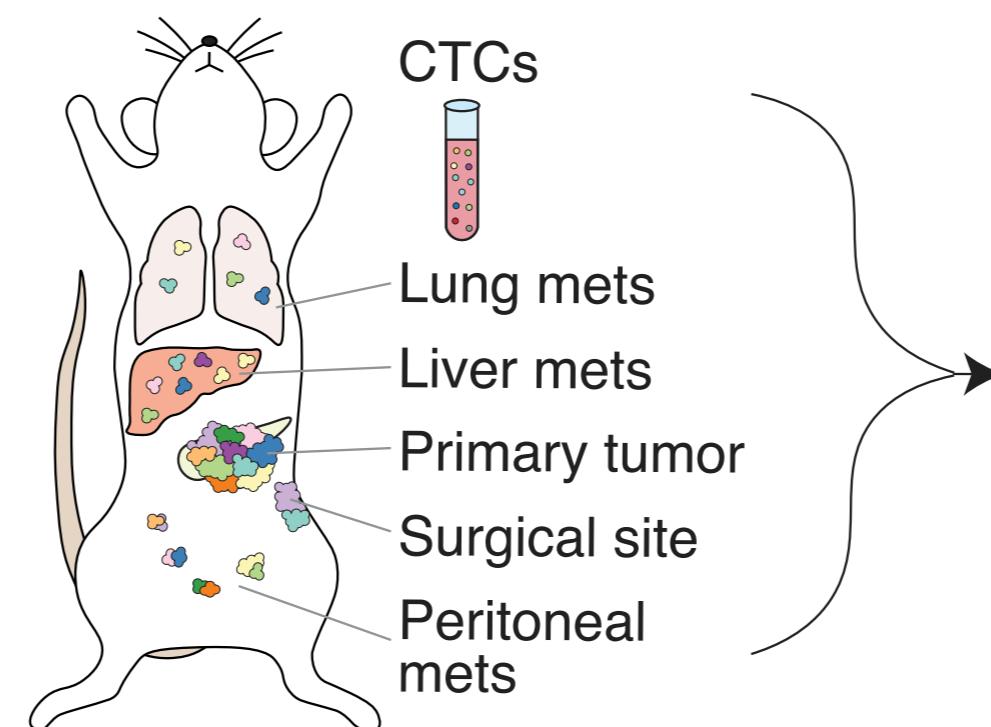


*Just two clones dominate most of the cell population!*

# Cells retain transcriptional identity after metastasis



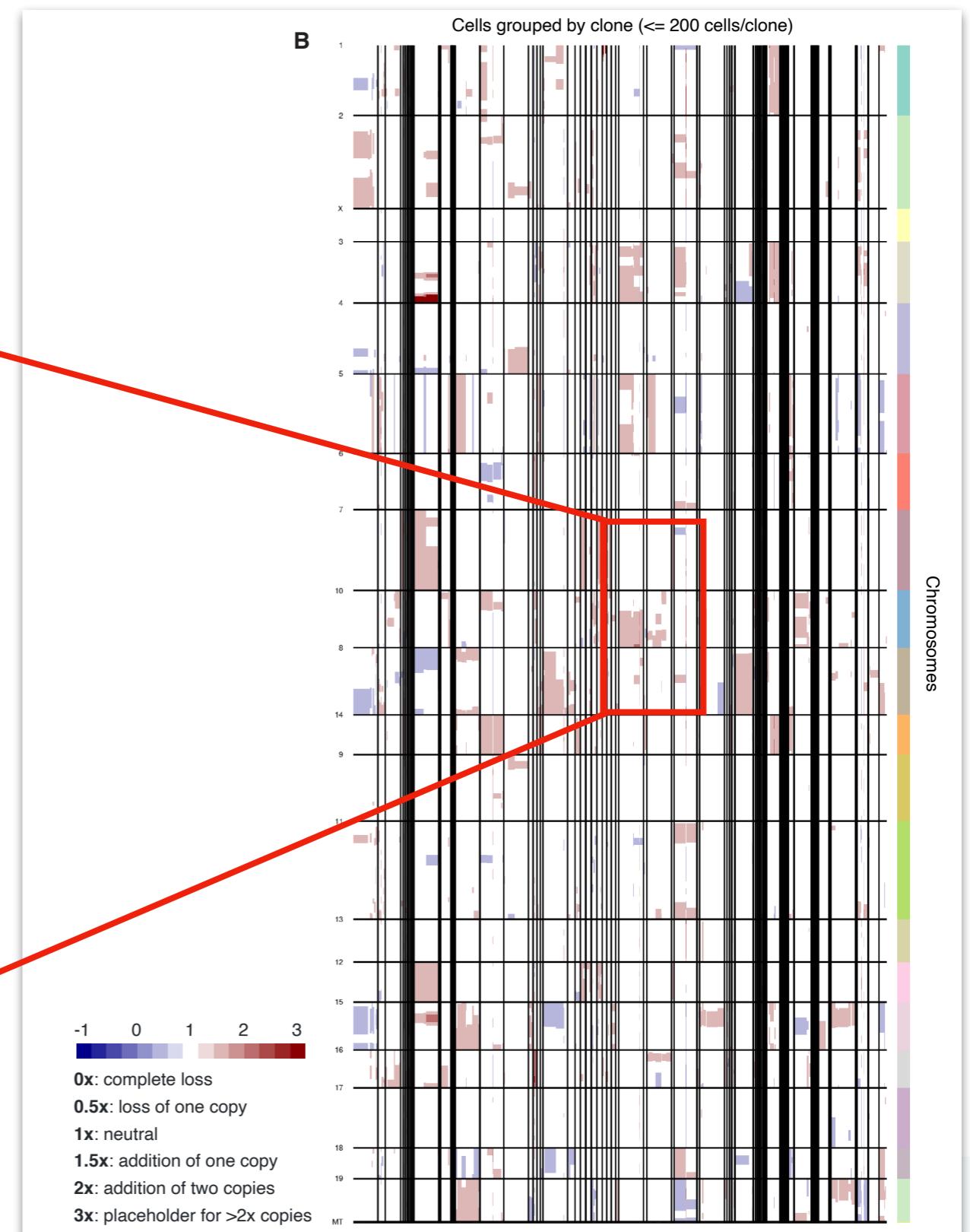
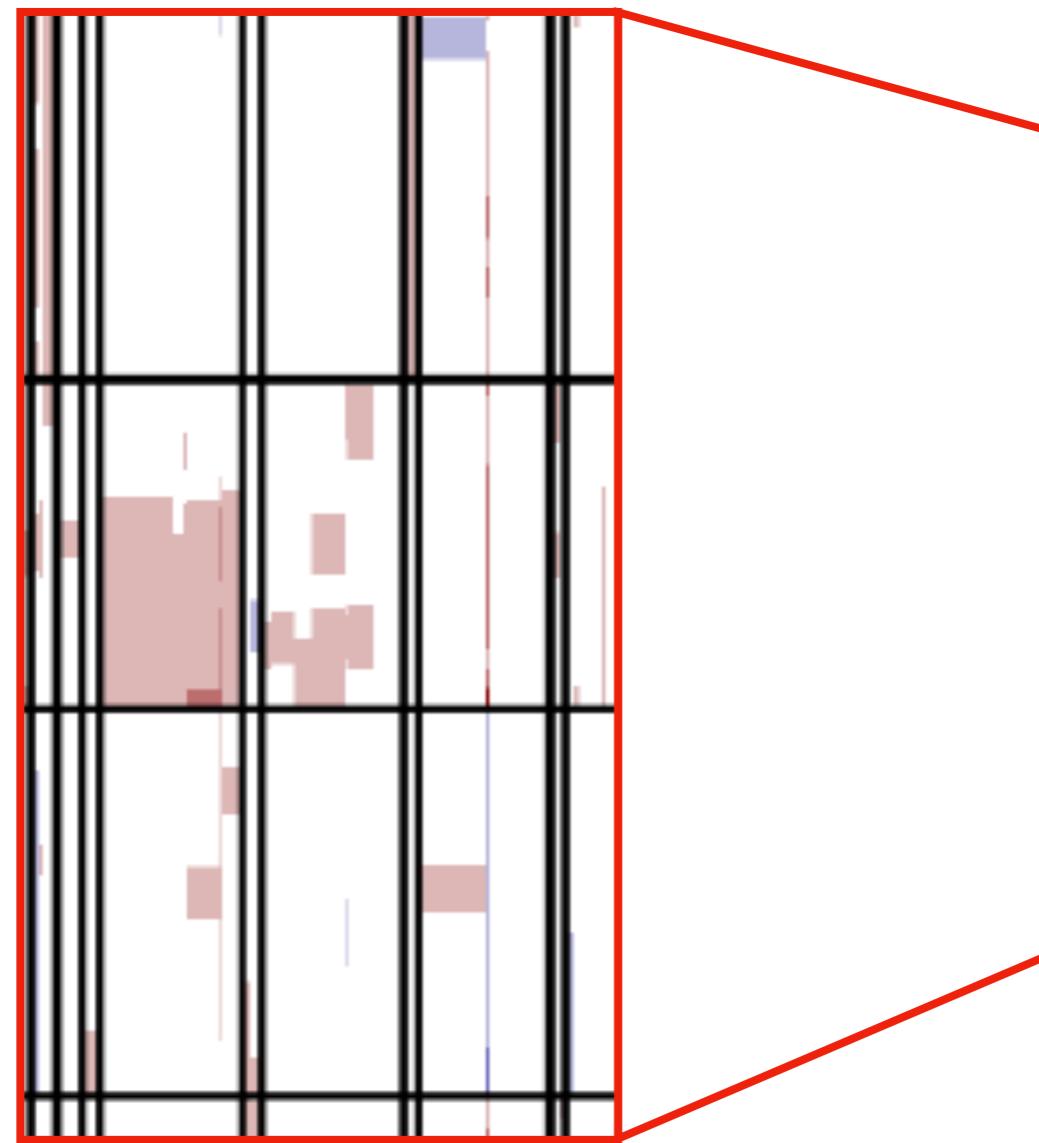
- Clone-site pseudobulk analysis - samples colored by clone
- Samples hierarchically clustered (based on scRNA-seq) - cluster preferentially by clone rather than harvest site



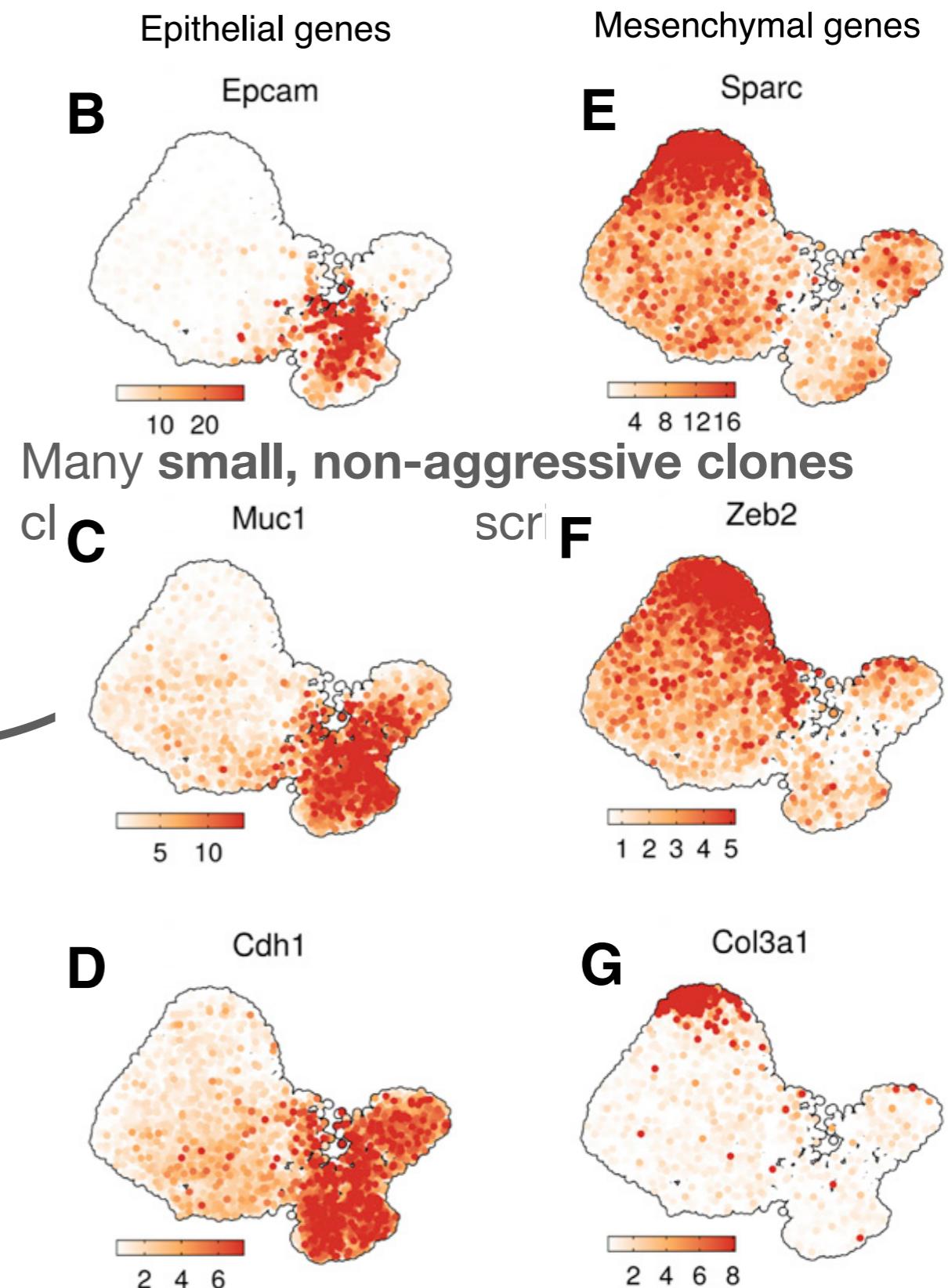
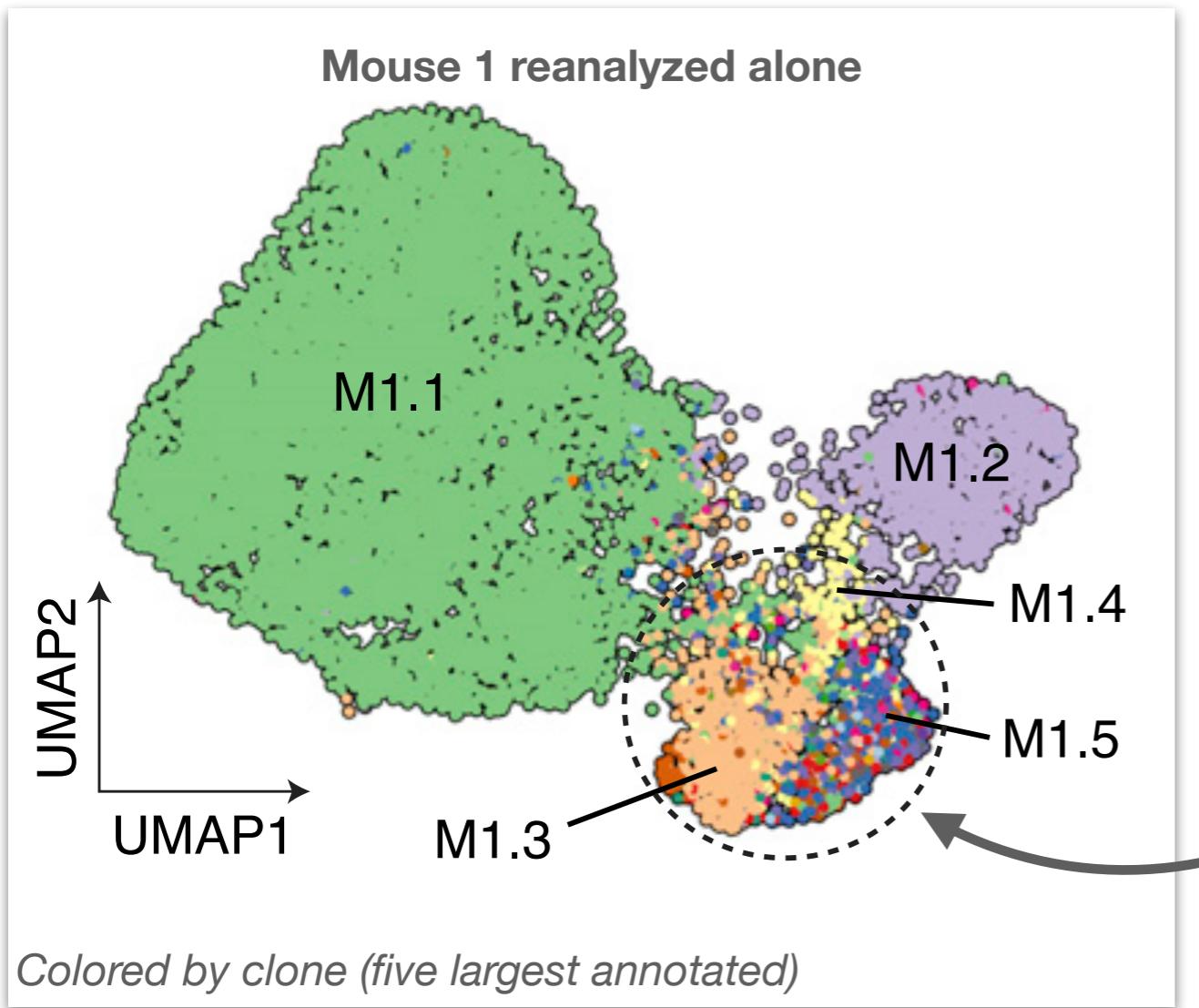
- ① FACS sort GFP+ PDAC cells
- ② sc-Seq transcriptome & barcodes

# Cells retain transcriptional identity after metastasis

- Performed *InferCNV* analysis

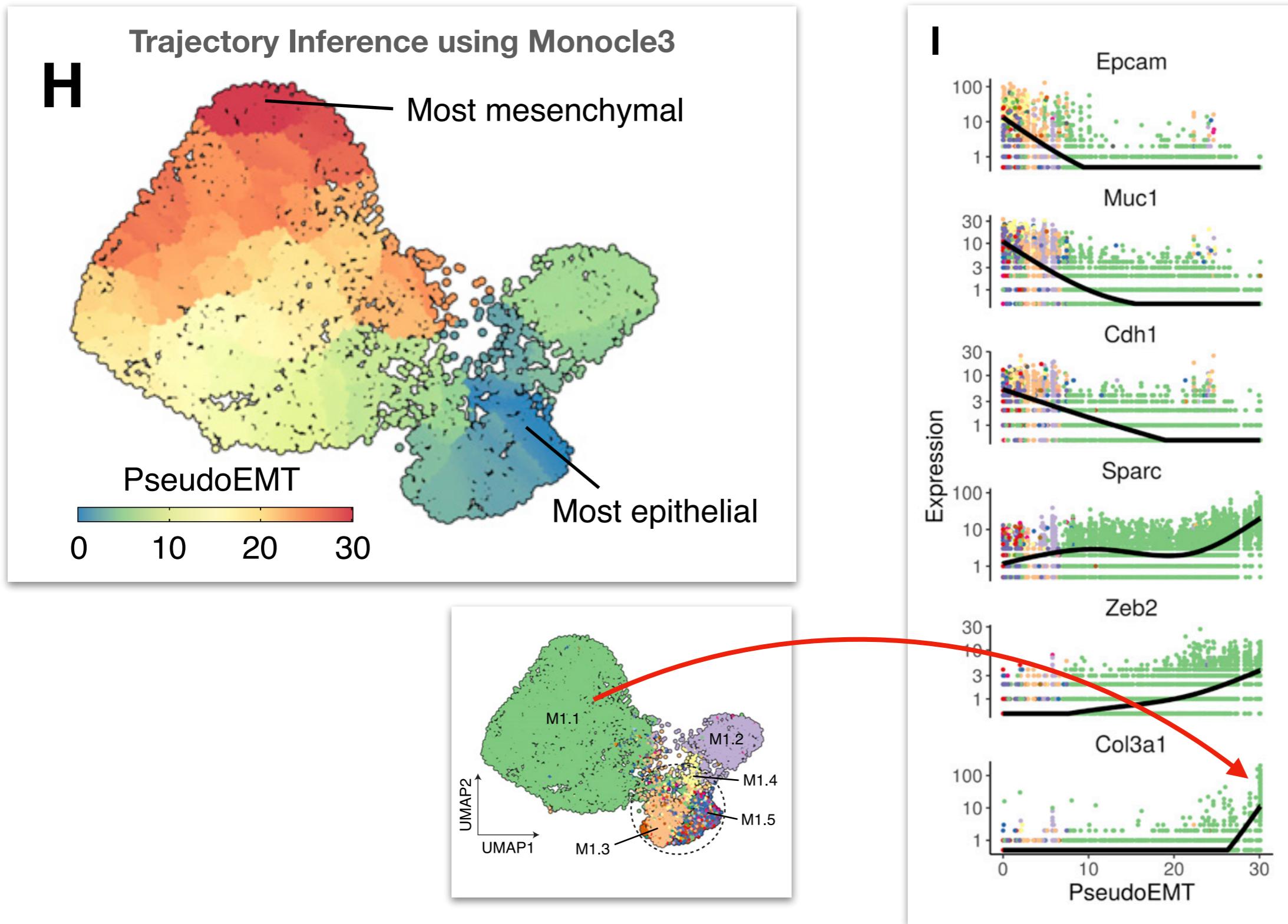


# Transcriptional EMT continuum *in vivo*



From Figure 3 of Simeonov et al., Cancer Cell (2021)

# Transcriptional EMT continuum *in vivo*

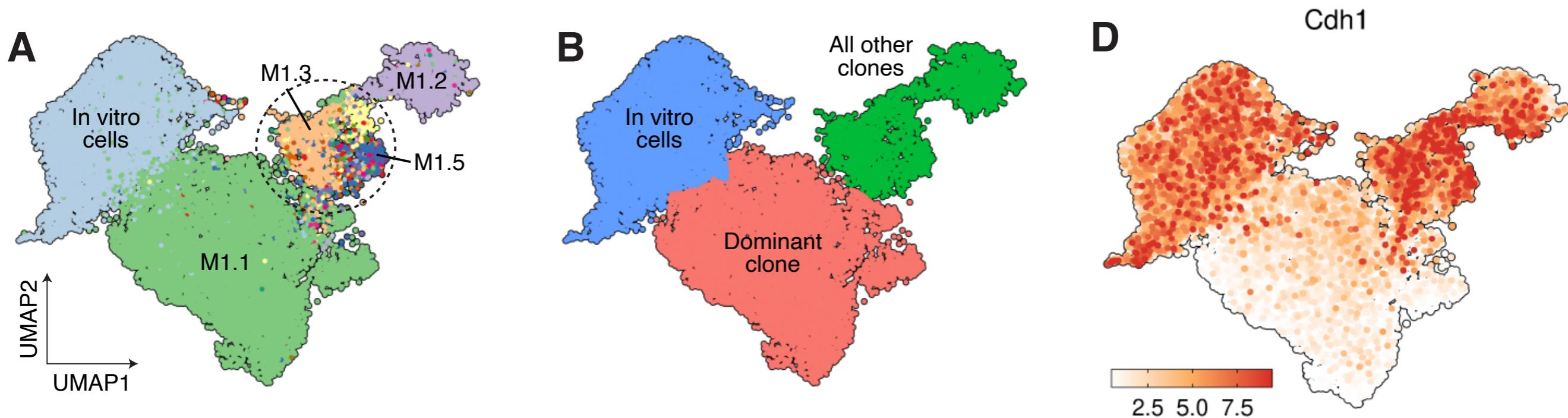


From Figure 3 of Simeonov et al., Cancer Cell (2021)

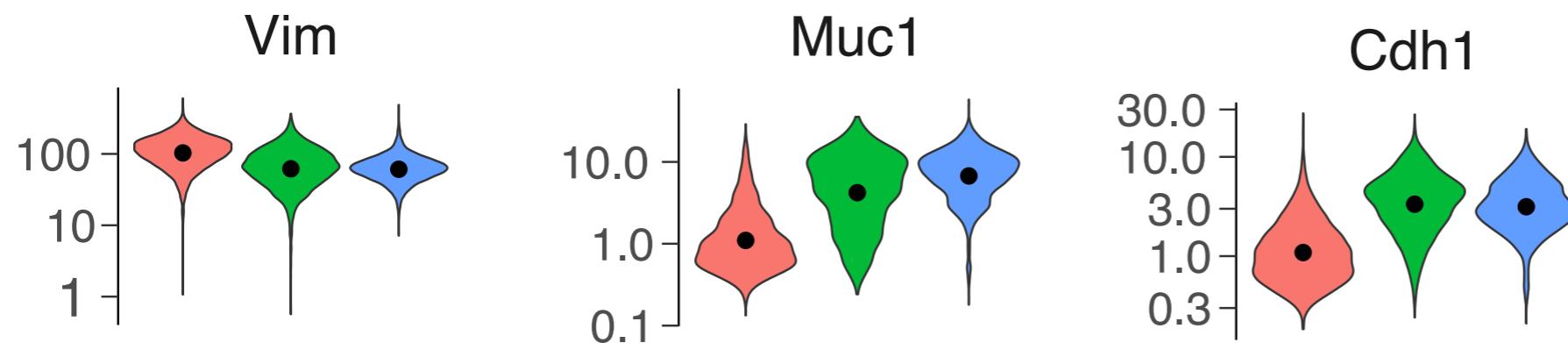


# Transcriptional EMT continuum *in vivo*

- 27/29 clones are epithelial - is this the default transcriptional state?
- scRNA-seq of *in vitro* cultured cells -> 40 distinct clones



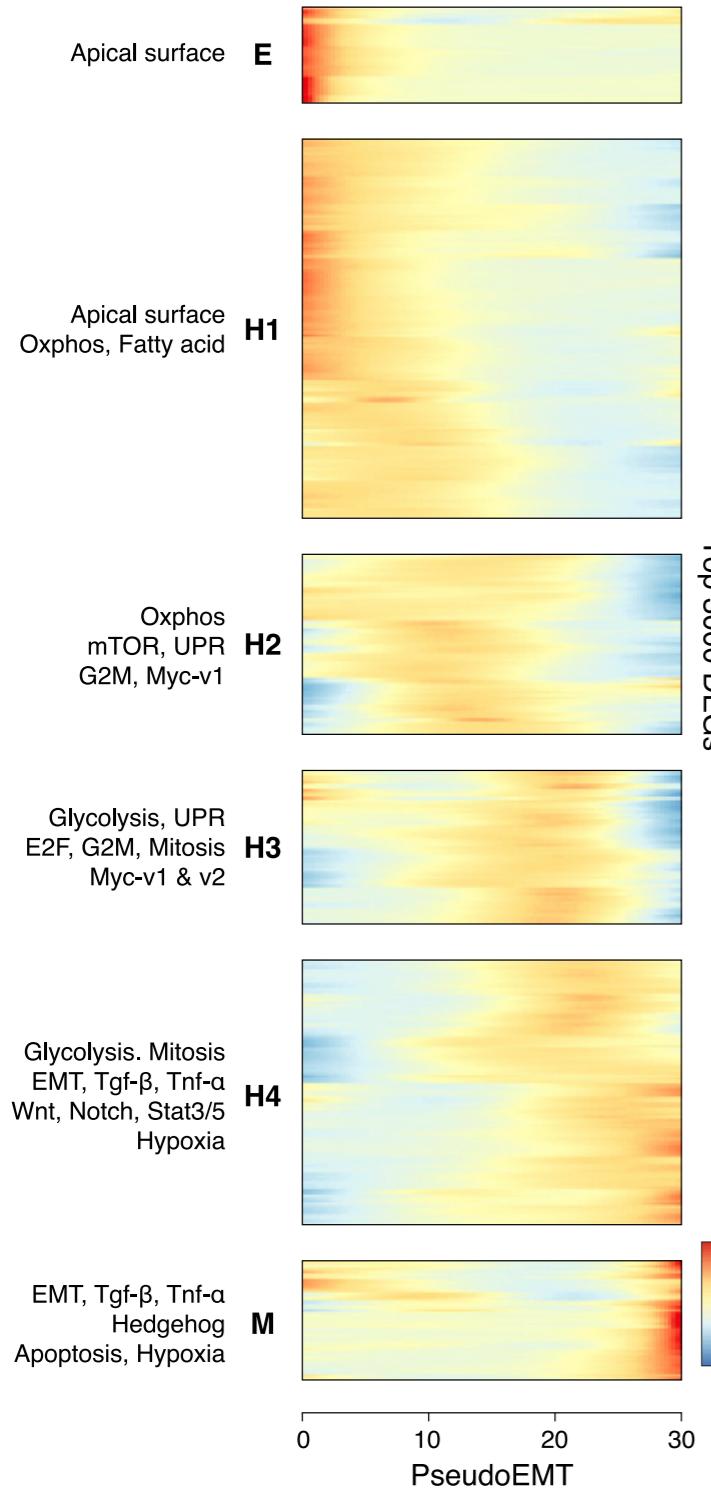
- Compared to PDAC / EMT, *in vitro* cultured cells **strikingly epithelial**



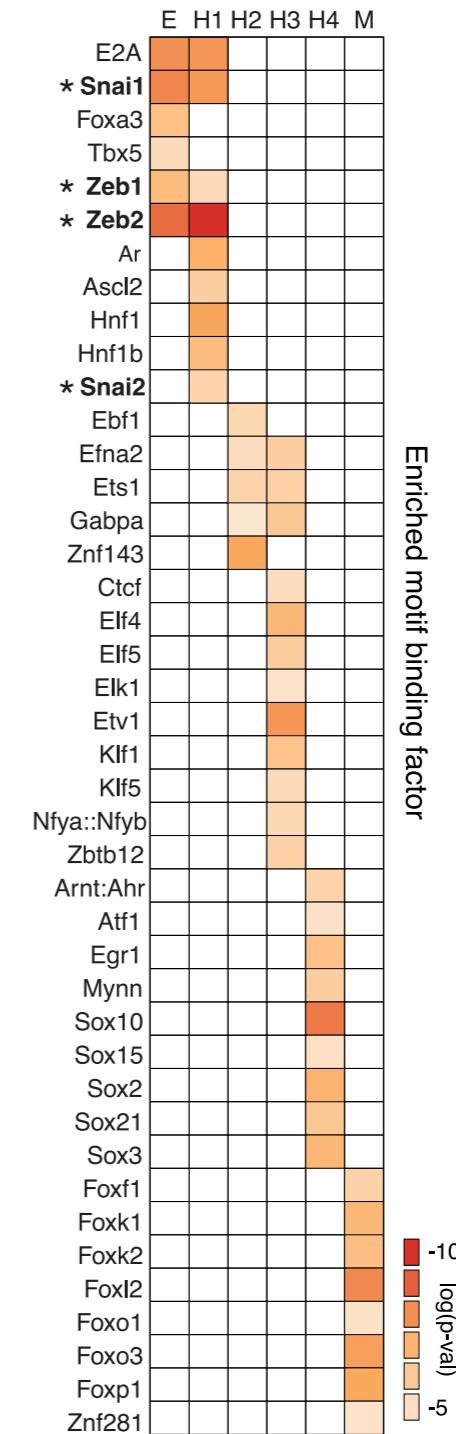
From Figure S5 of Simeonov et al., Cancer Cell (2021)

# Transcriptional EMT continuum *in vivo*

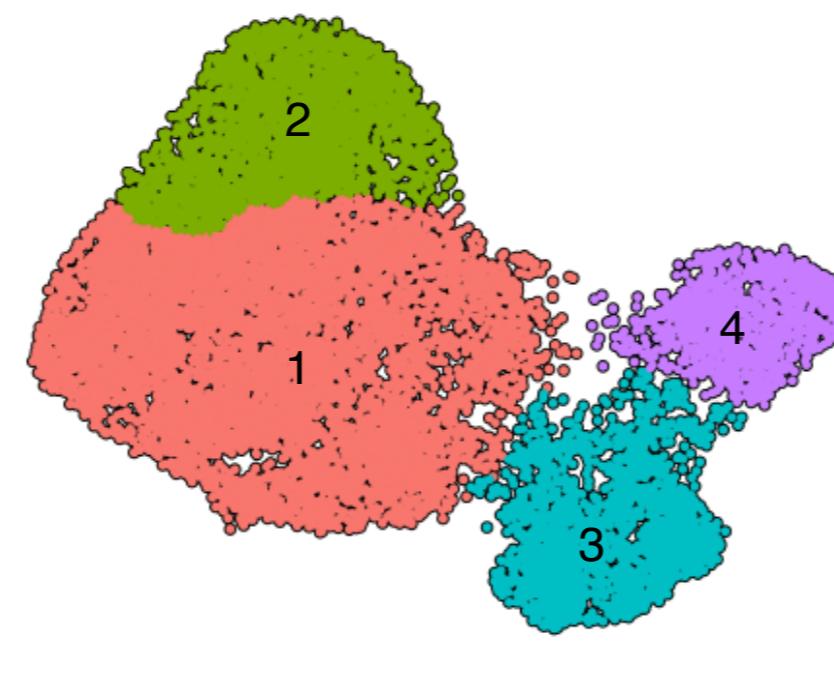
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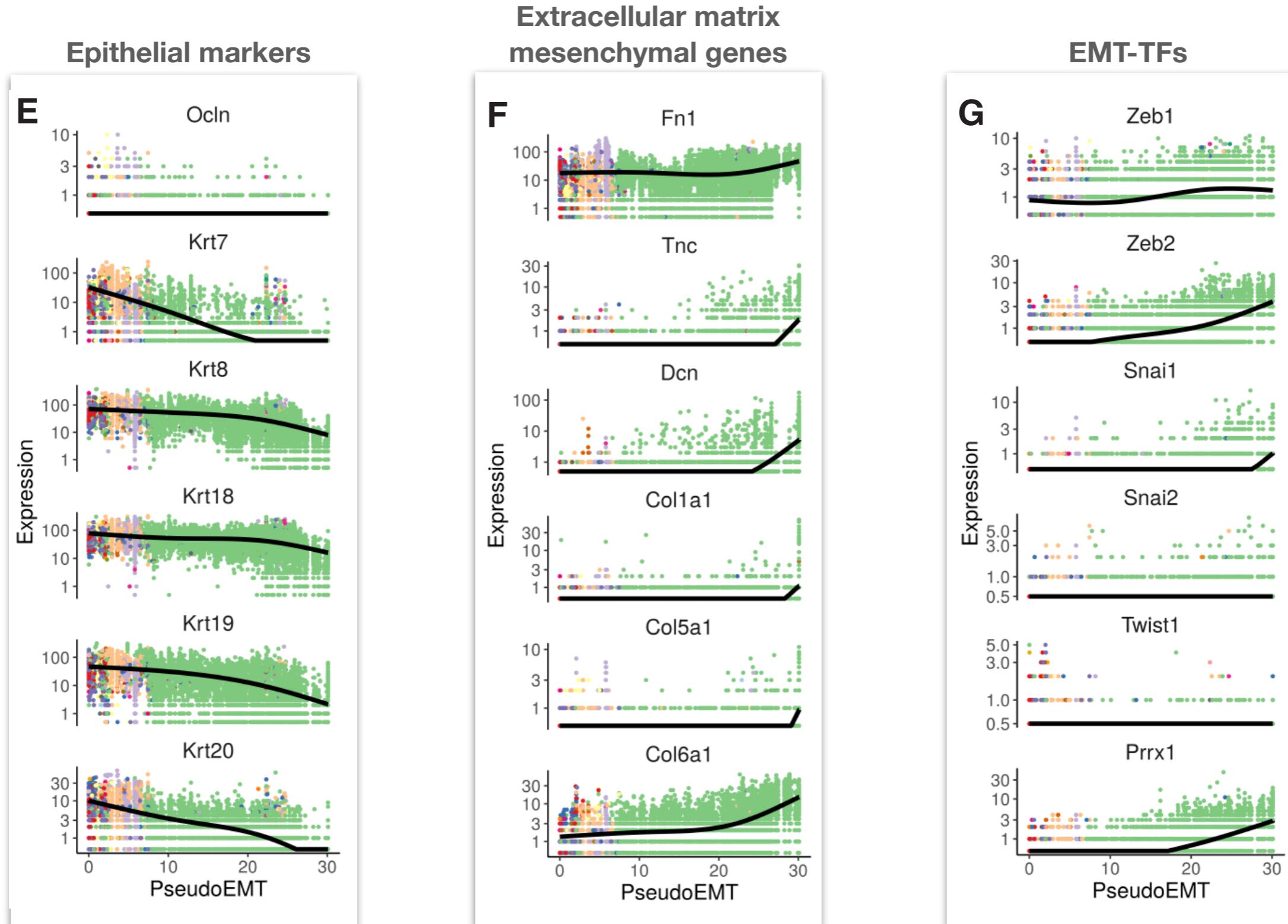
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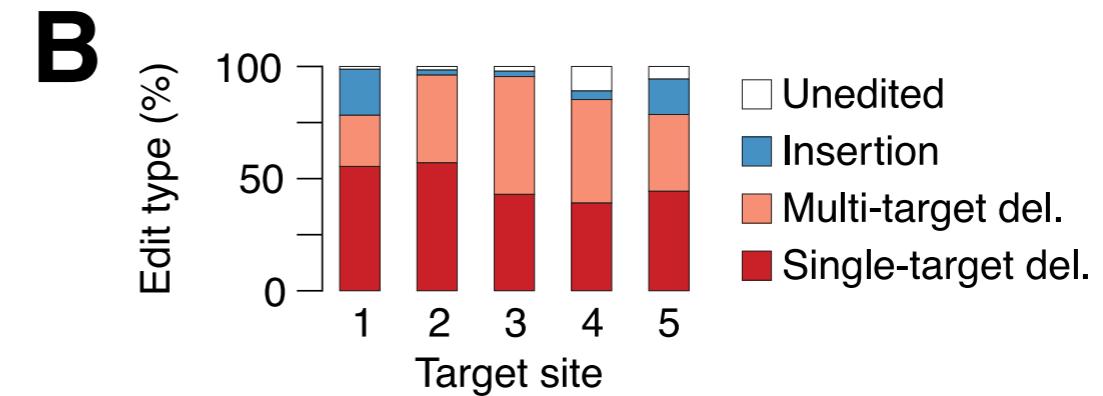
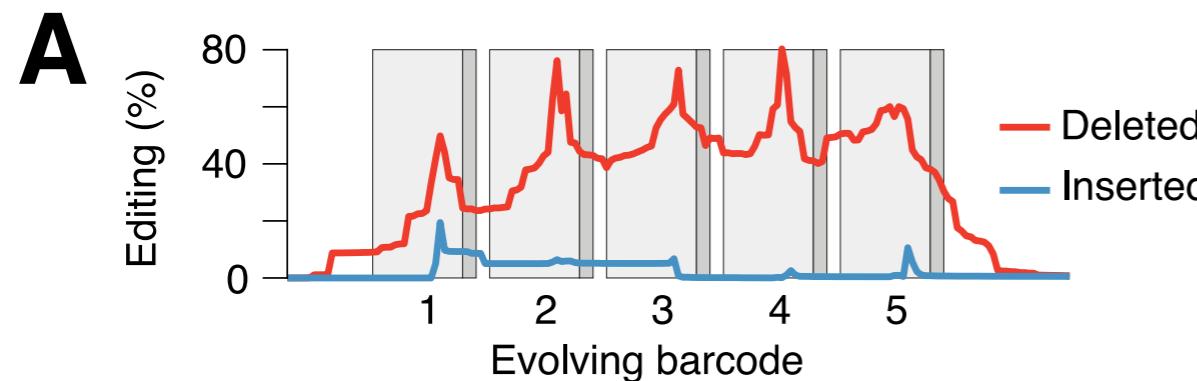
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# Transcriptional EMT continuum *in vivo*

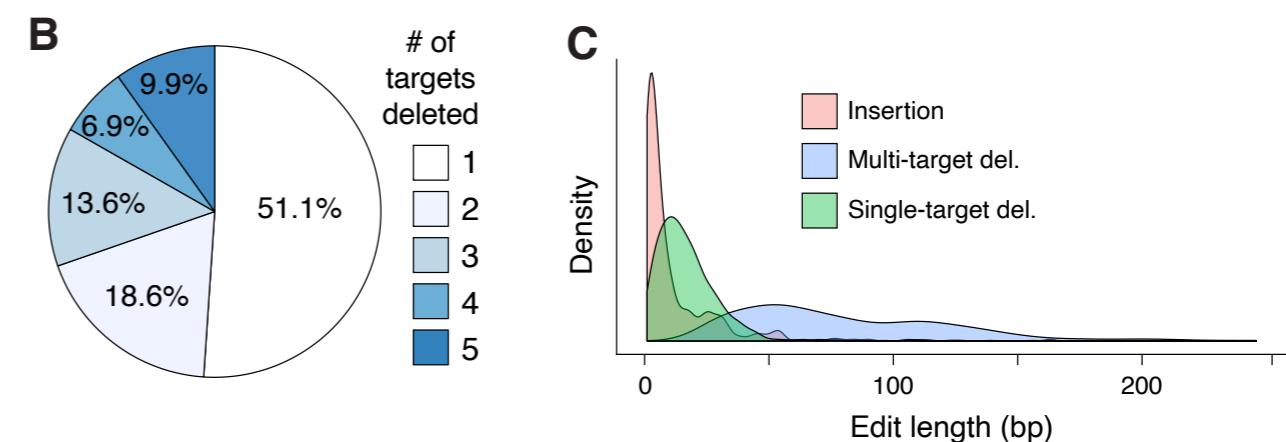


# High-res subclonal lineage reconstruction



**A**

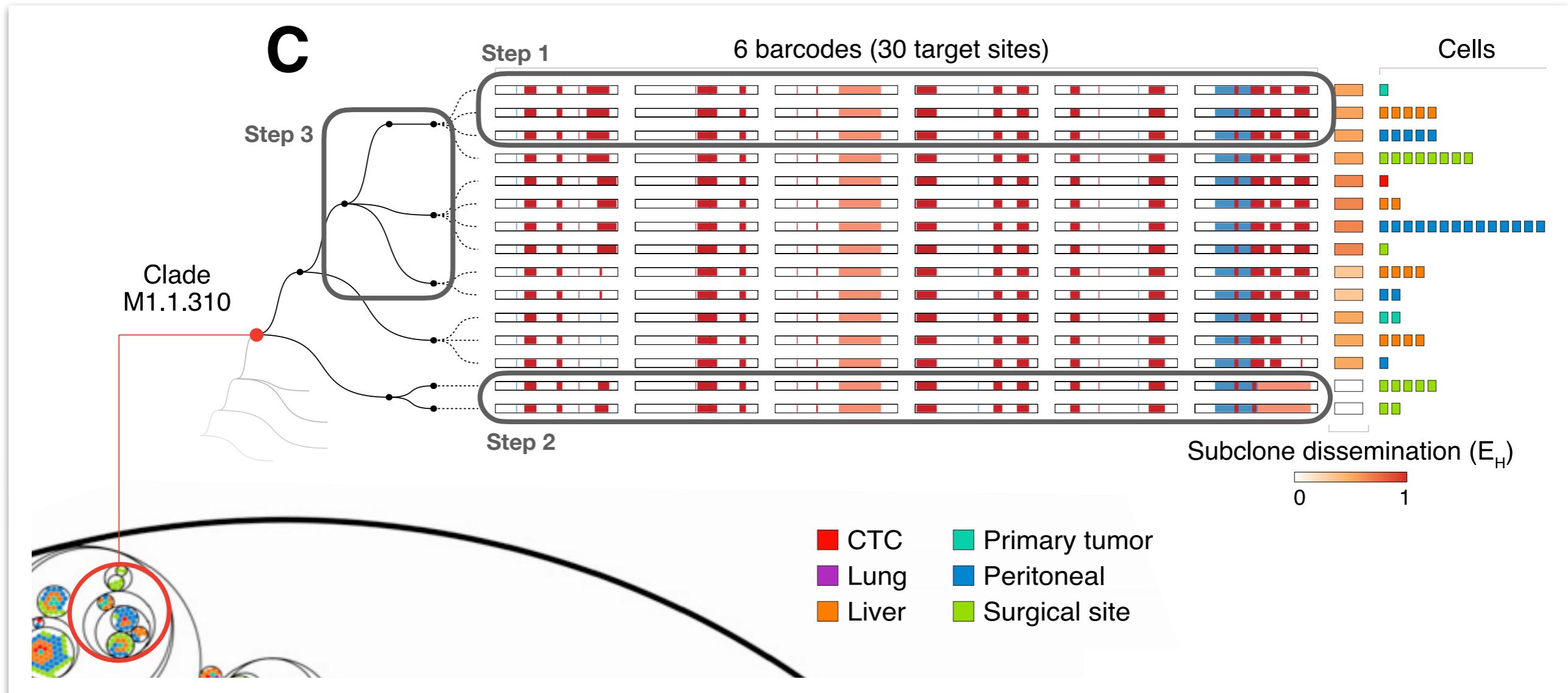
	Mouse 1	Mouse 2	Both mice
Total barcodes	50,269	26,705	76,974
Total barcodes with edit	50,132	25,968	76,100
% of barcodes with edit	99.7%	97.2%	98.9%
Total targets	251,345	133,525	384,870
Total targets with edit	246,232	122,013	368,245
% of targets with edit	98.0%	91.4%	95.7%
Distinct edits	2,104	1,383	3,487 (633)
Distinct evolving barcodes	3,886	1,696	5,582 (61)
Distinct barcode-of-barcodes (subclones)	3,997	2,062	6,059 (4)



Recovered on average:

- **Mouse 1:** 18.5 target sites (3.7 barcodes) / cell
- **Mouse 2:** 8.5 target sites (1.7 barcodes) / cell

# High-res subclonal lineage reconstruction

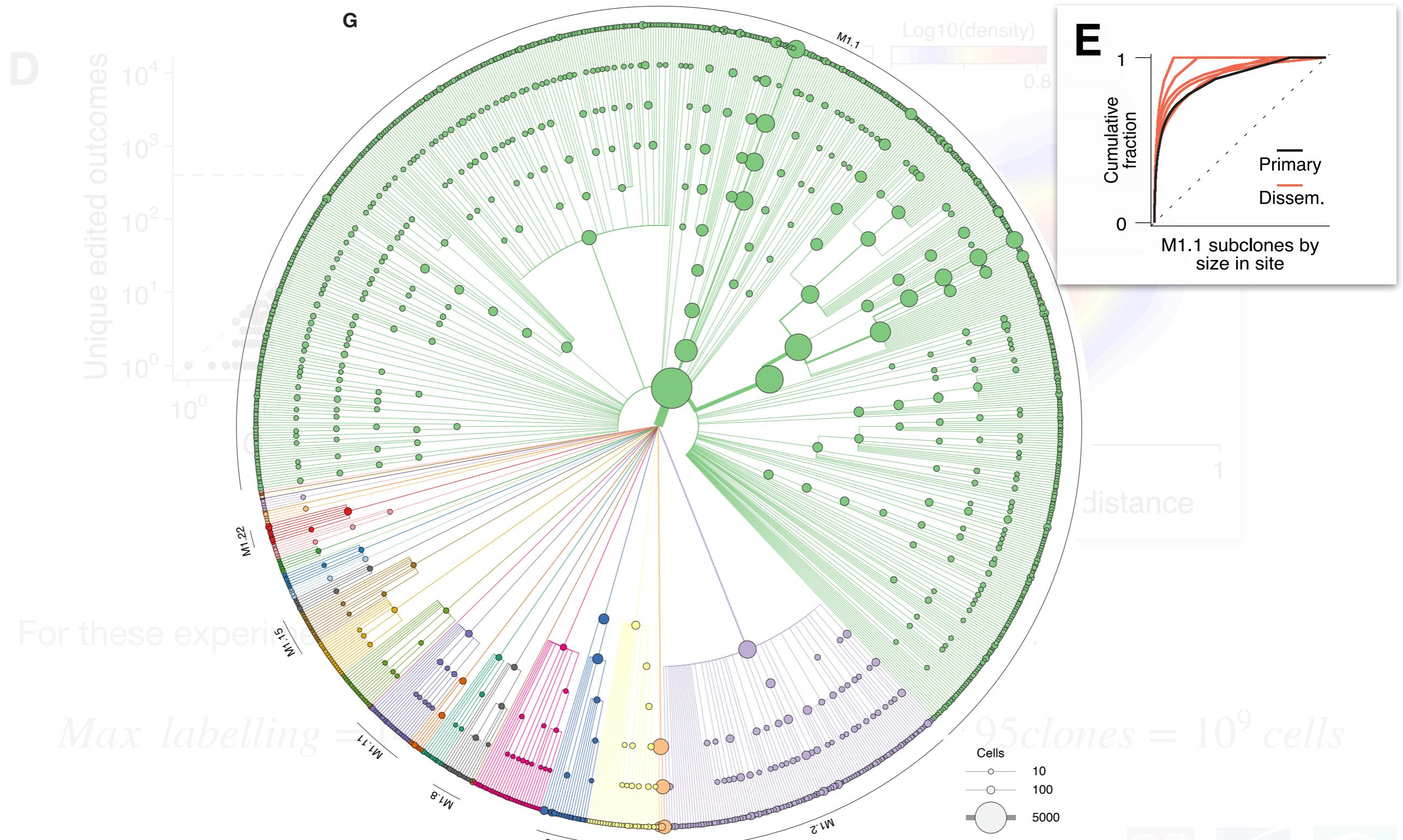


**Step 1:** assemble “barcode of barcodes”

**Step 2:** group cells into subclones

**Step 3:** reconstruct phylogenetic relationships between subclones

# High-res subclonal lineage reconstruction

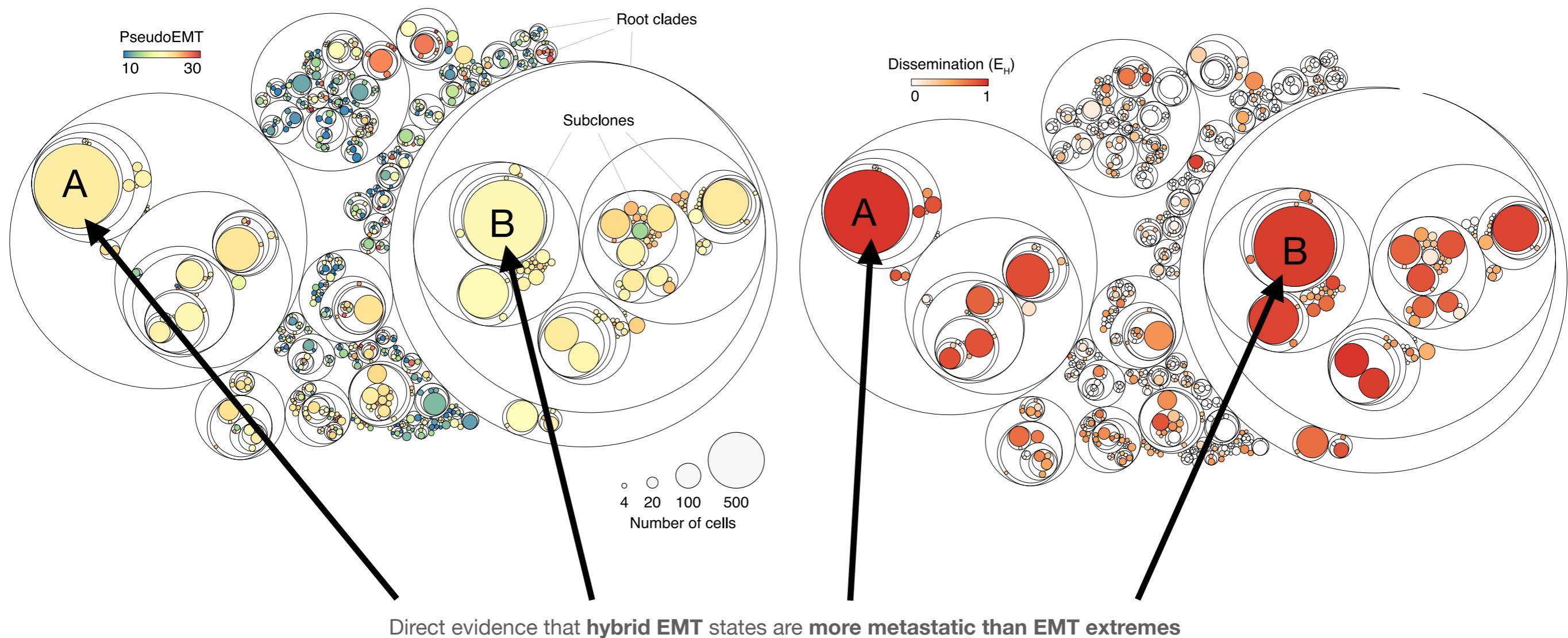


From Figures 4, S6 of Simeonov et al., Cancer Cell (2021)



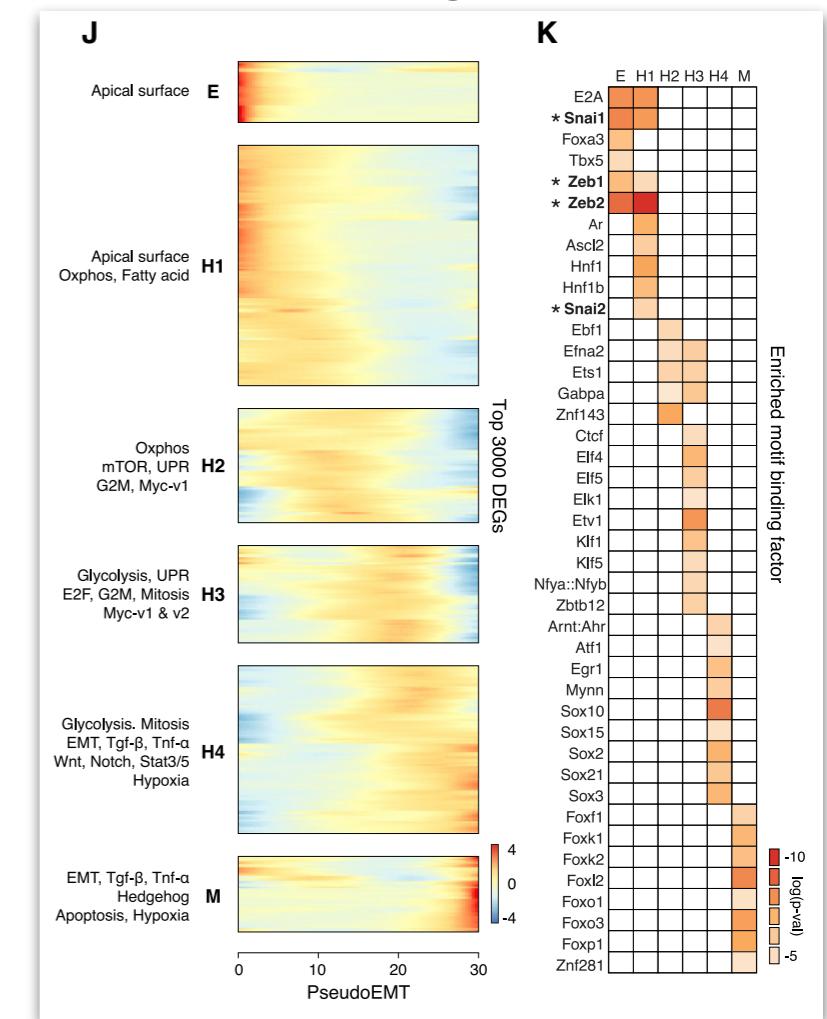
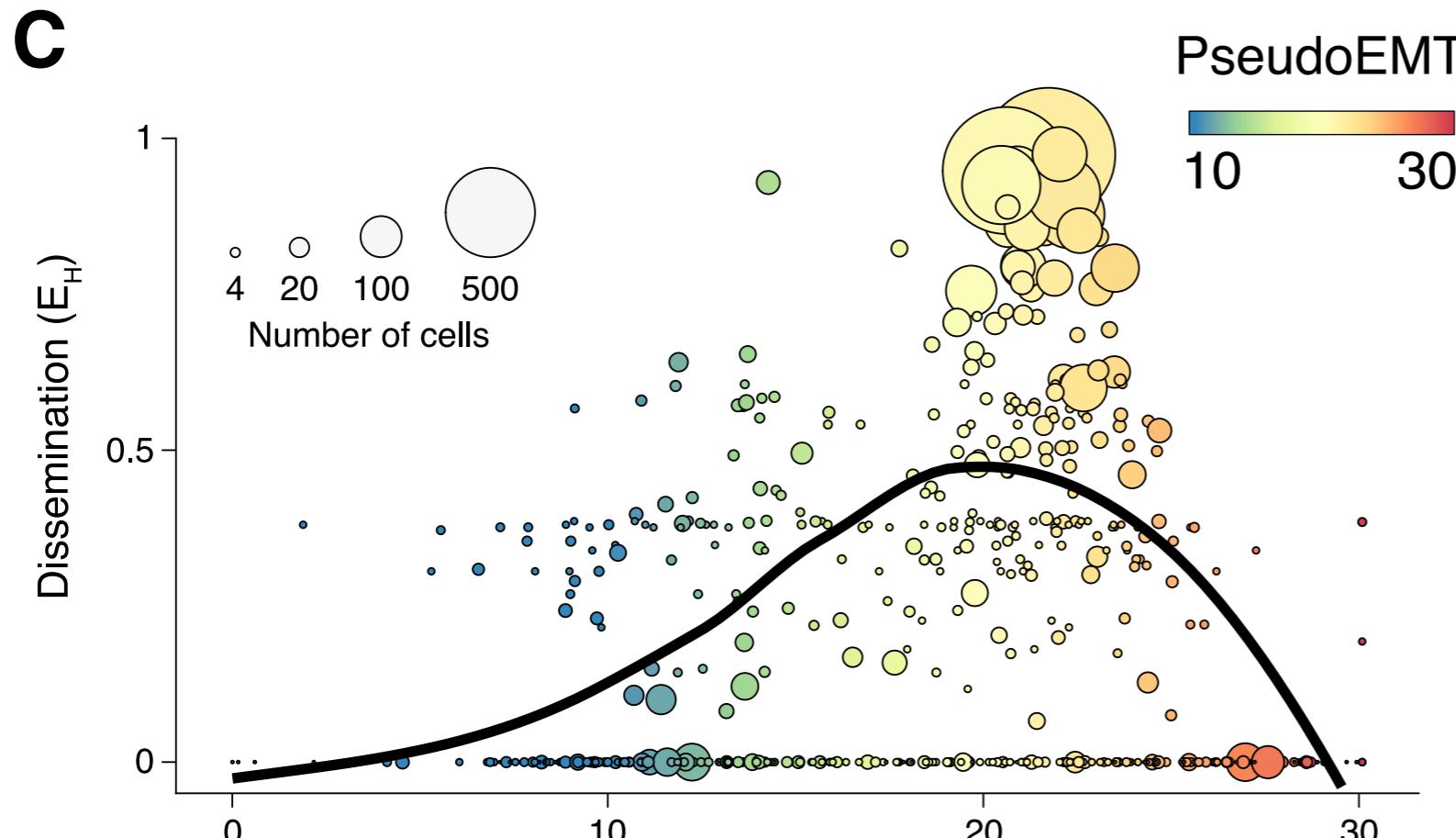
# Peak metastatic aggression corresponds to late-hybrid EMT states

How does the range of **intraclonal EMT states** relate to **subclonal behavior?**



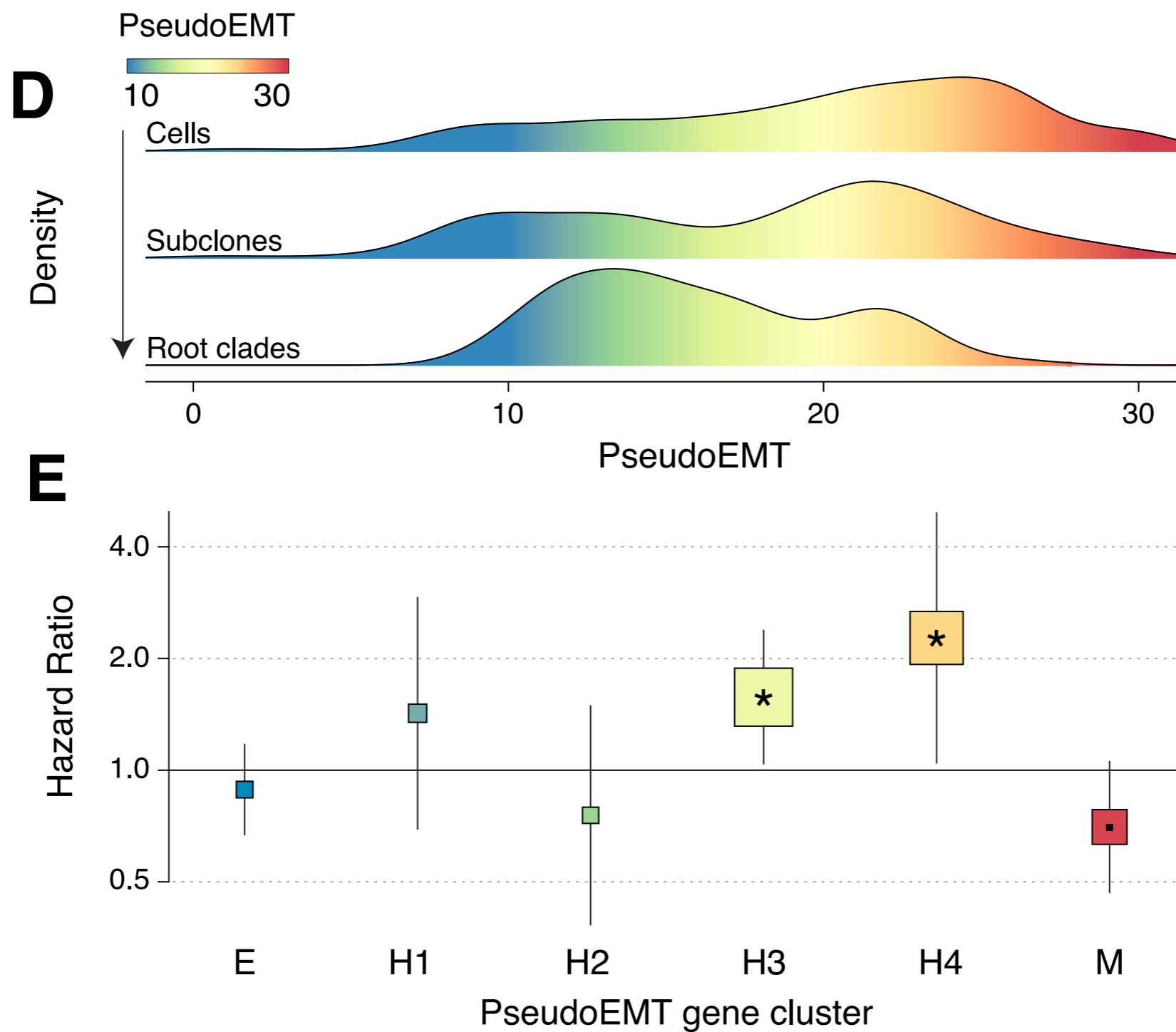
# Peak metastatic aggression corresponds to late-hybrid EMT states

*Remember from Figure 3....*

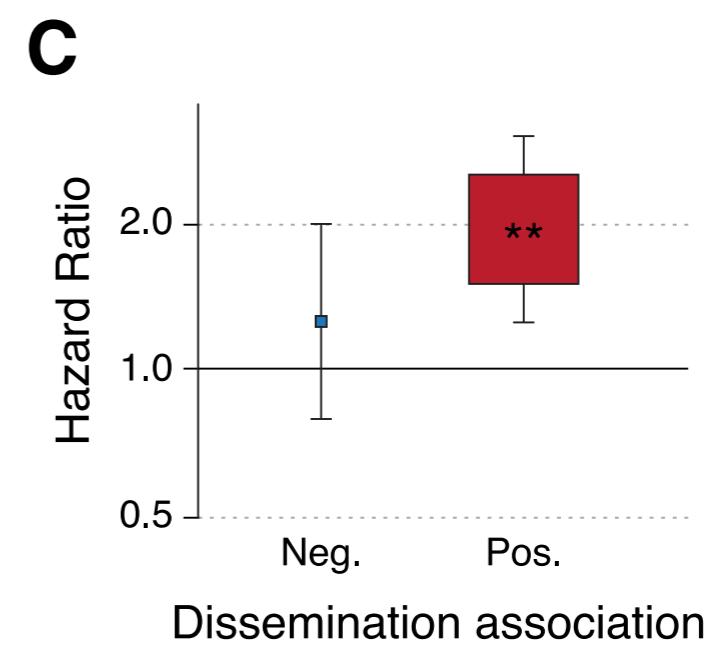
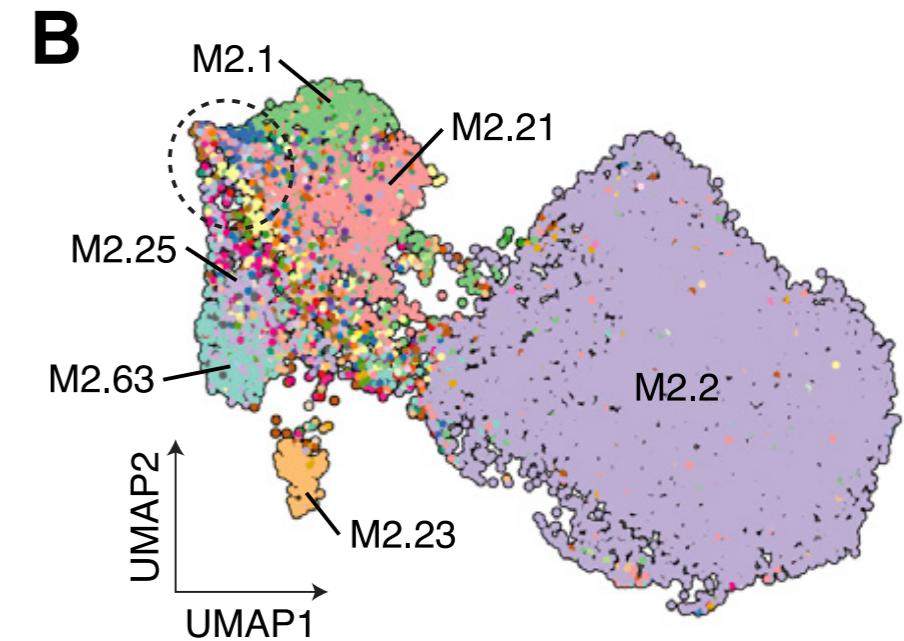
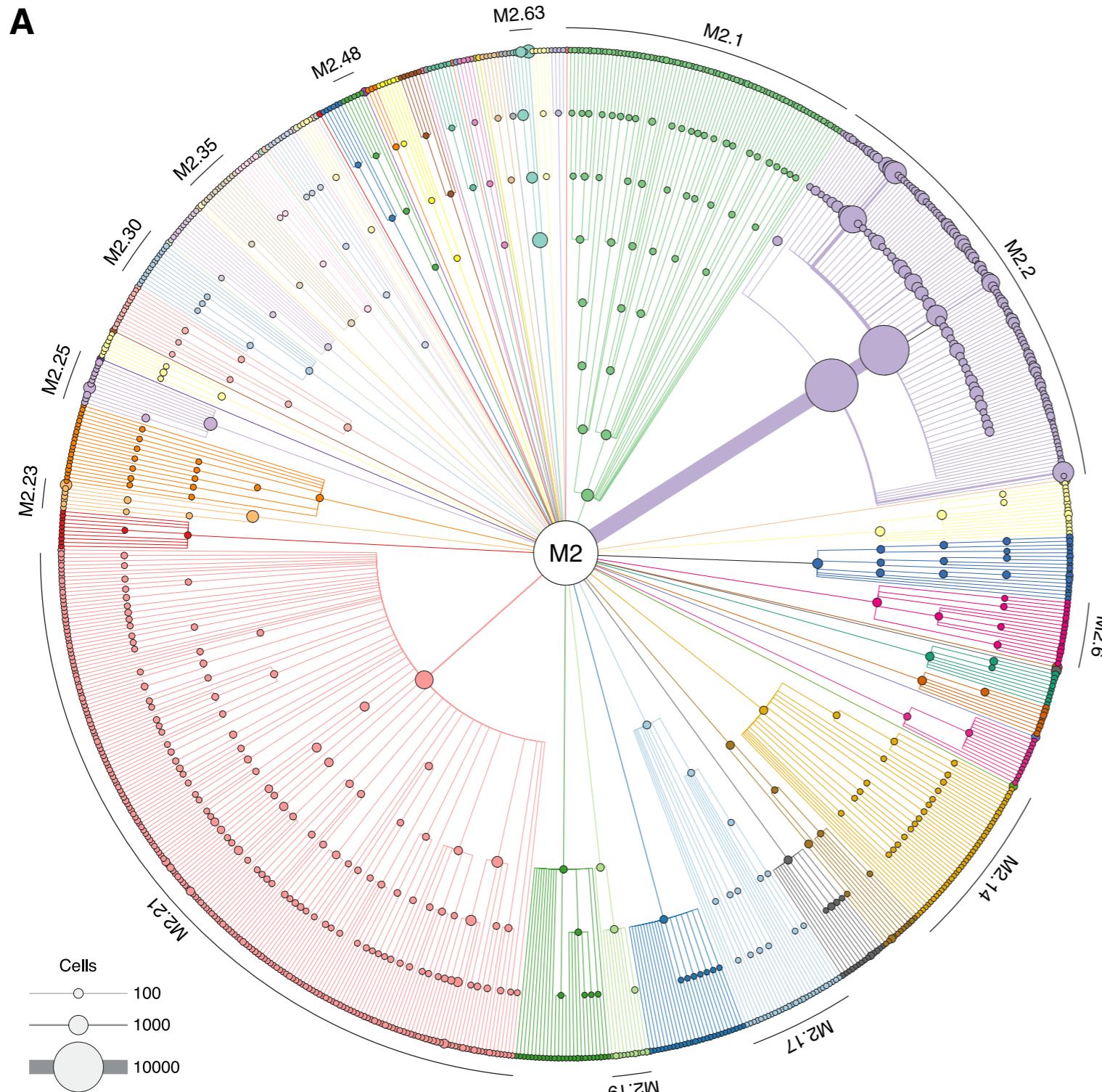


**Important theme:** this paper provides a lot of evidence that EMT, while its own process is very inherent to cancer metastasis - remember we're studying a cancer model not an EMT model

# Peak metastatic aggression corresponds to late-hybrid EMT states

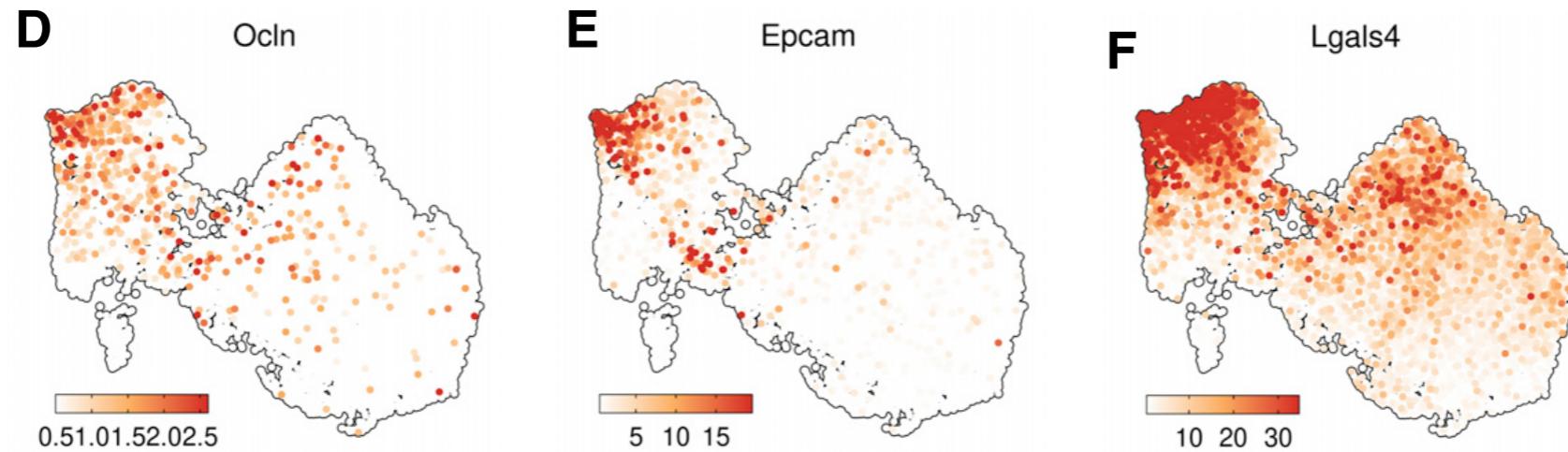


# Complementary process to canonical EMT

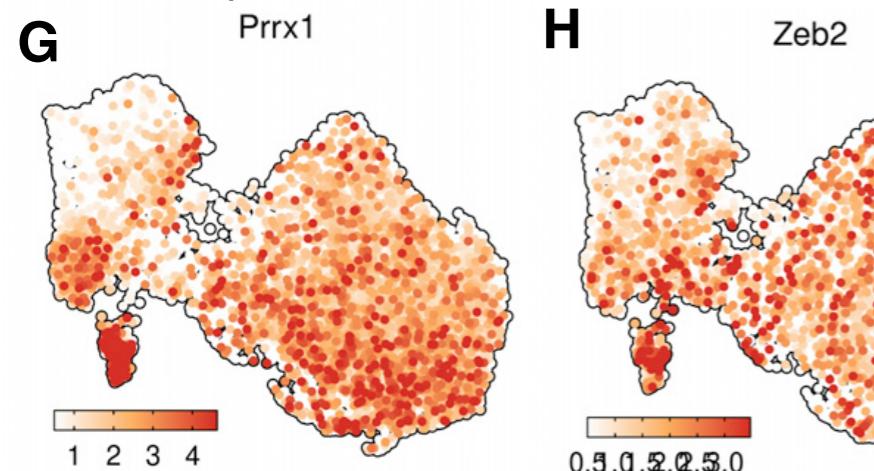


From Figure 6 of Simeonov et al., Cancer Cell (2021)

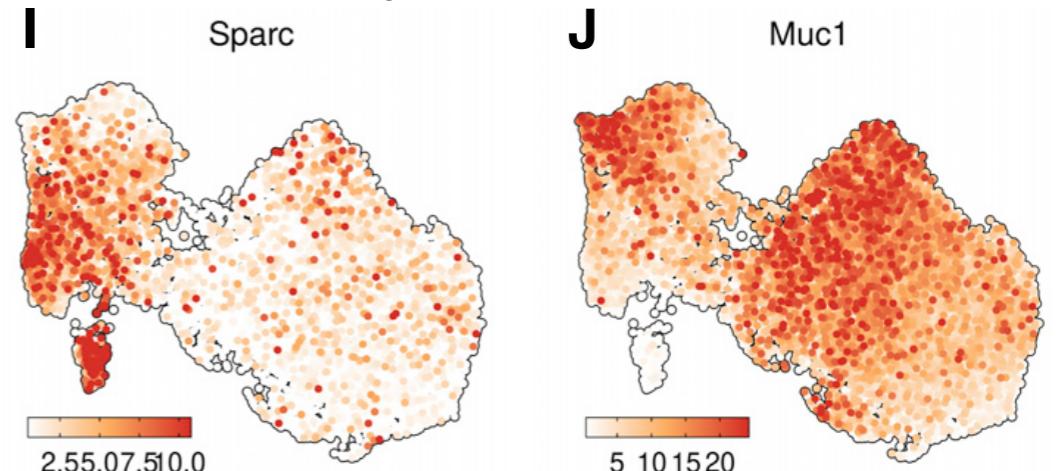
# Complementary process to canonical EMT



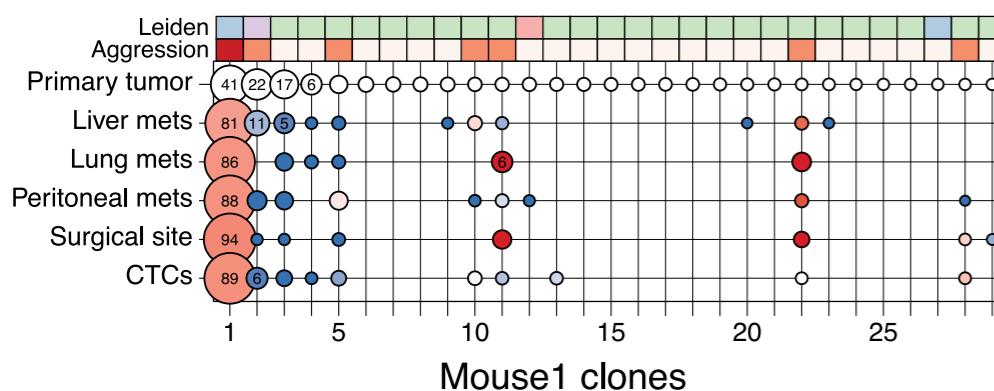
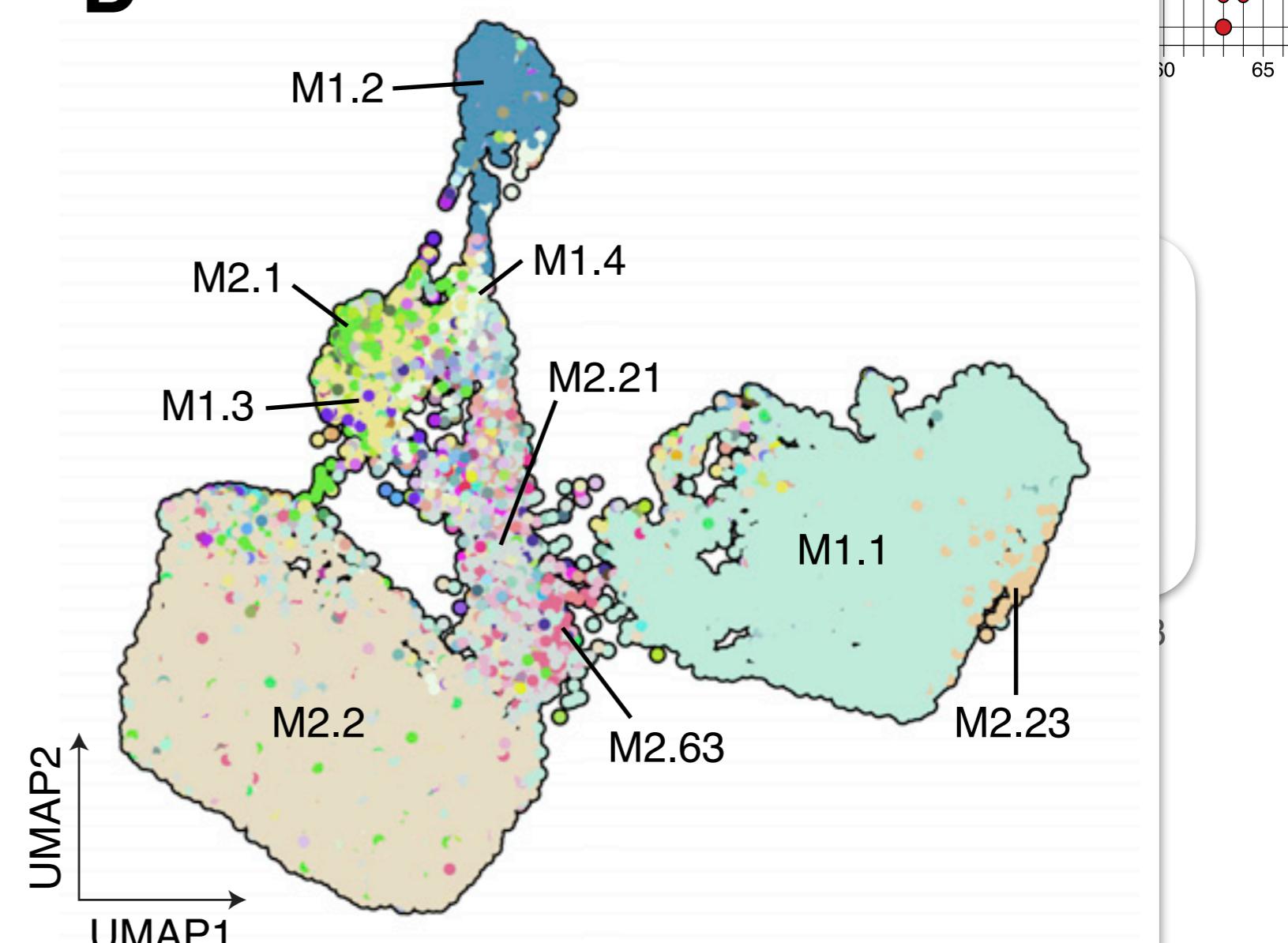
*GEX as expected*



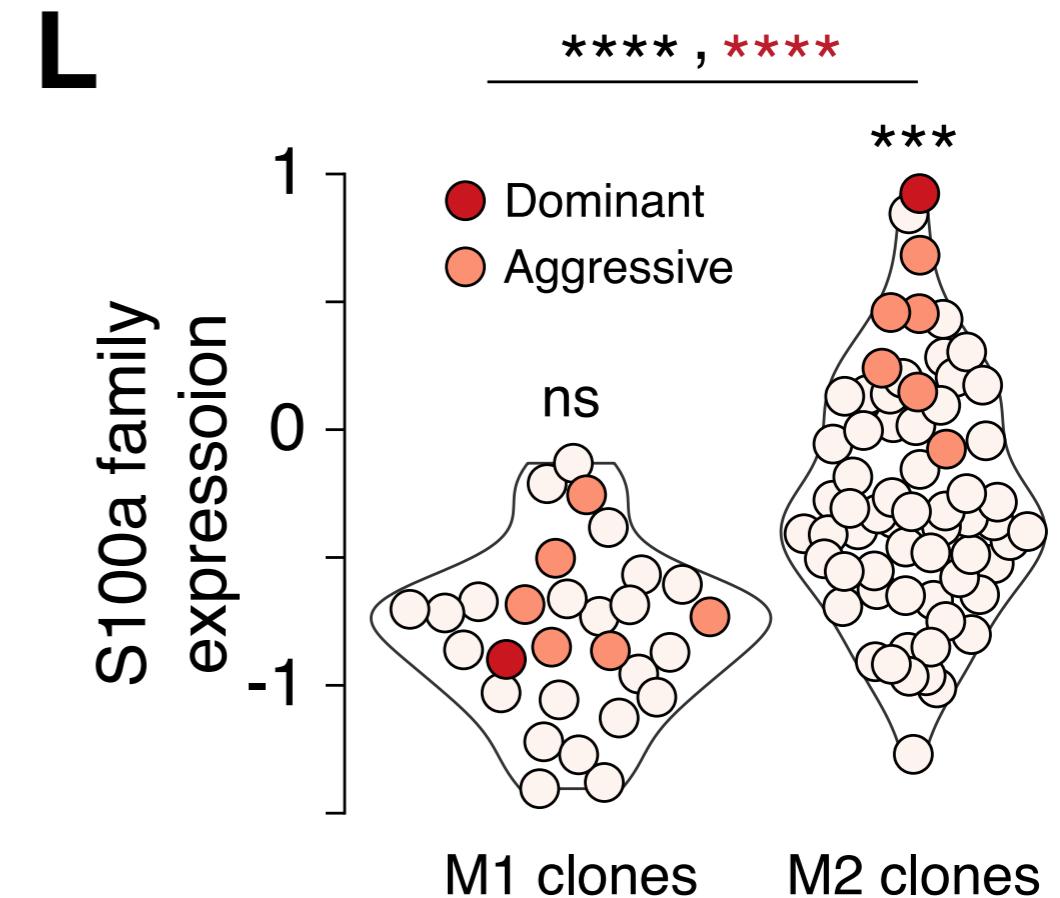
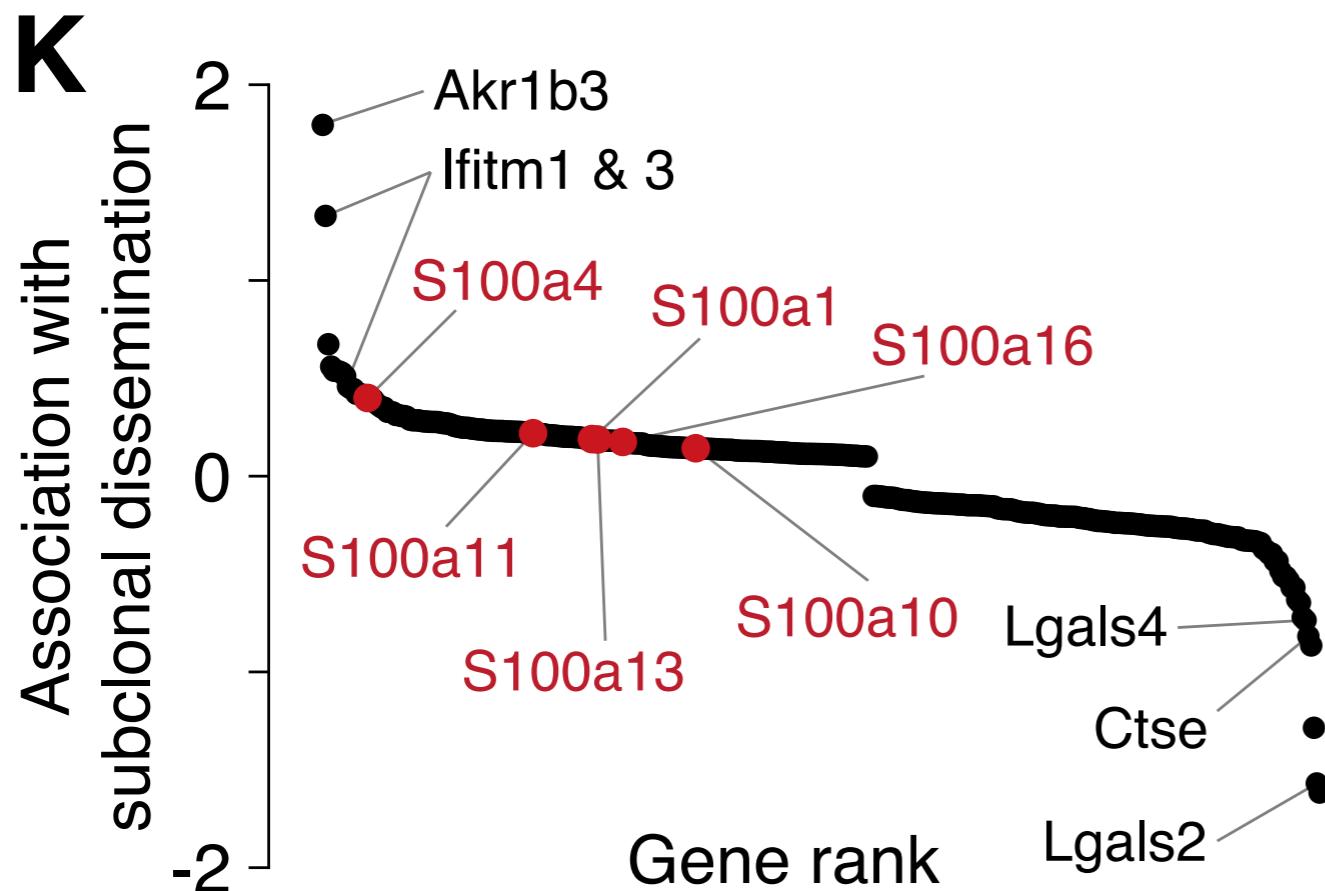
*GEX contradictory*



# Complementary process to canonical EMT

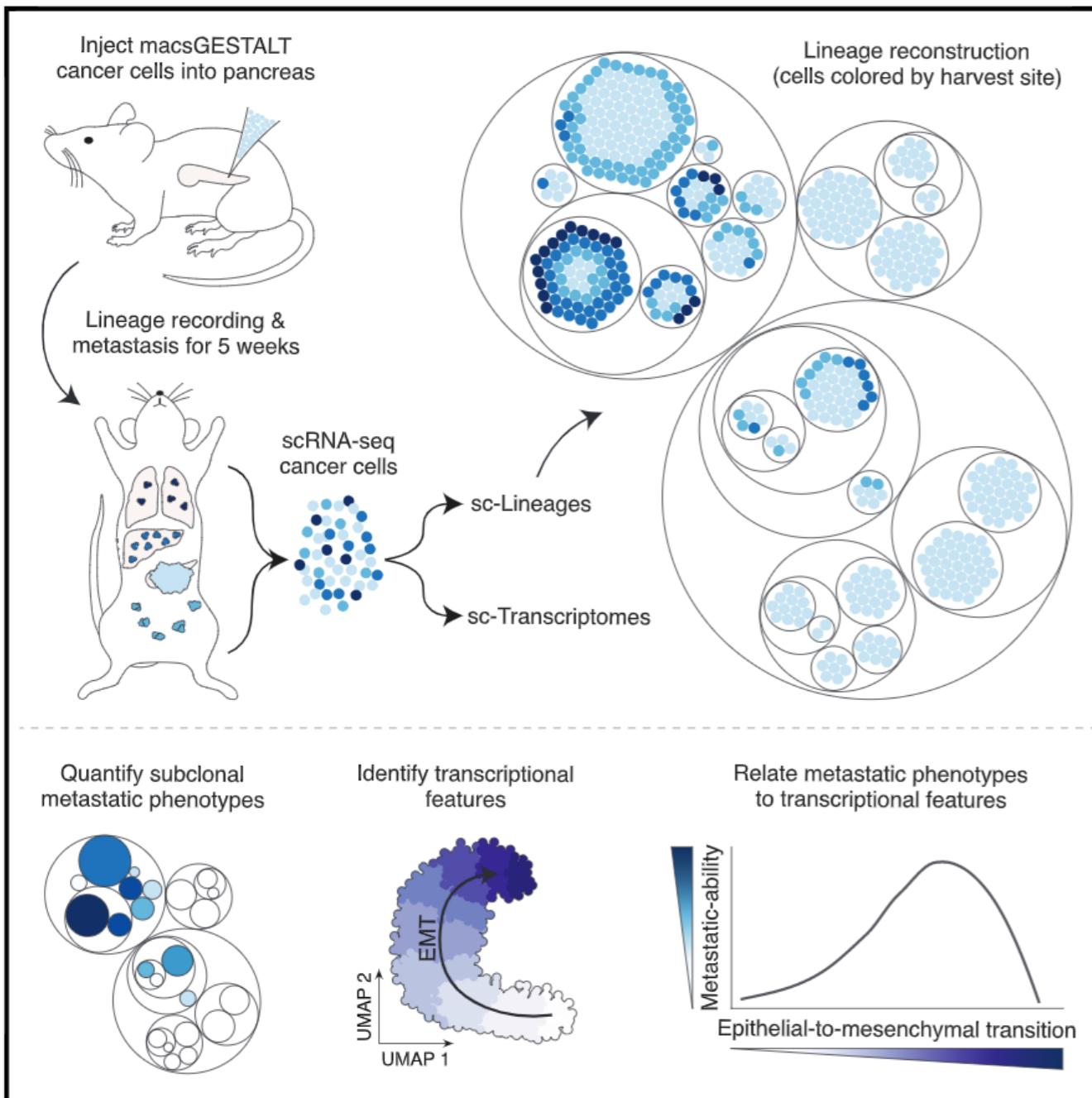
**B****D**

# Complementary process to canonical EMT



# Major contributions of macsGESTALT

## Graphical abstract

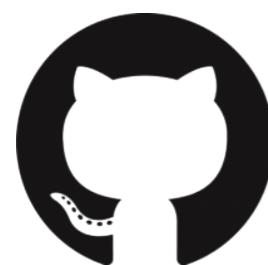


- macsGESTALT is an inducible lineage-recorder
- In vivo model of pancreatic cancer metastasis
- Finding recurrent drivers across cancers remains elusive
- Metastatically competent model wherein most clones do NOT metastasize
- In EMT, metastatic aggression rises and peaks during a late intermediate hybrid stage
- ID gene sets within EMT hybrid stages that are predictive of human survival outcome (in 2 cancers, not in 3)
- S100 genes were found across metastatic subpopulations

# Thank you for listening!

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Access all sides with links  
to papers and notes:



**GitHub.com/mvinyard/vintools**

