Assignment 3: Data Exploration

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

- 1. Change "Student Name, Section #" on line 3 (above) with your name and section number.
- 2. Work through the steps, creating code and output that fulfill each instruction.
- 3. Be sure to **answer the questions** in this assignment document.
- 4. When you have completed the assignment, **Knit** the text and code into a single PDF file.
- 5. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai. Add your last name into the file name (e.g., "FirstLast_A03_DataExploration.Rmd") prior to submission.

The completed exercise is due on <>.

```
library(knitr)
opts_chunk$set(tidy.opts=list(width.cutoff=60),tidy=TRUE, echo = T)
```

Set up your R session

1. Check your working directory, load necessary packages (tidyverse), and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets "Neonics" and "Litter", respectively. Be sure to add the stringsAsFactors = TRUE parameter to the function when reading in the CSV files.

```
# Checking working directory
getwd()
```

[1] "/Users/admin/Desktop/ENV872_EDA/Environmental_Data_Analytics_2022/Assignments"

```
# Loading tidyverse
library(tidyverse)
```

```
## -- Attaching packages ------ tidyverse 1.3.1 --

## v ggplot2 3.3.5 v purrr 0.3.4

## v tibble 3.1.6 v dplyr 1.0.7

## v tidyr 1.1.4 v stringr 1.4.0

## v readr 2.1.1 v forcats 0.5.1
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicologoy of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Neonicotinoids are a new class of insecticides chemically related to nicotine. Like nicotine, the neonicotinoids act on certain kinds of receptors in the nerve synapse. They are much more toxic to invertebrates, like insects, than they are to other animals. Neonicotinoids are water soluble which enables easy uptake by plants. New research done on Neonicotinoids points to potential toxicity to bees and other beneficial insects through low level contamination of nectar and pollen with neonicotinoid insecticides used in agriculture. Although these low level exposures do not normally kill bees directly, they may impact some bees' ability to foraging for nectar, learn and remember where flowers are located, and possibly impair their ability to find their way home to the nest or hive. Despite the controlled studies completed to date, the actual impact of neonicotinoid insecticides on honey bees in the field are difficult to measure.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Trees that fall and decay in the forest add nutrients to the forest soil and retain moisture in the forest. Fallen wood greater than 7 cm diameter is referred to as coarse woody debris. The time coarse woody litter takes to decompose is dependent upon moisture and temperature. Whereas, Fine woody material dries quickly and therefore decays slowly. Fine woody debris may act as a tinder that promotes the start and spreading of forest fires. Coarse woody debris adds long-lasting unique habitat structure and resources to both terrestrial and aquatic habitats, by being an important source of energy and nutrients for microorganisms and detritivores, trapping sediments, offering protection against harsh environmental conditions, and serving as refugium from predation to both consumers and inhabitants of decomposing wood. Coarse woody debris increases the diversity of biological communities in aquatic and terrestrial habitats.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON_Litterfall_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: * Litter and fine woody debris sampling is executed at terrestrial NEON sites that contain woody vegetation >2m tall. * Along with most of NEON's plant productivity measurements, litter and fine woody debris sampling occurs only in tower plots. Locations of tower plots are selected randomly within the 90% flux footprint of the primary and secondary airsheds * Ground traps are sampled once per year. Target sampling frequency for elevated traps varies by vegetation present at the site, with frequent sampling (1x every 2weeks) in deciduous forest sites during senescence, and in- frequent year-round sampling (1x every 1-2 months) at evergreen sites.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
# Using the dim function to view the dimension of the
# dataset
dim(Neonics) #The dataset has 4623 rows and 30 columns
```

[1] 4623 30

6. Using the summary function on the "Effect" column, determine the most common effects that are studied. Why might these effects specifically be of interest?

summary(Neonics\$Effect)

##	Accumulation	Avoidance	Behavior	Biochemistry
##	12	102	360	11
##	Cell(s)	Development	Enzyme(s)	Feeding behavior
##	9	136	62	255
##	Genetics	Growth	Histology	Hormone(s)
##	82	38	5	1
##	Immunological	Intoxication	Morphology	Mortality
##	16	12	22	1493
##	Physiology	Population	Reproduction	
##	7	1803	197	

Answer: The most common effects that are studied are Population and Mortality. These effects are commonly studied to monitor the effect of the Neonicotinoids on the insects.

7. Using the summary function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.

summary(Neonics\$Species.Common.Name)

##	Honey Bee	Parasitic Wasp
##	667	285
##	Buff Tailed Bumblebee	Carniolan Honey Bee
##	183	152
##	Bumble Bee	Italian Honeybee
##	140	113

##	Japanese Beetle	Asian Lady Beetle
##	94	76
##	Euonymus Scale	Wireworm
## ##	75	Minute Directo Pur
##	European Dark Bee 66	Minute Pirate Bug 62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
## ##	45 Aphid Family	39 Cabbage Looper
##	Aprilu Family 38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
## ##	Thrin Order	Cround Post la Family
##	Thrip Order 29	Ground Beetle Family 27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
## ##	Ladybird Beetle 23	Mason Bee 22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
## ##	19 Calico Scale	18 Fairwfly Paragitaid
## ##	Carrico Scare	Fairyfly Parasitoid 18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18

Vedalia Beetle	Silkworm	##
18	18	##
Bee Order	Araneoid Spider Order	##
17	17	##
Insect Class	Egg Parasitoid	##
17	17	##
Oystershell Scale Parasitoid	Moth And Butterfly Order	##
17	17	##
Hemlock Wooly Adelgid	Hemlock Woolly Adelgid Lady Beetle	##
16	16	##
Onion Thrip	Mite	##
16	16	##
Corn Earworm	Western Flower Thrips	##
14	15	##
House Fly	Green Peach Aphid	##
14	14	##
Red Scale Parasite	Ox Beetle	##
14	14	##
Armoured Scale Family	Spined Soldier Bug	##
13	14	##
Eulophid Wasp	Diamondback Moth	##
13	13	##
Predatory Bug	Monarch Butterfly	##
13	13	##
Braconid Parasitoid	Yellow Fever Mosquito	##
12	13	##
Eastern Subterranean Termite	Common Thrip	##
12	12	##
Mite Order	Jassid	##
12	12	##
Pond Wolf Spider	Pea Aphid	##
12	12	##
Glasshouse Potato Wasp	Spotless Ladybird Beetle	##
10	11	##
Southern House Mosquito	Lacewing	##
10	10	##
Ant Family	Two Spotted Lady Beetle	##
9	10	##
(Other)	Apple Maggot	##
670	9	##

Answer: The six most commonly studied species in the dataset are Honey Bee, Parasitic Wasp, Buff Tailed Bumblebee, Carniolan Honey Bee, Bumble Bee and Italian Honeybee. These insects which belong to the same order 'Hymenoptera' are at greater risk since new research done on Neonicotinoids points to potential toxicity to bees and other beneficial insects through low level contamination of nectar and pollen with neonicotinoid insecticides used in agriculture.

8. Concentrations are always a numeric value. What is the class of Conc.1..Author. in the dataset, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

```
## [1] "factor"
```

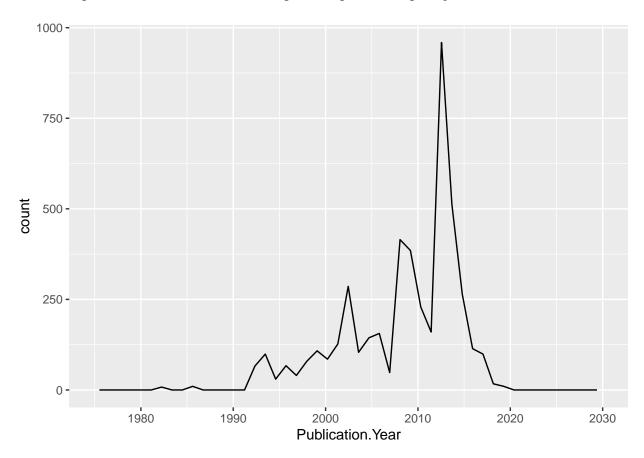
Answer: The dataset is not identified as numeric since there are other characters such as "NR", " \sim " and "/" present in the dataset which are not numeric values.

Explore your data graphically (Neonics)

9. Using geom_freqpoly, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) + geom_freqpoly(aes(x = Publication.Year), bins = 50) +
    scale_x_continuous(limits = c(1975, 2030))
```

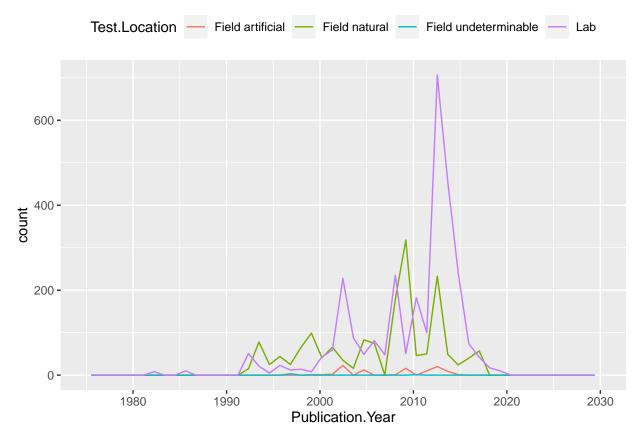
Warning: Removed 3 row(s) containing missing values (geom_path).



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics) + geom_freqpoly(aes(x = Publication.Year, color = Test.Location),
    bins = 50) + scale_x_continuous(limits = c(1975, 2030)) +
    theme(legend.position = "top")
```

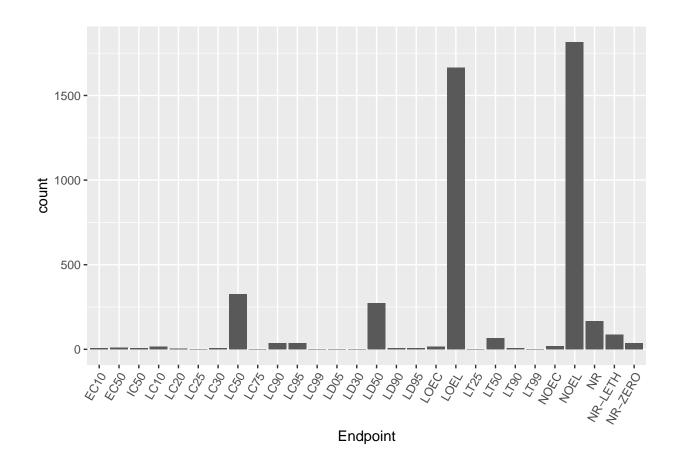
Warning: Removed 12 row(s) containing missing values (geom_path).



Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test locations are Field Natural and Lab. In the earlier years (1990's), Field Natural seems to have been a common test location. Around early 2000's during the boon of technology, labs were used as common test locations and we can observe a sharp peak at around 2013.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.



unique(Veonics\$Endpoint))
---------	--------------------	---

##	[1]	LD50	LC5	50	IC50	L	OEL	NR		NR-ZER	O LCS	95	LC90	NO	EL		
##	[10]	LOEC	ECS	50	NOEC	N	R-LETH	LD90)	LC10	LT5	50	LT90	LT	25		
##	[19]	LC75	LD3	30	EC10	L	C25	LC20)	LD05	LCS	9	LT99	LC	30		
##	[28]	LD95															
##	28 Le	evels:	EC10	EC50	IC50	LC10	LC20	LC25	LC30	LC50	LC75	LC90	LC95	LC99		NR-ZE	RO.

summary(Neonics\$Endpoint)

##	EC10	EC50	IC50	LC10	LC20	LC25	LC30	LC50	LC75	LC90
##	6	11	6	15	5	1	6	327	1	37
##	LC95	LC99	LD05	LD30	LD50	LD90	LD95	LOEC	LOEL	LT25
##	36	2	1	1	274	6	7	17	1664	1
##	LT50	LT90	LT99	NOEC	NOEL	NR	NR-LETH	NR-ZERO		
##	65	7	2	19	1816	167	86	37		

Answer: The two most common endpoints are LOEL and NOEL. LOEL - Lowest observable effect level, NOEL - No observable effect level.

Explore your data (Litter)

12. Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the unique function, determine which dates litter was sampled in August 2018.

```
# Determining the format
class(Litter$collectDate)
## [1] "factor"
# Changing format from factor to date
DateFormat <- as.Date(Litter$collectDate, format = "%Y-%m-%d")
DateFormat
     [1] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
##
     [6] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
    [11] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
##
    [16] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
    [21] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [26] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [31] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [36] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [41] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [46] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [51] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [56] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [61] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [66] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
##
   [71] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
    [76] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [81] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [86] "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02" "2018-08-02"
##
    [91] "2018-08-02" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
   [96] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
##
   [101] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
  [106] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [111] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
  [116] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
   [121] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [126] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [131] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [136] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [141] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [146] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [151] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [156] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [161] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [166] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [171] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [176] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [181] "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30" "2018-08-30"
## [186] "2018-08-30" "2018-08-30" "2018-08-30"
# Determining the new format
```

[1] "Date"

class(DateFormat)

```
# Using unique function to determine which dates litter was
# sampled in August 2018
unique(Litter$collectDate)
```

```
## [1] 2018-08-02 2018-08-30
## Levels: 2018-08-02 2018-08-30
```

13. Using the unique function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from unique different from that obtained from summary?

```
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051 ## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057 ## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

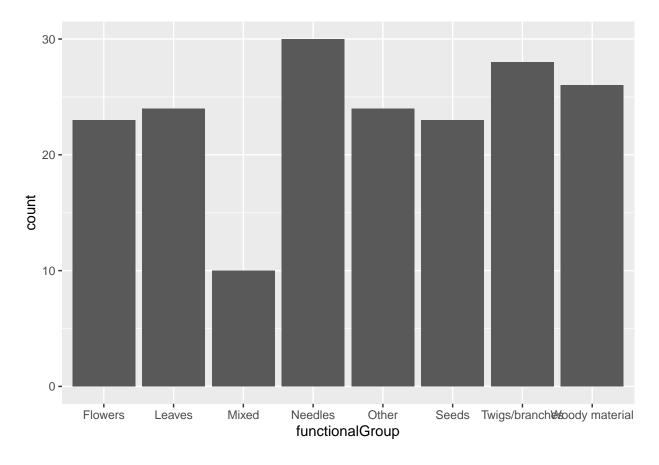
summary(Litter\$plotID)

```
## NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 NIWO_058 NIWO_061 ## 20 19 18 15 14 8 16 17 ## NIWO_062 NIWO_063 NIWO_064 NIWO_067 ## 14 14 16 17
```

Answer: Twelve plots were sampled at Niwot Ridge. While the unique function provies the plot ID and the total number of plots that were sampled at Niwot Ridge, the summary function provides information on the number of each individual plots among the twelve plots.

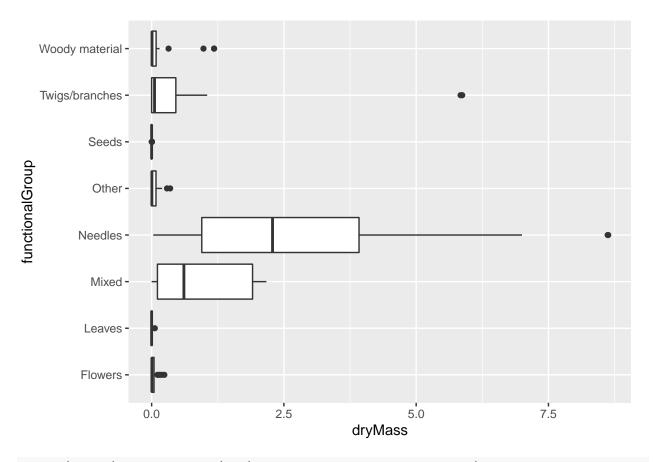
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter, aes(x = functionalGroup)) + geom_bar()
```



15. Using geom_boxplot and geom_violin, create a boxplot and a violin plot of dryMass by functional-Group.

```
ggplot(Litter) + geom_boxplot(aes(x = dryMass, y = functionalGroup,
    group = cut_width(functionalGroup, 1)))
```



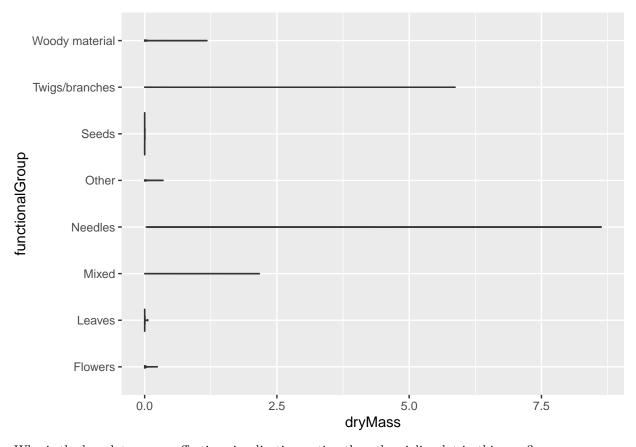
```
ggplot(Litter) + geom_violin(aes(x = dryMass, y = functionalGroup),
    draw_quantiles = c(0.25, 0.5, 0.75))

## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values

## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
## collapsing to unique 'x' values

## Warning in regularize.values(x, y, ties, missing(ties), na.rm = na.rm):
```

collapsing to unique 'x' values



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: The boxplot is a more effective visualization option than the violin plot in this case since we can clearly identify the Interquartile range and the median of our data along with the outliers. Whereas, in the violin plot which displays density distributions, we are not able to view the violins since the data is not distributed evenly.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles seems to have the highest biomass, followed by mixed.