# Lecture 4

Visualizing and Statistics

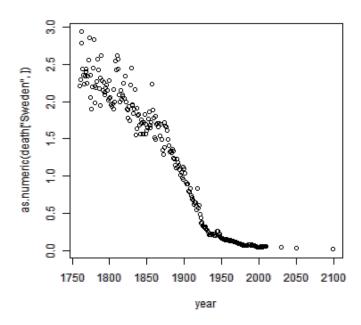
Andrew Jaffe Instructor

## **Today**

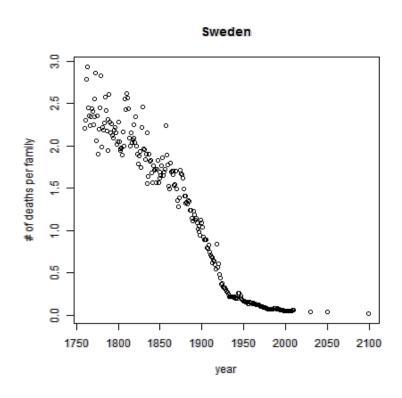
Today we are going to cover more advanced visualization and some useful statistics functions

We covered some basic plots on Tuesday, but we are going to expand the ability to customize these basic graphics first.

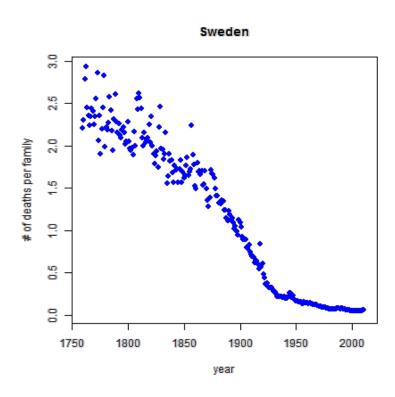
```
> death = read.csv("http://biostat.jhsph.edu/~ajaffe/files/indicatordeadkids35.csv",
+    as.is = T, header = TRUE, row.names = 1)
> year = as.integer(gsub("X", "", names(death)))
> plot(as.numeric(death["Sweden", ]) ~ year)
```



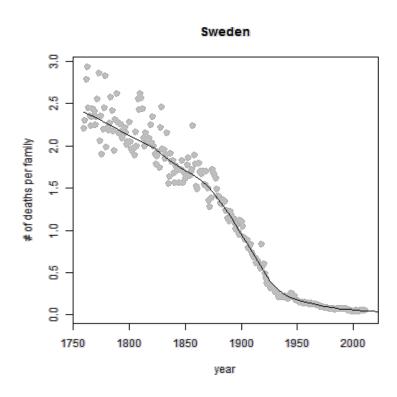
```
> plot(as.numeric(death["Sweden", ]) ~ year, ylab = "# of deaths per family",
+ main = "Sweden")
```



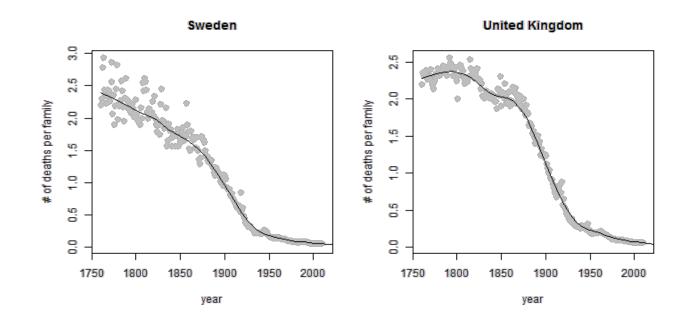
```
> plot(as.numeric(death["Sweden", ]) ~ year, ylab = "# of deaths per family",
+ main = "Sweden", xlim = c(1760, 2012), pch = 19, cex = 1.2, col = "blue")
```



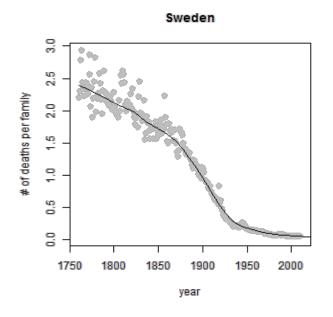
```
> scatter.smooth(as.numeric(death["Sweden", ]) ~ year, span = 0.2, ylab = "# of deaths per family",
+ main = "Sweden", lwd = 3, xlim = c(1760, 2012), pch = 19, cex = 0.9, col = "grey")
```

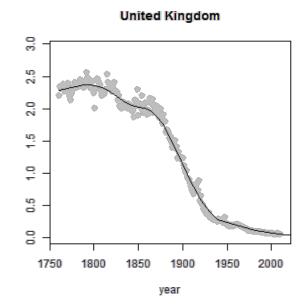


```
> par(mfrow = c(1, 2))
> scatter.smooth(as.numeric(death["Sweden", ]) ~ year, span = 0.2, ylab = "# of deaths per family",
+ main = "Sweden", lwd = 3, xlim = c(1760, 2012), pch = 19, cex = 0.9, col = "grey")
> scatter.smooth(as.numeric(death["United Kingdom", ]) ~ year, span = 0.2, ylab = "# of deaths per famil
+ main = "United Kingdom", lwd = 3, xlim = c(1760, 2012), pch = 19, cex = 0.9,
+ col = "grey")
```



```
> par(mfrow = c(1, 2))
> yl = range(death[c("Sweden", "United Kingdom"), ])
> scatter.smooth(as.numeric(death["Sweden", ]) ~ year, span = 0.2, ylim = yl,
+ ylab = "# of deaths per family", main = "Sweden", lwd = 3, xlim = c(1760,
+ 2012), pch = 19, cex = 0.9, col = "grey")
> scatter.smooth(as.numeric(death["United Kingdom", ]) ~ year, span = 0.2, ylab = "",
+ main = "United Kingdom", lwd = 3, ylim = yl, xlim = c(1760, 2012), pch = 19,
+ cex = 0.9, col = "grey")
```





### Graphical parameters

par() can be used to set or query graphical parameters. Parameters can be set by specifying them as arguments to par in tag = value form, or by passing them as a list of tagged values.

Note that some parameters are passed from plot(,...) calls whereas others need to be explicitly set using par() - like above with par(mfrow = c(nrow,ncol))

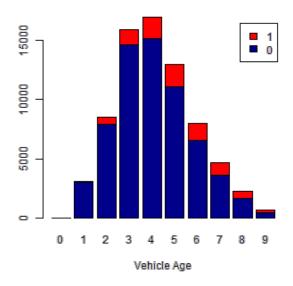
Note that some parameters are both very flexible but also very finicky, especially margins.

#### **Bar Plots**

· Stacked Bar Charts are sometimes wanted to show how

```
> ## Stacked Bar Charts
> cars = read.csv("http://biostat.jhsph.edu/~ajaffe/files/kaggleCarAuction.csv",
+ as.is = T)
> counts <- table(cars$IsBadBuy, cars$VehicleAge)
> barplot(counts, main = "Car Distribution by Age and Bad Buy Status", xlab = "Vehicle Age",
+ col = c("darkblue", "red"), legend = rownames(counts))
```

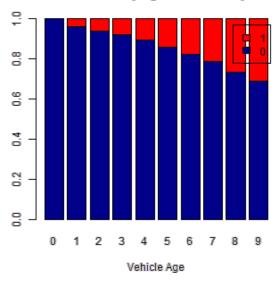
#### Car Distribution by Age and Bad Buy Status



#### **Bar Plots**

```
> ## Use percentages (column percentages)
> barplot(prop.table(counts, 2), main = "Car Distribution by Age and Bad Buy Status",
+ xlab = "Vehicle Age", col = c("darkblue", "red"), legend = rownames(counts))
```

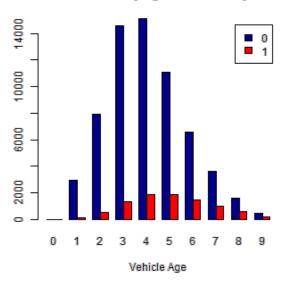
#### Car Distribution by Age and Bad Buy Status



#### **Bar Plots**

```
> # Stacked Bar Plot with Colors and Legend
> barplot(counts, main = "Car Distribution by Age and Bad Buy Status", xlab = "Vehicle Age",
+ col = c("darkblue", "red"), legend = rownames(counts), beside = TRUE)
```

#### Car Distribution by Age and Bad Buy Status



### **Graphics parameters**

Set within most plots in the base 'graphics' package:

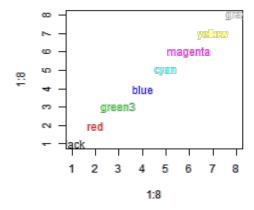
- pch = point shape, http://voteview.com/symbols\_pch.htm
- · cex = size/scale
- · xlab, ylab = labels for x and y axes
- · main = plot title
- · lwd = line density
- · col = color
- · cex.axis, cex.lab, cex.main = scaling/sizing for axes marks, axes labels, and title

R relies on color 'palettes'.

```
> palette()
```

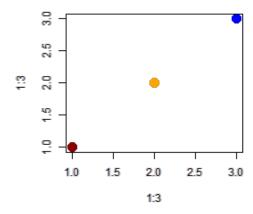
```
[1] "black" "red" "green3" "blue" "cyan" "magenta" "yellow"
[8] "gray"
```

```
> plot(1:8, 1:8, type = "n")
> text(1:8, 1:8, lab = palette(), col = 1:8)
```



The default color palette is pretty bad, so you can try to make your own.

```
> palette(c("darkred", "orange", "blue"))
> plot(1:3, 1:3, col = 1:3, pch = 19, cex = 2)
```



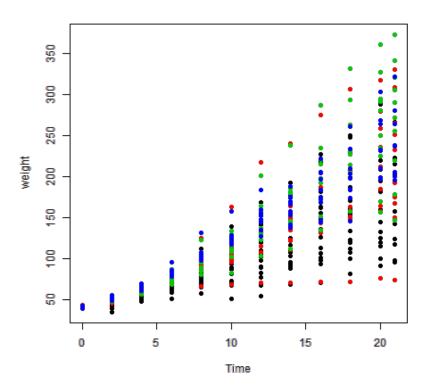
It's actually pretty hard to make a good color palette. Luckily, smart and artistic people have spent a lot more time thinking about this. The result is the 'RColorBrewer' package

RColorBrewer::display.brewer.all() will show you all of the palettes available. You can even print it out and keep it next to your monitor for reference.

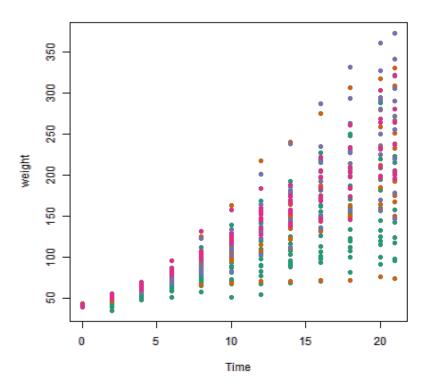
The help file for brewer.pal() gives you an idea how to use the package.

You can also get a "sneak peek" of these palettes at: <a href="www.colorbrewer2.com">www.colorbrewer2.com</a>. You would provide the number of levels or classes of your data, and then the type of data: sequential, diverging, or qualitative. The names of the RColorBrewer palettes are the string after 'pick a color scheme:'

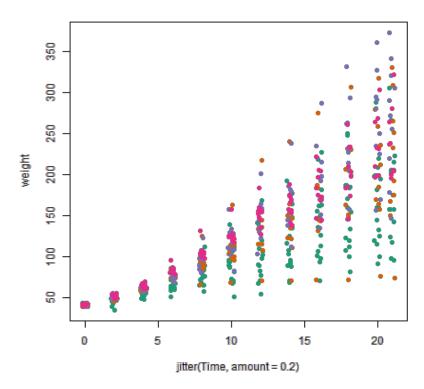
```
> palette("default")
> with(ChickWeight, plot(weight ~ Time, pch = 19, col = Diet))
```



```
> library(RColorBrewer)
> palette(brewer.pal(5, "Dark2"))
> with(ChickWeight, plot(weight ~ Time, pch = 19, col = Diet))
```



```
> library(RColorBrewer)
> palette(brewer.pal(5, "Dark2"))
> with(ChickWeight, plot(weight ~ jitter(Time, amount = 0.2), pch = 19, col = Diet),
+ xlab = "Time")
```



### Adding legends

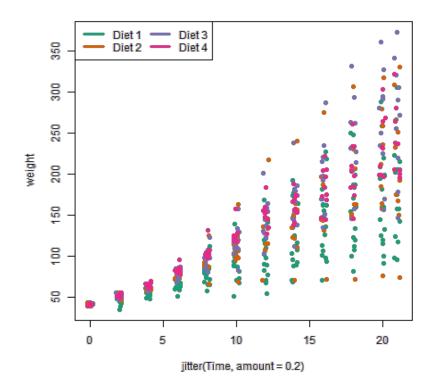
The legend() command adds a legend to your plot. There are tons of arguments to pass it.

x, y=NULL: this just means you can give (x,y) coordinates, or more commonly just give x, as a character string: "top","bottom","topleft","bottomleft","topright","bottomright".

legend: unique character vector, the levels of a factor

pch, lwd: if you want points in the legend, give a pch value. if you want lines, give a lwd value.

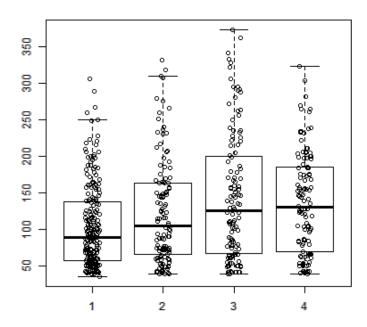
col: give the color for each legend level



### Boxplots, revisited

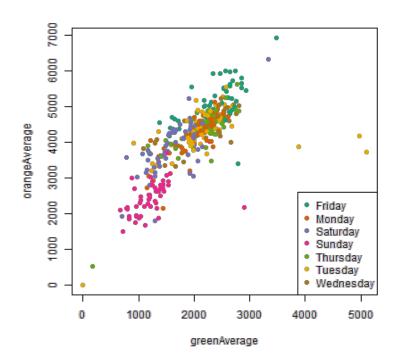
These are one of my favorite plots. They are way more informative than the barchart + antenna...

```
> with(ChickWeight, boxplot(weight ~ Diet, outline = FALSE))
> points(ChickWeight$weight ~ jitter(as.numeric(ChickWeight$Diet), 0.5))
```



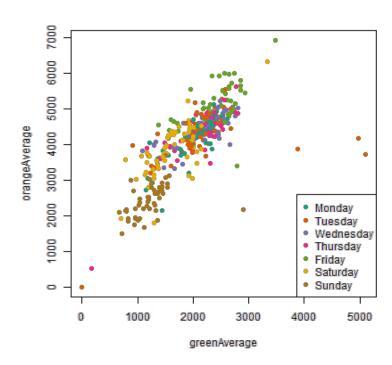
### Coloring by variable

```
> load("../lecture2/charmcirc.rda")
> palette(brewer.pal(7, "Dark2"))
> dd = factor(dat$day)
> with(dat, plot(orangeAverage ~ greenAverage, pch = 19, col = as.numeric(dd)))
> legend("bottomright", levels(dd), col = 1:length(dd), pch = 19)
```



## Coloring by variable

```
> dd = factor(dat$day, levels = c("Monday", "Tuesday", "Wednesday", "Thursday",
+    "Friday", "Saturday", "Sunday"))
> with(dat, plot(orangeAverage ~ greenAverage, pch = 19, col = as.numeric(dd)))
> legend("bottomright", levels(dd), col = 1:length(dd), pch = 19)
```



#### **Devices**

By default, R displays plots in a separate panel. From there, you can export the plot to a variety of image file types, or copy it to the clipboard.

However, sometimes its very nice to save many plots made at one time to one pdf file, say, for flipping through. Or being more precise with the plot size in the saved file.

R has 5 additional graphics devices: bmp(), jpeg(), png(), tiff(), and pdf()

The syntax is very similar for all of them:

```
pdf("filename.pdf", width=8, height=8) # inches
plot() # plot 1
plot() # plot 2
# etc
dev.off()
```

Basically, you are creating a pdf file, and telling R to write any subsequent plots to that file. Once you are done, you turn the device off. Note that failing to turn the device off will create a pdf file that is corrupt, that you cannot open.

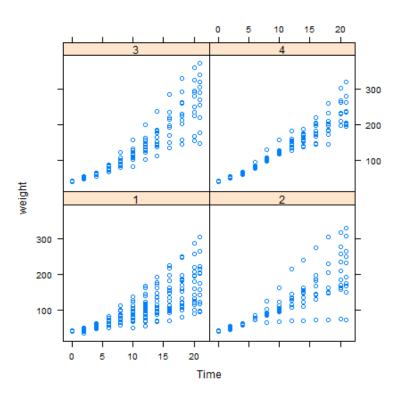
### More powerful graphics

There are two very common packages for making very nice looking graphics.

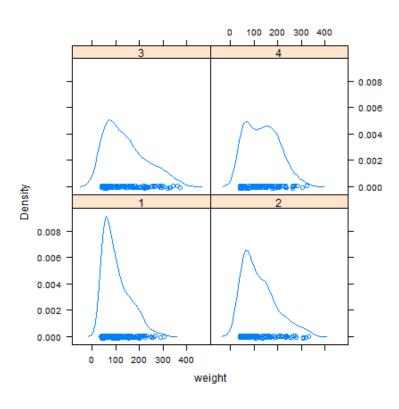
lattice: http://lmdvr.r-forge.r-project.org/figures/figures.html

ggplot2: http://docs.ggplot2.org/current/index.html

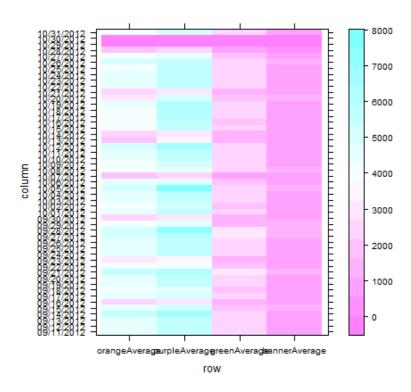
```
> library(lattice)
> xyplot(weight ~ Time | Diet, data = ChickWeight)
```



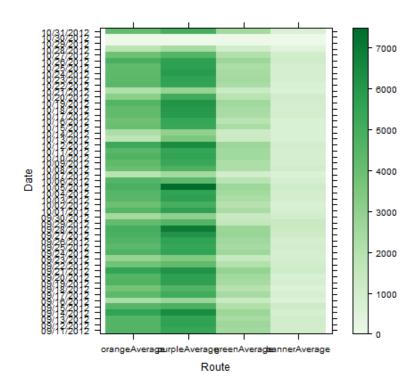
> densityplot(~weight | Diet, data = ChickWeight)



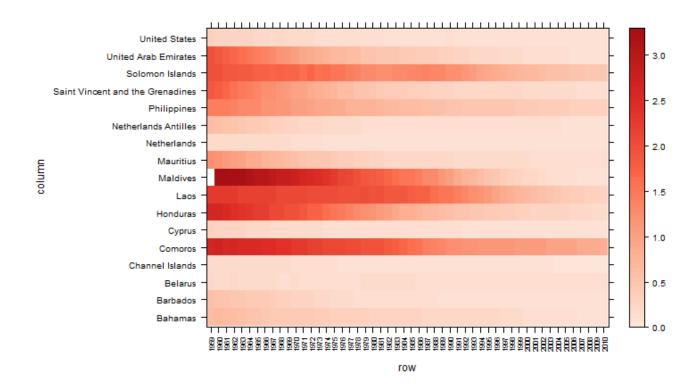
```
> rownames(dat2) = dat2$date
> mat = as.matrix(dat2[975:nrow(dat2), 3:6])
> levelplot(t(mat), aspect = "fill")
```



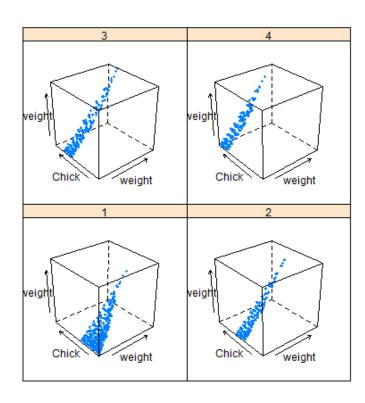
```
> theSeq = seq(0, max(mat), by = 50)
> my.col <- colorRampPalette(brewer.pal(5, "Greens")) (length(theSeq))
> levelplot(t(mat), aspect = "fill", at = theSeq, col.regions = my.col, xlab = "Route",
+ ylab = "Date")
```



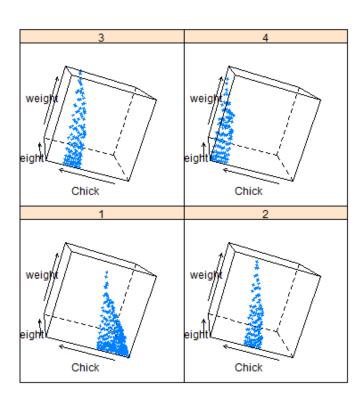
```
> tmp = death[grep("s$", rownames(death)), 200:251]
> yr = gsub("X", "", names(tmp))
> theSeq = seq(0, max(tmp, na.rm = TRUE), by = 0.05)
> my.col <- colorRampPalette(brewer.pal(5, "Reds"))(length(theSeq))
> levelplot(t(tmp), aspect = "fill", at = theSeq, col.regions = my.col, scales = list(x = list(label = y rot = 90, cex = 0.7)))
```



> cloud(weight ~ weight \* Chick | Diet, data = ChickWeight)



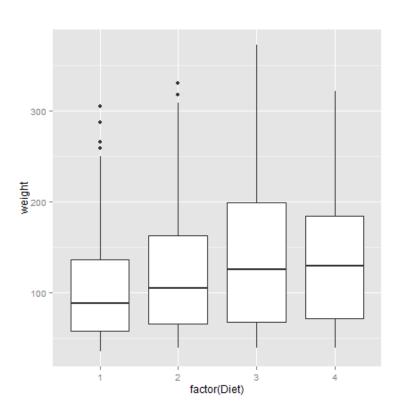
```
> cloud(weight ~ weight * Chick | Diet, data = ChickWeight, screen = list(z = 40, + x = -70, y = 60))
```



# ggplot2

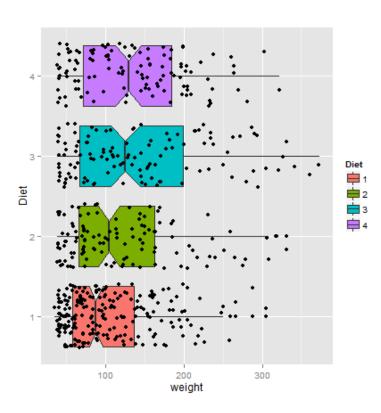
```
> library (ggplot2)
```

> qplot(factor(Diet), weight, data = ChickWeight, geom = "boxplot")



## ggplot2

```
> p = ggplot(ChickWeight, aes(Diet, weight))
> p + geom_boxplot(notch = TRUE, aes(fill = Diet)) + geom_jitter() + coord_flip()
```



### **Statistics**

Now we are going to cover how to perform a variety of basic statistical tests in R.

- · Correlation
- · T-tests
- · Proportion tests
- · Chi-squared
- · Fisher's Exact Test
- · Linear Regression

Note: We will be glossing over the statistical theory and "formulas" for these tests. There are plenty of resources online for learning more about these tests, as well as dedicated Biostatistics series at the School of Public Health

cor() performs correlation in R

```
cor(x, y = NULL, use = "everything",
    method = c("pearson", "kendall", "spearman"))

> cor(dat2$orangeAverage, dat2$purpleAverage)

[1] NA

> cor(dat2$orangeAverage, dat2$purpleAverage, use = "complete.obs")

[1] 0.9208
```

You can also get the correlation between matrix columns

```
> signif(cor(dat2[, grep("Average", names(dat2))], use = "complete.obs"), 3)
```

	orangeAverage	purpleAverage	greenAverage	bannerAverage
orangeAverage	1.000	0.889	0.837	0.441
purpleAverage	0.889	1.000	0.843	0.441
greenAverage	0.837	0.843	1.000	0.411
bannerAverage	0.441	0.441	0.411	1.000

Or between columns of two matrices, column by column.

```
> signif(cor(dat2[, 3:4], dat2[, 5:6], use = "complete.obs"), 3)
```

```
greenAverage bannerAverage
orangeAverage 0.837 0.441
purpleAverage 0.843 0.441
```

You can also use cor.test() to test for whether correlation is significant (ie non-zero). Note that linear regression is probably your better bet.

```
> ct = cor.test(dat2$orangeAverage, dat2$purpleAverage, use = "complete.obs")
> ct
```

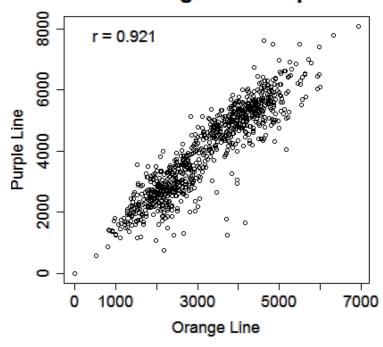
```
Pearson's product-moment correlation

data: dat2$orangeAverage and dat2$purpleAverage
t = 69.65, df = 871, p-value < 2.2e-16
alternative hypothesis: true correlation is not equal to 0
95 percent confidence interval:
0.9100 0.9303
sample estimates:
cor
0.9208
```

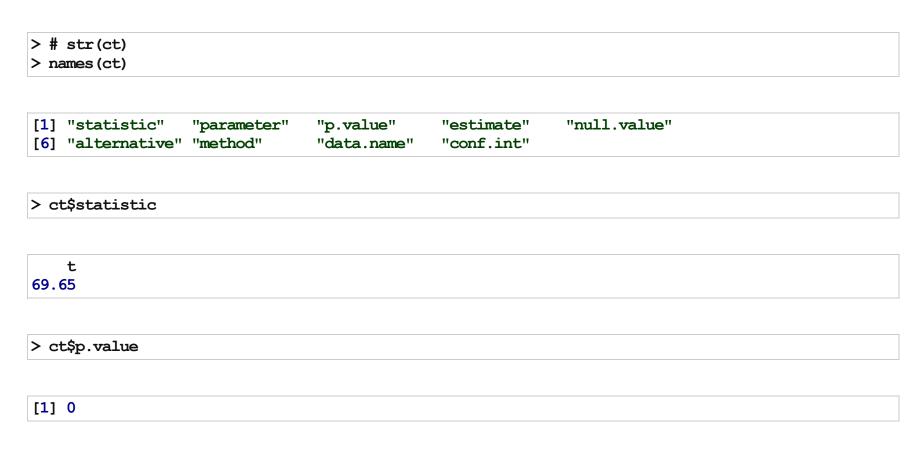
Note that you can add the correlation to a plot, via the legend() function.

```
> plot(dat2$orangeAverage, dat2$purpleAverage, xlab = "Orange Line", ylab = "Purple Line",
+ main = "Average Ridership", cex.axis = 1.5, cex.lab = 1.5, cex.main = 2)
> legend("topleft", paste("r =", signif(ct$estimate, 3)), bty = "n", cex = 1.5)
```

#### **Average Ridership**



For many of these testing result objects, you can extract specific slots/results as numbers, as the 'ct' object is just a list.



#### **T-tests**

The T-test is performed using the t.test() function, which essentially tests for the difference in means of a variable between two groups.

```
> tt = t.test(dat2$orangeAverage, dat2$purpleAverage)
> tt
```

```
Welch Two Sample t-test

data: dat2$orangeAverage and dat2$purpleAverage

t = -16.22, df = 1745, p-value < 2.2e-16

alternative hypothesis: true difference in means is not equal to 0

95 percent confidence interval:
-1141.5 -895.2

sample estimates:
mean of x mean of y
2994 4013
```

```
> names(tt)
```

```
[1] "statistic" "parameter" "p.value" "conf.int" "estimate"
[6] "null.value" "alternative" "method" "data.name"
```

## **T-tests**

You can also use the 'formula' notation.

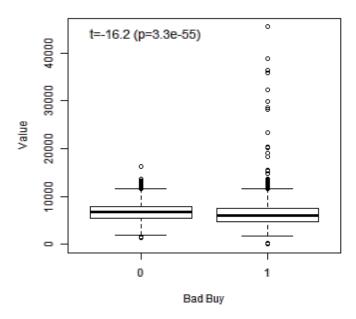
```
> tt2 = t.test(VehBCost ~ IsBadBuy, data = cars)
> tt2$estimate
```

```
mean in group 0 mean in group 1
6797 6259
```

## **T-tests**

You can add the t-statistic and p-value to a boxplot.

```
> boxplot(VehBCost ~ IsBadBuy, data = cars, xlab = "Bad Buy", ylab = "Value")
> leg = paste("t=", signif(tt$statistic, 3), " (p=", signif(tt$p.value, 3), ")",
+ sep = "")
> legend("topleft", leg, cex = 1.2, bty = "n")
```



## **Proportion tests**

prop.test() can be used for testing the null that the proportions (probabilities of success) in several groups are the same, or that they equal certain given values.

```
> prop.test(x = 15, n = 32)
```

```
1-sample proportions test with continuity correction

data: 15 out of 32, null probability 0.5

X-squared = 0.0312, df = 1, p-value = 0.8597

alternative hypothesis: true p is not equal to 0.5

95 percent confidence interval:
    0.2951 0.6497

sample estimates:
    p

0.4688
```

chisq.test() performs chi-squared contingency table tests and goodness-of-fit tests.

```
> tab = table(cars$IsBadBuy, cars$IsOnlineSale)
> tab
```

```
0 1
0 62375 1632
1 8763 213
```

```
> cq = chisq.test(tab)
> cq
   Pearson's Chi-squared test with Yates' continuity correction
data: tab
X-squared = 0.9274, df = 1, p-value = 0.3356
> names (cq)
[1] "statistic" "parameter" "p.value"
                                       "method"
                                                   "data.name" "observed"
[7] "expected" "residuals" "stdres"
> cq$p.value
[1] 0.3356
```

> prop.test(tab)

Note that does the same test as prop.test, for a 2x2 table.

```
> chisq.test(tab)

Pearson's Chi-squared test with Yates' continuity correction

data: tab
X-squared = 0.9274, df = 1, p-value = 0.3356
```

```
2-sample test for equality of proportions with continuity correction

data: tab

X-squared = 0.9274, df = 1, p-value = 0.3356
alternative hypothesis: two.sided

95 percent confidence interval:
-0.005208  0.001674

sample estimates:
prop 1 prop 2
0.9745  0.9763
```

Note that does the same test as prop.test, for a 2x2 table.

chisq.test(tab) prop.test(tab)

Now we will briefly cover linear regression. I will use a little notation here so some of the commands are easier to put in the proper context.

```
y_i = \alpha + \beta * x_i + \epsilon_i
```

#### where:

- · y\_i is the outcome for person i
- · \alpha is the intercept
- · \beta is the slope
- · x\_i is the predictor for person i
- · \epsilon\_i is the residual variation for person i

The 'R' version of the regression model is:

y ~ x

#### where:

- · y is your outcome
- · x is/are your predictor(s)

```
> fit = lm(VehOdo ~ VehicleAge, data = cars)
> fit
```

```
Call:
lm(formula = VehOdo ~ VehicleAge, data = cars)
Coefficients:
(Intercept) VehicleAge
60127 2723
```

'(Intercept)' is \alpha

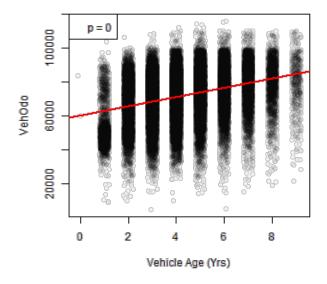
'VehicleAge' is \beta

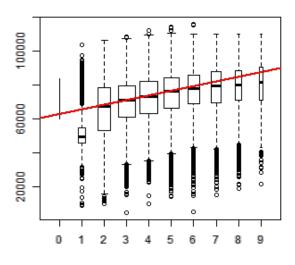
```
> summary(fit)
```

```
Call:
lm(formula = VehOdo ~ VehicleAge, data = cars)
Residuals:
  Min
          10 Median
                       3Q
                             Max
-71097 -9500 1383 10323 41037
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 60127.2
                        134.8 446.0 <2e-16 ***
VehicleAge 2722.9 29.9 91.2 <2e-16 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 13800 on 72981 degrees of freedom
Multiple R-squared: 0.102, Adjusted R-squared: 0.102
F-statistic: 8.31e+03 on 1 and 72981 DF, p-value: <2e-16
```

```
> summary(fit)$coef
```

```
Estimate Std. Error t value Pr(>|t|)
(Intercept) 60127 134.80 446.04 0
VehicleAge 2723 29.86 91.18 0
```





Note that you can have more than 1 predictor in regression models.

The interpretation for each slope is change in the predictor corresponding to a one-unit change in the outcome, holding all other predictors constant.

```
> fit2 = lm(VehOdo ~ VehicleAge + WarrantyCost, data = cars)
> summary(fit2)
```

```
Call:
lm(formula = VehOdo ~ VehicleAge + WarrantyCost, data = cars)
Residuals:
  Min
          10 Median
                       3Q
                             Max
-67895 -8673
                940 9305 45765
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
                                 359.1
(Intercept) 5.24e+04
                      1.46e+02
                                        <2e-16 ***
VehicleAge 1.94e+03 2.89e+01 67.4 <2e-16 ***
WarrantyCost 8.58e+00 8.25e-02 104.0 <2e-16 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 12900 on 72980 degrees of freedom
Multiple R-squared: 0.218, Adjusted R-squared: 0.218
F-statistic: 1.02e+04 on 2 and 72980 DF, p-value: <2e-16
```

Factors get special treatment in regression models - lowest level of the factor is the comparison group, and all other factors are relative to its values.

```
> fit3 = lm(VehOdo ~ factor(TopThreeAmericanName), data = cars)
> summary(fit3)
```

```
Call:
lm(formula = VehOdo ~ factor(TopThreeAmericanName), data = cars)
Residuals:
  Min
          10 Median
                        3Q
                              Max
-71947 -9634 1532 10472 45936
Coefficients:
                                 Estimate Std. Error t value Pr(>|t|)
(Intercept)
                                    68249
                                                  93 733.98 < 2e-16 ***
                                                 158 53.83 < 2e-16 ***
factor (TopThreeAmericanName) FORD
                                     8524
factor(TopThreeAmericanName)GM
                                     4952
                                                      38.39 < 2e-16 ***
                                                 129
factor (TopThreeAmericanName) NULL
                                    -2005
                                                6362 -0.32 0.75267
                                                 160 3.66 0.00026 ***
factor(TopThreeAmericanName)OTHER
                                      585
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 14200 on 72978 degrees of freedom
Multiple R-squared: 0.0482, Adjusted R-squared: 0.0482
F-statistic: 924 on 4 and 72978 DF, p-value: <2e-16
```

## Lab

Load the property taxes dataset from the website: http://biostat.jhsph.edu/~ajaffe/files/propertyTaxes.rda

- 1. How many houses were assessed?
- 2. Make boxplots using a) default and b) ggplot2 graphics showing cityTax by whether the property is a principal residence or not.
- 3. Subset the data to only retain those houses that are principal residences. How many such houses are there?
- 4. Describe the distribution of property taxes on these residences.
- 5. Convert the 'lotSize' variable to a numeric square feet variable. Assume hyphens represent decimal places within measurements. Also, 1 acre = 43560 square feet
- 6. Plot this numeric lotSize versus cityTax on principal residences. What is the correlation between these two variables? Fit a linear regression model with cityTax as the outcome what is the coefficient for lotSize, as well as its t-statistic and p-value?