GLMM

Nick Syring

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Introduction

Introduction

Poisson Regression

3.1 Children Ever Born Data

The "Children Ever Born" (CEB) dataset consists of grouped data on the number of births of Fijian women. The women are described according to their marriage duration in years in ordinal levels: (0-4, 5-9, 10-14, 15-19, 20-24, 25-29); their place of residence (Suva—the capital city—Urban, or Rural); and, their level of education (none, lower primary, upper primary, secondary or greater). The count, mean, and variance of the number of children ever born, and the group size, is given for each group of women by cross-classified factorial level. These summaries are sufficient to model counts of children ever born by a Poisson distribution (each individual woman's count is not needed).

The CEB data is an example of an observational dataset — the characteristics of the individuals are inherent rather than set by experimenters as in an experimental/controlled trial—but, that should be clear from the context. Several interesting questions may be answered using this data, such as: are more or fewer born children associated with higher or lower education among Fijian women; does an urban versus rural living location influence the number of children ever born; and, do the number of children ever born steadily increase with marrige duration, or tend to plateau?

```
ceb <- read.table('ceb.dat')
head(ceb)</pre>
```

```
## dur res educ mean var n y
## 1 0-4 Suva none 0.50 1.14 8 4.00
## 2 0-4 Suva lower 1.14 0.73 21 23.94
## 3 0-4 Suva upper 0.90 0.67 42 37.80
## 4 0-4 Suva sec+ 0.73 0.48 51 37.23
```

5 0-4 urban none 1.17 1.06 12 14.04 ## 6 0-4 urban lower 0.85 1.59 27 22.95

A statistician (or student statistician) familiar with multiple linear regression and/or ANOVA for factorial experiments may instinctively choose to fit a Gauss-Markov linear model to the CEB data, treating the responses as independent normal random variables. However, since the responses are counts, a Poisson model is more reasonable. But, just how does one perform *Poisson regression?*— as opposed to the familiar multiple linear regression described by the Gauss-Markov model:

$$Y = X\beta + \epsilon, \quad \epsilon \sim N_n(0_{n \times 1}, \sigma^2 I_n).$$

That is the motivation for this chapter, in which we will explore the family of *Generalized Linear Models* from defining and fitting the model, to performing inference and model diagnostics, all within the context of the CEB example.

3.2 Defining GLMs

For the CEB data we naturally want to model the CEB grouped counts as realizations of Poisson r.v.'s with means $n_j x_j^{\mathsf{T}} \beta$ where n_j is the number of women in the j^{th} factorial group, x_j is the vector of their common covariates, and β is the common regression coefficient vector. Then, the likelihood of the model is

$$L(\beta; \text{data}) = \prod_{j=1}^{70} \frac{(n_j x_j^\top \beta)^{y_j} e^{-n_j x_j^\top \beta}}{y_j!}$$

and the loglikelihood is given by

$$\ell(\beta; \text{data}) = \text{constant} + \sum_{j=1}^{70} y_j \log(n_j x_j^\top \beta) - n_j x_j^\top \beta.$$

The Poisson likelihood is a member of the Exponential Family, which contains all distributions with PDFs that may be expressed as

$$f(y; \theta, \phi) = \exp\{[y\theta - b(\theta)]/a(\phi) + c(y, \phi)\}.$$

Looking ahead, we will apply the exponential family model above to independent but not identically distributed responses, similar to the data we encounter in multiple linear regression and the Gauss-Markov model, so we will allow θ as well as the forms of the a,b, and c functions to vary over observations, but we will fix ϕ , so that the loglikelihood for a sample of size n may be written as follows:

$$\ell(\beta; \text{data}) = \sum_{i=1}^{n} \{ [y_i \theta_i - b_i(\theta_i)] / a_i(\phi) + c_i(\phi, y_i) \}.$$

The Poisson regression model for grouped data is a fairly simple member of this family, having $\theta = \log(n_j x_j^{\top} \beta)$, $\phi = a(\phi) = 1$, and $b(\theta) = \exp(\theta) = n_j x_j^{\top} \beta$. In fact, it is very often the case that GLMs satisfy $a(\phi) \propto \phi$ up to a known constant. In general, GLMs satisfy

$$E(Y) = b'(\theta)$$
 and $V(Y) = b''(\theta)a(\phi)$.

For the Poisson regression model, in particular, we have

$$\theta = \log(n_j x_j^{\top} \beta); \quad b(\theta) = \exp(\theta); \quad \text{and} \quad a(\phi) = 1$$

$$E(Y) = b'(\theta) = \frac{\partial}{\partial \theta} \exp(\theta) = \exp(\theta) = n_j x_j^{\mathsf{T}} \beta; \text{ and,}$$

$$V(Y) = b''(\theta)a(\phi) = \frac{\partial^2}{\partial \theta^2} \exp(\theta) = \exp(\theta) = n_j x_j^\top \beta.$$

3.3 Fitting GLMs

Like any other model defined by a likelihood, GLMs may be fit by maximizing the (log)likelihood. But, it is generally not the case that the maximizers (MLEs) are available in closed form. Instead, they are computed iteratively using Newton's method or a similar iterative procedure. Refer again to the exponential family loglikelihood using the usual representation $a_i(\phi) = \phi/w_i$ where w_i are known constants:

$$\ell(\beta; \mathrm{data}) = \sum_{i=1}^n \{w_i[y_i\theta_i - b_i(\theta_i)]/\phi + c_i(\phi, y_i)\}.$$

Let $\mu_i = E(Y_i)$. Then, $b'(\theta_i) = \mu_i$, or, equivalently, $g_c(\mu_i) = \theta_i$ where g_c is termed the *canonical link*; for example, $g_c := \log$ for the Poisson distribution. Additionally, let g link the mean to the linear function of covariates, i.e., $g(\mu_i) = \eta_i = x_i^{\top} \beta$; e.g., g is the identity function for the Poisson model. Since $b'_i(\theta_i)$ is also equal to μ_i in the exponential family, we may differentiate the loglikelihood with respect to the regression parameter β using the chain rule:

$$\frac{\partial \ell}{\partial \beta_j} = \sum_{i=1}^n \left\{ \frac{w_i}{\phi} \left[y_i \frac{\partial \theta_i}{\partial \beta_j} - \frac{\partial b_i(\theta_i)}{\partial \beta_j} \right] + c_i(\phi, y_i) \right\}$$

using

$$\frac{\partial \theta_i}{\partial \beta_j} = \frac{\partial \theta_i}{\partial \mu_i} \frac{\partial \mu_i}{\partial \beta_j}.$$

Since $\mu_i = b_i'(\theta_i)$ we have $\partial \theta_i / \partial \mu_i = 1/b_i''(\theta_i)$. But, in light of $\mu_i = b'(\theta_i)$ we may always write $b_i''(\theta_i)$ as a function of μ_i , i.e., $V(\mu_i) = b_i''(\theta_i)/w$ so that $V(Y_i) = b_i''(\theta_i)/w$

 $V(\mu_i)\phi$. Moreover, since $\mu_i = g^{-1}(x_i^{\top}\beta)$ we have $\partial \mu_i/\partial \beta_j = x_{ij}/g'[g^{-1}(x_i^{\top}\beta)]$. Substituting, we can write the score function using only μ_i as follows:

$$\frac{\partial \ell}{\partial \beta_j} = \frac{1}{\phi} \sum_{i=1}^n \frac{y_i - \mu_i}{g'(\mu_i) V(\mu_i)} x_{ij}.$$

The second (mixed partial) derivative may be written

$$\frac{\partial^2 \ell}{\partial \beta_j \partial \beta_k} = -\frac{1}{\phi} \sum_{i=1}^n \frac{x_{ij} x_{ik} h(\mu_i)}{g'(\mu_i)^2 V(\mu_i)}$$

where $h(\mu_i) = 1 + (y_i - \mu_i)\{V'(\mu_i)/V(\mu_i) + g''(\mu_i)/g'(\mu_i)\}$. The expectation of the second derivative (which when multiplied by -1 appears in the Fisher information matrix) is the same quantity with $h(\mu_i)$ replaced by $E[h(\mu_i)]$, which simply equals 1 because $E(Y_i - \mu_i) = 0$. The Hessian of the loglikelihood is clearly a quadratic form $\phi^{-1}X^\top WX$ where X is the $n \times p$ design matrix of covariates and $W = [h(\mu_i)/\{g'(\mu_i)^2V(\mu_i)\}]$ is an $n \times n$ diagonal matrix of "weights". Less obvious, we may define $G = \mathrm{diag}\{g'(\mu_i)/h(\mu_i)\}$ so that the gradient of the loglikelihood equals $\phi^{-1}X^\top WG(y-\mu)$. With this clever rewriting, Newton's method updates take on the form of a weighted least squares solution:

$$\begin{split} \beta^{[k+1]} &= \beta^{[k]} + (X^\top W X)^{-1} X^\top W G(y-\mu) \\ &= (X^\top W X)^{-1} X^\top W \{G(y-\mu)X + \beta^{[k]}\} \\ &= (X^\top W X)^{-1} X^\top W z \end{split}$$

where $z := G(y - \mu) + X\beta^{[k]}$ is sometimes referred to as the "pseudo-data". Repeating the weighted least squares update, iteratively, until convergence, is termed *iteratively re-weighted least squares* (IRLS) since, of course, the weights in W are updating with each iteration.

For our Poisson regression based on the grouped CEB data we have the following likelihood, gradient, and Hessian:

$$\begin{split} \ell(\beta; \text{data}) &= \sum_{j=1}^{70} \left[y_j x_j^\top \beta - n_j e^{x_j^\top \beta} \right] \\ \nabla_s \ell &= \sum_{j=1}^{70} \left[y_j x_{js} - n_j x_{js} e^{x_j^\top \beta} \right] \\ \nabla_{s,t}^2 \ell &= -\sum_{i=1}^{70} n_j x_{js} x_{jt} e^{x_j^\top \beta}. \end{split}$$

Rewriting the Hessian and gradient as above for the general exponential family GLM we have

$$W_{k,k} = n_k \mu_k$$
 and $G_k = (n_k \mu_k)^{-1}$

so that the IRLS updates are given by

$$(X^\top W X)^{-1} X^\top W z$$
 with $z_k = (n_k \mu_k)^{-1} (y_k - n_k \mu_k) + x_k^\top \beta.$

3.3.1 IRLS for the CEB data

Below we compute the MLEs for the Poisson regression of the grouped CEB data "by hand" using IRLS—and, also compare to the glm function in R. For our calculation we initialize the elements of the parameter vector μ by the sample means $\mu_j = y_j/n_j$. We set the pseudo data equal to $z_j = -(1/\mu_j)(y_j/n_j - \mu_j) + log(\mu_j)$ and iterate the computation of least squares estimates $\hat{\beta}$.

Note that the CEB data contains grouped "counts" computed as $y_j = \mu_j n_j$ where the μ_j values are rounded. And, as a result, the y_j counts are not integers. This does not affect our "by hand" calculation of $\hat{\beta}$ whatsoever because we never use the full Poisson PMF in our computations; the glm function in R, on the other hand, will throw many warnings if the y_j values are not rounded, apparently because it uses the PMF via dpois "under the hood". The only differences between our fitted $\hat{\beta}$ and glm's are due to rounding y_j 's.

```
n <- nrow(ceb)
group.sizes <- ceb$n
Y <- ceb$y
# IRLS - factor coding
# initialize with mu = Y/group.sizes
options(contrasts = c('contr.treatment', 'contr.treatment'))
X <- model.matrix(y~dur+res+educ, data = ceb)</pre>
mu <- Y/group.sizes
XB <- log(mu)
W <- diag(as.numeric(mu))</pre>
z \leftarrow -(1/mu)*(Y/group.sizes-mu) + XB
beta <- solve(t(X)%*%W%*%X)%*%t(X)%*%W%*%z
tol <- 0.0001
difference <- 1
maxiter <- 100
iter <- 1
while((difference > tol) & (iter < maxiter)){</pre>
  XB <- X%*%beta
  mu \leftarrow exp(XB)
  W <- diag(as.numeric(group.sizes*mu))</pre>
  z \leftarrow (Y/diag(W) - rep(1,n)) + XB
  beta.old <- beta
  beta <- solve(t(X)%*%W%*%X)%*%t(X)%*%W%*%z
```

```
difference <- max(abs(beta - beta.old))</pre>
  iter<-iter+1
}
beta
##
                    [,1]
## (Intercept) 0.05695417
## dur10-14
               1.37053208
## dur15-19
              1.61423104
## dur20-24
              1.78548879
## dur25-29
              1.97679469
## dur5-9
              0.99765038
## resSuva
             -0.15121728
## resurban
           -0.03895822
## educnone
             -0.02308034
## educsec+
              -0.33266474
## educupper -0.12474575
## the glm function can be used with offset equal to logarithm of the group sizes
my.glm <- glm(round(y)~dur+res+educ, family = poisson(link = 'log'), data = ceb, offse
summary(my.glm)
##
## Call:
## glm(formula = round(y) ~ dur + res + educ, family = poisson(link = "log"),
      data = ceb, offset = log(n)
##
## Deviance Residuals:
##
      Min
            1Q Median
                                 3Q
                                        Max
## -2.2960 -0.6641
                    0.0725
                             0.6336
                                     3.6782
##
## Coefficients:
##
              Estimate Std. Error z value Pr(>|z|)
## (Intercept) 0.05754
                        0.04803 1.198
                                           0.231
## dur10-14
                         0.05107 26.815 < 2e-16 ***
              1.36940
## dur15-19
              1.61376
                         0.05119 31.522 < 2e-16 ***
## dur20-24
                         0.05121 34.852 < 2e-16 ***
              1.78491
## dur25-29
             1.97641
                         0.05003 39.501 < 2e-16 ***
## dur5-9
             ## resSuva
             -0.15166
                         0.02833 -5.353 8.63e-08 ***
## resurban
             -0.03924
                         0.02463 -1.594
                                           0.111
## educnone -0.02297
                         0.02266 -1.014
                                           0.311
                         0.05390 -6.180 6.41e-10 ***
## educsec+ -0.33312
## educupper -0.12425 0.03000 -4.142 3.44e-05 ***
```

```
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## (Dispersion parameter for poisson family taken to be 1)
##
## Null deviance: 3731.852 on 69 degrees of freedom
## Residual deviance: 70.665 on 59 degrees of freedom
## AIC: 522.14
##
## Number of Fisher Scoring iterations: 4
```

3.4 Inference on GLMs

No doubt you noticed the glm function output produces standard errors, "z" values, and p-values for each fitted coefficient, just as you would find accompanying summarized lm output. But, what is the justification for these p-values?

Since $\hat{\beta}$ is an MLE, standard likelihood theory holds that $\hat{\beta} \sim N_p(\beta, I^{-1}(\beta))$ for "large" n where $I^{-1}(\beta)$ is the Fisher information. As discussed above, the "observed information" (which is the Hessian) is equal to $-\phi^{-1}(X^\top WX)^{-1}$ where W is the weight matrix in the final iteration of IRLS, and $-\phi^{-1}(X^\top WX)^{-1}$ coincides with the Fisher Information when we replace $h(\mu_i)$ by $E(h(\mu_i)) = 1$ in the IRLS (then called Fisher scoring) updates. Therefore, $\hat{\beta} \sim N_p(\beta, \phi^{-1}(X^\top WX)^{-1})$ for "large" n, but this is only useable if ϕ is known (which it is, and equals 1, for the Poisson and Binomial models). Otherwise, we replace ϕ by its MLE and use the corresponding Student's t distribution with n-p degrees of freedom rather than the standard normal for inference on β_i .

The upshot is that we may base tests for, e.g., $H_0: \beta_j = 0$, on Student's t with n-p df; i.e.,

$$\text{Reject } H_0: \beta_j = 0 \text{ if } \left| \frac{\hat{\beta}_j}{\sqrt{\hat{\phi}^{-1}(X^\top W X)_{j,j}^{-1}}} \right| > t_{1-\alpha/2,n-p}.$$

Multivariate Wald simultaneous $100(1-\alpha)\%$ confidence regions are given by the eliiptical contours:

$$\phi \text{ known:} \quad \{\beta: (\hat{\beta}-\beta)^\top \phi^{-1} (X^\top W X)^{-1} (\hat{\beta}-\beta) < \chi^2_{1-\alpha,p} \}$$

$$\phi \text{ unknown:} \quad \{\beta: (\hat{\beta}-\beta)^\top \hat{\phi}^{-1} (X^\top W X)^{-1} (\hat{\beta}-\beta) < F_{1-\alpha,p,n-p} \}$$

Moreover, an approximate 95% CI for the mean response $\mu = g^{-1}(x^{\top}\beta)$ for covariate vector x is given by the Delta method interval:

$$g^{-1}(\boldsymbol{x}^{\intercal}\hat{\boldsymbol{\beta}}) \pm t_{1-\alpha/2,n-p} \sqrt{\hat{\phi}^{-1} \left[\nabla_{\boldsymbol{\beta}} g^{-1}(\boldsymbol{x}^{\intercal}\boldsymbol{\beta}) \right]^{\intercal} (\boldsymbol{X}^{\intercal} W \boldsymbol{X})^{-1} \left[\nabla_{\boldsymbol{\beta}} g^{-1}(\boldsymbol{x}^{\intercal}\boldsymbol{\beta}) \right]}.$$

In multiple linear regression (Gauss-Markov) models we use partial F tests (which are likelihood ratio tests) to test for significance of sets of covariates, i.e., $H_0: \beta_j = \beta_{j+1} = \dots = \beta_{j+\ell} = 0$. For GLMs, similar tests are available. For models where ϕ is known, we have

$$-2\{\ell(\hat{\beta}_{h_0})-\ell(\hat{\beta})\} \overset{H_0}{\sim} F_{\ell,n-p}$$

where $\hat{\beta}_{h_0}$ is the MLE under the null hypothesis where ℓ coefficients are set equal to 0.

There are several methods to estimate ϕ when unknown. Pearson's method observes that

$$\phi^{-1}X^2\stackrel{.}{\sim} \chi^2_{n-p}\quad \text{where}\quad X^2:=\sum_{i=1}^n\frac{(Y_i-\hat{\mu}_i)^2}{\phi V(\hat{\mu}_i)}$$

if the model fits the data adequately. Hence, $\hat{\phi}_P = X^2/(n-p)$ ought to be a good estimate of ϕ . For certain data sets, such as Poisson data with low counts, the Pearson estimate may behave badly, and a modified version (Fletcher's estimator) is preferred:

$$\begin{split} \hat{\phi}_F &= \frac{\hat{\phi}_P}{1-\overline{s}}, \quad \text{where} \\ \overline{s} &:= n^{-1} \sum_{i=1}^n V'(\hat{\mu}_i) \frac{(y_i - \hat{\mu}_i)}{V(\hat{\mu}_i)}. \end{split}$$

The Deviance difference for models A and B where $A \subset B$ is given by $D_A - D_B = -2\{\ell(\hat{\beta}_A) - \ell(\hat{\beta}_B)\}\phi$. The scaled deviance difference is

$$D_A^* - D_B^* = -2\{\ell(\hat{\beta}_A) - \ell(\hat{\beta}_B)\} \stackrel{\cdot}{\sim} \chi_\ell^2$$

where the difference in number of fitted parameters is ℓ . Despite the notation, the scaled deviance difference does depend on ϕ , whereas the deviance difference does not. Two alternative tests make use of the scaled deviance to compare nested GLMs. The first is analogous to the partial F test:

$$F = \frac{(D_A^* - D_B^*)/\ell}{D_B^*/(n-p)} \stackrel{\cdot}{\sim} F_{\ell,n-p}$$

but the approximation to the F distribution is very rough. Alternatively, we can replace the scale parameter in the scaled deviance by its Pearson (or Fletcher) estimator and obtain

$$\hat{D}_A^* - \hat{D}_B^* \stackrel{\cdot}{\sim} \chi_\ell^2,$$

where the "hat" on the scaled deviances indicates their dependence on $\hat{\phi}$.

3.4.1 Inference and prediction for the CEB data using Poisson regression

Next, we'll demonstrate computations of confidence intervals and tests for significance of sets of covariates within the Poisson regression model for the CEB data. We can either do these computations "by hand" using the results of our IRLS or we can use built-in R functions like glm and confint.

For Wald-type confidence intervals for regression coefficients we require the inverse Hessian (the estimate of the inverse Fisher information, equal when Fisher scoring is used). We compute this from the final iteration of IRLS (assuming the algorithm converged). For most GLMs we will need to estimate the scale parameter ϕ and we include the Pearson and Fletcher estimates below—but in the Poisson model $\phi=1$. Note both estimates are close to 1. The p-values included in the summarized glm output imply the intercept is not significantly different from zero, but that several other coefficients are different from zero, including, e.g., β_1 , β_2 , and β_3 . Our Wald-type CIs and p-values agree with the glm p-values; for example, our 95% CI for β_0 computed "by hand" is (-0.0372 0.1511), suggesting the intercept is not significantly different from zero, and our 95% CI for β_1 is (1.2704, 1.4706) with a p-value for the test of $\beta_1=0$ that is indistinguishable from 0.

We computed the deviance difference (which is minus twice the difference in log-likelihood) between the intercept-only model and the full model. The deviance difference is about 3661 on ten degrees of freedom (the difference in number of fitted coefficients between the models). If the intercept only model fits, then this deviance difference should be comparable to a Chi-squared r.v. with 10 degrees of freedom, but the corresponding p-value is basically zero, supporting the claim that the full model fits much better than the intercept-only model. Compare our deviance difference calculation to the output of glm: glm includes the null deviance and residual deviance, the difference of these two gives the deviance difference statistic used to compare the intercept-only and full models. It is about 3661, agreeing almost exactly with our "by hand" calculation.

```
Hessian <- t(X)%*%W%*%X
inv.Hessian <- solve(Hessian)
p <- length(beta)

# Just for illustration, phi = 1 for Poisson model
Pearson.X2 <- sum(((Y - group.sizes * mu)^2) / (group.sizes * mu))
Pearson.phi <- Pearson.X2 / (n-p)
s.bar <- mean((Y - group.sizes * mu) / (group.sizes * mu))
Fletcher.phi <- Pearson.phi/(1-s.bar)
Pearson.phi</pre>
```

```
Fletcher.phi
## [1] 1.194283
# CIs for the first 4 regression coefficients
# If phi were unknown, it's estimate would appear in the estimated standard error of
# estimated coefficient
# beta[1] + qt(c(0.025, 0.975), n-p)*sqrt((1/Pearson.phi)*inv.Hessian[1,1])
beta[1] + qnorm(c(0.025, 0.975))*sqrt(inv.Hessian[1,1])
## [1] -0.03721264 0.15112097
\texttt{beta[2]} + \texttt{qnorm}(\texttt{c(0.025,0.975)}) * \texttt{sqrt(inv.Hessian[2,2])}
## [1] 1.270425 1.470639
2*(1-pnorm(abs(beta[2]/sqrt(inv.Hessian[2,2]))))
## [1] 0
beta[3] + qnorm(c(0.025,0.975))*sqrt(inv.Hessian[3,3])
## [1] 1.513870 1.714592
beta[4] + qnorm(c(0.025,0.975))*sqrt(inv.Hessian[4,4])
## [1] 1.685091 1.885886
# The R function confint can also be used with GLMs to provide confidence intervals fo
confint(my.glm, 'dur10-14')
## Waiting for profiling to be done...
##
      2.5 % 97.5 %
## 1.270141 1.470370
```

```
# "Model F Test" - testing that all (non-intercept) coefficients equal zero
# for Poisson model, since phi is known, we have a LRT equivalent to a partial F test
# based on deviance difference
Ybar <- sum(Y)/sum(group.sizes)
D <- -2*(sum(Y*log(group.sizes*Ybar))-sum(group.sizes*Ybar) - sum(Y*log(group.sizes*mu))+sum(group.sizes*Ybar)
Ybar
## [1] 3.960497
## [1] 3660.872
1-pchisq(D,p-1)
## [1] 0
# with rounded Ys and using glm output
Yr <- round(Y)
mu.glm <- exp(X%*%matrix(my.glm$coefficients,p,1))</pre>
Ybar <- sum(Yr)/sum(group.sizes)
D <- -2*(sum(Yr*log(group.sizes*Ybar))-sum(group.sizes*Ybar) - sum(Yr*log(group.sizes*mu.glm))+sum(group.sizes*Ybar)
Ybar
## [1] 3.960403
## [1] 3661.186
1-pchisq(D,p-1)
## [1] 0
```

3.5 Model Checking/Diagnostics

It is essential statistical practice to check whether the model adequately fits the data. If the model fits poorly, then inferences/predictions garnered from the model are suspect. In multiple linear regression we assess model fit by analyzing residuals. When the multiple linear regression model fits, the residuals should

be approximately standard normal. Lack of fit manifests in residuals that are skewed or heavy-tailed, contain outliers, or tend to increase in variability with one or more covariates and/or the predicted responses.

Model-checking for GLMs can be done in essentially the same manner—the key is to find a quantity that reasonably fills the role of residuals in multiple linear regression. For GLMs, there are two choices, the Pearson residuals and the deviance residuals.

Pearson residuals are defined $e_i^P=\frac{Y_i-\hat{\mu}_i}{\sqrt{V(\mu_i)}},$ or, sometimes, $e_i^P=\frac{Y_i-\hat{\mu}_i}{\sqrt{\hat{\phi}V(\mu_i)}}.$ The

first definition results in quantities that should be approximately zero-mean normal random variates with variance ϕ whereas the second definition provides standard normal quantities. Some practitioners prefer the deviance residuals to the Pearson residuals as the latter are often observed to be asymmetric and, hence, not as "normal" as expected. The deviance is equal to

Deviance =
$$-2\phi\{\ell(\hat{\beta}) - \sup \ell\}$$

where $\ell(\hat{\beta})$ is the loglikelihood evaluated at the MLEs and $\sup \ell$ is the loglikelihood with $\mu_i = y_i$, i.e., a totally saturated model. Multiplying by ϕ removes the dependence of the loglikelihood on the scale parameter. The deviance can be written as a sum of terms, say, Deviance $= \sum_{i=1}^n d_i$, and each observation's contribution d_i to the deviance is used to define the deviance residuals as follows:

$$e_i^D = \mathrm{sign}(y_i - \mu_i) \sqrt{d_i}.$$

3.5.1 Residual analysis for CEB data

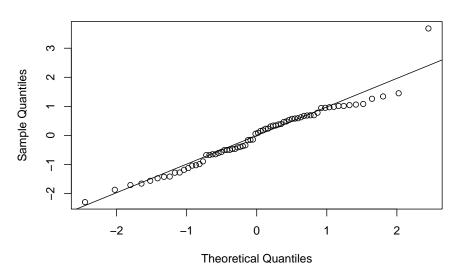
Using either the deviance residuals or the Pearson residuals shows a few important things. First, the residuals are approximately standard normal, with the exception of one "outlier", observation 17. Second, if we sort the observations by fitted mean response $\hat{\mu}_i$ from least to greatest, we see absolutely no trend up or down, or any pattern at all, in the residuals. This implies we have correctly modeled the mean-variance relationship, and that we have also correctly modeled the mean as a linear function of the covariates.

We obtain similar plots by simply running "plot(glm object)", but there are some default differences. Plotting a glm object will provide plots of "residuals" versus "fitted values", but in fact, the labels on these plots are slightly misleading. The residuals vs. fitted values plots uses either Pearson or deviance residuals (cannot tell for sure from the plot or the documentation) versus the logarithm of the fitted responses $\log(\hat{y}_i) = \log(n_i\hat{\mu}_i)$.

```
dev <- -2*((Y*log(group.sizes*mu))-(group.sizes*mu) - ((Y*log(Y))-(Y)))
deviance.resids <- ifelse((Y-group.sizes*mu) < 0,-1,1)*sqrt(dev)
pearson.resids <- (Y - group.sizes*mu)/sqrt(group.sizes*mu)</pre>
```

```
# residual plots using deviance residuals
qqnorm(deviance.resids)
qqline(deviance.resids)
```

Normal Q-Q Plot

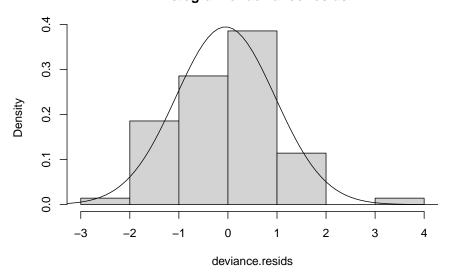


shapiro.test(deviance.resids)

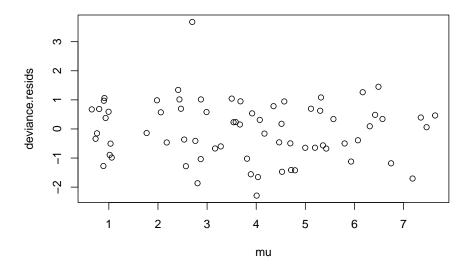
```
##
## Shapiro-Wilk normality test
##
## data: deviance.resids
## W = 0.96065, p-value = 0.02719
```

```
hist(deviance.resids, freq = FALSE)
dev.norm <- function(x) dnorm(x,mean(deviance.resids), sd(deviance.resids))
curve(dev.norm, -5,5, add = TRUE)</pre>
```

Histogram of deviance.resids

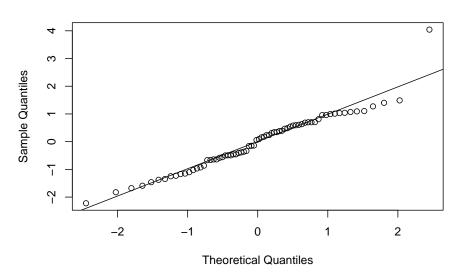


plot(mu, deviance.resids)



```
# residual plots using pearson residuals
qqnorm(pearson.resids)
qqline(pearson.resids)
```

Normal Q-Q Plot

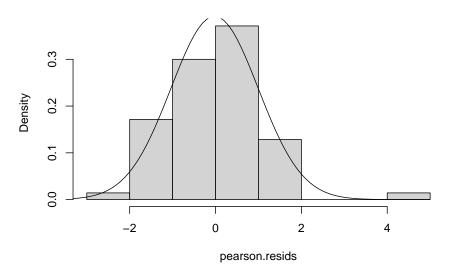


```
shapiro.test(pearson.resids)
```

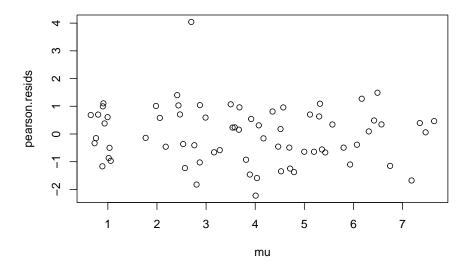
```
##
## Shapiro-Wilk normality test
##
## data: pearson.resids
## W = 0.94988, p-value = 0.007147

hist(pearson.resids, freq = FALSE)
pearson.norm <- function(x) dnorm(x,mean(pearson.resids), sd(pearson.resids))
curve(pearson.norm, -5,5, add = TRUE)</pre>
```

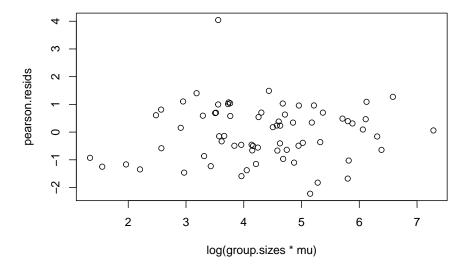
Histogram of pearson.resids

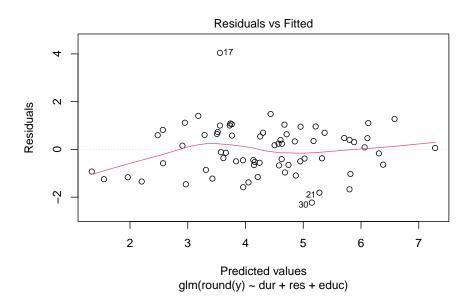


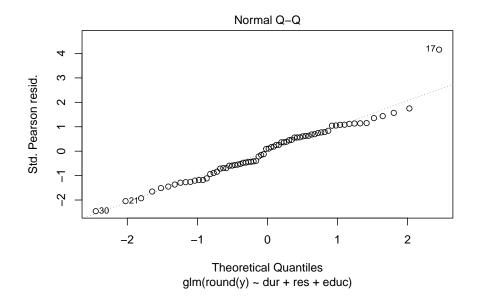
plot(mu, pearson.resids)

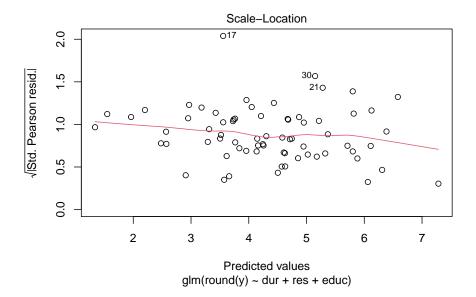


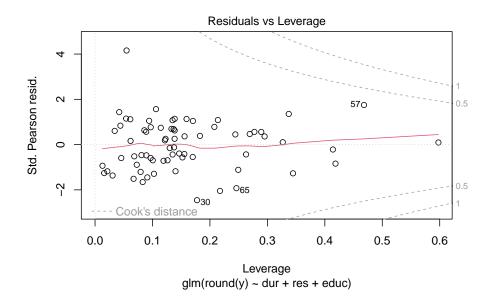
plots are using what look like Pearson (or maybe deviance) residuals versus predicted values un plot(log(group.sizes*mu), pearson.resids)











3.6 Outlier analysis using Cook's distance

Outliers are observations corresponding to large residuals. They may occur due to chance, or, more likely, indicate a lack of model fit for a specific observation. The lack of fit may be due to something innocuous like a mistake made in recording data; or, it may be that the observation in question is very different from the others in the sample, and does not follow the same response-covariate relationship.

Outliers are not a problem, however, unless their inclusion causes the fitted model to be substantially different than had they been excluded. Therefore, we do not necessarily care that a particular residual is large, but that it is influential on the model fit.

In multiple linear regression we may measure this influence by Cook's distance of the data point, which is related to both the magnitude of its residual and its leverage, as measured by the hat (or influence) matrix. For GLMs we can define the Cook's distance in the same manner: the Cook's distance of data point k is given by

$$\begin{split} C_k &= \frac{1}{(p+1)} \sum_{i=1}^n \frac{(\hat{\mu}_i^{[k]} - \hat{\mu}_i)^2}{\hat{\phi} V(\hat{\mu}_i)}; \text{ or } \\ C_k &= \frac{(e_k^P)^2}{\hat{\phi} (p+1)} \frac{h_k}{(1-h_k)^2} \end{split}$$

where $H=W^{1/2}X(X^\top WX)^{-1}X^\top W^{1/2}$ is the hat matrix and h_k is its k^{th} diagonal entry. Large Cook's distance implies the model predictions change substantially when the data point in question is removed.

3.6.1 Outlier analysis for the CEB data

Typically data points with Cook's distance > 1 are considered highly influential and the case for their exclusion is considerable. In this case, our apparent outlier is not influential, nor is it even the most influential observation sampled.

Below we have computed Cook's distance "by hand" and by using the corresponding built-in R function; the only (very slight) difference between the two seems to be caused by the fact the built-in function uses the glm object, which we fitted after rounding the responses, whereas our "by hand" calculation uses Pearson residuals based on the original (unrounded) responses.

```
ceb2 <- ceb[-17,]
my.glm2 <- glm(round(y)~dur+res+educ, family = poisson(link = 'log'), data = ceb2, off

W2 <- sqrt(W)
h <- W2%*%X%*%solve(t(X)%*%W%*%X)%*%t(X)%*%W2</pre>
```

```
((pearson.resids[17]^2)/p)*(h[17,17]/((1-h[17,17])^2))
## [1] 0.0912897
cd <- cooks.distance(my.glm)</pre>
cd[17]
          17
## 0.09123744
sort(cd)
                                                                             26
            44
                         12
                                      16
                                                   43
                                                                58
## 0.0001578983 0.0002173653 0.0003181272 0.0004391595 0.0004872775 0.0008276467
                         48
                                      69
                                                    5
                                                                40
## 0.0009604834 0.0010416260 0.0011600877 0.0011813567 0.0015364693 0.0017374273
            63
                         15
                                      59
                                                   64
                                                                8
## 0.0017913034 0.0020157684 0.0022231406 0.0023307859 0.0024333875 0.0027257717
             1
                         29
                                      54
                                                   56
                                                                34
## 0.0027795217 0.0028483882 0.0029171612 0.0029318194 0.0030296972 0.0030322783
            55
                         36
                                      27
                                                   33
                                                                 9
## 0.0033869538 0.0034063422 0.0048780994 0.0049440311 0.0052402581 0.0053485413
                         6
                                      24
                                                    3
## 0.0055965679 0.0056512722 0.0056712755 0.0057553959 0.0059235574 0.0061112712
                                                   49
                                      14
## 0.0062842199 0.0063288779 0.0064608723 0.0067079644 0.0067366794 0.0068678049
                                                    7
##
            66
                         25
                                      13
                                                                61
## 0.0072889100 0.0074308461 0.0082351514 0.0104259702 0.0110105154 0.0114006236
            70
                         42
                                      52
                                                   20
                                                                28
## 0.0115384460 0.0145603375 0.0149361729 0.0164709118 0.0173145140 0.0189913643
            51
                         38
                                      53
                                                   19
                                                                39
## 0.0191422888 0.0203317679 0.0205784895 0.0220776790 0.0225077753 0.0265192825
            35
                         10
                                      45
                                                   22
                                                                46
## 0.0290870153 0.0375227510 0.0462157923 0.0771290592 0.0847041708 0.0912374357
            21
                         65
                                      30
                                                   57
## 0.1058278940 0.1102296604 0.1184297114 0.2448544534
```

Linear Mixed Models

4.1 ANOVA with random factors

The simplest linear mixed models are used to analyze linear models for one-way ANOVA, randomized complete block designs, and two-way ANOVA. *Mixed* models as opposed to *fixed* models (the linear models you have heretofore studied) are needed when factors have levels that are **random**. Random levels occur whenever the units making up those levels behave like random samples from a population. Two examples are given below. And, we discuss how to perform ANOVA-like tests for factors that are random rather than fixed. The upshot is that (at least for balanced experiments/datasets) the tests for fixed effects are identical to those for random effects, only the interpretation is different (and importantly so!).

4.1.1 Strength of metallic bonds

The dataset below, called "Bonds", contains responses for 21 samples of metals, 7 each for iron, nickel, and copper, that quantify the strength of metallic bonds. One sample from each metal was extracted from each of 7 ingots. We expect ingots to act like blocks—differences in ingots account for a substantial amount of variability in the responses, but the precise block effects are of no inferential/scientific inference. We only include the blocks in order to reduce the residual variance after accounting for block variance. A randomized controlled block design describes how this data was collected, but, if we repeated the experiment, the blocks (ingots) would be completely different. That is, the blocks are not fixed but random. Rather than estimating block effects that would surely change experiment to experiment, we should focus on estimating the amount of variability explained by the blocks, which should remain about

the same experiment to experiment. This suggests a different model than used to analyze RCBD experiments with fixed blocks.

The usual linear model for fixed blocks is

$$y_{ijk} = \mu + \alpha_i + \beta_j + \epsilon_{ij},$$

where y_{ij} is the response for treatment (metal) i in block (ingot) j; the α_i 's are the metal (treatment) effects; the β_j 's are the ingot (block) effects; and, $\epsilon_{ij} \stackrel{iid}{\sim} N(0, \sigma^2)$ are the random residuals.

The above linear model is the wrong model for this data because the block effects (and, hence, also the interaction effects) are meaningless outside of the given data set; these are not population-level parameters because the blocks are random rather than fixed. The appropriate model (given normality and independence of random residuals is reasonable) is the following *mixed effects model*:

$$y_{ij} = \mu + \alpha_i + \beta_j + \epsilon_{ij}, \tag{4.1}$$

where $\beta_j \stackrel{iid}{\sim} N(0, \sigma_b^2)$ and, independently, $\epsilon_{ij} \stackrel{iid}{\sim} N(0, \sigma^2)$.

For balanced experiments (the number of replicates is equal across each combination of factor levels) we can test for block and treatment effects by comparing nested/aggregated models. Let \overline{Y}_i denote the mean response for metal i averaged over ingots. We can write down the following aggregated model from (4.1) as

$$\overline{y}_{i} = \mu + \alpha_{i} + \epsilon_{i}, \tag{4.2}$$

where $\epsilon_i = \frac{1}{J} \sum_{j=1}^{J} \epsilon_{ij}$. Then, ϵ_j has variance $\sigma_b^2 + \sigma^2/J$. The F statistic

$$F = \frac{J \cdot MSE_{agg}}{MSE_{full}}$$

where MSE_{agg} and MSE_{full} are the mean squared errors from the models in (4.2) and (4.1) can be used to test the hypothesis $H_0: \sigma_b^2 = 0$.

4.1.2 Machine productivity

The dataset given below contains the results of a designed experiment to evaluate worker productivity using 3 different industrial machines. The goal is to determine which machine is most productive while controlling for natural variation in worker productivity. The observed workers represent a random sample from a population of workers (blocks), analogous to the ingots in the previous example. The difference between the two examples (besides the context) is that the machine treatments are replicated within workers, so that there are three

observations of a productivity score fore each worker for each type of machine. This means that we can fit a model with *interaction terms* capable of capturing changes in machine effects on productivity between different workers (if those changes are present):

$$y_{ijk} = \mu + \alpha_i + \beta_j + (\alpha \beta)_{ij} + \epsilon_{ijk}, \tag{4.3}$$

where k denotes the k^{th} replicate within machine i and worker j; and where $(\alpha\beta)_{ij}$ denote the machine-worker interaction effects.

Let \overline{Y}_{ij} be the mean response averaging over replicates for the treatment i and block j combination. Then,

$$\begin{split} V(\overline{Y}_{ij\cdot}) &= V\left(K^{-1}\sum_{k=1}^K Y_{ijk}\right) \\ &= \frac{1}{K^2}V\left(\sum_{k=1}^K \{\mu + \alpha_i + \beta_j + (\alpha\beta)_{ij} + \epsilon_{ijk}\}\right) \\ &= \frac{1}{K^2}V\left(K\mu + K\alpha_i + K\beta_j + K(\alpha\beta)_{ij} + \sum_{k=1}^K \epsilon_{ijk}\right) \\ &= \sigma_b^2 + \sigma_{ab}^2 + K^{-1}\sigma^2. \end{split}$$

If we rewrite the model for the cell mean responses as

$$\overline{y}_{ii} = \mu + \alpha_i + \beta_i + \epsilon_{ii}, \tag{4.4}$$

then the aggregated error term follows $\epsilon_{ij} \stackrel{iid}{\sim} N(0,\sigma_{ab}^2+\sigma^2/K)$. The residual mean square (or called the mean squared error) of model (4.3) (let's call it $MSE_{\rm full}$) has mean σ^2 with $n-p_1$ degrees of freedom where n is the sample size and p is the number of coefficients in the fitted model (p_1 equals the number of crossed factor levels, the number of blocks times the number of treatments). The residual mean square for the aggregated model in (4.4) (let's call it $MSE_{\rm agg}$) has mean $\sigma_{ab}^2+\sigma^2/K$ with $n/K-p_2$ degrees of freedom where p_2 is the number of treatments plus the number of blocks minus 1. An unbiased estimate of σ_{ab}^2 is given by $MSE_{\rm agg}-\frac{1}{K}MSE_{\rm full}$. Consider testing the null hypothesis $H_0:\sigma_{ab}^2=0$. The statistic

$$F := \frac{K \cdot MSE_{agg}}{MSE_{full}} \overset{H_0}{\sim} F_{n/K-p_2, n-p_1},$$

that is, under the null hypothesis. The test that rejects H_0 for $F > F_{1-\alpha,n/K-p_2,n-p_1}$ is exactly equivalent to the partial F test between the full model and the full model without the interaction terms (the additive model).

Below we use R to compute ANOVA tables for the full model, full model without interaction, and the aggregated model. The F test statistic for the aggregated model is about 46.13 on 10 and 36 degrees of freedom, which exactly matches the partial F test between the full and additive models.

```
library(nlme)
# aggregated model
Mach.agg <- aggregate(score~Machine*Worker, data = Machines, FUN=mean)</pre>
m2 <- lm(score~Machine+Worker, data = Mach.agg)</pre>
anova(m2)
## Analysis of Variance Table
##
## Response: score
            Df Sum Sq Mean Sq F value
## Machine 2 585.09 292.544 20.5761 0.0002855 ***
            5 413.97 82.793 5.8232 0.0089495 **
## Worker
## Residuals 10 142.18 14.218
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
# full model with interaction
m0 <- lm(score~Machine*Worker, data = Machines)</pre>
anova(m0)
## Analysis of Variance Table
##
## Response: score
##
                 Df Sum Sq Mean Sq F value
                                               Pr(>F)
## Machine
                 2 1755.26 877.63 949.17 < 2.2e-16 ***
## Worker
                 5 1241.89 248.38 268.63 < 2.2e-16 ***
## Machine:Worker 10 426.53
                             42.65
                                     46.13 < 2.2e-16 ***
## Residuals
                 36
                      33.29
                               0.92
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
(142.18*3/10)/(33.29/36)
## [1] 46.12628
1-pf((142.18*3/10)/(33.29/36), 10, 36)
```

[1] 0

```
# additive model (no interaction)
m1 <- lm(score~Machine+Worker, data = Machines)
anova(m1)
## Analysis of Variance Table
## Response: score
##
            Df Sum Sq Mean Sq F value
             2 1755.26 877.63 87.798 < 2.2e-16 ***
## Worker
             5 1241.89
                        248.38
                                24.848 4.867e-12 ***
## Residuals 46 459.82
                         10.00
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
anova(m1,m0)
## Analysis of Variance Table
##
## Model 1: score ~ Machine + Worker
## Model 2: score ~ Machine * Worker
##
     Res.Df
              RSS Df Sum of Sq
                                        Pr(>F)
## 1
        46 459.82
## 2
        36 33.29 10
                        426.53 46.13 < 2.2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

4.2 A general linear mixed model

For experiments comparing responses between factors the ANOVA-type analyses above are sufficient. But, for more general models with random effects, e.g., those including continuous covariates, general-purpose methods are needed. The general mixed effects model may be written

$$Y = X\beta + Z\alpha + \epsilon$$

where Y is an $n \times 1$ response, X is an $n \times p$ design matrix for fixed (non-random) effects; Z is and $n \times a$ matrix for random effects; β is a $p \times 1$ non-random coefficient vector; $\alpha \sim N_a(0,\psi_\theta)$ is an $a \times 1$ multivariate normal random coefficient vector with mean 0 and covariance matrix ψ_θ indexed by a parameter θ ; and $\epsilon \sim N_n(0,\Lambda_\theta)$ is a multivariate normal random residual vector with covariance matrix Λ_θ . An alternative way of writing the model (quite succintly) is

$$Y \sim N_n(X\beta, Z\psi_\theta Z^\top + \Lambda_\theta). \tag{4.5}$$

4.2.1 Parameter estimation using PML

The linear mixed model in (4.5) may be fit using maximum likelihood estimation (MLE), but the (potentially) complicated covariance structure poses challenges to numerical maximization of the likelihood function. Rather than straightforward MLE, linear mixed models are usually fit using either restricted maximum likelihood estimation (REML, also called residual MLE) or profile maximum likelihood estimation (PML). Both approaches aim to simplify the computation of the maximum while retaining the good asymptotic properties of maximum likelihood estimators. While MLE is a frequentist concept, it turns out that REML can be derived most intuitively using a Bayesian approach.

If you have previously covered general linear models, i.e., $Y = X\beta + \epsilon$ where $Cov(\epsilon) = \Sigma$ for some known Σ or some matrix $\Sigma = \sigma^2 V$ for an unknown scalar σ^2 and a known matrix V, then you are familiar with weighted least squares (WLS) estimation. If the linear mixed model covariance parameter θ were known, then the model could be fit using WLS:

$$\hat{\beta}(\theta) = (X^\top W^{-1} X)^{-1} X^\top W^{-1} Y, \quad \text{where} \quad W = Z \psi_{\theta} Z^\top + \Lambda_{\theta}.$$

The above WLS solution inspires the PML technique: maximize the likelihood with respect to θ after plugging in $\beta = \hat{\beta}(\theta)$. PML reproduces MLEs exactly; its advantage is its simplified formulation of the likelihood maximization problem.

Nevertheless there are two problems with the PML strategy suggested above: one computational and the other statistical.

The first problem is that PML/WLS requires inverting the $n\times n$ matrix $W=Z\psi_{\theta}Z^{\top}+\Lambda_{\theta}$. For even moderate sample sizes this matrix inversion can be both computationally demanding and computationally unstable. Fortunately, some clever linear algebra resolves this problem. Define matrix $A:=\psi^{-1}+Z^{\top}\Lambda^{-1}Z$. Observe that the matrix $\Lambda^{-1}-\Lambda^{-1}ZA^{-1}Z^{\top}\Lambda^{-1}$ equals the inverse of $Z\psi Z^{\top}+\Lambda$:

$$\begin{split} &(\Lambda^{-1} - \Lambda^{-1}ZA^{-1}Z^{\top}\Lambda^{-1})(Z\psi Z^{\top} + \Lambda) \\ &= \Lambda^{-1}Z\psi Z^{\top} + I - \Lambda^{-1}ZA^{-1}Z^{\top}\Lambda^{-1}Z\psi Z^{\top} - \Lambda^{-1}ZA^{-1}Z^{\top} \\ &= I + \Lambda^{-1}Z(\psi - A^{-1}(Z^{\top}\Lambda^{-1}Z\psi + I))Z^{\top} \\ &= I + \Lambda^{-1}Z(\psi - \psi(Z^{\top}\Lambda^{-1}A\psi + I)^{-1}(Z^{\top}\Lambda^{-1}A\psi + I))Z^{\top} \\ &= I \end{split}$$

The key property of this inverse is that it only requires computing the inverse of $p \times p$ and $a \times a$ matrices, and does not require the computation of any $n \times n$ matrix inverse. In many practical applications both p and a are much smaller than n, and the matrices Λ and ψ often have simple structures, so that the above matrix inversions may be performed very quickly and reliably.

The second issue with PML cannot as easily be overcome. Think back to the Gauss-Markov model $Y = X\beta + \epsilon$ where $Cov(\epsilon) = \sigma^2 I_n$. Recall that the MLE

 $\hat{\sigma}^2$ is biased, i.e., $E(\hat{\sigma}^2) = \frac{n-p}{n}\sigma^2$. In practice we may have a substantial number of covariates relative to sample size so that this bias can be quite significant; and, importantly, the bias results in an underestimate which causes over-optimism with respect to inferences concerning β . The same phenomenon occurs in ML/PML estimation: the PML estimate of θ is biased. The desire for an estimation strategy that constructively produces unbiased estimates is the motivation for REML, covered next.

4.2.2 REML - frequentist approach

There are two formulations both leading to the residual (also called restircted/reduced) maximum likelihood approach (REML), one frequentist (non-Bayesian) and the other Bayesian. Let's explore both approaches.

The motivation for REML is to produce unbiased (or at least less biased than MLE) estimates of the covariance parameters. The REML approach to doing this may be interpreted as a likelihood approach based on residuals, or as a marginal likelihood approach to covariance parameter estimation.

Begin with the linear mixed model in (4.5). Assume X has full rank p and let $L = [L_1 \ L_2]$ denote a block matrix of $n \times p$ and $n \times (n-p)$ blocks with the properties

$$L_1^{\top}X = I_n$$
 and $L_2^{\top}X = 0_{n-n}$,

such that L has full rank n.

This setup is a bit abstract, so it helps to find at least one concrete example of such an L. Start with the projection matrix $P_X := X(X^\top X)^{-1}X^\top$. Since P_X is a symmetric, idempotent matrix of rank p it has exactly p eigenvalues of value 1 and n-p 0 eigenvalues, and its eigenvectors are orthonormal. Hence,

$$P_X = \begin{bmatrix} V_1 & V_2 \\ V_3 & V_4 \end{bmatrix} \begin{bmatrix} \mathbf{1}_{p \times p} & \mathbf{0}_{p \times n-p} \\ \mathbf{0}_{n-p \times p} & \mathbf{0}_{n-p \times n-p} \end{bmatrix} \begin{bmatrix} V_1 & V_2 \\ V_3 & V_4 \end{bmatrix}^\top.$$

It follows from this representation that

$$P_X = \begin{bmatrix} V_1 \\ V_3 \end{bmatrix} \begin{bmatrix} V_1 \\ V_3 \end{bmatrix}^\top =: L_1 L_1^\top$$

Furthermore, if v is an eigenvector of P_X with eigenvalue 1, then v is an eigenvector of $I-P_X$ with eigenvalue 0 just by the definition of an eigenvalue $(Av=\lambda v)$. It follows that

$$I - P_X = \begin{bmatrix} V_2 \\ V_4 \end{bmatrix} \begin{bmatrix} V_2 \\ V_4 \end{bmatrix}^\top =: L_2 L_2^\top.$$

By construction, $L_1L_2^{\top}=0$, $L_1^{\top}L_1=I_p$ and $L_2^{\top}L_2=I_{n-p}$. The columns of L are orthogonal, and so L is full rank.

In what follows, note that the REML estimates are invariant to the choice of L, so long as it has the above three properties.

Now, consider the full rank linear transformation $Y \mapsto LY = [Y_1 \quad Y_2]^\top$. By the properties of L, we have

$$LY = \begin{bmatrix} Y_1 \\ Y_2 \end{bmatrix} \sim N \left(\begin{bmatrix} \beta \\ 0 \end{bmatrix}, \begin{bmatrix} L_1^\top \Sigma L_1 & L_1^\top \Sigma L_2 \\ L_2^\top \Sigma L_1 & L_2^\top \Sigma L_2 \end{bmatrix} \right)$$

where
$$\Sigma = Cov(Y) = Z\psi_{\theta}Z^{\top} + \Lambda_{\theta}$$
.

The next step is to consider an equivalent characterization of the distribution of (Y_1, Y_2) as the product of the conditional distribution of $Y_1|Y_2 = y_2$ and the marginal distribution of Y_2 . Using the general formulas for the conditional distribution of a multivariate normal random vector, we get

$$Y_1|y_2 \sim N(\beta - L_1^{\intercal} \Sigma L_2 (L_2^{\intercal} \Sigma L_2)^{-1} y_2, \ L_1^{\intercal} \Sigma L_1 - L_1^{\intercal} \Sigma L_2 (L_2^{\intercal} \Sigma L_2)^{-1} L_2^{\intercal} \Sigma L_1).$$

Some tedious linear algebra (omitted here) can be used to show that the above covariance matrix is actually equal to $(X^{\top}\Sigma^{-1}X)^{-1}$, regardless of the specific choice of L.

Therefore, the conditional likelihood is given by

$$\ell_c := \text{const.} - \tfrac{1}{2} \log |(X^\top \Sigma^{-1} X)^{-1}| - \tfrac{1}{2} (y_1 - \beta - y_2^\star) X^\top \Sigma^{-1} X (y_1 - \beta - y_2^\star)^\top,$$

where
$$y_2^{\star} = L_1^{\top} \Sigma L_2 (L_2^{\top} \Sigma L_2)^{-1} y_2$$
.

Again, some tedious linear algebra computations can be used to show that

$$L_2(L_2^{\top}\Sigma L_2)^{-1}L_2^{\top} = \Sigma^{-1} - \Sigma^{-1}X(X^{\top}\Sigma^{-1}X)^{-1}X^{\top}\Sigma^{-1},$$

which is an improvement computationally because it requires only inversion of a $p \times p$ matrix and Σ^{-1} , which has an efficient formula; see above. Furthermore, the block matrix determinant identity given by

$$\det \begin{bmatrix} A & B \\ B^\top & C \end{bmatrix} = |C||A - B^\top C^{-1}B|,$$

applied to $L^{\top}\Sigma L$ can be used to show that

$$\log |L_2^\top \Sigma L_2| = \log |L^\top \Sigma L| + \log |X^\top \Sigma^{-1} X|.$$

Further, rules for determinants imply

$$\log |L^{\top} \Sigma L| = \log |L^{\top} L \Sigma| = \log |L^{\top} L| + \log |\Sigma|.$$

Using these simplified expressions, and noting that $\log |L^{\top}L|$ is constant in parameters, we may write tge marginal (or residual) likelihood as

$$\ell_r := \text{const.} - \tfrac{1}{2} \log |\Sigma| + \tfrac{1}{2} \log |X^\top \Sigma^{-1} X| - \tfrac{1}{2} y^\top (\Sigma^{-1} - \Sigma^{-1} X (X^\top \Sigma^{-1} X)^{-1} X^\top \Sigma^{-1}) y.$$

What is the reason for referring to ℓ_r as a residual loglikelihood? Recall ℓ_r is the marginal loglikelihood of $L_2^{\top}Y$ where $L_2^{\top}X = 0$. Therefore,

$$E(L_2^{\top}Y) = L_2^{\top}X\beta = (L_2^{\top}X)\beta = 0\beta = 0.$$

We say $L_2^{\top}Y$ behaves as a residual because $L_2^{\top}X = 0$.

The REML estimate of θ is given by the maximizer of ℓ_r with respect to θ . Note also that this is simple the MLE of θ with respect to the marginal likelihood of $L_2^{\top}Y$, i.e., the likelihood of "part of the data".

Next, consider maximizing the conditional likelihood ℓ_c with respect to β and where $\theta = \hat{\theta}$ is fixed at its REML estimate, i.e., $\Sigma = \hat{\Sigma}$. Take the first derivative with respect to β , equate to zero, and observe

$$\begin{split} \hat{\beta} &= y_1 - L_1^\top \Sigma L_2 (L_2^\top \Sigma L_2)^{-1} y_2 \\ &= L_1^\top y - L_1^\top \Sigma L_2 (L_2^\top \Sigma L_2)^{-1} L_2^\top y \\ &= L_1^\top \Sigma \Sigma^{-1} y - L_1^\top \Sigma L_2 (L_2^\top \Sigma L_2)^{-1} L_2^\top \Sigma \Sigma^{-1} y \\ &= (L_1^\top \Sigma - (L_1^\top \Sigma L_2) (L_2^\top \Sigma L_2)^{-1} L_2^\top \Sigma) \Sigma^{-1} y \\ &= (L_1^\top \Sigma L_1^\top X - (L_1^\top \Sigma L_2) (L_2^\top \Sigma L_2)^{-1} L_2^\top \Sigma L_1^\top X) \Sigma^{-1} y \\ &= [(L_1^\top \Sigma L_1) - (L_1^\top \Sigma L_2) (L_2^\top \Sigma L_2)^{-1} (L_2^\top \Sigma L_1)] X \Sigma^{-1} y \\ &= (X^\top \Sigma^{-1} X)^{-1} X^\top \Sigma^{-1} y. \end{split}$$

using that $L_1^{\top}X = I$ and the equivalence between the conditional covariance of $Y_1|y_2$ and $(X^{\top}\Sigma^{-1}X)^{-1}$ shown above.

We conclude that the REML estimator of β is a weighted least squares estimator in which the weight matrix is given by the REML-based plug-in estimator $\hat{\Sigma}^{-1}$.

4.2.3 REML - Bayesian approach

Suppose we model (in a Bayesian sense) the parameter β with an improper constant prior, and the random component α with a multivariate normal prior with covariance ψ . For whatever parameters define θ , endow those with constant/improper priors as well. Then, combining priors and multivariate normal likelihood we have the following posterior:

$$\log \Pi_n(\beta) = \mathrm{const.} - \tfrac{1}{2} \left(y - X\beta - Z\alpha \right)^\top \Lambda^{-1} \left(y - X\beta - Z\alpha \right) - \tfrac{1}{2} \alpha^\top \psi^{-1} \alpha.$$

A nice (but somewhat hard to find) factorization of the posterior is due to Searle et al. (Variance Components, Section 9.2). Expand the exponent:

$$\begin{split} \log \Pi_n(\beta) &= \text{const.} - \tfrac{1}{2} \left(y - X \beta \right)^\top \Lambda^{-1} \left(y - X \beta \right) \\ &- \tfrac{1}{2} \alpha^\top (\psi^{-1} + Z^\top \Lambda^{-1} Z) \alpha + \alpha^\top Z^\top \Lambda^{-1} (y - X \beta) \end{split}$$

Let $A := \psi^{-1} + Z^{\top} \Lambda^{-1} Z$ and complete the square in α :

$$\begin{split} \log \Pi_n(\beta) &= \mathrm{const.} - \tfrac{1}{2} \left(y - X \beta \right)^\top \Lambda^{-1} \left(y - X \beta \right) \\ &- \tfrac{1}{2} (\alpha - A^{-1} Z^\top \Lambda^{-1} (y - X \beta))^\top A (\alpha - A^{-1} Z^\top \Lambda^{-1} (y - X \beta)) \\ &+ \tfrac{1}{2} (y - X \beta)^\top \Lambda^{-1} Z A^{-1} Z^\top \Lambda^{-1} (y - X \beta). \end{split}$$

Combine the terms quadratic in $y - X\beta$. The resulting matrix $\Lambda^{-1} - \Lambda^{-1}ZA^{-1}Z^{\top}\Lambda^{-1}$ is equal to $(Z\psi Z^{\top} + \Lambda)^{-1}$, which is a fact we used above in the frequentist approach to REML. Therefore, the posterior factorizes into

$$\begin{split} \log \Pi_n(\beta) &= \text{const.} - \tfrac{1}{2} \left(y - X \beta \right)^\top (Z \psi Z^\top + \Lambda)^{-1} (y - X \beta) \\ &- \tfrac{1}{2} (\alpha - A^{-1} Z^\top \Lambda^{-1} (y - X \beta))^\top A (\alpha - A^{-1} Z^\top \Lambda^{-1} (y - X \beta)) \end{split}$$

Integrate over α to obtain the posterior for β . Marginally, α is multivariate normal; therefore, we obtain

$$\Pi_n(\beta) = (2\Pi)^{-n/2} |\Lambda|^{-1/2} |\psi|^{-1/2} |A|^{-1/2} \exp\left\{-\tfrac{1}{2} \left(y - X\beta\right)^\top (Z\psi Z^\top + \Lambda)^{-1} (y - X\beta)\right\}.$$

It can be verified that $|Z\psi Z^{\top} + \Lambda| = |\Lambda||\psi||A|$ by using the following identity

$$|D^{-1} - CA^{-1}B| = |D||A^{-1}||A - BD^{-1}C|$$

where $Z\psi Z^{\top} + \Lambda = D^{-1} - CA^{-1}B = (\Lambda^{-1} - \Lambda^{-1}ZA^{-1}Z^{\top}\Lambda^{-1})^{-1}$; and, see, e.g., Appendix M equation 31 in Searle et al. (Variance Components). As a result, the posterior for β is multivariate normal with covariance $Z\psi Z^{\top} + \Lambda$.

Let $V := (Z\psi Z^\top + \Lambda)$. In the exponent, add and subtract $X\hat{\beta} = X(X^\top V^{-1}X)^{-1}X^\top V^{-1}y$ (which depends on θ) in the quadratic to obtain

$$\begin{split} \left(y - X\beta\right)^\top V^{-1}(y - X\beta) &= \left(y - X\hat{\beta}\right)^\top V^{-1}(y - X\hat{\beta}) \\ &- 2\left(y - X\hat{\beta}\right)^\top V^{-1}(X\hat{\beta} - X\beta) \\ &+ \left(X\hat{\beta} - X\beta\right)^\top V^{-1}(X\hat{\beta} - X\beta). \end{split}$$

It is straightforward to show the middle term is zero. Thereforen, the joint posterior for (β, θ) may be written

$$\log \Pi_n(\beta,\theta) = \text{const.} - \tfrac{1}{2} \log |V| - \tfrac{1}{2} (y - X \hat{\beta})^\top V^{-1} (y - X \hat{\beta}) - \tfrac{1}{2} (X \hat{\beta} - X \beta) V^{-1} (X \hat{\beta} - X \beta)$$

If we integrate out β (w.r.t. a multivariate normal density), then we obtain the following marginal posterior for the variance parameters:

$$\log \Pi_n(\theta) = \mathrm{const.} - \tfrac{1}{2} \log |V| + \tfrac{1}{2} \log |X^\top V^{-1} X| - \tfrac{1}{2} \left(y - X \hat{\beta}\right)^\top V^{-1} (y - X \hat{\beta}).$$

4.3 Inference for fixed effects

The most common inferential questions concern the regression coefficient vector β . Wald confidence regions for β are defined by the sets

$$C_{\alpha} := \{b: (b - \hat{\beta})(X^{\top}V^{-1}X)^{-1}(b - \hat{\beta}) < \chi^2_{1-\alpha,p}\}$$

where $\chi^2_{1-\alpha,p}$ is the upper α quantile of the Chi-squared distribution with p degrees of freedom. Unfortunately, these regions are only exact when θ is known and can severely undercover for small sample sizes. Software packages typically perform an adjustment, such as the Kenward-Rogers scaling, which is a generalization of Satterthwaite's approximation.

For a point null hypothesis $H_0: \beta = \beta_0$ the Wald test statistic is

$$W = (\beta_0 - \hat{\beta})(X^\top V^{-1}X)^{-1}(\beta_0 - \hat{\beta})$$

and H_0 is rejected if $W > \chi^2_{1-\alpha,p}$. More generally, let D denote an $r \times p$ matrix r < p of full rank. Reject the null hypothesis $H_0: D\beta = b_0$ if $W > \chi^2_{1-\alpha,r}$ where

$$W = (b_0 - D\hat{\beta})(DX^\top V^{-1}XD^\top)^{-1}(b_0 - D\hat{\beta})$$