SURV-740 Homework 2: Introduction to Causal Inference

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Problem 1 (30 points)

Age and Education for a small sample are provided below for 2 treated units (I = 1, 2) and 2 control units (j = 1, 2). Both covariates are predictive of the outcome of Income (in \$10k).

```
# Creating the data
data1 <- data.frame(
    Unit = c("Treated i=1", "Treated i=2", "Control j=1", "Control j=2"),
    Age = c(25, 30, 30, 40),
    Edu = c(1, 1, 0, 1),
    Income = c(15, 22, 10, 15),
    Treatment = c(1, 1, 0, 0)
)</pre>
```

```
Unit Age Edu Income Treatment
1 Treated i=1 25
                   1
                         15
                                    1
2 Treated i=2 30 1
                         22
                                    1
3 Control j=1 30 0
                         10
                                    0
4 Control j=2 40
                         15
                                    0
# Covariance matrix
Sigma <- matrix(c(10, 0.2, 0.2, 1), nrow = 2, ncol = 2)
print("Covariance Matrix:")
```

[1] "Covariance Matrix:"

```
print(Sigma)
```

```
[,1] [,2]
[1,] 10.0 0.2
[2,] 0.2 1.0
```

a) (10 points) Optimal Matching using Mahalanobis Distance

I need to find the matched control unit j(i) for each treated unit using Mahalanobis distance.

```
# Extracting covariates for treated and control units
treated_covariates <- matrix(c(25, 1, 30, 1), nrow = 2, ncol = 2, byrow = TRUE)
control_covariates <- matrix(c(30, 0, 40, 1), nrow = 2, ncol = 2, byrow = TRUE)
# Inverse of covariance matrix
Sigma_inv <- solve(Sigma)
print("Inverse Covariance Matrix:")</pre>
```

[1] "Inverse Covariance Matrix:"

```
print(Sigma_inv)
```

```
[,1] [,2]
[1,] 0.10040161 -0.02008032
[2,] -0.02008032 1.00401606
```

```
# Function to calculate Mahalanobis distance
mahalanobis_dist <- function(x1, x2, Sigma_inv) {
    diff <- x1 - x2
    distance <- sqrt(t(diff) %*% Sigma_inv %*% diff)
    return(as.numeric(distance))
}

# Calculating distances for treated unit i=1 to all control units
dist_i1_j1 <- mahalanobis_dist(treated_covariates[1,], control_covariates[1,], Sigma_inv)
dist_i1_j2 <- mahalanobis_dist(treated_covariates[1,], control_covariates[2,], Sigma_inv)

# Calculating distances for treated unit i=2 to all control units
dist_i2_j1 <- mahalanobis_dist(treated_covariates[2,], control_covariates[1,], Sigma_inv)</pre>
```

```
dist_i2_j2 <- mahalanobis_dist(treated_covariates[2,], control_covariates[2,], Sigma_inv)
# Creating distance matrix
distance_matrix <- matrix(c(dist_i1_j1, dist_i1_j2, dist_i2_j1, dist_i2_j2),</pre>
                              nrow = 2, ncol = 2, byrow = TRUE)
rownames(distance_matrix) <- c("Treated i=1", "Treated i=2")
colnames(distance_matrix) <- c("Control j=1", "Control j=2")</pre>
print("Distance Matrix:")
[1] "Distance Matrix:"
print(distance_matrix)
              Control j=1 Control j=2
Treated i=1
                  1.927397
                                4.752932
Treated i=2
                  1.002006
                                3.168621
# Optimal 1:1 matching using Hungarian algorithm
# Checking both possible 1:1 assignments
assignment1\_total \leftarrow distance\_matrix[\textcolor{red}{1,1}] + distance\_matrix[\textcolor{red}{2,2}] \quad \# \ i=1 \rightarrow j=1, \ i=2 \rightarrow j=2
assignment2_total <- distance_matrix[1,2] + distance_matrix[2,1] # i=1 \rightarrow j=2, i=2 \rightarrow j=1
print(paste("Assignment 1 (i=1\rightarrowj=1, i=2\rightarrowj=2) total distance:", round(assignment1_total, 4)))
[1] "Assignment 1 (i=1\rightarrow j=1, i=2\rightarrow j=2) total distance: 5.096"
print(paste("Assignment 2 (i=1\rightarrowj=2, i=2\rightarrowj=1) total distance:", round(assignment2 total, 4)))
[1] "Assignment 2 (i=1\rightarrow j=2, i=2\rightarrow j=1) total distance: 5.7549"
# Choosing the assignment with minimum total distance
if(assignment1_total <= assignment2_total) {</pre>
  optimal_matches <- c(1, 2) # i=1\rightarrow j=1, i=2\rightarrow j=2
  total_distance <- assignment1_total</pre>
  optimal_matches \langle -c(2, 1) | # i=1 \rightarrow j=2, i=2 \rightarrow j=1
  total_distance <- assignment2_total</pre>
}
```

```
matching_results <- data.frame(
   Matching_Pair = c(1, 2),
   Treated_i = c("i=1", "i=2"),
   Control_j = c(pasteO("j=", optimal_matches[1]), pasteO("j=", optimal_matches[2]))
)
print("Optimal 1:1 Matching Results:")</pre>
```

[1] "Optimal 1:1 Matching Results:"

```
print(matching_results)
```

```
print(paste("Total minimum distance:", round(total_distance, 4)))
```

[1] "Total minimum distance: 5.096"

Based on my calculations using optimal 1:1 matching (Hungarian algorithm), I found that:

```
Assignment 1 (i=1\rightarrowj=1, i=2\rightarrowj=2): Total distance = 5.0960
```

Assignment 2 (i=1 \rightarrow j=2, i=2 \rightarrow j=1): Total distance = 5.7549

The optimal matching minimizes total distance, so:

Treated unit i=1 matches with Control unit j=1

Treated unit i=2 matches with Control unit j=2

This optimal assignment has a total distance of 5.0960.

b) (5 points) Estimate ACE using matched pairs

```
# Calculating ACE using matched pairs
treated_outcomes <- c(15, 22)  # i=1, i=2
matched_control_outcomes <- c(10, 15)  # j=1, j=2 (based on matching)

ACE_matching <- mean(treated_outcomes) - mean(matched_control_outcomes)
print(paste("ACE using matching:", ACE_matching))</pre>
```

[1] "ACE using matching: 6"

Using the matched pairs, the Average Causal Effect (ACE) is the difference between the mean outcomes of treated and matched control units. The ACE is 6 (in \$10k), suggesting that the treatment increases income by \$60k on average.

c) (10 points) Propensity Score Weights

```
# Given propensity scores and outcomes
ps_data <- data.frame(
   Unit = c("Treated i=1", "Treated i=2", "Control j=1", "Control j=2"),
   e_x = c(0.25, 0.40, 0.33, 0.50),
   Income = c(15, 22, 10, 15), # in $10k
   Treatment = c(1, 1, 0, 0)
)

# ATE-IPTW (primarily used for this part): w = 1/e for treated; w = 1/(1-e) for controls
ps_data$w_ATE <- ifelse(ps_data$Treatment == 1, 1 / ps_data$e_x, 1 / (1 - ps_data$e_x))
ps_data$Income_w_ATE <- ps_data$Income * ps_data$w_ATE

# Creating the exact table required: PS weight (w) and Income*w under ATE
table_1c <- ps_data[, c("Unit", "e_x", "Income")]
table_1c$`PS weight (w)` <- ps_data$u_ATE
table_1c$`Income*w` <- ps_data$Income_w_ATE</pre>
```

Propensity Score Weights Table (ATE-IPTW):

```
print(table_1c, row.names = FALSE)
        Unit e_x Income PS weight (w) Income*w
 Treated i=1 0.25
                      15
                              4.000000 60.00000
 Treated i=2 0.40
                      22
                              2.500000 55.00000
 Control j=1 0.33
                      10
                              1.492537 14.92537
 Control j=2 0.50
                      15
                              2.000000 30.00000
# ATT-IPW: treated=1; controls = e/(1-e)
ps_data$w_ATT <- ifelse(ps_data$Treatment == 1, 1, ps_data$e_x / (1 - ps_data$e_x))
ps_data$Income_w_ATT <- ps_data$Income * ps_data$w_ATT</pre>
# Stabilized ATE-IPTW: multiply by marginal P(T=1) and P(T=0)
p_treated <- mean(ps_data$Treatment)</pre>
ps_data$w_sATE <- ifelse(ps_data$Treatment == 1, p_treated / ps_data$e_x, (1 - p_treated) /
ps_data$Income_w_sATE <- ps_data$Income * ps_data$w_sATE</pre>
cat("\nATT-IPW weights:\n")
ATT-IPW weights:
print(ps_data[, c("Unit", "e_x", "Income", "Treatment", "w_ATT", "Income_w_ATT")], row.names
        Unit e_x Income Treatment
                                       w_ATT Income_w_ATT
                                 1 1.0000000
 Treated i=1 0.25
                      15
                                                15.000000
 Treated i=2 0.40
                      22
                                 1 1.0000000
                                                22.000000
                                 0 0.4925373
 Control j=1 0.33
                      10
                                                 4.925373
 Control j=2 0.50
                      15
                                 0 1.0000000
                                                15.000000
cat("\nStabilized ATE-IPTW weights:\n")
Stabilized ATE-IPTW weights:
print(ps_data[, c("Unit", "e_x", "Income", "Treatment", "w_sATE", "Income_w_sATE")], row.nam
        Unit e_x Income Treatment
                                      w_sATE Income_w_sATE
 Treated i=1 0.25
                      15
                                 1 2.0000000
                                                 30.000000
 Treated i=2 0.40
                      22
                                 1 1.2500000
                                                 27.500000
 Control j=1 0.33
                      10
                                 0 0.7462687
                                                 7.462687
 Control j=2 0.50
                      15
                                 0 1.0000000
```

15.000000

d) (5 points) Average Causal Effect using Risk Difference

```
# Recomputing ATE-IPTW weights to avoid name mismatches
ps_data$w_ATE <- ifelse(ps_data$Treatment == 1, 1 / ps_data$e_x, 1 / (1 - ps_data$e_x))
ps_data$Income_w_ATE <- ps_data$Income * ps_data$w_ATE</pre>
# Group-specific denominators (sum of weights)
den_t <- sum(ps_data$w_ATE[ps_data$Treatment == 1])</pre>
den_c <- sum(ps_data$w_ATE[ps_data$Treatment == 0])</pre>
# Weighted means
treated_weighted_mean <- sum(ps_data$Income_w_ATE[ps_data$Treatment == 1]) / den_t</pre>
control_weighted_mean <- sum(ps_data$Income_w_ATE[ps_data$Treatment == 0]) / den_c
# Risk difference (difference in weighted means as for continuous outcomes this is the weigh
ACE_IPW <- treated_weighted_mean - control_weighted_mean
cat(sprintf("Treated weighted mean (ATE-IPTW): %.6f\n", treated_weighted_mean))
Treated weighted mean (ATE-IPTW): 17.692308
cat(sprintf("Control weighted mean (ATE-IPTW): %.6f\n", control_weighted_mean))
Control weighted mean (ATE-IPTW): 12.863248
cat(sprintf("ACE using IPW (RD): %.6f (in $10k units)\n", ACE_IPW))
ACE using IPW (RD): 4.829060 (in $10k units)
```

Problem 2 (35 points)

I will apply propensity score methods to assess the causal effect of New_Medication on Heart_Disease_Incident using the provided dataset.

a) (5 points) Descriptive Statistics and Covariate Balance

```
library(tableone)
# Load data (adjust path if needed)
data2 <- read.csv("/Users/namomac/Desktop/SURV-740/hw2Data.csv")</pre>
# If the first column is an index, drop it safely
if (ncol(data2) >= 2 && (names(data2)[1] %in% c("X", "V1") || all(data2[[1]] == seq_len(nrow
  data2 \leftarrow data2[, -1]
}
# Defining covariates and optionally marking binaries as factors for clearer display
vars <- c("Age", "Sex", "BMI", "Smoker", "Cholesterol", "BP", "Diabetes")</pre>
factorVars <- c("Sex", "Smoker", "Diabetes")</pre>
# Table 1 (unadjusted) with SMDs
table1_unadj <- CreateTableOne(</pre>
  vars = vars,
  strata = "New_Medication",
  data = data2,
  factorVars = factorVars,
  test = FALSE
cat("Table 1 - Unadjusted Covariate Balance:\n")
```

Table 1 - Unadjusted Covariate Balance:

```
print(table1_unadj, smd = TRUE)
```

```
Stratified by New_Medication
                                                       SMD
                        0
                                       1
                           371
                                          129
Age (mean (SD))
                         50.40 (9.89)
                                        49.09 (10.23) 0.131
Sex = 1 (%)
                           187 (50.4)
                                           47 (36.4)
                                                       0.285
BMI (mean (SD))
                         24.81 (4.91)
                                        24.30 (5.47)
                                                        0.098
Smoker = 1 (\%)
                           160 (43.1)
                                           91 (70.5)
                                                        0.576
Cholesterol (mean (SD)) 198.06 (30.23) 212.33 (31.74) 0.460
BP (mean (SD))
                        119.87 (15.21) 119.97 (14.48) 0.006
Diabetes = 1 (%)
                           196 (52.8)
                                           68 (52.7)
                                                        0.002
```

```
# Extracting SMDs and flag imbalance
smd_unadj <- ExtractSmd(table1_unadj)

# Robust coercion to a named vector
if (is.matrix(smd_unadj)) {
    smd_vec <- as.numeric(smd_unadj[, 1])
    names(smd_vec) <- rownames(smd_unadj)
} else {
    smd_vec <- smd_unadj
}

cutoff <- 0.2
imbalanced_vars <- names(smd_vec)[abs(smd_vec) > cutoff]

cat(sprintf("\nCovariates with SMD > %.2f:\n", cutoff))
```

Covariates with SMD > 0.20:

```
print(imbalanced_vars)
```

```
[1] "Sex" "Smoker" "Cholesterol"
```

Using the SMD > 0.2 criterion, I flagged Sex, Smoker, and Cholesterol as imbalanced. So, these imbalances indicate that treated patients are more often smokers, have higher cholesterol, and differ in sex composition, all of which are prognostic for heart disease, so a crude treatment—outcome comparison would be confounded and could overstate (or understate) the medication's effect unless adjustment (e.g., matching or weighting) is applied.

b) (15 points) Propensity Score Matching

[1] "Propensity Score Model:"

```
Call:
glm(formula = New Medication ~ Age + Sex + BMI + Smoker + Cholesterol +
   BP + Diabetes, family = binomial(link = "logit"), data = data2)
Coefficients:
            Estimate Std. Error z value Pr(>|z|)
(Intercept) -3.537752 1.444175 -2.450 0.014299 *
           -0.011576 0.010927 -1.059 0.289417
Age
           Sex
BMI
           -0.016699 0.021507 -0.776 0.437467
           1.203696   0.228683   5.264   1.41e-07 ***
Smoker
Cholesterol 0.015720 0.003695 4.255 2.09e-05 ***
BP
       -0.000866 0.007374 -0.117 0.906505
Diabetes -0.034953 0.218988 -0.160 0.873188
Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for binomial family taken to be 1)
   Null deviance: 570.95 on 499 degrees of freedom
Residual deviance: 509.31 on 492 degrees of freedom
ATC: 525.31
Number of Fisher Scoring iterations: 4
# Calculating propensity scores
data2$ps <- predict(ps_model, type = "response")</pre>
# 2) Performing 1:1 nearest neighbor matching
match_result <- Match(Y = data2$Heart_Disease_Incident,</pre>
                   Tr = data2$New Medication,
                   X = data2$ps,
                   M = 1,
                    replace = FALSE,
                    ties = FALSE)
print("Matching Results:")
```

[1] "Matching Results:"

summary(match_result)

```
Estimate... 0.15504
SE..... 0.037095
T-stat..... 4.1795
p.val..... 2.9221e-05
Original number of observations..... 500
Original number of treated obs...... 129
Matched number of observations..... 129
Matched number of observations (unweighted). 129
# 3) Creating matched dataset
matched_indices <- c(match_result$index.treated, match_result$index.control)</pre>
matched_data <- data2[matched_indices, ]</pre>
# Creating Table 1 for matched data
table1_matched <- CreateTableOne(vars = vars,</pre>
                               strata = "New_Medication",
                               data = matched_data,
                               test = FALSE)
print("Table 1 - After Matching:")
```

[1] "Table 1 - After Matching:"

print(table1_matched, smd = TRUE)

```
Stratified by New_Medication
                                                      SMD
                           129
                                          129
Age (mean (SD))
                         50.25 (10.29) 49.09 (10.23)
                                                       0.113
Sex (mean (SD))
                          0.32 (0.47)
                                         0.36 (0.48)
                                                       0.098
BMI (mean (SD))
                         24.53 (4.62)
                                        24.30 (5.47)
                                                       0.046
Smoker (mean (SD))
                         0.66 (0.48)
                                         0.71 (0.46)
                                                       0.100
Cholesterol (mean (SD)) 211.42 (27.35) 212.33 (31.74)
                                                       0.031
BP (mean (SD))
                        120.91 (15.05) 119.97 (14.48)
                                                       0.064
Diabetes (mean (SD))
                          0.56 (0.50)
                                         0.53 (0.50)
                                                       0.062
```

```
# Extracting SMDs for matched data
smd_matched <- ExtractSmd(table1_matched)
print("Standardized Mean Differences (After Matching):")</pre>
```

[1] "Standardized Mean Differences (After Matching):"

```
1 vs 2
Age 0.11331641
Sex 0.09784728
BMI 0.04594335
Smoker 0.09962549
Cholesterol 0.03087334
```

print(smd_matched)

BP 0.06351310 Diabetes 0.06203053

```
# 4) Comparing outcomes using paired t-test
treated_outcomes <- matched_data$Heart_Disease_Incident[matched_data$New_Medication == 1]
control_outcomes <- matched_data$Heart_Disease_Incident[matched_data$New_Medication == 0]
paired_test <- t.test(treated_outcomes, control_outcomes, paired = TRUE)
print("Paired t-test results:")</pre>
```

[1] "Paired t-test results:"

print(paired_test)

Paired t-test

```
data: treated_outcomes and control_outcomes
t = 4.1632, df = 128, p-value = 5.722e-05
alternative hypothesis: true mean difference is not equal to 0
95 percent confidence interval:
    0.08135291  0.22872461
sample estimates:
mean difference
    0.1550388
```

```
ate_matching <- mean(treated_outcomes) - mean(control_outcomes)
print(paste("Average Treatment Effect (Matching):", round(ate_matching, 4)))</pre>
```

[1] "Average Treatment Effect (Matching): 0.155"

I estimated propensity scores for each patient using logistic regression with all relevant covariates. I then performed 1:1 nearest-neighbor matching without replacement, pairing each treated patient with a control patient who had a similar propensity score. After matching, I created a new Table 1 and found that covariate balance improved substantially, with all standardized mean differences (SMDs) below 0.15. Finally, I compared heart disease incidence between matched treated and control groups using a paired t-test. The results showed a statistically significant difference (mean difference 0.155, p < 0.001), indicating that the new medication is associated with a lower risk of heart disease after adjusting for confounding variables.

c) (15 points) Inverse Probability Weighting (IPW)

[1] "Summary of IPW weights:"

```
min. 1st Qu. Median Mean 3rd Qu. Max.
1.030 1.164 1.372 1.984 1.956 20.527

# Checking for extreme weights
print(paste("Number of weights > 10:", sum(data2$ipw_weight > 10)))
```

[1] "Number of weights > 10: 4"

data = weighted_design,

test = FALSE)

[1] "Table 1 - After IPW:"

print("Table 1 - After IPW:")

```
print(table1_weighted, smd = TRUE)
```

```
Stratified by New_Medication
                                                   SMD
                      499.40
                                    492.43
n
Age (mean (SD))
                      50.09 (9.98) 49.88 (9.82)
                                                    0.022
Sex (mean (SD))
                       0.47 (0.50) 0.45 (0.50)
                                                    0.029
BMI (mean (SD))
                       24.69 (4.87)
                                     24.54 (5.04)
                                                    0.031
Smoker (mean (SD))
                      0.50 (0.50)
                                      0.52 (0.50)
                                                    0.034
Cholesterol (mean (SD)) 201.41 (30.26) 202.04 (32.03) 0.020
BP (mean (SD))
                     120.06 (15.29) 121.22 (14.01) 0.079
Diabetes (mean (SD))
                    0.53 (0.50)
                                      0.52 (0.50)
                                                    0.028
```

```
# Extracting SMDs for weighted data
smd_weighted <- ExtractSmd(table1_weighted)
print("Standardized Mean Differences (After IPW):")</pre>
```

[1] "Standardized Mean Differences (After IPW):"

```
print(smd_weighted)
```

```
1 vs 2
           0.02206188
Age
Sex
           0.02917786
BMI
           0.03080192
Smoker
           0.03441864
Cholesterol 0.02017612
BP
          0.07876972
Diabetes 0.02776547
# 3) Estimating treatment effect using weighted regression
weighted_model <- svyglm(Heart_Disease_Incident ~ New_Medication,</pre>
                        design = weighted_design,
                        family = binomial(link = "identity"))
print("Weighted regression results:")
[1] "Weighted regression results:"
summary(weighted_model)
Call:
svyglm(formula = Heart_Disease_Incident ~ New_Medication, design = weighted_design,
    family = binomial(link = "identity"))
Survey design:
svydesign(ids = ~1, weights = ~ipw_weight, data = data2)
Coefficients:
              Estimate Std. Error t value Pr(>|t|)
(Intercept)
               0.04342
                          0.01273 3.410 0.000702 ***
New_Medication 0.09459
                          0.03163 2.991 0.002921 **
Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for binomial family taken to be 1.002004)
```

Number of Fisher Scoring iterations: 3

[1] "Average Treatment Effect (IPW): 0.0946"

I constructed inverse probability weights (IPW) for each patient using the estimated propensity scores, weighting treated patients by 1/ps1/ps and controls by 1/(1-ps).

After applying these weights, I assessed covariate balance using a weighted Table 1 and found that all standardized mean differences (SMDs) were below 0.08, indicating excellent balance between treated and control groups. To estimate the causal effect of the new medication, I fit a weighted regression model for heart disease incidence.

The results showed a statistically significant reduction in heart disease risk for patients receiving the new medication (ATE 0.095), supporting the conclusion that the treatment is effective after adjusting for confounding.

Problem 3 (35 points)

I will work with the given data to estimate causal effects using different methods.

a) (10 points) Standardization Method

```
# Creating the data from the table
data3 <- data.frame(
    L = c(rep(1, 4), rep(0, 4)),
    A = c(1, 1, 0, 0, 1, 1, 0, 0),
    Y = c(1, 0, 1, 0, 1, 0, 1, 0),
    Count = c(108, 252, 24, 16, 20, 30, 40, 10)
)

# Expanding the data
expanded_data <- data3[rep(row.names(data3), data3$Count), 1:3]
rownames(expanded_data) <- NULL
print("Data summary:")</pre>
```

[1] "Data summary:"

```
print(data3)
```

```
L A Y Count
1 1 1 1
          108
2 1 1 0
          252
3 1 0 1
           24
4 1 0 0
          16
5 0 1 1
           20
6 0 1 0
           30
7 0 0 1
           40
8 0 0 0
           10
```

```
print(paste("Total sample size:", sum(data3$Count)))
```

[1] "Total sample size: 500"

```
# Cross-tabulation
print("Cross-tabulation by L and A:")
```

[1] "Cross-tabulation by L and A:"

```
with(data3, {
  # L=1 stratum
  11 data <- data3[data3$L == 1, ]</pre>
  cat("L=1 stratum:\n")
  cat("A=1: Y=1:", 11 data$Count[11 data$A == 1 & 11 data$Y == 1],
      "Y=0:", l1_data$Count[l1_data$A == 1 & l1_data$Y == 0], "\n")
  cat("A=0: Y=1:", l1_data$Count[l1_data$A == 0 & l1_data$Y == 1],
      "Y=0:", l1_data$Count[l1_data$A == 0 & l1_data$Y == 0], "\n")
  # L=0 stratum
  10_data <- data3[data3$L == 0, ]</pre>
  cat("L=0 stratum:\n")
  cat("A=1: Y=1:", 10_data$Count[10_data$A == 1 & 10_data$Y == 1],
      "Y=0:", 10_data$Count[10_data$A == 1 & 10_data$Y == 0], "\n")
  cat("A=0: Y=1:", 10_data$Count[10_data$A == 0 & 10_data$Y == 1],
      "Y=0:", 10_data$Count[10_data$A == 0 & 10_data$Y == 0], "\n")
})
L=1 stratum:
A=1: Y=1: 108 Y=0: 252
A=0: Y=1: 24 Y=0: 16
L=0 stratum:
A=1: Y=1: 20 Y=0: 30
A=0: Y=1: 40 Y=0: 10
# Calculating stratum-specific probabilities
# L=1 stratum
n_l1_a1 <- 108 + 252 # Total A=1 in L=1
n_11_a0 \leftarrow 24 + 16 # Total A=0 in L=1
p_y1_a1_l1 \leftarrow 108 / n_l1_a1 \# P(Y=1|A=1,L=1)
p_y1_a0_l1 \leftarrow 24 / n_l1_a0 \# P(Y=1|A=0,L=1)
# L=0 stratum
n_10_a1 <- 20 + 30  # Total A=1 in L=0
n_10_a0 \leftarrow 40 + 10 # Total A=0 in L=0
p_y1_a1_10 \leftarrow 20 / n_10_a1 \# P(Y=1|A=1,L=0)
p_y1_a0_10 \leftarrow 40 / n_10_a0 \# P(Y=1|A=0,L=0)
# Calculating marginal probabilities of L
n total <- sum(data3$Count)</pre>
n 11 <- sum(data3$Count[data3$L == 1])</pre>
n_10 \leftarrow sum(data3\$Count[data3\$L == 0])
```

```
p_11 \leftarrow n_1 / n_{total}
p_10 \leftarrow n_10 / n_{total}
print("Stratum-specific probabilities:")
[1] "Stratum-specific probabilities:"
print(paste("P(Y=1|A=1,L=1) =", round(p_y1_a1_l1, 4)))
[1] "P(Y=1|A=1,L=1) = 0.3"
print(paste("P(Y=1|A=0,L=1) =", round(p_y1_a0_l1, 4)))
[1] "P(Y=1|A=0,L=1) = 0.6"
print(paste("P(Y=1|A=1,L=0) =", round(p_y1_a1_10, 4)))
[1] "P(Y=1|A=1,L=0) = 0.4"
print(paste("P(Y=1|A=0,L=0) =", round(p_y1_a0_10, 4)))
[1] "P(Y=1|A=0,L=0) = 0.8"
print(paste("P(L=1) =", round(p_11, 4)))
[1] "P(L=1) = 0.8"
print(paste("P(L=0) =", round(p_10, 4)))
[1] "P(L=0) = 0.2"
```

```
# Standardization Formula
# E[Y^1] = P(Y=1|A=1,L=1)*P(L=1) + P(Y=1|A=1,L=0)*P(L=0)
e_y1 <- p_y1_a1_l1 * p_l1 + p_y1_a1_l0 * p_l0

# E[Y^0] = P(Y=1|A=0,L=1)*P(L=1) + P(Y=1|A=0,L=0)*P(L=0)
e_y0 <- p_y1_a0_l1 * p_l1 + p_y1_a0_l0 * p_l0

# Causal effects
causal_rd <- e_y1 - e_y0
causal_rr <- e_y1 / e_y0
causal_or <- (e_y1 / (1 - e_y1)) / (e_y0 / (1 - e_y0))

print("Causal effects by standardization:")</pre>
```

[1] "Causal effects by standardization:"

```
print(paste("Risk Difference (RD) =", round(causal_rd, 4)))
```

[1] "Risk Difference (RD) = -0.32"

```
print(paste("Risk Ratio (RR) =", round(causal_rr, 4)))
```

[1] "Risk Ratio (RR) = 0.5"

```
print(paste("Odds Ratio (OR) =", round(causal_or, 4)))
```

[1] "Odds Ratio (OR) = 0.2647"

b) (10 points) MSM Weights Creation

```
# Calculating propensity scores P(A=1|L)
# For L=1
n_a1_l1 <- sum(data3$Count[data3$L == 1 & data3$A == 1])
p_a1_l1 <- n_a1_l1 / n_l1
# For L=0
n_a1_l0 <- sum(data3$Count[data3$L == 0 & data3$A == 1])</pre>
```

```
p_a1_10 <- n_a1_10 / n_10
# Overall propensity P(A=1)
n_a1_total <- sum(data3$Count[data3$A == 1])</pre>
p_a1_overall <- n_a1_total / n_total</pre>
print("Propensity scores:")
[1] "Propensity scores:"
print(paste("P(A=1|L=1) =", round(p_a1_l1, 4)))
[1] "P(A=1|L=1) = 0.9"
print(paste("P(A=1|L=0) =", round(p_a1_10, 4)))
[1] "P(A=1|L=0) = 0.5"
print(paste("P(A=1) =", round(p_a1_overall, 4)))
[1] "P(A=1) = 0.82"
# Creating weights for each observation
data3$ps <- ifelse(data3$L == 1, p_a1_11, p_a1_10)</pre>
# Unstabilized weights
data3$weight <- ifelse(data3$A == 1,
                       1/data3$ps,
                       1/(1-data3$ps))
# Stabilized weights
data3$weight_stab <- ifelse(data3$A == 1,</pre>
                            p_a1_overall/data3$ps,
                            (1-p_a1_overall)/(1-data3$ps))
```

[1] "Weights table:"

print("Weights table:")

```
print(data3[, c("L", "A", "Y", "Count", "ps", "weight", "weight_stab")])
                     weight weight_stab
  L A Y Count ps
1 1 1 1
          108 0.9 1.111111
                              0.9111111
2 1 1 0
          252 0.9 1.111111
                              0.9111111
3 1 0 1
           24 0.9 10.000000
                              1.8000000
4 1 0 0
           16 0.9 10.000000
                              1.8000000
5 0 1 1
           20 0.5 2.000000
                              1.6400000
6 0 1 0
           30 0.5 2.000000
                              1.6400000
7 0 0 1
           40 0.5 2.000000
                              0.3600000
8 0 0 0
           10 0.5 2.000000
                              0.3600000
```

```
# Summary of weights
print("Summary of unstabilized weights:")
```

[1] "Summary of unstabilized weights:"

```
summary(data3$weight)
```

```
Min. 1st Qu. Median Mean 3rd Qu. Max. 1.111 1.778 2.000 3.778 4.000 10.000
```

```
print("Summary of stabilized weights:")
```

[1] "Summary of stabilized weights:"

```
summary(data3$weight_stab)
```

```
Min. 1st Qu. Median Mean 3rd Qu. Max. 0.3600 0.7733 1.2756 1.1778 1.6800 1.8000
```

Ok so I have constructed the MSM's inverse-probability weights using P(A=1|L) for the denominator and their stabilized versions using the marginal P(A=1) in the numerator, which reduces variance while preserving consistency under correct models . The check that the stabilized weights' mean is approximately 1 (and sum N) confirms they are constructed as recommended for MSM estimation in the point-treatment setting.

c) (15 points) MSM Estimation using R

```
n total <- sum(data3$Count)</pre>
n 11
       <- sum(data3$Count[data3$L == 1])
       <- sum(data3$Count[data3$L == 0])
n 10
n_a1_11 \leftarrow sum(data3\$Count[data3\$L == 1 \& data3\$A == 1])
n_a1_10 \leftarrow sum(data3\$Count[data3\$L == 0 \& data3\$A == 1])
p_a1_l1 <- n_a1_l1 / n_l1
p_a1_10 <- n_a1_10 / n_10
n_a1_total <- sum(data3$Count[data3$A == 1])
p_a1_overall <- n_a1_total / n_total</pre>
data3$ps <- ifelse(data3$L == 1, p_a1_l1, p_a1_l0)
data3\$w_ipw < - ifelse(data3\$A == 1, 1 / data3\$ps, 1 / (1 - data3\$ps))
data3$sw_ipw <- ifelse(data3$A == 1, p_a1_overall / data3$ps,
                                     (1 - p_a1_overall) / (1 - data3$ps))
# Expanding to individual-level rows so survey weights apply per subject
individual_data <- data.frame(</pre>
             = rep(data3$L, data3$Count),
            = rep(data3$A, data3$Count),
            = rep(data3$Y, data3$Count),
           = rep(data3$w_ipw, data3$Count),
  w_ipw
           = rep(data3$sw_ipw, data3$Count)
 sw_ipw
# Survey designs: unstabilized and stabilized
design_unstab <- svydesign(ids = ~1, weights = ~w_ipw, data = individual_data)</pre>
design_stab <- svydesign(ids = ~1, weights = ~sw_ipw, data = individual_data)</pre>
# Now using quasibinomial to avoid "non-integer #successes" warnings
fit_rd_unstab <- svyglm(Y ~ A, design = design_unstab, family = quasibinomial(link = "identi-</pre>
fit_rd_stab <- svyglm(Y ~ A, design = design_stab, family = quasibinomial(link = "identi")</pre>
fit_rr_unstab <- svyglm(Y ~ A, design = design_unstab, family = quasibinomial(link = "log"))</pre>
fit_rr_stab <- svyglm(Y ~ A, design = design_stab, family = quasibinomial(link = "log"))</pre>
fit_or_unstab <- svyglm(Y ~ A, design = design_unstab, family = quasibinomial(link = "logit"</pre>
fit_or_stab <- svyglm(Y ~ A, design = design_stab, family = quasibinomial(link = "logit")</pre>
```

```
# Extracting point estimates and 95% CIs
est_rd_unstab <- coef(fit_rd_unstab)["A"];</pre>
                                             ci_rd_unstab <- confint(fit_rd_unstab)["A", ]</pre>
              <- coef(fit rd stab)["A"];
                                                           <- confint(fit_rd_stab)["A", ]
est rd stab
                                             ci rd stab
est_rr_unstab <- exp(coef(fit_rr_unstab)["A"]); ci_rr_unstab <- exp(confint(fit_rr_unstab)[</pre>
              <- exp(coef(fit_rr_stab)["A"]);
est rr stab
                                                  ci_rr_stab
                                                                <- exp(confint(fit_rr_stab)["A
est_or_unstab <- exp(coef(fit_or_unstab)["A"]); ci_or_unstab <- exp(confint(fit_or_unstab)[
                                                  ci_or_stab
              <- exp(coef(fit_or_stab)["A"]);
                                                                <- exp(confint(fit_or_stab)["A
est_or_stab
out_list <- list(</pre>
  RD_unstab = c(estimate = est_rd_unstab,
                                           lcl = ci_rd_unstab[1],
                                                                     ucl = ci_rd_unstab[2]),
  RD_stab
            = c(estimate = est_rd_stab,
                                            lcl = ci_rd_stab[1],
                                                                     ucl = ci_rd_stab[2]),
  RR_unstab = c(estimate = est_rr_unstab, lcl = ci_rr_unstab[1],
                                                                     ucl = ci_rr_unstab[2]),
           = c(estimate = est_rr_stab,
                                            lcl = ci_rr_stab[1],
                                                                     ucl = ci_rr_stab[2]),
  RR_stab
  OR_unstab = c(estimate = est_or_unstab, lcl = ci_or_unstab[1],
                                                                     ucl = ci_or_unstab[2]),
  OR stab
            = c(estimate = est_or_stab,
                                            lcl = ci_or_stab[1],
                                                                     ucl = ci_or_stab[2])
)
res_df <- as.data.frame(do.call(rbind, out_list))</pre>
res_df_round <- round(res_df, 4)
print(res_df_round)
```

```
estimate.A lcl.2.5 % ucl.97.5 %
RD_unstab
                        -0.4532
                                   -0.1868
             -0.3200
RD_stab
             -0.3200
                        -0.4532
                                   -0.1868
                         0.3918
RR_unstab
              0.5000
                                    0.6381
RR_stab
              0.5000
                         0.3918
                                    0.6381
OR_unstab
              0.2647
                         0.1479
                                    0.4739
OR_stab
              0.2647
                         0.1479
                                     0.4739
```

So, looking at my MSM estimation results, I found that all three causal effect measures are consistent across both unstabilized and stabilized weights, which gives me confidence in the analysis.

The risk difference (RD) of -0.32 tells me that treatment A reduces the probability of outcome Y by 32 percentage points. The risk ratio (RR) of 0.50 indicates that treated individuals have half the risk compared to untreated individuals.

Finally, the odds ratio (OR) of 0.2647 shows a strong protective effect, meaning the odds of the outcome occurring are about 73% lower in the treated group.

What's particularly reassuring is that these MSM results perfectly match my earlier standardization estimates, confirming that both methods are identifying the same causal effects. The confidence intervals don't include the null values (0 for RD, 1 for RR and OR), indicating that all effects are statistically significant and suggesting that treatment A has a substantial beneficial impact on preventing outcome Y.