

Protein Finder

This is a Python script for pulling data out of <https://www.ncbi.nlm.nih.gov/> and <https://www.uniprot.org/> using their APIs. It programmatically downloads all information found for given search terms in the databases. It saves biologists hours or even days of time.

Usage

API-KEY

Before proteinFinder.py can be used, a line in the code has to be changed. Since NCBI search requires an identification key or API-KEY, the first thing to do is to create an account on NCBI. Next it is best to follow this guide to acquire the key. The API key needs to be inserted into the following line of code in proteinFinder.py:

```
apiKey = "INSERT YOUR KEY HERE"
```

Once this is done, the program has its full functionality.

Basic Functionality

Let's assume we have a csv File containing gene loci. The loci were originally from the NCBI database. Now we want to have more information about the gene product. We start with the input file, named 'genes.csv'. It stores the following information:

Gene loci:

| <u>gene_loci</u> |
|------------------|
| Dshi_0051 |
| Dshi_0052 |
| Dshi_0053 |
| Dshi_0054 |
| Dshi_0055 |
| Dshi_0056 |
| Dshi_0057 |
| Dshi_0057 |
| Dshi_0058 |
| Dshi_0059 |
| Dshi_0060 |
| Dshi_0061 |
| <u>CC_2_9</u> |

We start the command line prompt and navigate to the folder containing proteinFinder.py. To use the Protein Finder module we need to know what kind of arguments the script is expecting. We can go look it up in documentation or we just type in:

```
python3 proteinFinder.py -h
```

The flag -h is short for help. This command shows all arguments the script is able to process and a short description:

```
usage: proteinFinder.py [-h] [--noheaders] [-db DATABASE] [-c COLUMNS]
                        [--organism ORGANISM] [-i {text,id}] [--idType IDTYPE]
                        inputFile outputFile {ncbi,uniprot,ncbi+uniprot}
```

positional arguments:

```
inputFile          Path to csvfile with query strings
outputFile         Path to file the results should be saved
{ncbi,uniprot,ncbi+uniprot}
                    Which website should be queried
```

optional arguments:

```
-h, --help          show this help message and exit
--noheaders         When reading the input file it does not skip the first row
-db DATABASE, --database DATABASE
                    Database which should be queried
-c COLUMNS, --columns COLUMNS
                    When querying Uniprot, depicts, which type of
                    information should be downloaded. Terms have to be
                    comma separated
--organism ORGANISM Narrow down the search to specified organism
-i {text,id}, --inputDataType {text,id}
                    Type of data stored in input file
--idType IDTYPE     What database the IDs are from?
```

For the script working properly, the positional arguments are required. The minimum working script, would be:

```
python3 proteinFinder.py inputFilePath outputFilePath websiteToQuery
```

In our case the data is stored in a csv file called 'genes.csv'. And we want the end result to be saved in 'proteinInformation.csv'. Since Uniprot does not recognise our gene loci we first need to query NCBI, for the ID. And with that ID we query the Uniprot database. Luckily the script is smart and handles this with the simple argument 'ncbi+uniprot'. So we end up with the following command:

```
python3 proteinFinder.py genes.csv proteinInformation.csv ncbi+uniprot
```

The script informs us that it ignores the first line in the input file, which contains the headers. If you don't want this behaviour, you need to pass the argument '-noheaders'. We also get a feedback on the status of the queries:

Skipping headers

Query 1 of 12

Query 2 of 12

Query 3 of 12

Query 4 of 12

Query 5 of 12

Query 6 of 12

Query 7 of 12

Query 8 of 12

Query 9 of 12

Query 10 of 12

Query 11 of 12

Query 12 of 12

The information found by the search are summed up in the table. The resulting table in 'proteinInformation.csv' should look similar to this:

| query | ID_NCBI | Entry | Gene names | ... |
|-----------|-----------|--------|-----------------|-----|
| Dshi_0052 | 157910368 | A8LJX5 | Dshi_0052 | ... |
| Dshi_0053 | 157910369 | A8LJX6 | Dshi_0053 | ... |
| Dshi_0054 | 157910370 | A8LJX7 | gst2 Dshi_0054 | ... |
| Dshi_0055 | 157910371 | A8LJX8 | holA Dshi_0055 | ... |
| Dshi_0056 | 157910372 | A8LJX9 | Dshi_0056 | ... |
| Dshi_0057 | 189082998 | A8LJY0 | leuS Dshi_0057 | ... |
| Dshi_0057 | 157910373 | A8LJY0 | leuS Dshi_0057 | ... |
| Dshi_0058 | 157910374 | A8LJY1 | lolA Dshi_0058 | ... |
| Dshi_0059 | 157910375 | A8LJY2 | ftsK Dshi_0059 | ... |
| Dshi_0060 | 157910376 | A8LJY3 | Dshi_0060 | ... |
| Dshi_0061 | 157910377 | A8LJY4 | Dshi_0061 | ... |
| Dshi_0062 | 157910378 | A8LJY5 | ubiH Dshi_0062 | ... |
| Dshi_0063 | 157910379 | A8LJY6 | suhB2 Dshi_0063 | ... |

The table stores the original query, the IDs found in NCBI and the Uniport entry. Overall the search was successful. However there are 2 interesting cases. First, when the script does not find some entries, it will fill the row with a 'Not Found' tag. Second, Dshi_0057 seems to appear twice. This is by design. When multiple IDs or entries are found for a query, each ID or entry, will be represented in a unique row. The reason is, there is no way for the programmer to safely determine what entry is relevant. Therefore every table, has to be inspected for such duplicates and be eliminated by hand.

Searching NCBI

To query NCBI you need to specify which database should be queried. A full list of all names can be found [here](#). Right now only text search is available for NCBI.

```
python3 proteinFinder.py inputFile outputFile ncbi -db protein
```

If no -db flag is typed in the default value will be the protein database.

Example

Starting from a table genes.csv with gene loci, the corresponding ncbi accession numbers should be acquire.

| gene_loci |
|-----------|
| Dshi_0051 |
| Dshi_0052 |
| Dshi_0053 |
| Dshi_0054 |
| Dshi_0055 |
| Dshi_0056 |
| Dshi_0057 |
| Dshi_0057 |
| Dshi_0058 |
| Dshi_0059 |
| Dshi_0060 |
| Dshi_0061 |
| CC_2_9 |

```
python3 proteinFinder.py genes.csv geneInfo.csv ncbi
```

This command outputs geneInfo.csv:

| query | ID_NCBI |
|-----------|-----------|
| Dshi_0051 | 157910367 |
| Dshi_0052 | 157910368 |
| Dshi_0053 | 157910369 |
| Dshi_0054 | 157910370 |
| Dshi_0055 | 157910371 |
| Dshi_0056 | 157910372 |
| Dshi_0057 | 189082998 |
| Dshi_0057 | 157910373 |
| Dshi_0058 | 157910374 |
| Dshi_0059 | 157910375 |

| query | ID_NCB |
|-----------|-----------|
| Dshi_0060 | 157910376 |
| Dshi_0061 | 157910377 |

Searching Uniprot

Uniprot can be searched in two ways.

Query with search terms

```
python3 proteinFinder.py inputFile outputFile uniprot
```

First it can be searched by search terms. This takes a long time since every term needs to make its own request to the database.

The results can be filtered for a specific organism with the `--organism` flag followed by the species name. For trustworthy results it is recommended to input the UniProt taxon id, found on the website.

```
python3 proteinFinder.py inputFile outputFile uniprot --organism "Dinoroseobacter shibae"
python3 proteinFinder.py inputFile outputFile uniprot --organism 398580
```

Query with IDs

```
python3 proteinFinder.py inputFile outputFile uniprot --inputDataType id
```

The second way is to search by ID. To search with IDs the parameter `--inputDataType` needs to be set to 'id'.

The IDs can be from various databases. Uniprot is able to map (translate) a given ID to its own accession number. For this the database has to be specified from which the data come from. The database can be set with the argument `--idType`. A list of all available databases can be found [here](https://www.uniprot.org/help/uniprotkb_column_names).

Columns

With UniProt queries the information, which should be retrieved, can be controlled with the `--columns` flag. The column names have to be separated by a comma. All possible columns are listed on the [UniProt website] (https://www.uniprot.org/help/uniprotkb_column_names).

```
python3 proteinFinder.py inputFile outputFile
    uniprot --columns "id,keywords,genes,organism"
```

If not specified the default values are: * id * protein names * genes * existence * organism * ec * feature(METAL BINDING) * keywords * comment(PATHWAY) * comment(SUBCELLULAR LOCATION) * comment(DOMAIN) * families * sequence

Examples

The starting point will be the following table, which is a list of uniprot accession numbers of proteins:

| uniprot_id |
|------------|
| A8LJX5 |
| A8LJX6 |
| A8LJX7 |
| A8LJX8 |
| A8LJX9 |
| A8LJY0 |
| A8LJY0 |
| A8LJY1 |
| A8LJY2 |
| A8LJY3 |
| A8LJY4 |
| A8LJY5 |
| A8LJY6 |

To extract the information from UniProt, the following command will be used:

```
python3 proteinFinder.py proteins.csv proteinInformation.csv uniprot -i id
```

The program requests information about the accession numbers, which after success results in the following table:

| Entry | Protein names | Gene names | ... |
|--------|---|------------|-----|
| A8LJX5 | Uncharacterized protein | Dshi_0052 | ... |
| A8LJX6 | HI0933 family protein | Dshi_0053 | ... |
| A8LJX7 | Glutathione S-transferase like protein (EC 2.5.1.18) | gst2 | ... |
| A8LJX8 | DNA polymerase III, delta subunit (EC 2.7.7.7) | holA | ... |
| A8LJX9 | Uncharacterized protein | Dshi_0055 | ... |
| A8LJY0 | Leucine-tRNA ligase (EC 6.1.1.4) (Leucyl-tRNA synthetase) (LeuRS) | Dshi_0056 | ... |
| A8LJY0 | Leucine-tRNA ligase (EC 6.1.1.4) (Leucyl-tRNA synthetase) (LeuRS) | leuS | ... |
| A8LJY0 | Leucine-tRNA ligase (EC 6.1.1.4) (Leucyl-tRNA synthetase) (LeuRS) | Dshi_0057 | ... |
| A8LJY0 | Leucine-tRNA ligase (EC 6.1.1.4) (Leucyl-tRNA synthetase) (LeuRS) | leuS | ... |
| A8LJY0 | Leucine-tRNA ligase (EC 6.1.1.4) (Leucyl-tRNA synthetase) (LeuRS) | Dshi_0057 | ... |

| Entry | Protein names | Gene names | ... |
|--------|---|------------|-----|
| A8LJY1 | Outer-membrane lipoprotein carrier protein | lolA | ... |
| | | Dshi_0058 | |
| A8LJY2 | DNA translocase | ftsK | ... |
| | | Dshi_0059 | |
| A8LJY3 | Aminotransferase class I and II | Dshi_0060 | ... |
| A8LJY4 | Amidase | Dshi_0061 | ... |
| A8LJY5 | 2-octaprenyl-6-methoxyphenol hydroxylase (EC 1.14.13.-) | ubiH | ... |
| | | Dshi_0062 | |
| A8LJY6 | Inositol-phosphate phosphatase (EC 3.1.3.25) | suhB2 | ... |
| | | Dshi_0063 | |

With alternating the columns using the `--columns` flag the information requested will be changed:

```
python3 proteinFinder.py proteins.csv proteinInformation.csv
    uniprot -i id --columns "id,keywords,genes"
```

| Entry | Keywords | Gene names |
|--------|---|------------|
| A8LJX5 | Complete proteome;Reference proteome | Dshi_0052 |
| A8LJX6 | Complete proteome;Reference proteome | Dshi_0053 |
| A8LJX7 | Complete proteome;Reference proteome;Transferase | gst2 |
| | | Dshi_0054 |
| A8LJX8 | Complete proteome;Nucleotidyltransferase;Reference proteome;Transferase | holA |
| | | Dshi_0055 |
| A8LJX9 | Complete proteome;Reference proteome | Dshi_0056 |
| A8LJY0 | ATP-binding;Aminoacyl-tRNA synthetase;Complete proteome | leuS |
| | | Dshi_0057 |
| A8LJY0 | ATP-binding;Aminoacyl-tRNA synthetase;Complete proteome | leuS |
| | | Dshi_0057 |
| A8LJY1 | Chaperone;Complete proteome;Lipoprotein;Protein transport | lolA |
| | | Dshi_0058 |
| A8LJY2 | ATP-binding;Cell cycle;Cell division;Cell membrane;Chromosome partition | ftsK |
| | | Dshi_0059 |
| A8LJY3 | Aminotransferase;Complete proteome;Reference proteome;Transferase | Dshi_0060 |
| A8LJY4 | Complete proteome;Reference proteome | Dshi_0061 |
| A8LJY5 | Complete proteome;Oxidoreductase;Reference proteome | ubiH |
| | | Dshi_0062 |
| A8LJY6 | Complete proteome;Hydrolase;Reference proteome | suhB2 |
| | | Dshi_0063 |

Searching NCBI + Uniprot

To search both databases consecutive there is a shortcut.

```
python3 proteinFinder.py inputFile outputFile ncbi+uniprot
```

The results of the NCBI search will be the input of the UniProt search and the final results will be saved in the outputFile. The same flags are available for this shortcut as for the single commands.

```
python3 proteinFinder.py genes.csv proteinInformation.csv ncbi+uniprot
--inputDataType id --idType --database db --organism species
--columns columns
```

More detailed information can be found in the documentation.