

OceanBioME.jl: A flexible environment for modelling the coupled interactions between ocean biogeochemistry and physics

Jago Strong-Wright^{1,2¶}, Si Chen^{1,2*}, Navid C Constantinou^{3,4*}, Simone Silvestri^{5*}, Gregory LeClaire Wagner^{5*}, and John R Taylor^{1,2}

1 Department of Applied Mathematics and Theoretical Physics, University of Cambridge, Cambridge, United Kingdom 2 Centre for Climate Repair, Cambridge, United Kingdom 3 Australian National University, Australia 4 Australian Research Council Centre for Climate Extremes 5 Massachusetts Institute of Technology, USA ¶ Corresponding author * These authors contributed equally.

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Statement of Need

To date, about 25% of anthropogenic carbon emissions have been taken up by the ocean (Friedlingstein et al., 2022). This occurs through complex interactions between physics, chemistry, and biology, much of which is poorly understood. Due to the vast size of the ocean and the sparsity of data; modelling and data assimilation play a vital role in quantifying the ocean carbon cycle. Traditionally ocean biogeochemical (BGC) modelling involves large and inflexible code bases written in high-performance but low-level languages. Using and modifying these models usually requires knowledge of the details of the numerical implementation (e.g. the grid layout and time-stepping schemes). Most traditional BGC models also require CPU-based supercomputers to run quickly, which presents a barrier to experimentation and innovation.

One area where novel ideas must be explored with BGC codes is assessing ocean carbon dioxide removal (OCDR) strategies. Quantifying the effectiveness and identifying the impacts of OCDR is challenging due to the aforementioned complexity of the ocean BGC system. Moreover, field trials of OCDR interventions are generally small-scale and targeted, while the intervention required to have a climate-scale impact is regional or global. This necessitates adaptable, easy-to-use, and verifiable BGC modelling tools which can be used to assess OCDR strategies at the fast pace with which they are being developed (National Academies of Sciences & Medicine, 2022). We have built OceanBioME.jl to meet these challenges by creating a tool that provides a modular interface to the different components, within the ocean modelling framework provided by Oceananigans.jl. Previously implementing biogeochemical models in Oceananigans.jl required the user to add forcing terms and boundary conditions to generic tracers (e.g. Simoes-Sousa et al., 2022). OceanBioME.jl provides a suite of biogeochemical models ranging from simple idealized to full-complexity models and models for boundary fluxes (e.g. air-sea gas exchange). Oceananigans.jl and OceanBioME.jl are built from the ground-up to exploit the power of graphical processor units (GPUs), while also retaining the ability to run on CPUs. The flexibility of the Oceananigans.jl framework allows OceanBioME.jl to be applied across a wide range of scales and use cases, including small-scale large-eddy simulations and regional and global models.

Summary

OceanBioME.jl is a flexible modelling environment written in Julia (Bezanson et al., 2017) for simulating the coupled interactions between ocean biogeochemistry, carbonate chemistry,

and physics. OceanBioME.jl can be used as a stand-alone box model, or integrated into Oceananigans.jl (Ramadhan et al., 2020) for coupled physical-biogeochemical simulations in one, two, or three dimensions. As a result, OceanBioME.jl and Oceananigans.jl can be used to simulate the biogeochemical response across an enormous range of scales: from surface boundary layer turbulence at the sub-meter scale to eddying global ocean simulations at the planetary scale, and on computational systems ranging from laptops to supercomputers. An example of a problem involving small-scale flow features is showcased in Figure 1, which shows a simulation of a sub-mesoscale eddy in a $1\text{ km} \times 1\text{ km}$ horizontal domain with an intermediate complexity biogeochemical model and a kelp growth model solved along the trajectories of drifting buoys (a list of examples shown in this paper and links to source code are given at the end of the paper). OceanBioME.jl leverages Julia's multiple dispatch and effective inline capabilities to fuse its computations directly into existing Oceananigans.jl kernels, thus maintaining Oceananigans.jl's bespoke performance, memory- and cost-efficiency on GPUs in OceanBioME.jl-augmented simulations.

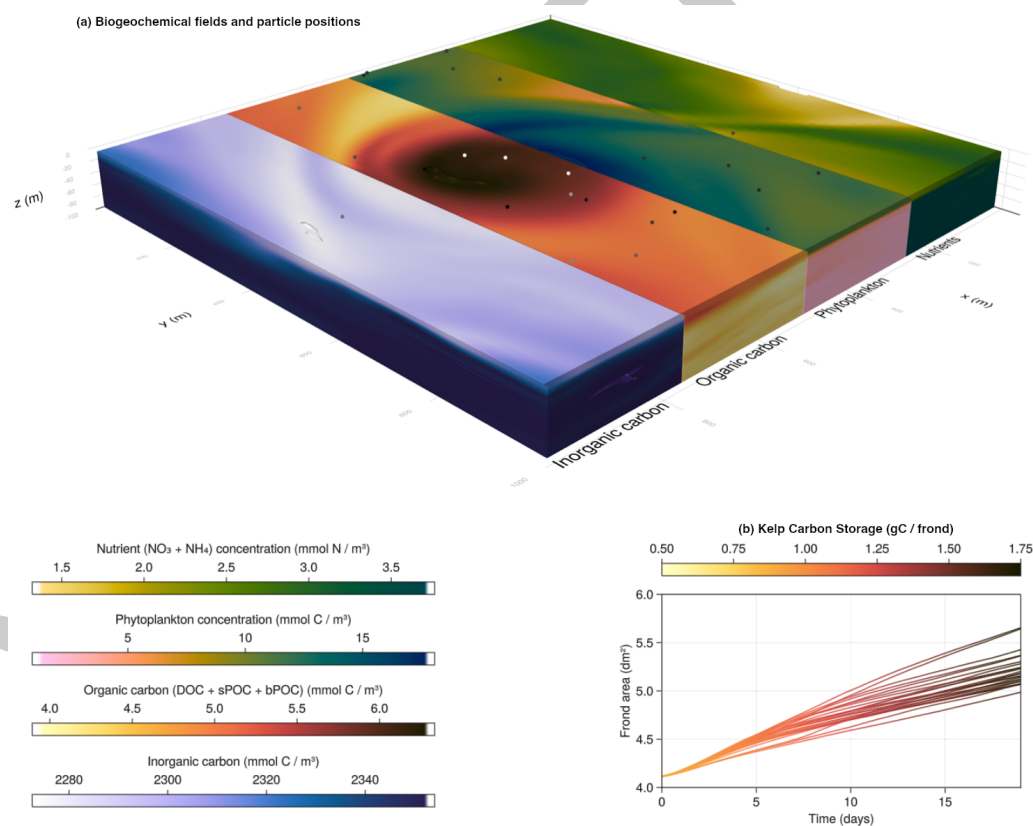


Figure 1: In this simulation of baroclinic instability in the Eady problem, a background buoyancy gradient and corresponding thermal wind generates a sub-mesoscale eddy, roughly following the setup of Taylor (2016). To this physical setup, we added a medium complexity (9 tracers) biogeochemical model, some components of which are shown above. On top of this, we added particles modelling the growth of sugar kelp, which are free-floating and advected by the flow, and carbon dioxide exchange from the air. Thanks to Julia's speed and efficiency the above model ($1\text{ km} \times 1\text{ km} \times 100\text{ m}$ with $64 \times 64 \times 16$ grid points) took about 30 minutes of computing time to simulate 10 days of evolution on an Nvidia P100 GPU. Panel (a) shows the domain with the colour representing the concentration of various biogeochemical tracer fields: inorganic carbon, organic carbon (dissolved and particulate), phytoplankton, and nutrients. The increase in organic carbon concentration in the centre of the eddy can be seen, as well as carbon being subducted (most visible in the xz face in the organic carbon). Points on the surface represent the kelp particle positions, with the colour representing the range of frond size. Panel (b) shows the carbon stored in each kelp frond, highlighting the variability depending on the nutrition availability in each particle's location history. Figure made with Makie.jl (Danisch & Krumbiegel, 2021).

55 OceanBioME.jl is built with a highly modular design that allows user control and customization.
56 There are three distinct module types implemented in OceanBioME.jl:

- 57 ▪ First, we provide tracer-based ecosystem modules in AdvectedPopulations as a set
58 of coupled ordinary differential equations (ODEs) which evolve the concentration of
59 the tracer. These equations can be solved by OceanBioME.jl as box models. This is
60 useful for both testing and running simple educational models and for understanding
61 the behaviour of biogeochemical models without the effects of the physical dynamics.
62 The same tracer equations can then be seamlessly integrated into Oceananigans.jl to
63 further include the effects of advection and diffusion.
- 64 ▪ The second module type is Individual “biologically active” particles. These consist of
65 individual-based models solved along particle paths, which can be coupled with the tracer-
66 based modules and physics from Oceananigans.jl. The biologically active particles can
67 be advected by the currents, and/or they can move according to prescribed dynamics.
68 For example, migrating zooplankton or fish can be modelled with biologically active
69 particles and OceanBioME.jl allows these to interact with tracer-based components such
70 as phytoplankton or oxygen.
- 71 ▪ The AdvectedPopulations are supported by Boundaries modules which are easy to
72 apply and provide information at the top and bottom of the ocean. We have implemented
73 comprehensive air-sea flux models (e.g. [Wanninkhof, 1992](#)) within the GasExchange
74 submodule to calculate carbon dioxide and oxygen flux at the sea surface, and sediment
75 models (e.g. [Soetaert et al., 2000](#)) which calculate fluxes of carbon and oxygen at the
76 seafloor.

77 We currently provide a simple Nutrient-Phytoplankton-Zooplankton-Detritus (NPZD) model
78 ([Kuhn et al., 2015](#)), and an intermediate complexity model, LOBSTER ([Lévy et al., 2005](#))
79 and we have created a straightforward “plug and play” framework to add additional tracers
80 such as carbonate and oxygen chemistry systems and additional forcing. A key feature of
81 this package is the ability to easily modify the model equations or add different formulations,
82 allowing exploration and experimentation. If a user wanted to implement a different model they
83 could use the existing ones as a template and modify only a few lines of code where the ODEs
84 are defined as functions. The user can then insert their model into our abstracted framework
85 to couple the model with the other components such as light attenuation and sediments.
86 We provide a detailed tutorial describing how to do this, which also serves as a description
87 of how our models are created. This framework is made possible by our contributions to
88 Oceananigans.jl, adding a streamlined user interface to swap biogeochemical models with no
89 modification to other model configurations. Our interface also facilitates rapid prototyping, as
90 models can be implemented and swapped easily by just extending a few key functions. This
91 flexibility and ease-of-use is unmatched in existing biogeochemical models.

92 Oceananigans.jl includes several dynamical cores which include a fully non-hydrostatic model
93 capable of large-eddy simulations (LES) and a free-surface hydrostatic model. This allows
94 OceanBioME.jl to be used to simulate biogeochemistry across a vast range of scales using the
95 same BGC model formulation. As an example, [Figure 2](#) shows the annual average surface
96 phytoplankton concentration from a near-global model NPZD model.

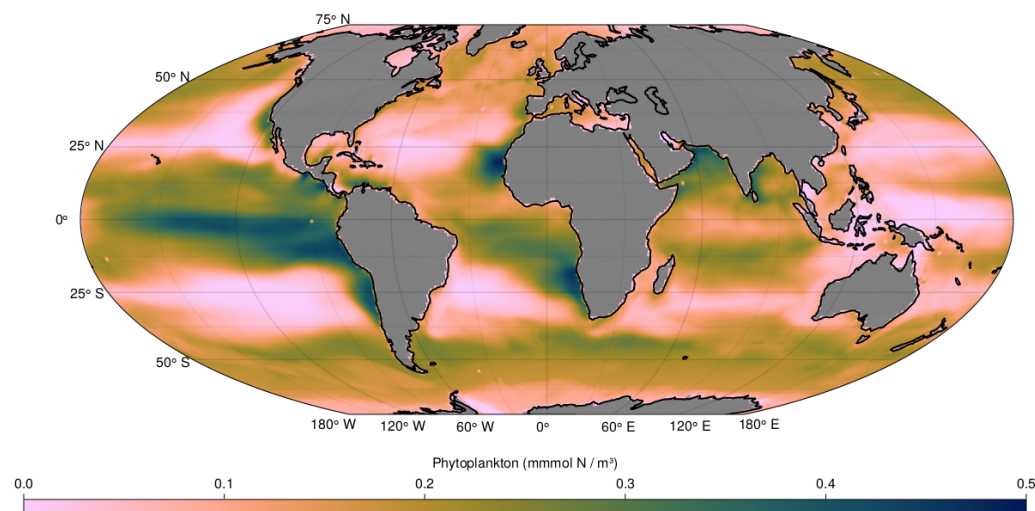


Figure 2: The annual average surface phytoplankton concentration from a near-global NPZD model run. Although the model is uncalibrated and is missing some important processes (e.g. river input), it reproduces the large-scale patterns reasonably well. This simulation used 1° horizontal resolution and 48 (irregularly spaced) vertical points. It took around 45 minutes per year to run on an Nvidia A100 GPU when integrating the physics, or less than 5 minutes per year when using pre-calculated velocity fields. Figure made with Makie.jl (Danisch & Krumbiegel, 2021).

97 The biologically active particles built into OceanBioME.jl are particularly useful for OCCR
 98 applications. Accurate carbon accounting is essential for assessing the effectiveness of OCCR
 99 strategies. Biologically active particles can be used to track carbon from a particular source while
 100 accounting for interactions with its surroundings. Biologically active particles can also be used
 101 to model OCCR deployment strategies including seaweed cultivation, alkalinity enhancement,
 102 and marine biomass regeneration. OceanBioME.jl currently includes an extended version of
 103 the sugar kelp model presented by Broch & Slagstad (2012) as an example of the utility and
 104 implementation of these features. Figure 3 shows a simple column model with an OCCR
 105 intervention (macroalgae growth) added after a warm-up period, which increases the carbon
 106 export of the system.

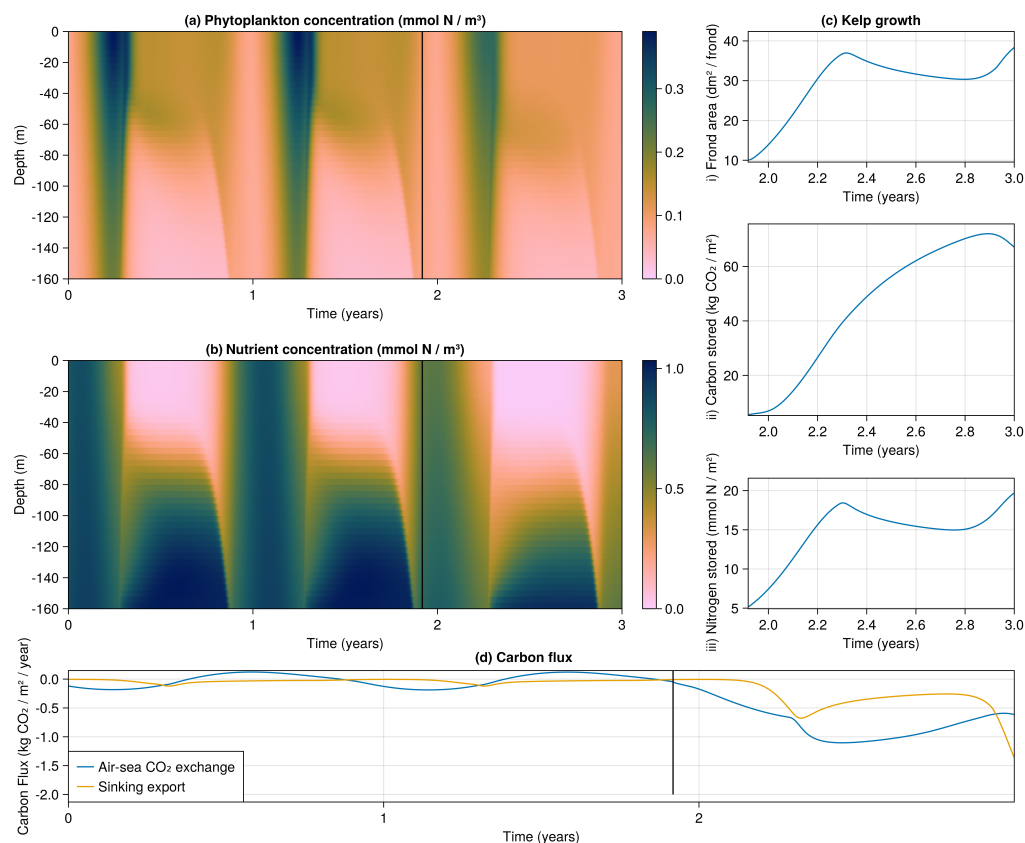


Figure 3: Results of a 1D model, forced by idealised light and mixing, which qualitatively reproduces the biogeochemical cycles in the North Atlantic. Kelp (500 frond / m² in the top 50 m of water) is added in December of the 2nd year (black vertical line) which causes an increase in air-sea carbon dioxide exchange and sinking export, as shown in panel (d). Panel (a) shows the phytoplankton growth cycle which also changes in response to the altered nutrient dynamics visible in panel (b). Panel (c) shows the kelp growth with the front size in (i), the carbon storage in (ii), and the nitrogen storage in (iii). Figure made with Makie.jl (Danisch & Krumbiegel, 2021).

107 The implementation of OceanBioME.jl models allows for seamless integration with data
108 assimilation packages, such as EnsembleKalmanProcesses.jl (Dunbar et al., 2022). This
109 enables rapid calibration of model parameters and provides a powerful utility for integrating
110 observations and models, with the potential to improve model skill and identify key sources of
111 uncertainty.

112 A key metric for the validity of biogeochemical systems is the conservation of elements such as
113 carbon and nitrogen in the system. We therefore continuously test the implemented models in
114 a variety of simple scenarios (i.e. isolated, with/without air-sea flux, with/without sediment) to
115 ensure that conservation conditions are met, and we will continue to add tests for any models.
116 Additionally, we check OceanBioME.jl utilities through standard tests such as comparison
117 to analytical solutions for light attenuation, and conservation of tracers for active particle
118 exudation and sinking. We plan to expand the features of OceanBioME.jl in the future and
119 very much welcome user contributions.

120 Finally, this software is currently facilitating multiple research projects into ocean CDR which
121 would have been significantly harder with other models. For example, Chen et al. (in prep.)
122 uses the active particle coupling provided to investigate the effects of location and planting
123 density of kelp in the open ocean on their carbon drawdown effect, as in the example above.
124 Strong-Wright et al. (in prep.) uses the coupling of both the biogeochemistry and easy interface

to couple the physics to study flow interactions with a fully resolved giant kelp forest model including the effects on nutrient transport and distribution.

Examples

Example	OceanBioME features utilised	Code location
Sub-mesoscale eddy (Figure 1)	LOBSTER biogeochemical model ¹ with carbonate model active, CO ₂ exchange with the air ² , Light attenuation ³ , mass conserving negativity protection ⁴	examples/eady.jl with resolution increased to 64x64x16
Near-global (Figure 2) proof of concept	Light attenuation ³ , NPZD model ⁵	https://github.com/OceanBioME/GlobalOceanBioME.jl/releases/tag/v0.0.1
Idealised 1D model with kelp individuals (Figure 3)	LOBSTER biogeochemical model ¹ with carbonate model and variable Redfield ratio for organic components active, CO ₂ exchange with the air ² , light attenuation ³ , mass conserving negativity protection ⁴ , and Saccharina Latissima (sugar kelp) model ⁶	paper/figures/column.jl, similar to examples/column.jl and examples/kelp.jl

¹ LOBSTER

² GasExchange

³ TwoBandPhotosyntheticallyActiveRadiation

⁴ ScaleNegativeTracers

⁵ NutrientPhytoplanktonZooplanktonDetritus

⁶ SLatissima

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