Pipeline QA

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Sequence Processing

```
Loading sequencing data
mrexp filenames <- list(mothur = "../data/mrexp mothur.RDS",</pre>
                        qiime_denovo_chimerafilt = "../data/mrexp_qiime_denovo_chimera_filt.RDS",
                        qiime_denovo_nochimerafilt = "../data/mrexp_qiime_denovo_nochimera.RDS",
                        qiime_openref_chimerafilt = "../data/mrexp_qiime_refclus_chimera_filt.RDS",
                        qiime_openref_nochimerafilt = "../data/mrexp_qiime_refclus_nochimera.RDS")
mrexp obj <- mrexp filenames %>% map(readRDS)
fvarLabels(mrexp_obj$qiime_openref_chimerafilt) <- paste0("taxonomy",1:7)</pre>
## Loading required package: metagenomeSeq
## Loading required package: limma
##
## Attaching package: 'limma'
## The following object is masked from 'package:BiocGenerics':
##
##
       plotMA
## Loading required package: glmnet
## Loading required package: Matrix
##
## Attaching package: 'Matrix'
## The following object is masked from 'package:S4Vectors':
##
##
       expand
  The following object is masked from 'package:tidyr':
##
##
       expand
## Loading required package: foreach
## Attaching package: 'foreach'
## The following objects are masked from 'package:purrr':
##
       accumulate, when
## Loaded glmnet 2.0-5
## Loading required package: RColorBrewer
```

Rename gime samples for consistent set of ids

```
get_new_ids <- function(mr_qiime, sample_sheet){</pre>
      qiime_id_set <- pData(mr_qiime) %>% rownames()
      id_fix_df <- sample_sheet %>%
            filter(seq lab == "JHU", barcode lab == "JHU") %>%
            select(id, pcr_16S_plate, pos) %>%
            mutate(pos = str replace(pos, " ",""),
                    qiime_id = str_c(pcr_16S_plate, pos, sep = "-")) %>%
            filter(qiime_id %in% qiime_id_set) %>%
            group_by(id) %>%
            mutate(id2 = if_else(grepl(x = id, pattern = "B0_M0"),
                                   paste(id,1:n(),sep = "_"),id))
      id_fix_df$id2[match(id_fix_df$qiime_id, qiime_id_set)]
}
id_set <- get_new_ids(mrexp_obj$qiime_denovo_chimerafilt, sample_sheet)</pre>
rownames(pData(mrexp_obj$qiime_denovo_chimerafilt)) <- id_set</pre>
colnames(assayData(mrexp_obj$qiime_denovo_chimerafilt)$counts) <- id_set</pre>
id_set <- get_new_ids(mrexp_obj$qiime_denovo_nochimerafilt, sample_sheet)</pre>
rownames(pData(mrexp_obj$qiime_denovo_nochimerafilt)) <- id_set</pre>
colnames(assayData(mrexp_obj$qiime_denovo_nochimerafilt)$counts) <- id_set</pre>
id_set <- get_new_ids(mrexp_obj$qiime_openref_chimerafilt, sample_sheet)</pre>
rownames(pData(mrexp_obj$qiime_openref_chimerafilt)) <- id_set</pre>
colnames(assayData(mrexp_obj$qiime_openref_chimerafilt)$counts) <- id_set</pre>
id_set <- get_new_ids(mrexp_obj$qiime_openref_nochimerafilt, sample_sheet)</pre>
rownames(pData(mrexp_obj$qiime_openref_nochimerafilt)) <- id_set</pre>
colnames(assayData(mrexp_obj$qiime_openref_nochimerafilt)$counts) <- id_set</pre>
```

Pipeline characteristics

- Section objectives
 - make non-quantitative statements
 - capturing differences in quality across samples
- Characterization of different pipelines
 - number of clusters
 - different taxonomic assignments
- Statements/ Figures showing how datasets behave
- number of assigned vs. non-assigned
- TODO difference in richness
 - need to figure out how I want to normalize/ transform the data prior to calculating diversity values
- number of features found across samples and replicates
- TODO Table pipeline sequence budget
 - number of reads filtered due to low quality
 - number of reads merged
 - number of chimeras

Developing Code for characterizing pipeline results

Number of OTUs

mrexp_obj %>% map(nrow)

```
## $mothur
## Features
##
     25739
##
## $qiime_denovo_chimerafilt
## Features
     14326
##
##
## $qiime_denovo_nochimerafilt
## Features
     24617
##
##
## $qiime_openref_chimerafilt
## Features
##
       2832
##
## $qiime_openref_nochimerafilt
## Features
     11381
```