

summary_file

My Linh Thibodeau

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Library and reading tables

```
suppressMessages(suppressWarnings(library(tidyverse)))
knitr::opts_chunk$set(fig.width=12, fig.height=9)
library(knitr)
library(kableExtra)
```

```
## Warning: package 'kableExtra' was built under R version 3.4.2
```

```
options(knitr.table.format = "html")
mut_sig <- read.table("mut_sig.tsv", header = TRUE, sep = "\t")
ref_mut_sig_ordered_by_sig3 <- readRDS("somatic_mutations_formated_files/ref_mut_sig_ordered_by_sig3.rds")
mut_sig_gather <- read.table("mut_sig_gather.tsv", header = TRUE, sep="\t")
ALL_mut_gather <- read.table("somatic_mutations_formated_files/ALL_mut_gather.tsv", header = TRUE, sep = "\t")
aml_mut_gather <- read.table("somatic_mutations_formated_files/aml_mut_gather.tsv", header = TRUE, sep = "\t")
breast_mut_gather <- read.table("somatic_mutations_formated_files/breast_mut_gather.tsv", header = TRUE, sep = "\t")
medullo_mut_gather <- read.table("somatic_mutations_formated_files/medulloblastoma_mut_gather.tsv", header = TRUE, sep = "\t")
pancreas_mut_gather <- read.table("somatic_mutations_formated_files/pancreas_mut_gather.tsv", header = TRUE, sep = "\t")
all_cancer_types_mut <- read.table("somatic_mutations_formated_files/all_cancer_types_mut.tsv", header = TRUE, sep = "\t")
all_cancer_types_mut_with_ref_sig <- read.table("somatic_mutations_formated_files/all_cancer_types_mut_with_ref_sig.tsv", header = TRUE, sep = "\t")
all_cancer_mutations_per_snv_sig_score <- read.table("somatic_mutations_formated_files/all_cancer_mutations_per_snv_sig_score.tsv", header = TRUE, sep = "\t")
all_cancer_types_mut_proportion_signatures <- read.table("somatic_mutations_formated_files/all_cancer_types_mut_proportion_signatures.tsv", header = TRUE, sep = "\t")
all_cancer_types_stats <- read.table("statistics/all_cancer_types_stats.tsv", header = TRUE, sep = "\t")
```

DATA INFORMATION

For this homework, I will be using open access complete set of cancer somatic mutations from:

Alexandrov, L. B. et al. Signatures of mutational processes in human cancer. 500, 415–421 (2013).

Please note that I deliberately broke down each step of this homework in very small chunks/steps/scripts to make this material more “generalizable”, which will allow me to use them for my research work as well.

This files will contain a narrative of homework 7 and some helpful notes regarding my process.

Download the data

I started by using curl shell script in my Makefile to download the complete set of cancer somatic mutations (see Makefile for details):

```
mut_sig_raw.txt:
    curl -o mut_sig_raw.txt ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/signatures.txt
```

I am also using my `mut_sig_clean.R` script to format the data. I performed that step to remove empty spaces from the column names because this would create problems with variable manipulation later.

Perform exploratory analyses

Tasks:

- Bring the data in as data frame.
- Save a couple descriptive plots to file with highly informative names.
- Reorder the continents based on life expectancy. You decide the details. Sort the actual data in a deliberate fashion. You decide the details, but this should at least implement your new continent ordering.
- Write the Gapminder data to file(s), for immediate and future reuse.

Please note that I did not complete the 5 tasks above in the order suggested because it made more sense to proceed in the order outlined below with my dataset !

The original format of the reference mutation signature is:

```
mut_sig %>% arrange(Somatic.Mutation.Type) %>% head(5) %>% kable()
```

Substitution.Type

Trinucleotide

Somatic.Mutation.Type

Signature.1A

Signature.1B

Signature.2

Signature.3

Signature.4

Signature.5

Signature.6

Signature.7

Signature.8

Signature.9

Signature.10

Signature.11

Signature.12

Signature.13

Signature.14

Signature.15

Signature.16

Signature.17

Signature.18

Signature.19

Signature.20
Signature.21
Signature.R1
Signature.R2
Signature.R3
Signature.U1
Signature.U2
C>A
ACA
A[C>A]A
0.0112
0.0104
0.0105
0.0240
0.0365
0.0149
0.0017
4e-04
0.0368
0.0120
0.0007
2e-04
0.0077
0.0007
0.0001
0.0013
0.0161
0.0018
0.0500
0.0107
0.0013
1e-04
0.0210
0.0137
0.0044
0.0105

0.0221
C>A
ACC
A[C>A]C
0.0092
0.0093
0.0061
0.0197
0.0309
0.0089
0.0028
5e-04
0.0287
0.0067
0.0010
1e-03
0.0047
0.0001
0.0042
0.0040
0.0097
0.0003
0.0076
0.0074
0.0024
7e-04
0.0065
0.0046
0.0047
0.0005
0.0123
C>A
ACG
A[C>A]G
0.0015
0.0016

0.0013
0.0019
0.0183
0.0022
0.0005
0e+00
0.0017
0.0005
0.0003
0e+00
0.0017
0.0001
0.0005
0.0000
0.0022
0.0000
0.0017
0.0005
0.0000
0e+00
0.0000
0.0048
0.0003
0.0000
0.0028
C>A
ACT
A[C>A]T
0.0063
0.0067
0.0037
0.0172
0.0243
0.0092
0.0019
4e-04

0.0300
0.0068
0.0092
2e-04
0.0046
0.0002
0.0296
0.0057
0.0088
0.0032
0.0181
0.0074
0.0029
6e-04
0.0058
0.0081
0.0034
0.0112
0.0118
C>G
ACA
A[C>G]A
0.0018
0.0051
0.0048
0.0216
0.0097
0.0117
0.0013
0e+00
0.0085
0.0048
0.0005
7e-04
0.0031
0.0018

0.0001
 0.0011
 0.0048
 0.0016
 0.0014
 0.0058
 0.0005
 5e-04
 0.0038
 0.0024
 0.0012
 0.0044
 0.0108

- (1) I am using my `mut_sig_tables.Rmd` script bring the mutational signatures in as a data frame. I am using the `gather` function of `tidyverse` to create a new dataframe table which will facilitate making plots later. I saved the output of this new data frame to `mut_sig_gather.tsv`.

```
mut_sig_gather %>% arrange(Somatic.Mutation.Type) %>% head(5) %>% kable()
```

Substitution.Type	Trinucleotide	Somatic.Mutation.Type	Signature	Score
C>A	ACA	A[C>A]A	Signature.1A	0.0112
C>A	ACA	A[C>A]A	Signature.1B	0.0104
C>A	ACA	A[C>A]A	Signature.2	0.0105

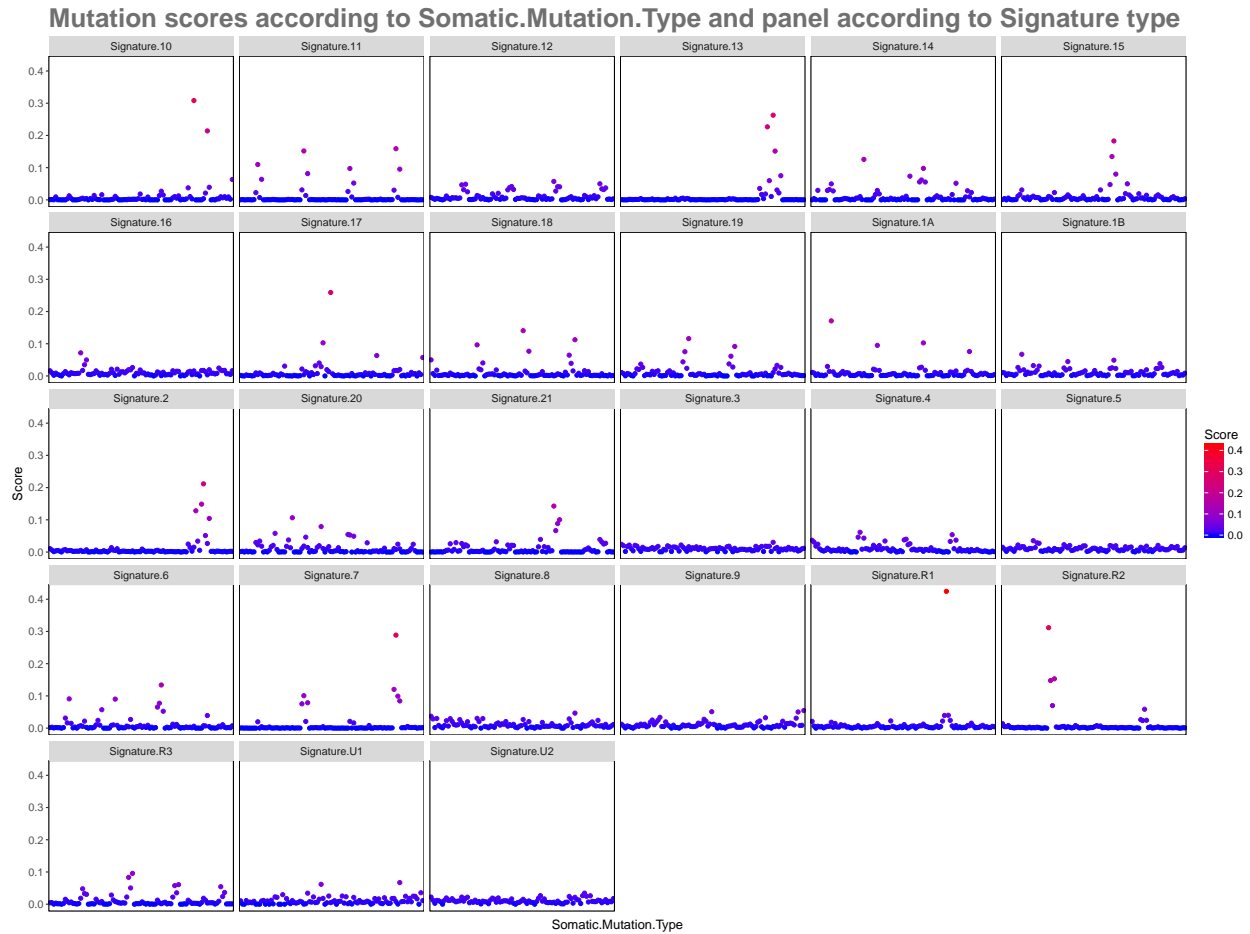


Figure 1: mutation_scores_for_all_snv_facet_signature

C>A

ACA

A[C>A]A

Signature.3

0.0240

C>A

ACA

A[C>A]A

Signature.4

0.0365

Note. This allows us to know how much each Somatic.Mutation.Type contributes to each signature.

(2) Here are some initial plots

(3) In this section, I am using the mut_sig_plot.Rmd script to create some plots, which I save in the subfolder called plots. I am reordering the Somatic.Mutation.Type (e.g. A[C>A]A, A[C>A]C, etc.)

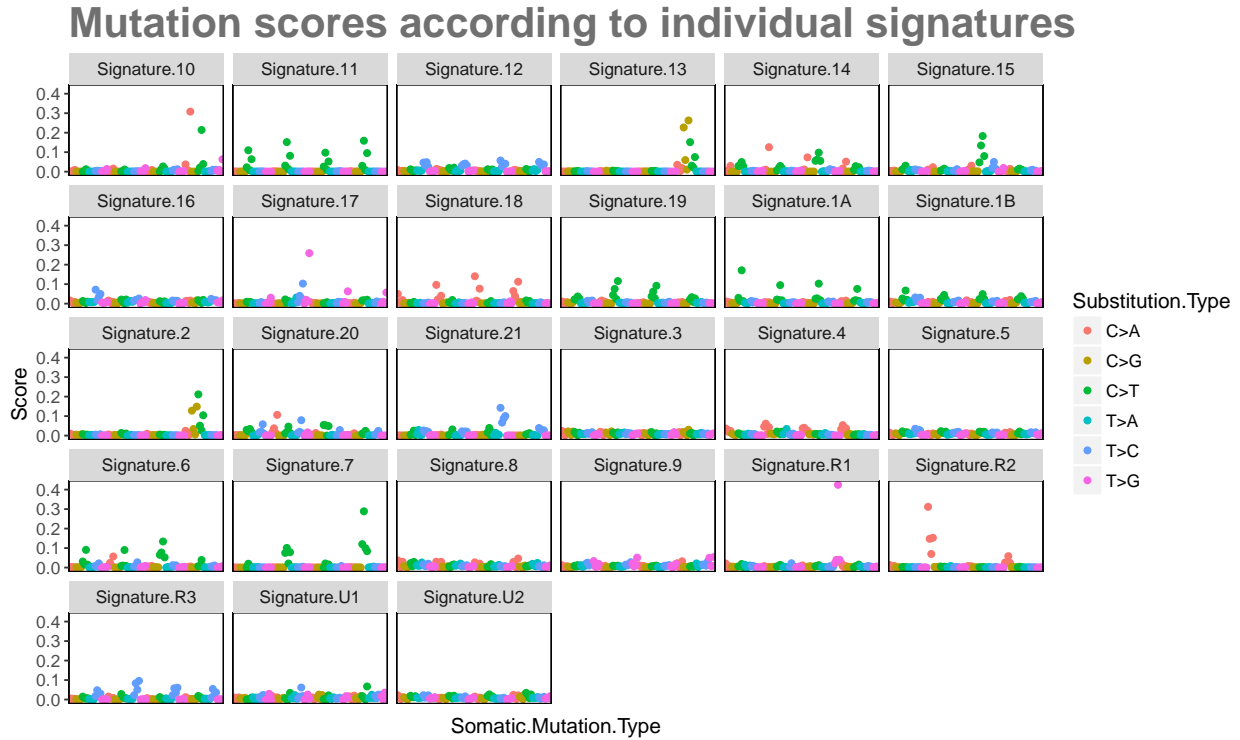


Figure 2: mutation_scores_for_all_snv_type_facet_signature_colored_snv

according to their Signature.3 Score.

```
ref_mut_sig_ordered_by_sig3 %>% head(5) %>% kable()
```

```
Substitution.Type
Trinucleotide
Somatic.Mutation.Type
Signature.1A
Signature.1B
Signature.2
Signature.3
Signature.4
Signature.5
Signature.6
Signature.7
Signature.8
Signature.9
Signature.10
Signature.11
```



Figure 3: cancer_type_sig_compare_plots

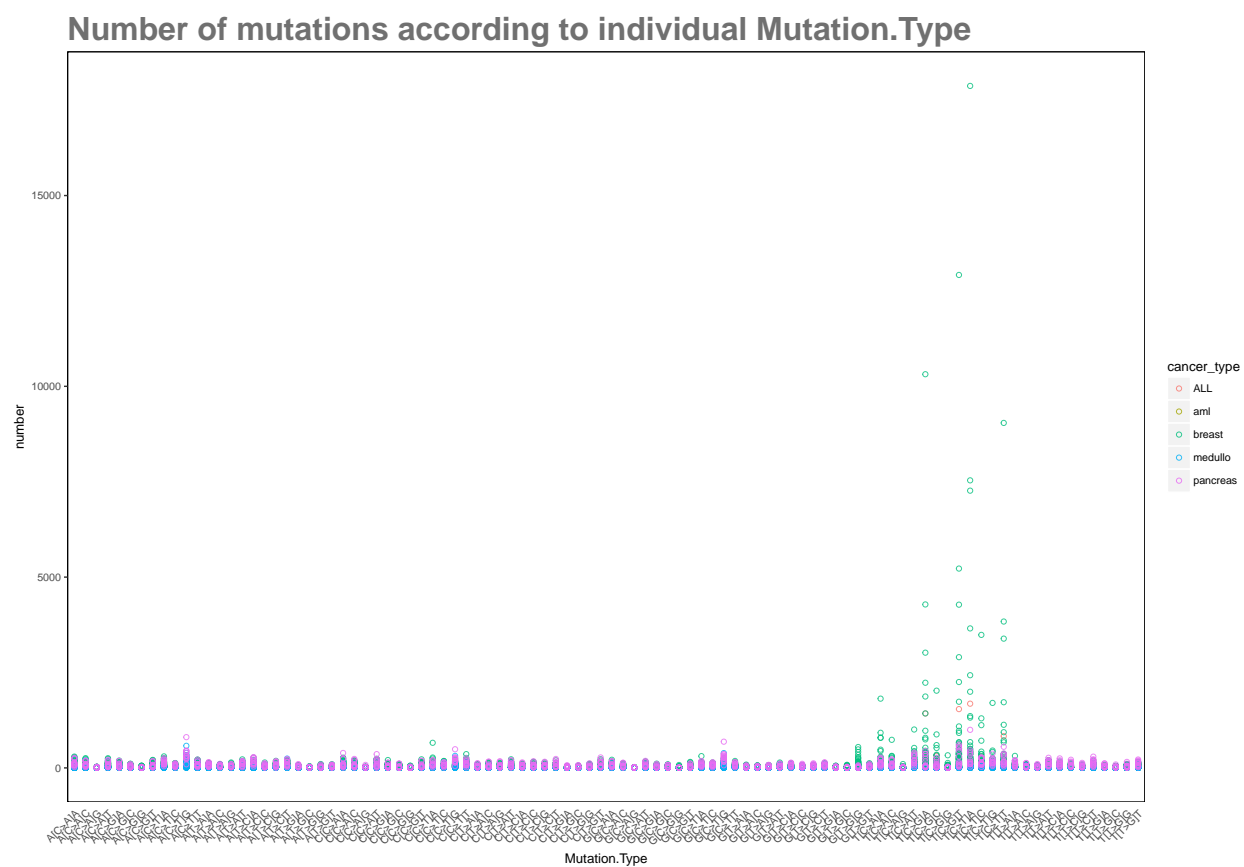


Figure 4: all_cancer_types_mut_geompoint

Signature.12
Signature.13
Signature.14
Signature.15
Signature.16
Signature.17
Signature.18
Signature.19
Signature.20
Signature.21
Signature.R1
Signature.R2
Signature.R3
Signature.U1
Signature.U2
C>A
ACA
A[C>A]A
0.0112
0.0104
0.0105
0.0240
0.0365
0.0149
0.0017
0.0004
0.0368
0.0120
0.0007
2e-04
0.0077
0.0007
0.0001
0.0013
0.0161
0.0018

0.0500
0.0107
0.0013
1e-04
0.0210
0.0137
0.0044
0.0105
0.0221
C>A
ACC
A[C>A]C
0.0092
0.0093
0.0061
0.0197
0.0309
0.0089
0.0028
0.0005
0.0287
0.0067
0.0010
1e-03
0.0047
0.0001
0.0042
0.0040
0.0097
0.0003
0.0076
0.0074
0.0024
7e-04
0.0065
0.0046

0.0047
0.0005
0.0123
C>A
ACG
A[C>A]G
0.0015
0.0016
0.0013
0.0019
0.0183
0.0022
0.0005
0.0000
0.0017
0.0005
0.0003
0e+00
0.0017
0.0001
0.0005
0.0000
0.0022
0.0000
0.0017
0.0005
0.0000
0e+00
0.0000
0.0048
0.0003
0.0000
0.0028
C>A
ACT
A[C>A]T

0.0063
0.0067
0.0037
0.0172
0.0243
0.0092
0.0019
0.0004
0.0300
0.0068
0.0092
2e-04
0.0046
0.0002
0.0296
0.0057
0.0088
0.0032
0.0181
0.0074
0.0029
6e-04
0.0058
0.0081
0.0034
0.0112
0.0118
C>A
CCA
C[C>A]A
0.0067
0.0090
0.0061
0.0194
0.0461
0.0097

0.0101
0.0012
0.0303
0.0098
0.0031
7e-04
0.0135
0.0035
0.0056
0.0106
0.0159
0.0010
0.0965
0.0112
0.0178
2e-03
0.0076
0.3117
0.0156
0.0173
0.0057

Note. We are still with the original format of the reference signatures here. I have decided to present only one example of ordering the data according to a factor, for more examples, you may refer to previous work from homework 5.

- (4) Writing the mutational signatures input/output to files is performed (embedded) in the scripts mentioned above. I have read and written a lot of files in each script. Please refer to the section “Automate the pipeline” below.

Perform statistical analyses

Tasks:

- Import the data created in the first script.
- Make sure your new continent order is still in force. You decide the details.
- Fit a linear regression of life expectancy on year within each country. Write the estimated intercepts, slopes, and residual error variance (or sd) to file. The R package broom may be useful here.
- Find the 3 or 4 “worst” and “best” countries for each continent. You decide the details.

As you know, I am using genomic data, so I have to adapt the tasks to my data, so here is what I did:

- (1) I am importing and cleaning the data with my script `read_clean_genome_text_files.R`:

- reference mutation signatures (mut_sig.txt)
- somatic mutations for 5 types of cancer: ALL, AML, breast, medulloblastoma, pancreas

I have performed the same “gathering” steps as described in the previous section for each cancer type.

```
ALL_mut_gather %>% arrange(Mutation.Type) %>% head(5) %>% kable()
```

Mutation.Type

case_id

number

A[C>A]A

PD4020a

33

A[C>A]C

PD4020a

15

A[C>A]G

PD4020a

1

A[C>A]T

PD4020a

24

A[C>G]A

PD4020a

31

```
aml_mut_gather %>% arrange(Mutation.Type) %>% head(5) %>% kable()
```

Mutation.Type

case_id

number

A[C>A]A

X400220

8

A[C>A]A

X426980

11

A[C>A]A

X452198

1

A[C>A]A

X573988

3

A[C>A]A

X758168

11

```
breast_mut_gather %>% arrange(Mutation.Type) %>% head(5) %>% kable()
```

Mutation.Type

case_id

number

A[C>A]A

PD3851a

29

A[C>A]A

PD3890a

99

A[C>A]A

PD3904a

114

A[C>A]A

PD3905a

88

A[C>A]A

PD3945a

235

```
medullo_mut_gather %>% arrange(Mutation.Type) %>% head(5) %>% kable()
```

Mutation.Type

case_id

number

A[C>A]A

LFS_MB1

45

A[C>A]A

LFS_MB2

23

A[C>A]A

LFS_MB4

24

A[C>A]A

MB1

6

A[C>A]A

MB101

82

```
pancreas_mut_gather %>% arrange(Mutation.Type) %>% head(5) %>% kable()
```

Mutation.Type

case_id

number

A[C>A]A

APGI_1839

77

A[C>A]A

APGI_1840

156

A[C>A]A

APGI_1956

59

A[C>A]A

APGI_1992

117

A[C>A]A

APGI_2000

122

- (2) The Somatic.Mutation.Type of the mut_sig.tsv (reference data) are still ordered according to Signature.3 values, as illustrated in Signature3_compare_plots.pdf
- (3) Here, we are not looking at signatures of the reference somatic mutation (mut_sig.tsv) but we are looking at the Mutation.Type of 5 types of cancer: ALL, AML, breast, medulloblastoma, pancreas.

Summary statistics

We will be looking at the mean, median, standard deviation and count of each Mutation.Type in each dataset (we need the “gathered” versions of files for that step).

```
all_cancer_types_stats %>% head(10) %>% kable()
```

Mutation.Type

mean_aml

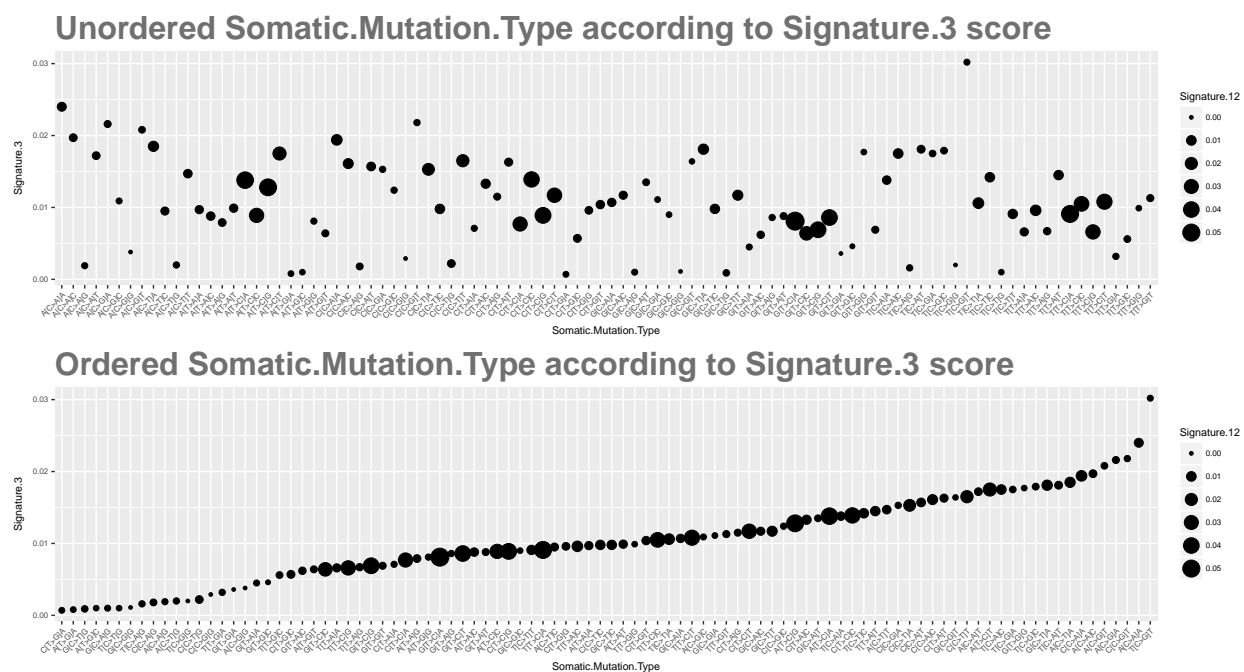


Figure 5: sig3

median_aml
sd_aml
total_aml
mean_breast
median_breast
sd_breast
total_breast
mean_medullo
median_medullo
sd_medullo
total_medullo
mean_pancreas
median_pancreas
sd_pancreas
total_pancreas
A[C>A]A
7.8571429
8
4.4131837

55
70.571429
48
65.041000
8398
24.73
16.5
28.889149
2473
121.866667
98
74.126211
1828
A[C>A]C
4.8571429
3
4.4880794
34
60.949580
39
51.024230
7253
19.70
12.5
24.634910
1970
90.600000
77
53.730545
1359
A[C>A]G
0.8571429
1
0.8997354
6
8.747899

7
6.891678
1041
2.80
2.0
3.305948
280
12.133333
10
9.203002
182
A[C>A]T
5.1428571
4
3.7161168
36
54.873950
33
50.542909
6530
17.72
11.0
23.407277
1772
81.466667
66
60.053389
1222
A[C>G]A
3.8571429
2
3.8047589
27
46.445378
22
45.246912

5527
13.18
10.0
13.096672
1318
64.866667
53
56.628951
973
A[C>G]C
1.8571429
2
1.6761634
13
28.512605
17
24.791287
3393
7.44
7.0
7.455349
744
37.000000
24
28.869163
555
A[C>G]G
0.7142857
0
1.1126973
5
11.159664
6
13.038394
1328
2.37

2.0
2.033333
237
9.466667
8
8.078779
142
A[C>G]T
3.5714286
3
3.1547394
25
47.663865
23
47.465648
5672
10.35
8.0
9.976108
1035
64.066667
32
59.666294
961
A[C>T]A
18.4285714
15
14.3742495
129
78.672269
57
55.095312
9362
31.86
22.5
29.770170

3186
141.933333
133
57.274361
2129
A[C>T]C
8.1428571
5
6.3358391
57
40.663865
33
24.169345
4839
16.07
11.5
14.968826
1607
61.600000
53
29.056103
924

Note. I wrote some stats tables for individual cancer types, then aggregated them (I peaked at how to make a loop here, but ended up performing the task on individual dataset because I couldn't figure out how to avoid "overwriting" at each loop iteration). The summary statistics tables are available [here](#)

Some linear regression modeling

I have used this website [here](#) discussing broom and variance, and this website [here](#) on some broom vignettes. And that's the moment I realized I didn't have two quantitative variables to perform linear regression modelling. This is a recurring problem with me: I always get into performing the analyses before making sure that the dataset format is appropriate (check this [hw04 readme file](#) here if you want an example).

So instead of fitting a linear regression of the lifeExp according to year, I will do something slightly different:

Merging and plotting linear regression

I will merge the cancer somatic data with the reference signature data, and I will fit a linear regression between Signature.3 and Signature.12 and then Signature.2 and Signature.13 (see [here](#) for more details on mutational signatures).

Note.1. It looks like the T>C somatic mutation group is the one differing the most between Signature.3 and Signature.12, but otherwise, these two signatures could fit "relatively well" a linear regression model

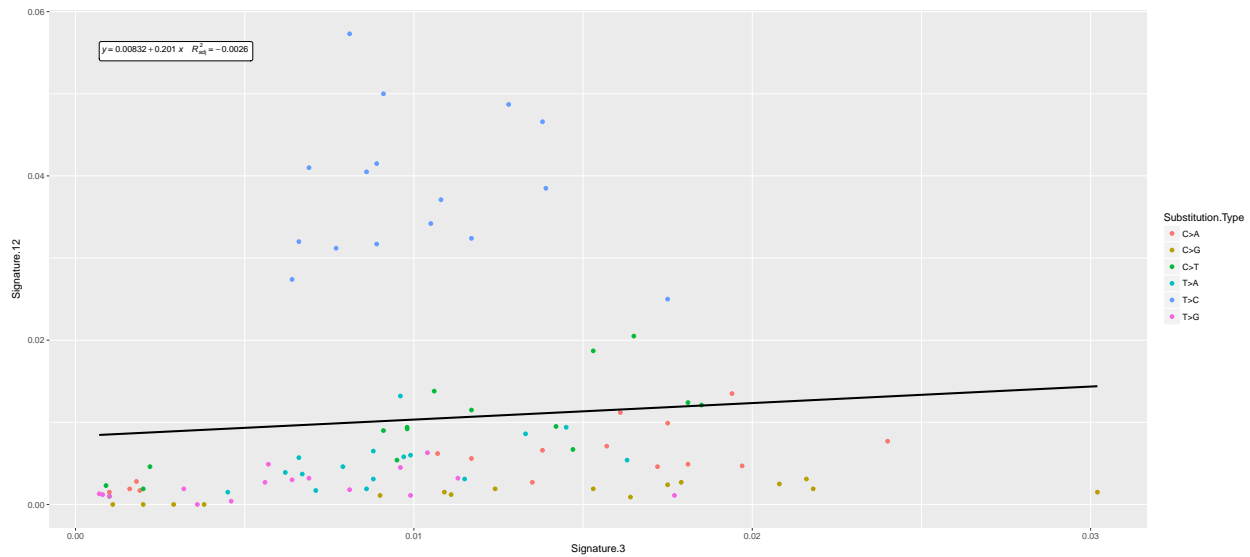


Figure 6: p3_linear_Sig3_Sig12

(although this is obviously not the best model for this type of data, because the adjusted R-squared value is below zero = the model does not fit the data very well).

Note.2. The code and format used above to add a label with the linear regression formula used the following references:

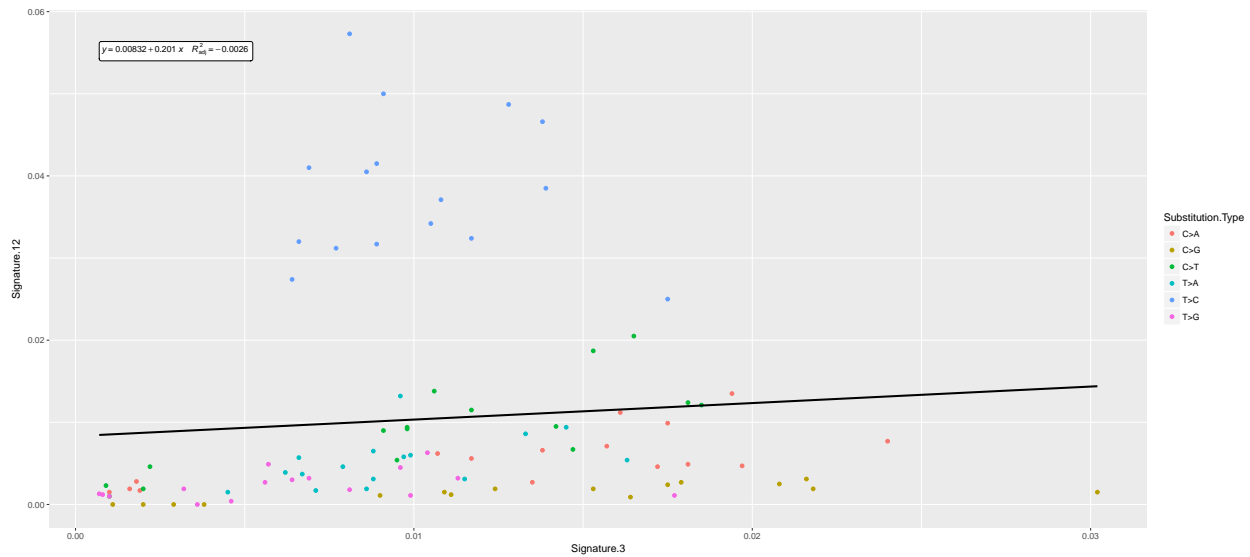
- A stack overflow discussion on Regression equation
- The stack overflow discussion on Label position discussion was also useful.
- This article of the cran rstudio website was also used.

```
lm(Signature.12 ~ Signature.3, mut_sig) %>% coef()
```

```
## (Intercept) Signature.3
## 0.008322249 0.201064066
```

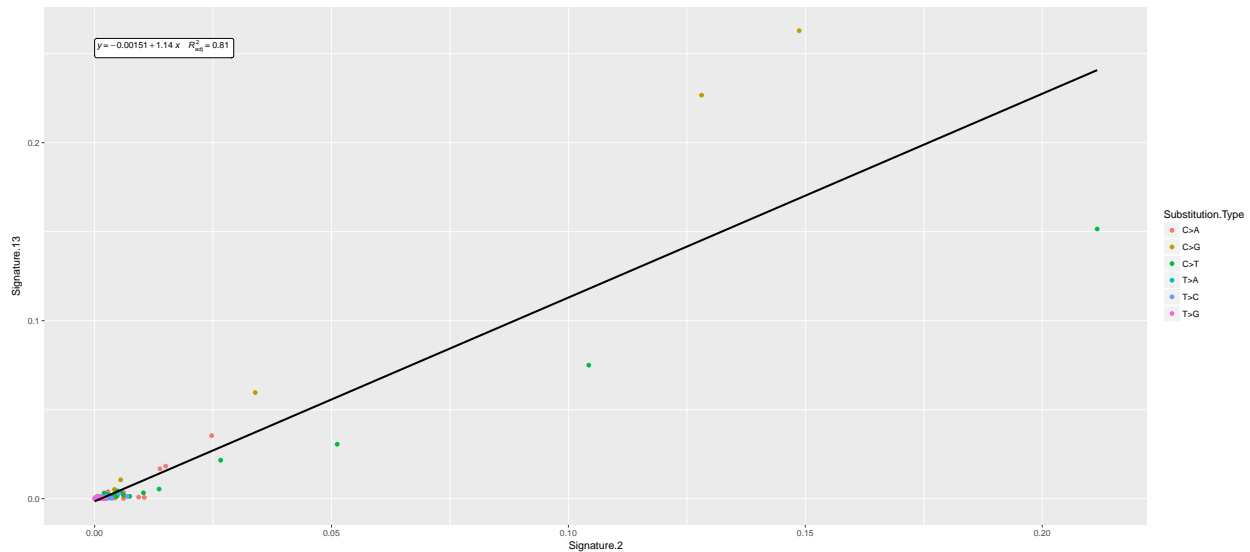
Note. We obtain the same Intercept and Signature.3 coefficient than in our plot (see label formula)

Just for fun, I want to check if a linear regression would be a better fit if I remove the Substitution.Type T>C.



Note. It does seem like the most different values between these two signatures are the T>C Substitution.Type, as our new adjusted R-squared tells us that 15% of the variance of Signature.12 can be explained by Signature.3 if we remove T>C.

Now, let's assess the linear regression model between Signature.2 and Signature.13.



Note. We do get a very high adjusted R-squared value, and this is because these two signatures are thought to be related to the same underlying mutational processes (see here for more details on mutational signatures). Therefore, we can deduce that the Mutation.Type profiles of Signature.2 and Signature.13 are quite similar.

```
lm(Signature.13 ~ Signature.2, mut_sig) %>% coef()
```

```
## (Intercept) Signature.2
## -0.001509612 1.144922739
```

I will also show a plot of the number of mutations for all cancer types according to Signature.3 and see what the linear regression looks like.

Find the 3 or 4 “worst” or “best” Signature.3 scores for each cancer_type.

Let's see the cases with the lowest and highest proportion of mutations caused by Signature.3 :

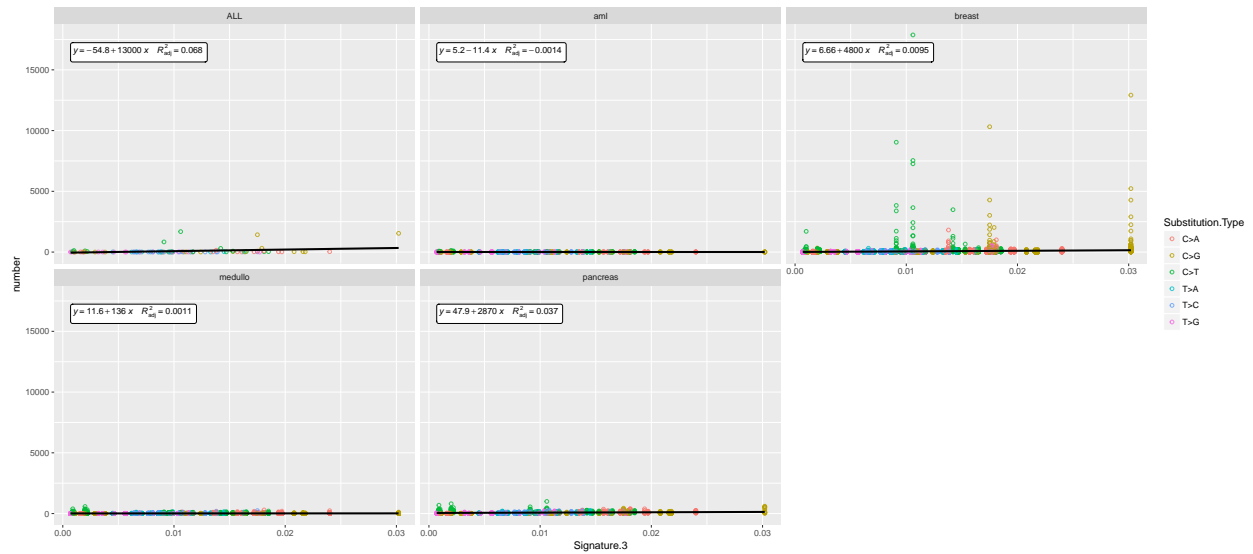


Figure 7: p2_linear_Sig3_all_cancer_mutations

```
all_cancer_types_mut_proportion_signatures %>%
  group_by(case_id, Signature) %>%
  filter(Signature=="Signature.3") %>%
  arrange(mutations_per_snv_sig_score) %>%
  head(3) %>% kable()
```

```
case_id
total_number_mutation_per_case
Mutation.Type
number
cancer_type
Substitution.Type
Trinucleotide
Signature
Score
mutations_per_snv_sig_score
proportion_number_each_snv_each_sig
MB28
44
C[T>G]A
0
medullo
T>G
```

CTA
 Signature.3
 7e-04
 0.0011407
 2.59e-05
 MB28
 44
 A[T>G]A
 0
 medullo
 T>G
 ATA
 Signature.3
 8e-04
 0.0013037
 2.96e-05
 MB28
 44
 G[C>T]G
 3
 medullo
 C>T
 GCG
 Signature.3
 9e-04
 0.0014667
 3.33e-05

```
all_cancer_types_mut_proportion_signatures %>%
  group_by(case_id) %>%
  filter(Signature=="Signature.3") %>%
  arrange(desc(mutations_per_snv_sig_score)) %>%
  head(3) %>% kable()
```

case_id
 total_number_mutation_per_case
 Mutation.Type
 number
 cancer_type

Substitution.Type
Trinucleotide
Signature
Score
mutations_per_snv_sig_score
proportion_number_each_snv_each_sig
PD4120a
67364
T[C>G]T
12919
breast
C>G
TCT
Signature.3
0.0302
75.34788
0.0011185
PD4120a
67364
A[C>A]A
114
breast
C>A
ACA
Signature.3
0.0240
59.87911
0.0008889
PD4120a
67364
C[C>G]T
225
breast
C>G
CCT
Signature.3

0.0218

54.39019

0.0008074

Note. This actually makes sense because breast cancer usually has higher Signature.3 due to homologous recombination repair deficiency, often secondary to *BRCA1/BRCA2* loss of function.

Generate figures

No worries here, I have generated plenty of figures.

Automate the pipeline (see Makefile)

- `mut_clean_genome_text_files.R` -> takes as input the file `mut_sig_raw.txt` and writes the cleaned up/formated version `mut_sig.txt`
 - `mut_sig_reorder.R` -> reorder the `mut_sig.txt` Somatic.Mutation.Type according to Signature.3
 - `mut_sig_tables.R` -> uses tidyverse function `gather` to go from a “wide” to a “long” dataset format and it is also used to perform aggregation tasks, so that the data can be ready for plots later one.
 - `mut_sig_stat.R` -> input the “gathered” dataset files of each cancer type, group by Mutation.Type and outputs files containing summary statistics (mean, median, sd, total)
 - `mut_sig_plot.R` -> input the “gathered” and/or “aggregated” dataset files and produces plots of two types: general data plots, linear modeling plots.
 - `summary_file.Rmd` -> this current file is a summary of this homework, and it has for input the output of the scripts previously mentioned.
 - `Makefile` -> this is the script that runs all the scripts above and coordinate the steps.
-

Additional material - optional

Here are some samples of the data files I have written:

```
head(all_cancer_types_mut) %>% kable()
```

Mutation.Type

case_id

number

cancer_type

A[C>A]A

PD4020a

33

ALL

A[C>A]C

PD4020a

15

ALL

A[C>A]G

PD4020a

1

ALL

A[C>A]T

PD4020a

24

ALL

A[C>G]A

PD4020a

31

ALL

A[C>G]C

PD4020a

8

ALL

```
head(all_cancer_types_mut_with_ref_sig) %>% kable()
```

Mutation.Type

case__id

number

cancer__type

Substitution.Type

Trinucleotide

Signature.1A

Signature.1B

Signature.2

Signature.3

Signature.4

Signature.5

Signature.6

Signature.7

Signature.8

Signature.9

Signature.10
Signature.11
Signature.12
Signature.13
Signature.14
Signature.15
Signature.16
Signature.17
Signature.18
Signature.19
Signature.20
Signature.21
Signature.R1
Signature.R2
Signature.R3
Signature.U1
Signature.U2
A[C>A]A
PD4020a
33
ALL
C>A
ACA
0.0112
0.0104
0.0105
0.0240
0.0365
0.0149
0.0017
4e-04
0.0368
0.0120
0.0007
2e-04
0.0077

0.0007
0.0001
0.0013
0.0161
0.0018
0.0500
0.0107
0.0013
1e-04
0.0210
0.0137
0.0044
0.0105
0.0221
A[C>A]C
PD4020a
15
ALL
C>A
ACC
0.0092
0.0093
0.0061
0.0197
0.0309
0.0089
0.0028
5e-04
0.0287
0.0067
0.0010
1e-03
0.0047
0.0001
0.0042
0.0040

0.0097
0.0003
0.0076
0.0074
0.0024
7e-04
0.0065
0.0046
0.0047
0.0005
0.0123
A[C>A]G
PD4020a
1
ALL
C>A
ACG
0.0015
0.0016
0.0013
0.0019
0.0183
0.0022
0.0005
0e+00
0.0017
0.0005
0.0003
0e+00
0.0017
0.0001
0.0005
0.0000
0.0022
0.0000
0.0017

0.0005
0.0000
0e+00
0.0000
0.0048
0.0003
0.0000
0.0028
A[C>A]T
PD4020a
24
ALL
C>A
ACT
0.0063
0.0067
0.0037
0.0172
0.0243
0.0092
0.0019
4e-04
0.0300
0.0068
0.0092
2e-04
0.0046
0.0002
0.0296
0.0057
0.0088
0.0032
0.0181
0.0074
0.0029
6e-04

0.0058
0.0081
0.0034
0.0112
0.0118
A[C>G]A
PD4020a
31
ALL
C>G
ACA
0.0018
0.0051
0.0048
0.0216
0.0097
0.0117
0.0013
0e+00
0.0085
0.0048
0.0005
7e-04
0.0031
0.0018
0.0001
0.0011
0.0048
0.0016
0.0014
0.0058
0.0005
5e-04
0.0038
0.0024
0.0012

0.0044
0.0108
A[C>G]C
PD4020a
8
ALL
C>G
ACC
0.0026
0.0043
0.0031
0.0109
0.0054
0.0073
0.0012
0e+00
0.0037
0.0023
0.0003
3e-04
0.0015
0.0014
0.0000
0.0001
0.0024
0.0016
0.0017
0.0019
0.0022
8e-04
0.0046
0.0018
0.0015
0.0065
0.0074

```
head(all_cancer_mutations_per_snv_sig_score) %>% kable()
```

case_id	total_number_mutation_per_case	Mutation.Type	number	cancer_type	Substitution.Type	Trinucleotide	Signature	Score	mutations_per_snv_sig_score
PD4020a	7741	A[C>A]A	33	ALL	C>A	ACA	Signature.1A	0.0112	3.2110815
PD4020a	7741	A[C>A]C	15	ALL	C>A	ACC	Signature.1A	0.0092	2.6376741
PD4020a	7741	A[C>A]G	1	ALL					

C>A
ACG
Signature.1A
0.0015
0.4300556
PD4020a
7741
A[C>A]T
24
ALL
C>A
ACT
Signature.1A
0.0063
1.8062333
PD4020a
7741
A[C>G]A
31
ALL
C>G
ACA
Signature.1A
0.0018
0.5160667
PD4020a
7741
A[C>G]C
8
ALL
C>G
ACC
Signature.1A
0.0026
0.7454296