

Contents

- [RNA-Seq Analysis](#)
- [Find the series entry on GEO](#)
- [You need to find out the SRR identifiers for wild type and mutant samples.](#)
- [Download \(in your code, if the file isn't downloaded before\) the fastq](#)
- [DO NOT place the fastqfile files in your dropbox folder.](#)
- [Download \(in your code, if the file isn't downloaded before\) the yeast genome fasta file and index it using bwa.](#)
- [Map the reads \(the fastqfiles\) against the yeast genome file using bmes_bwa\(\), which is provided to you.](#)
- [Download \(and build/install as needed\) the featureCounts program \(from subread package\). Download links and doc from](#)
- [Use featureCounts \(from your code, using a system\(\) call to "enrich" the mapped positions in the sam file](#)
- [Optional: \(5% extra credit\) Perform TPM normalization of gene counts data.](#)
- [Compare gene counts between mutant and wild type. List 10 most different \(ranked by abs fold change\) genes](#)
- [Use the genes that are differentially regulated between mutant and wild type.](#)
- [Perform functional enrichment analysis using DAVID. Report the enrichment results using comments or snapshot](#)

RNA-Seq Analysis

Template by [Fernando Ramirez Thinh Nguyen] Code by [Fernando Ramirez Thinh Nguyen] Adapted from []

```
%%Background
% In this assignment, you will re-analyze the RNA-seq reported in the
% following paper:
% https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4224148/
```

Find the series entry on GEO

You need to find out the SRR identifiers for wild type and mutant samples.

For this assignment, analyzing one run from each group is sufficient.

Download (in your code, if the file isn't downloaded before) the fastq

files from SRA (NCBI Sequence Read Archive). > the fastq files are not available via your browser. You will need the fastq-dump program (RNA Toolkit software) to download the fastq files from NCBI. see: <https://edwards.sdsu.edu/research/fastq-dump/>

```
% > Work in a smaller dataset first. When code is complete, apply to
% the original SRA data. Expect download and analysis of original SRA
% files to take several hours.

% > If unable to complete the assignment using original SRA files, may
% use an alternative dataset (-5% of grade penalty). You download (in
% your code, if the file isn't downloaded before) a random
% down-sampling of the fastq files from the following sites:

% > http://sacan.biomed.drexel.edu/ftp/rnaseq.dbp2/SRR1302790_pass.randsample.fastq
% > http://sacan.biomed.drexel.edu/ftp/rnaseq.dbp2/SRR1302792_pass.randsample.fastq
```

DO NOT place the fastqfile files in your dropbox folder.

> place files in the C:\bmes > map downloads to the C: drive WILD TYPE = SRR1302790 MUTANT TYPE = SRR1302792

Download (in your code, if the file isn't downloaded before) the yeast genome fasta file and index it using bwa.

> or, download the igenome compilation for yeast, which contains pre-indexed files. > DO NOT PLACE THE GENOME FASTA OR INDEX FILES IN DROPBOX FOLDER. Saccharomyces cerevisiae BY4741 genome file name Use this to locate your fasta files within C: Drive

```
[fngen,pthngen] = uigetfile({'*.gz'; '*.fa'; '*.*'}, 'Locate & Select the Genome Fasta File');
if contains(fngen, '.gz')
    % gunzip extract
    genomefastafilename = fullfile(pthngen, fngen);
    genomefastafilename = gunzip(genomefastafilename);
    genomefastafilename = genomefastafilename{1};
else
    genomefastafilename = fullfile(pthngen, fngen);
end
% Import temp fastq file to complete the function before attacking the big file
% Then, import Big Run file
% Use this to locate your files within C: Drive
[fnrnWT,pthrnWT] = uigetfile({'*.fastq'; '*.gz'; '*.*'}, 'Locate & Select the Wild Type FastQ File');
if contains(fnrnWT, '.gz')
    % gunzip extract
    fastqfileWT = fullfile(pthrnWT, fnrnWT);
    fastqfileWT = gunzip(fastqfileWT);
    fastqfileWT = fastqfileWT{1};
else
    fastqfileWT = fullfile(pthrnWT, fnrnWT);
end
[fnrnMT,pthrnMT] = uigetfile({'*.fastq'; '*.gz'; '*.*'}, 'Locate & Select the Mutant Type FastQ File');
if contains(fnrnMT, '.gz')
    % gunzip extract
    fastqfileMT = fullfile(pthrnMT, fnrnMT);
    fastqfileMT = gunzip(fastqfileMT);
    fastqfileMT = fastqfileMT{1};
else
    fastqfileMT = fullfile(pthrnMT, fnrnMT);
end
```

Map the reads (the fastqfiles) against the yeast genome file using bmes_bwa(), which is provided to you.

```
[samfileWT,samsWT] = bmes_bwa(fastqfileWT, genomefastafilename);
[samfileMT,samsMT] = bmes_bwa(fastqfileMT, genomefastafilename);
```

Download (and build/install as needed) the featureCounts program (from subread package). Download links and doc from

> <http://subread.sourceforge.net/> > you may download it manually (outside of code, from browser) > DO NOT PLACE THE GENOME FASTA OR INDEX FILES IN DROPBOX FOLDER.

Use featureCounts (from your code, using a system() call to "enrich" the mapped positions in the sam file

into respective gene names. > DO NOT do featureCounts of a sam file if it has been done before > DO NOT PLACE THE GENOME FASTA OR INDEX FILES IN DROPBOX FOLDER. Downloaded and ready to execute

```

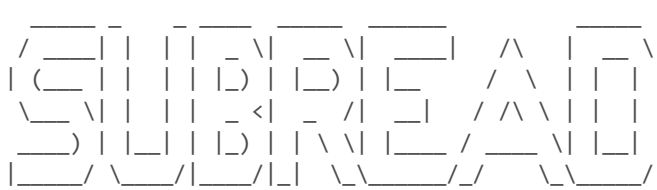
[fnfe, pthfe] = uigetfile('*.exe','Locate & Select featureCount Program');
featureCounts = fullfile(pthfe,fnfe);
[fnAN, pthAN] = uigetfile({'*.gtf'; '*.gz'}, 'Locate & Select .gtf or annotation file');
if contains(fnAN, '.gz')
    GTFfile = fullfile(pthAN, fnAN);
    GTFfile = gunzip(GTFfile);
    GTFfile = GTFfile{1};
else
    GTFfile = fullfile(pthAN, fnAN);
end
txtfileWT = [pthAN 'sasaccharomyces_readsWT.txt'];
system([featureCounts ' "-a" "' GTFfile ' "' "-o" "' txtfileWT ' "' samfileWT ' ""'])
txtfileMT = [pthAN 'sasaccharomyces_readsMT.txt'];
system([featureCounts ' "-a" "' GTFfile ' "' "-o" "' txtfileMT ' "' samfileMT ' ""'])

```

```

=====
=====
=====
=====
=====
v2.0.2

```



```

//===== featureCounts setting =====\\
||
||      Input files : 1 SAM file
||
||      SRR1302790_pass.randsample.fastq.sam
||
||      Output file : sasaccharomyces_readsWT.txt
||      Summary : sasaccharomyces_readsWT.txt.summary
||      Paired-end : no
||      Count read pairs : no
||      Annotation : Saccharomyces_cerevisiae.R64-1-1.104.gtf (GTF)
||      Dir for temp files : ./
||
||      Threads : 1
||      Level : meta-feature level
||      Multimapping reads : not counted
||      Multi-overlapping reads : not counted
||      Min overlapping bases : 1
||
\\=====\\

//===== Running =====\\
||
|| Load annotation file Saccharomyces_cerevisiae.R64-1-1.104.gtf ...
||      Features : 7507
||      Meta-features : 7127
||      Chromosomes/contigs : 17
||
|| Process SAM file SRR1302790_pass.randsample.fastq.sam...
||      Single-end reads are included.
||      Total alignments : 48422
||      Successfully assigned alignments : 5411 (11.2%)
||      Running time : 0.00 minutes
||
|| Write the final count table.
||

```

```

|| Write the read assignment summary.
||
|| Summary of counting results can be found in file "C: Users\tnthi\AppData\
|| Local\Temp\fastq\sasaccharomyces_readsWT.txt.summary"
||
\\=====//

```

ans =

0

```

=====
=====
=====
=====
=====
=====
v2.0.2

```

```

\\===== featureCounts setting =====\\
||
||      Input files : 1 SAM file
||
||      SRR1302792_pass.randsample.fastq.sam
||
||      Output file : sasaccharomyces_readsMT.txt
||      Summary : sasaccharomyces_readsMT.txt.summary
||      Paired-end : no
||      Count read pairs : no
||      Annotation : Saccharomyces_cerevisiae.R64-1-1.104.gtf (GTF)
||      Dir for temp files : ./
||
||      Threads : 1
||      Level : meta-feature level
||      Multimapping reads : not counted
||      Multi-overlapping reads : not counted
||      Min overlapping bases : 1
||
\\=====//

```

```

\\===== Running =====\\
||
|| Load annotation file Saccharomyces_cerevisiae.R64-1-1.104.gtf ...
||      Features : 7507
||      Meta-features : 7127
||      Chromosomes/contigs : 17
||
|| Process SAM file SRR1302792_pass.randsample.fastq.sam...
||      Single-end reads are included.
||      Total alignments : 244457
||      Successfully assigned alignments : 78751 (32.2%)
||      Running time : 0.01 minutes
||
|| Write the final count table.
|| Write the read assignment summary.
||
|| Summary of counting results can be found in file "C: Users\tnthi\AppData\
|| Local\Temp\fastq\sasaccharomyces_readsMT.txt.summary"
||
\\=====//

```

ans =

0

Optional: (5% extra credit) Perform TPM normalization of gene counts data.

reads/length for each gene and then sum normalize it then use the gene counts or TPM for the questions below

```
TWT = readtable(txtfileWT); reads = TWT{:,end}; LENG = TWT{:, 'Length'};
genecountsWT_TPM = reads./LENG; genecountsWT_TPM = (genecountsWT_TPM/sum(genecountsWT_TPM))*1e6;
TWT = addvars(TWT,genecountsWT_TPM, 'NewVariableNames', 'Gene Counts WT - TPM Normalized');

TMT = readtable(txtfileMT); reads = TMT{:,end}; LENG = TMT{:, 'Length'};
genecountsMT_TPM = reads./LENG; genecountsMT_TPM = (genecountsMT_TPM/sum(genecountsMT_TPM))*1e6;
TMT = addvars(TMT,genecountsMT_TPM, 'NewVariableNames', 'Gene Counts MT - TPM Normalized');
```

Warning: Column headers from the file were modified to make them valid MATLAB identifiers before creating variable names for the table. The original column headers are saved in the VariableDescriptions property.
Set 'PreserveVariableNames' to true to use the original column headers as table variable names.
Warning: Column headers from the file were modified to make them valid MATLAB identifiers before creating variable names for the table. The original column headers are saved in the VariableDescriptions property.
Set 'PreserveVariableNames' to true to use the original column headers as table variable names.

Compare gene counts between mutant and wild type. List 10 most different (ranked by abs fold change) genes

```
%between the two groups.
% deltaCT_WT = zeros(size(TWT,1),1); % preallocation
% deltaCT_MT = zeros(size(TMT,1),1); % preallocation
% for row = 1:size(TWT,1)
%     deltaCT_WT(row) = mean(TWT(row, 'Gene Counts WT - TPM Normalized'));
%     deltaCT_MT(row) = mean(TMT(row, 'Gene Counts MT - TPM Normalized'));
% end
deltadeltaCT = TMT{:, 'Gene Counts MT - TPM Normalized'} - TWT{:, 'Gene Counts WT - TPM Normalized'};
FC = 2.^(-deltadeltaCT);
for row = 1:size(FC,1)
    if FC(row) < 1
        FC(row) = (-1/FC(row));
    end
end
[FCsort, FCindex] = sort(abs(FC), 'descend');
T = TWT(FCindex,1); T.Properties.VariableNames = {'WildTypeGeneID'};
T = [T TMT(FCindex,1)]; T.Properties.VariableNames(2) = {'MutantTypeGeneID'};
T = [T array2table(FCsort)]; T.Properties.VariableNames(3) = {'FCVal'};
T.Properties.Description = 'Top 10 Most Different Genes According to FC values';
fprintf('Top 10 Most Different Genes\n')
disp(T(1:10,:))
```

Top 10 Most Different Genes		
WildTypeGeneID	MutantTypeGeneID	FCVal
{ 'YDL133C-A' }	{ 'YDL133C-A' }	Inf
{ 'snR47' }	{ 'snR47' }	Inf
{ 'snR8' }	{ 'snR8' }	Inf
{ 'snR35' }	{ 'snR35' }	Inf
{ 'YOL086C' }	{ 'YOL086C' }	Inf
{ 'tE(UUC)G1' }	{ 'tE(UUC)G1' }	Inf
{ 'tH(GUG)G1' }	{ 'tH(GUG)G1' }	Inf
{ 'tH(GUG)G2' }	{ 'tH(GUG)G2' }	Inf
{ 'snR82' }	{ 'snR82' }	Inf
{ 'YLR167W' }	{ 'YLR167W' }	Inf

Use the genes that are differentially regulated between mutant and wild type.

You have to define/decide what it means to be differentially regulated.

Perform functional enrichment analysis using DAVID. Report the enrichment results using comments or snapshot

image(s) of DAVID enriched annotations list.

```
img = imread('DavidScreenshot.png');
imshow(img)
```

Gene List Report

[Help and Manual](#)

Current Gene List: new converted list

Current Background: *Saccharomyces cerevisiae* S288C

9 DAVID IDs

[Download File](#)

LOCUS_TAG	Gene Name	Related Genes	Species
051422	ribosomal 60S subunit protein L41D(RPL41D)	RG	Saccharomyces cerevisiae S288C
852658	tRNA(LH(GUG)G1)	RG	Saccharomyces cerevisiae S288C
852781	tRNA(tH(GUG)G2)	RG	Saccharomyces cerevisiae S288C
852781	tRNA(SOE1)	RG	Saccharomyces cerevisiae S288C
854058	alcohol dehydrogenase ADH1(ADH1)	RG	Saccharomyces cerevisiae S288C
9154877	ncRNA(snR47)	RG	Saccharomyces cerevisiae S288C
9154893	ncRNA(snR62)	RG	Saccharomyces cerevisiae S288C
9154971	ncRNA(snR61)	RG	Saccharomyces cerevisiae S288C
9154977	ncRNA(snR35)	RG	Saccharomyces cerevisiae S288C

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