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The title of my thesis  
which should be split on  
several lines if it is too long

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[B]ecause we tend to reward others when they do well and punish them when they do badly, and because there is regression to the mean, it is part of the human condition that we are statistically punished for rewarding others and rewarded for punishing them — [Kahneman \(2012\)](#).



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## Preface

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First words and acknowledgements. And we add a lot of text to make sure that it spans more than one line, as otherwise it may not show up.



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## **Abstract**

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This example thesis briefly shows the main features of our thesis style, and how to use it for your purposes.



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## Abbreviations and Notation

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**AAV** adeno-associated virus.

**ABM** actin based motility.

**Ago** Argonaute.

**AIDS** acquired immune deficiency syndrome.

**AMR** antimicrobial resistance.

**API** application programming interface.

**Arp2/3** actin-related-protein 2 and 3.

**AS** application server.

**ATP** adenosine triphosphate.

**ATR** acid tolerance response.

**BaCV** *Bartonella*-containing vacuole.

**BrCV** *Brucella*-containing vacuole.

**cAMP** cyclic adenosine monophosphate.

**CAP** community-acquired pneumonia.

**CAR** coxsackievirus and adenovirus receptor.

**CCP** clathrin-coated pit.

**CCV** clathrin-coated vesicles.

**Cdc42** cell division control protein 42 homolog.

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**LIST OF ABBREVIATIONS AND NOTATION**

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- CDHR3** cadherin-related family member 3.
- COPII** coat protein complex II, involved in anterograde endoplasmic reticulum (ER)–Golgi transport.
- CSD** Cat Scratch Disease.
- DBP** DNA-binding protein.
- DGCR8** DiGeorge syndrome critical region gene 8.
- DMEM** Dulbecco Modified Eagle Medium.
- DSS** data store server.
- ECM** extracellular matrix.
- EHEC** enterohemorrhagic *E. coli*.
- Eis** enhanced intracellular survival protein.
- EPEC** enteropathogenic *E. coli*.
- ER** endoplasmic reticulum.
- ERAD** endoplasmic-reticulum-associated protein degradation.
- ERGIC** ER-Golgi intermediate compartment.
- EV** extracellular virion.
- FBS** fetal bovine serum.
- FCV** *Francisella*-containing vacuole.
- GAP** GTPase activating protein.
- GAPDH** glyceraldehyde-3-phosphate dehydrogenase.
- GDP** guanosine diphosphate.
- GEF** guanine nucleotide exchange factor.
- GFP** green fluorescent protein.
- GILT**  $\gamma$ -interferon-inducible lysosomal thiol reductase.
- GTP** guanosine triphosphate.
- HCS** high content screening.
- HDT** host directed therapeutics.
- HIV** human immunodeficiency virus.
- HPC** high performance computing.

- HSP** heat shock protein.
- HTS** high throughput screening.
- I(1,3,4,5,6)P<sub>5</sub>** inositol 1,3,4,5,6-pentakisphosphate.
- ICAM-1** interleukin-8.
- Ics** *Shigella* intracellular spread protein.
- ICU** intensive care unit.
- IL-8** intercellular adhesion molecule 1.
- ILK** integrin linked kinase.
- Ipa** *Shigella* invasion protein.
- KIF11** kinesin family member 11.
- LCV** *Legionella*-containing vacuole.
- LLO** listeriolysin O.
- LMNA** lamin A/C.
- LOF** loss-of-function.
- LPS** lipopolysaccharide.
- M cells** microfold cells.
- MAPK** mitogen-activated protein kinase.
- MHC** major histocompatibility complex.
- MID** Argonaute middle domain.
- miRNA** microRNA.
- MOI** multiplicity of infection.
- mRNA** messenger RNA.
- mTOR** mechanistic target of rapamycin.
- MV** mature virion.
- N-WASP** neural Wiskott-Aldrich syndrome protein.
- NF-κB** nuclear factor κ-light-chain-enhancer of activated B cells.
- NLS** nuclear localization signal.
- NPC** nuclear pore complex.

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**LIST OF ABBREVIATIONS AND NOTATION**

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- NPF** nucleation promoting factor.
- OTE** off-target effects.
- PACT** protein activator of protein kinase PKR.
- PAZ** PIWI-Argonaute-Zwille domain.
- PFA** paraformaldehyde.
- PI(3)P** phosphatidylinositol 3-phosphate.
- PI(4)P** phosphatidylinositol 4-phosphate.
- PI(4,5)P<sub>2</sub>** phosphatidylinositol 4,5-bisphosphate.
- PI(5)P** phosphatidylinositol 5-phosphate.
- PIWI** P-element-induced whimpy testes domain.
- PKR** RNA-dependent protein kinase pathway.
- PPIB** Peptidyl-prolyl cis-trans isomerase B.
- pre-microRNA (miRNA)** precursor miRNA.
- pri-miRNA** primary miRNA.
- PrPC** cellular prion protein.
- pTP** precursor terminal protein.
- Rac1** Ras-related C3 botulinum toxin substrate 1.
- RDRC** RNA-dependent RNA polymerase complex.
- RhoG** Ras homology growth-related.
- RISC** RNA-induced silencing complex.
- RITS** RNA-induced transcriptional silencing complex.
- ROS** reactive oxygen species.
- RTD** research, technology and development.
- SARS** severe acute respiratory syndrome.
- SCV** *Salmonella*-containing vacuole.
- shRNA** short hairpin RNA.
- SIF** *Salmonella*-induced filaments.
- Sip** *Salmonella* invasion protein.

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List of Abbreviations and Notation

- siRNA** short interfering RNA.
- Sop** *Salmonella* outer protein.
- SP41** *Brucella* surface protein.
- SPI** *Salmonella* pathogenicity island.
- SptP** secreted effector protein.
- T3SS** type III secretion system.
- T4SS** type IV secretion system.
- TAA** trimeric autotransporter adhesin.
- TAP** tapasin.
- TIR** toll-interleukin-1 receptor.
- Tir** translocated intimin receptor.
- TLR** toll-like receptor.
- TRBP** TRA RNA-binding protein.
- VEGF** vascular endothelial growth factor.
- VLDLR** very low-density lipoprotein receptor.
- WASP** Wiskott-Aldrich syndrome protein.
- WHO** World Health Organization.



## Chapter 1

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### Introduction

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Infectious diseases have played an undeniably important role in human history. With human populations becoming sufficiently aggregated to sustain direct life cycle viral and bacterial infectants around 2000 BC, devastating invasions of a growing number of pathogens started to occur ([Dobson and Carper, 1996](#)).

One of the earliest well documented incidence of a large-scale epidemic is known as the Plague of Athens. Starting in 430 BC and lasting roughly three years, a highly infectious disease killed 75'000 to 100'000 people or 25% of Athen's population. This catastrophic event is attributed either to smallpox, a viral infection with *Variola major* or typhus, caused by *Rickettsia* bacteria ([Littman, 2009](#)).

The bacterium *Yersinia pestis* is responsible for three major plague pandemics in the early and late middle ages, as well as in the late 19th century. Originating in northern Africa in 523 AD and spreading around the Mediterranean basin throughout the years 541–546, the Plague of Justinian is assumed to have killed up to half of the population of affected areas. The effect on cities was disproportionately severe. In Constantinople, for example, an estimated 230'000 people out of 375'000 lost their lives to the disease ([Treadgold, 1997](#)). Returning in the years 1347–1351, known today as the Black Death, a plague pandemic again wiped out around half of Europe's population. Death toll estimates range from 15 to 23.5 million ([Zietz and Dunkelberg, 2004](#)). Leaving behind a grim cultural heritage, this catastrophe had a lasting effect on economic and social structures in Europe. The third large-scale outbreak started around 1855 in southern China and quickly spread to Japan, Taiwan and India again wreaking havoc on the affected population.

Bringing diseases such as smallpox, measles (an infection with the Measles

## 1. INTRODUCTION

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virus) and typhus to the Americas during the European invasion of the New World had grave repercussions for the indigenous population, carrying no natural resistance towards the newly introduced pathogens. It is estimated that the population of present day Mexico fell from 20 million to 1.6 million over the course of the 16th century due to multiple disease epidemics, critically contributing to the successful colonization of the new continents (Dobson and Carper, 1996).

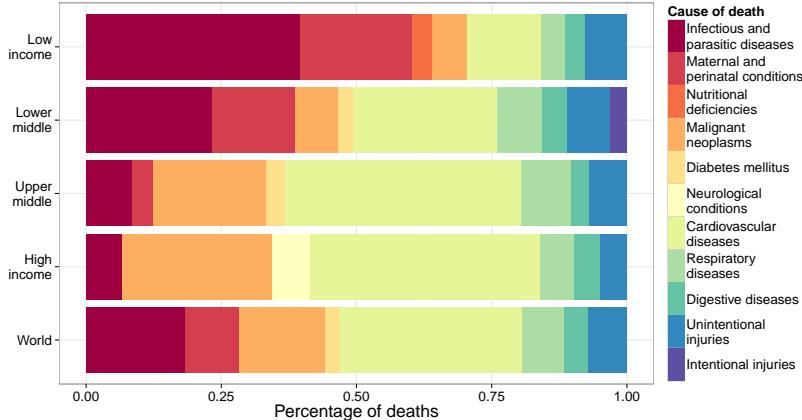
Cholera and influenza are further contagious diseases with high mortality rates, responsible for global epidemics. *Vibrio cholerae*, a bacterium which infects the intestine, became widespread in the early 19th century and led to seven pandemics since, the last of which only started in 1961. Antibacterial treatment of sewage and purification of drinking water greatly help to prevent and contain spreading of the disease but in areas with inadequate sanitation, such as Haiti after the 2010 earthquake, it remains a pathogen difficult to control. The influenza virus causes seasonal epidemics characterized by low lethality rates among people with intact immune systems<sup>1</sup>. Irregularly occurring influenza pandemics, initiated by zoonosis of new virus strains, against which no natural immunity exists, however, are accompanied by much higher lethality rates. The most significant such event is known today as the Spanish flu pandemic of 1918, costing the lives of 50–100 million, nearly half of which were young, healthy adults (Taubenberger and Morens, 2006).

In addition to diseases plaguing humanity for centuries, new ones continually emerge. Human immunodeficiency virus (HIV) is believed to have transferred from non-human primates in the early 20th century and the recent outbreaks of severe acute respiratory syndrome (SARS) and swine flu serve as reminders of such occurrences.

Despite development of means to treat and prevent many previously devastating diseases, infectious pathogens remain a serious threat to global health. In 2012, an estimated total of 58.3 million people died (20.1% in high, 29.4% in upper-middle, 36.5% in lower-middle and 14% in low income countries). Figure 1.1 partitions the total death count into World Bank income groups and causes. In low income countries, infective diseases are the most prevalent cause of death (39.6%), followed by maternal and perinatal complications with substantial margin (20.8%). In lower middle income countries, cardiovascular conditions catch up (26.5%), but are still almost matched in frequency by infectious diseases (23.3%). In upper middle (8.5%) and high income countries (6.7%), the importance of infectious disease while weakened remains account-

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<sup>1</sup>In spite of low lethality, these seasonal epidemics still incur significant economic damages. The World Health Organization (2003) estimates annual health care costs and loss of productivity due to influenza at US \$71–176 billion for the United States of America alone.



**Figure 1.1:** Relative frequencies of death causes in 2012 by World Bank income groups. Binning is based on Gross National Income (GNI) per capita and the thresholds are \$1045 or less for low income, \$1046 to \$4125 for lower-middle, \$4126 to \$12745 for upper-middle and \$12746 or more for high income economies. The data was obtained from the [World Health Organization \(2012\)](#).

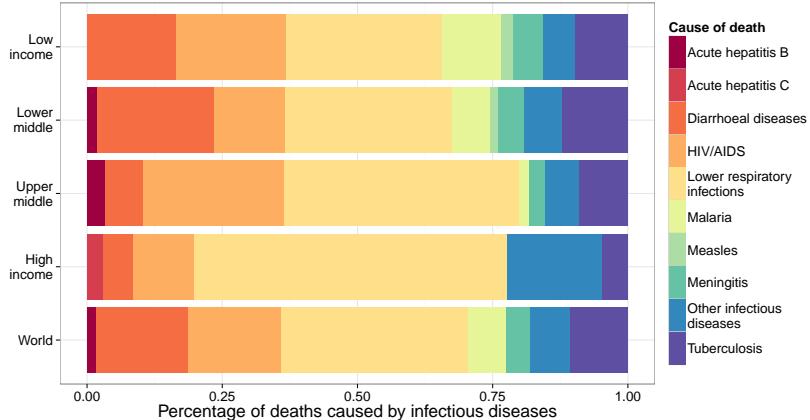
able for a significant number of deaths. Globally, infectious diseases are the second most frequent cause of death (18.3%), even more prevalent than all forms of cancer combined (15.8%) and only preceded by cardiovascular diseases (33.7%).

Focusing only on deaths caused by infectious disease, lower respiratory infections are most frequent (for each income region individually, low to high: 28.7%, 30.8%, 43.5% and 57.7% as well as worldwide: 34.5%; cf. figure 1.2). Diarrhoeal diseases and HIV/AIDS are the next most common worldwide (16.9% and 17.3%, respectively) where diarrhea is more prevalent in lower income regions (16.6% and 21.4% versus 7% and 5.6%), while HIV/AIDS plays a major role irrespective of income region (low to high: 20.4%, 13.3%, 26.2% and 11.3%).

Dealing with highly virulent pathogens and preventing their spreading requires a multi-pronged approach. First and foremost, etiology and routes of transmission have to be understood. Knowledge of vectors and natural reservoirs is of great importance as a first line of defense. In the case of plague, for example, insecticides killing fleas were successfully used as a prophylactic measure, as was controlling rat populations. Sanitary precautions including purification of drinking water, cooking foods well and the usage of disinfectants prevent initial infection, while measures such as sewage treatment, hand washing and wearing face masks help limiting spread among humans. Vaccination is the most important preventive measure. Exposing the immune system to a

## 1. INTRODUCTION

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**Figure 1.2:** Relative frequencies of deadly infectious diseases for 2012 by World Bank income groups. Binning is based on Gross National Income (GNI; see figure 1.1). The data was obtained from the [World Health Organization \(2012\)](#).

foreign antigen in a controlled manner artificially induces immunity. Among the great successes of widespread vaccination efforts is the global eradication of smallpox through a coordinated initiative lead by the World Health Organization in the 1970's.

Post-infection therapies include symptomatic treatments, as well as anti-infective drugs. Anti-bacterial or anti-viral agents exploit differences in proteomes between host and pathogen to selectively disable the invader with minimal toxicity to the host. This approach has been tremendously successful throughout most of the 20th century, leading to widespread application and prompting development of resistance towards the commonly used compounds. Adding to the severity of the problem is a lack of discovery of new drugs. No new class of anti-bacterial agents has been found since 1987, causing big pharmaceutical companies to withdraw from the area. The remaining research is mainly focused on improving on existing drugs, leading to a weak product pipeline, especially for the treatment of gram-negative bacteria ([Silver, 2011](#)). In its first global study on anti-microbial resistance, the [World Health Organization \(2014\)](#) notes:

Antimicrobial resistance (AMR) within a wide range of infectious agents is a growing public health threat of broad concern ... A post-antibiotic era—in which common infections and minor injuries can kill—far from being an apocalyptic fantasy, is instead a very real possibility for the 21st century.

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An alternative to pathogen directed search lies in targeting the set of host proteins necessary for infection. Many intracellular parasites subvert cellular functions to gain entry via host-mediated processes such as endocytosis. Upon entry, they move to a suitable niche and rely on host resources for proliferation. Challenges include evading host-cell defense mechanisms, generating sufficient space for replication, nutrient acquisition and keeping the host alive as long as possible, most of which require complex interactions between invader and host-based mechanisms. Finally, exiting the host cell again requires the parasite to successfully insert itself into existing signaling pathways ([Leirão et al., 2004](#)).

Host directed therapeutics (HDT) offer an escape from the conundrum of wanting to combat but not wanting to select for the surviving microbial parasites. The major challenge under this regimen is finding infectome components that are nonessential for cell survival, as orthogonality of host and infectant can no longer be exploited. Proving feasibility of the approach, [Czyż et al.](#) screened a library of 640 compounds already approved by the United States Food and Drug Administration (USFDA or FDA) for inducing resistance to four intracellular pathogens (*Coxiella burnetii*, *Legionella pneumophila*, *Brucella abortus*, and *Rickettsia conorii*). They found multiple drugs, not classified as antibiotics, that successfully inhibited intracellular bacterial growth while entailing only limited toxicity to THP-1 host cells. [Prussia et al.](#) review the usage of genome-wide screens to study host-pathogen interactions (for HIV and influenza) which in turn serve as basis for rational identification of drug targets for novel host-directed antivirals.

A detailed understanding of the human infectome is of crucial importance to the development of HDT and may even benefit the development of new antimicrobial agents. Feasibility of systematic loss of function screens using RNA interference methodology offers a unique opportunity to investigate complex cellular networks, making this an ideal tool for laying groundwork in combating infectious diseases. Of great importance, however, is ensuring reproducibility and comparability of such datasets, as well as ready availability to the scientific community.



## Chapter 2

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# Biological Background

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In order to better understand infectious diseases from a cell biological standpoint, this chapter reviews the current state of knowledge surrounding both bacterial and viral entry mechanisms. A sweeping overview of epidemiology and pathogenesis for several specific bacterial (*Bartonella henselae*, *Brucella abortus*, *Listeria monocytogenes*, *Salmonella enterica* and *Shigella flexneri*), as well as viral parasites (adenoviruses, rhinoviruses and *Vaccinia virus*) is given and the chapter concludes with a look at RNA interference as this mechanism is a cornerstone of genome-wide knockdown experiments.

### 2.1 Microbial Host-Cell Infection

Multi-layered keratinized skin is impenetrable for almost all microbial parasites. Instead they either require breaches such as cuts, scratches, puncture wounds and arthropod bites, or environmental interfaces which offer less impervious protection. Examples include respiratory, gastrointestinal and urogenital tracts, which all contain segments where only a single layer of epithelial cells has to be overcome. Although often protected by chemical defense mechanisms (acidity of the stomach and urogenital tract, as well as microbicidal factors in mucous secretions in the respiratory tract and small intestine), combined with frequent flushing (urination, peristalsis and the coordinated beating of cilia), some microbes have adapted to survive these hostile environments.

For extracellular pathogens to successfully colonize epithelial linings, they must avoid being removed by cleansing mechanisms of the host. Many bacteria accomplish this by expressing adhesins, protein complexes that recognize and bind to specific host-cell receptors, providing host and tissue tropism. Bacterial

## 2. BIOLOGICAL BACKGROUND

pili serve to extend reach and penetrate mucous secretions and therefore often carry adhesins. Enteropathogenic *E. coli* (EPEC) have extended this scheme by injecting their own receptor protein Tir (translocated intimin receptor) through the T3SS into the host cell to which it then attaches. This has entails the additional convenience that the intracellular domain of Tir can be used to modify host cell behavior ([Alberts et al., 2008](#)).

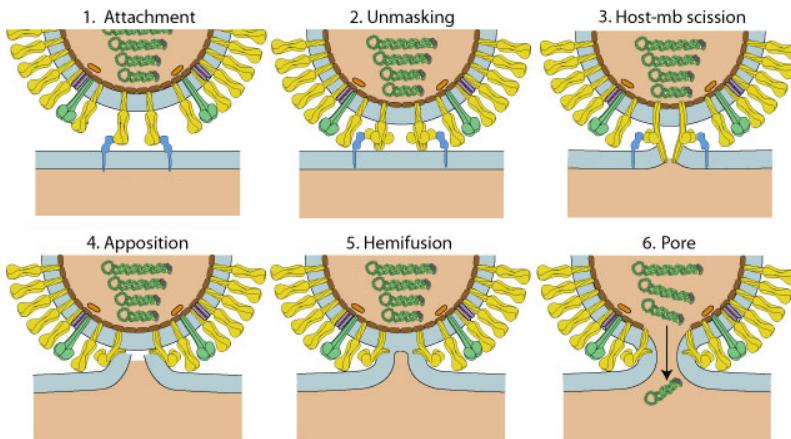
The outside of many epithelial barriers is covered in natural bacterial flora and crossing over into sterile cavities has the advantage of not having to compete with organisms well accustomed to that particular niche. Furthermore, intracellular pathogens are no longer accessible to antibodies and phagocytic cells and perhaps have a nutrient rich environment at their disposal.

### **2.1.1 Viral Infection Mechanisms**

The first step of any viral entering sequence is binding to the target surface. This can be mediated by attachment factors which simply serve to concentrate the virions on the cell surface or by virus receptors, which additionally act as communicators between host and pathogen. Common attachment factors include glycosaminoglycan chains and sialic acids and are comparatively unspecific. Glycoprotein spikes on enveloped and capsid proteins of non-enveloped viruses provide host specificity by binding cellular receptors. These cellular receptors typically serve other purposes and are exploited for infection. Binding affinity for individual interactions may be weak but aggregation of multiple interactions provide virtually irreversible avidity ([Smith, 2012](#)).

**Viral import.** For viral cell entry, different strategies exist. Enveloped viruses can either directly fuse with the plasma membrane (e.g. HIV) or be endocytosed by the host cell (e.g. influenza), while non-enveloped viruses either create a pore and directly inject their genome into the cytosol (e.g. polio virus) or are endocytosed (e.g. adenovirus). Endocytosis has major advantages over alternative strategies. Reaching its replicatory niche within the host cell is a difficult task for a microorganism having no means of locomotion and hijacking the endocytic system solves this problem elegantly. Furthermore, maturation of endosomes provides precise environmental cues to the invader for triggering uncoating and release. Both fusion with the cell membrane and injection of viral material into the cytosol leaves back traces of infection to be detected by the immune system. Being completely engulfed by the host, however, the intruders leave back no telltale traces. Additionally, lytic membrane penetration techniques are not as problematic to the host if only applied inside an endosome as opposed to the plasma membrane.

## 2.1. Microbial Host-Cell Infection



**Figure 2.1:** A generalized view of the steps necessary for viral–cellular membrane fusion. In pre-fusion conformation, fusion proteins have their hydrophobic fusion moieties tucked away. Upon attachment (1), they are unmasked (2), interact with the target membrane and ultimately penetrate it. Conformational change in fusion proteins induces membrane scission (3) and forces the two bilayers into close proximity (4), yielding a state of hemifusion (5). Finally a fusion pore is formed (6), stabilized by the post-fusion conformation which is lower in energy than the pre-fusion state. Adapted from [Hulo et al. \(2011\)](#)

Endocytic viruses trigger uptake either in a receptor mediated fashion (clathrin, caveolin or lipid raft dependent) or via non-specific macropinocytosis. The clathrin pathway is most widely used, for example by rhinoviruses, some adenoviruses, and coronaviruses and presents with characteristic invaginations, termed clathrin-coated pits (CCPs). The inwards facing pockets are subsequently pinched off by the membrane scission proteins dynamin-1 and dynamin-2, releasing clathrin-coated vesicles (CCVs) into the cytosol. Caveolin-mediated endocytosis is thought to be a more tightly regulated and low capacity pathway but is nevertheless exploited by several virus species, including picornaviruses and some retroviruses. Caveola formation is lipid raft dependent and recruitment of caveolin yields 50–70 nm flask-shaped pockets which are closed off by dynamin action. A lipid raft dependent, caveolin independent pathway has been described in simian virus 40 infection, but remains poorly understood. Lastly, larger virions such as poxviruses or herpesviruses initiate macropinocytosis, a mechanism typically employed by the cell for non-specific uptake of extracellular particles. An actin dependent membrane ruffling leads to formation of a lamellipodium which folds back onto the plasma membrane, enclosing an extraluminal volume and thus creating a macropinosome.

Upon endocytic uptake, viral pathogens need to uncoat and eject their genetic

## 2. BIOLOGICAL BACKGROUND

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**Table 2.1:** The Baltimore classification scheme is based on diversity of genetic system that have evolved in viruses. For each group, a selection of virus families capable of infecting humans, is provided, along with whether the virions are enveloped and the location of their replicatory niche. The data was obtained from [Hulo et al. \(2011\)](#)

	Genome based class	Examples	Enveloped	Replication site
DNA viruses	Group I: dsDNA	<i>Adenoviridae</i> <i>Poxviridae</i>	no yes	nucleus cytoplasm
	Group II: ssDNA(+)	<i>Parvovirinae</i>	no	nucleus
		<i>Anelloviridae</i>	no	nucleus
	Group III: dsRNA	<i>Reoviridae</i>	no	cytoplasm
RNA viruses	Group IV: ssRNA(+)	<i>Coronaviridae</i>	yes	cytoplasm
		<i>Picornaviridae</i>	no	cytoplasm
		<i>Hepeviridae</i>	no	cytoplasm
	Group V: ssRNA(-)	<i>Filoviridae</i> <i>Paramyxoviridae</i>	yes yes	cytoplasm cytoplasm
Retro	Group VI: ssRNA(+)-RT	<i>Orthoretrovirinae</i>	yes	nucleus
	Group VII: dsDNA-RT	<i>Hepadnaviridae</i>	yes	nucleus

material into the cytosol, as soon as their replicatory niche is reached. Escape timing is a critical issue, as late endosomes turn into lysosomes, capable of digesting their contents. Many enveloped viruses employ fusion mechanisms, which can be classified as type I or type II. For both types, increasing acidity associated with endosome maturation, initiates membrane fusion. Type I fusion proteins are forced into a metastable conformation prior to being added to the viral envelope and low pH triggers a conformational change to a state of lower energy. The energy released is used to force the two membranes close together resulting in their fusion (see figure 2.1). In type II fusion proteins, the critical transformation is not a conformational change but one in quaternary structure.

Non enveloped viruses cannot fuse with host membranes and have developed alternative approaches such as lysis (e.g. adenovirus) or ejecting their genome through pore-forming complexes (e.g. reovirus). Polyomaviruses need to pass through the ER because they rely on ER localized proteins to uncoat their capsid. For export from the ER into the cytosol, they exploit the endoplasmic-reticulum-associated protein degradation (ERAD) pathway, which serves as export mechanism for misfolded proteins from the endoplasmic reticulum to be degraded by proteasomes.

**Replication.** In contrast to larger intracellular parasites that carry the genetic information required for sustaining their own metabolism and replication, vi-

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ral pathogens typically rely almost exclusively on host machinery. Furthermore, viruses have developed strategies for interfering with host transcription and translation in order to promote synthesis of viral proteins at the expense of host gene expression. Even modulation of the host cell cycle is not uncommon, as some DNA viruses (including adenoviruses) are able to trigger a G1 to S phase transition, yielding an increased concentration of active DNA polymerase, while other species are capable of inducing a G2/M arrest, which again provides an optimized environment for those viruses. Further virus–host interactions include regulation of apoptosis, immune response modulation and interferon signaling.

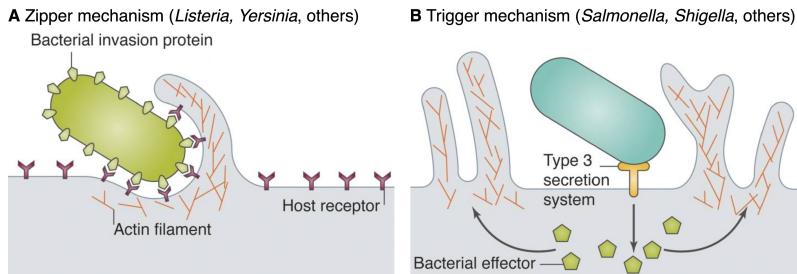
The remarkable diversity of genomic systems employed by viruses is captured by a classification system devised by [Baltimore \(1971\)](#). Table 2.1 lists the 7 types of viral genomes alongside examples of human viruses for each group, as well as whether those viruses are enveloped and where they replicate. Consequently, requirements for replication, transcription and translation vary widely among the different groups of viruses and due to the resulting mechanistic heterogeneity, viral propagation is not further explored within this general section. An excellent overview is provided by the online database of [Hulo et al.](#).

**Viral export.** The final stage of the viral life-cycle is concerned with virion assembly and exiting the host cell. Again, many strategies exist. Some nuclear replicating viruses (such as polyomaviruses) assemble their capsid proteins within the nucleus, requiring their structural proteins to target the nucleus via nuclear localization signals (NLSs) and leave the nucleus by disrupting the nuclear envelope, while others have their genome exported via nuclear pores and assemble progeny virions in the cytoplasm. Some cytoplasmic viruses (including poxviruses) replicate within special structures called viral factories or viroplasms, which increase efficiency of assembly and packaging and provides protection from host defense mechanisms. Other cytoplasmic viruses localize to organelles such as the ER (e.g. flaviviruses) where they are assembled and enter the secretory pathway via the Golgi apparatus. For intracellular motility, large virions such as poxviruses or herpesviruses have to rely on microtubule dependent transport whereas particles smaller than 20 nm can freely diffuse within the cytosol.

Once the host cell resources are depleted and replication is completed, progeny virions trigger their release. Viral shedding may occur via cell lysis, apoptosis, exocytosis or virion budding. Most non-enveloped and few enveloped viruses disrupt the plasma membrane with lytic viroproteins leading to cell death and release of cytoplasmic contents. While many viruses inhibit apoptosis, typically employed as a host defense measure, some (including hepeviruses and lentiviruses) have been implicated in exploiting this mechanism for expulsion

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**Figure 2.2:** Both the zipper (A) and trigger mechanisms (B) are actin dependent and lead to phagocytosis by usually non-phagocytic host cells. Zippering bacteria display an invasion protein on their surface that recruits actin filaments via a host receptor, while triggering bacteria inject an effector into the host cytosol by means of a syringe like type III secretion system leading to their uptake. Adapted from [Haglund and Welch \(2011\)](#).

and possibly subsequent infection of macrophages. Exocytosis and virion budding are two release strategies that are non-lethal to the host cell. Enveloped viruses acquire host-derived membrane either within the cell, typically at ER or Golgi exit sites or directly from the plasma membrane. In the latter case, envelopment coincides with host exit, whereas in the former case, virions are expelled via fusion of exocytic vesicles with the plasma membrane.

### 2.1.2 Bacterial Entry Mechanisms

Due to the much larger size of bacterial pathogens, endocytosis is not a feasible mechanism for entry, whereas phagocytosis can deal with uptake of particles this large. While phagocytosis is a function usually only available to macrophages, some bacteria have evolved mechanisms of inducing phagocytosis in other cell types. Species explicitly targeting macrophages, such as *Mycobacterium tuberculosis* and *Legionella pneumophila*, have to be able to escape phagosomes or deal with resisting digestion.

Two recurring patterns for inducing phagocytosis in non-phagocytic cells have been described by [Cossart and Sansonetti \(2004\)](#) as zipper (encountered in *Yersinia pseudotuberculosis* and *Listeria monocytogenes*) and trigger mechanisms (used by *Salmonella enterica* and *Shigella flexneri*). Not all entry strategies can be assigned to these two classes and several additional, unrelated pathways have been uncovered.

**Zipper mechanism.** Host entry by zippering bacteria is characterized by bacterial surface proteins binding to cellular receptors, thereby inducing signaling cascades that lead to limited, localized actin rearrangements. This extends

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the plasma membrane alongside the entering bacteria in a zipper-like fashion. [Cossart and Sansonetti](#) divide the scheme into three successive steps:

- (a) Contact and adherence. Independent of the actin cytoskeleton, bacterial adhesins interact with host cell surface proteins leading to receptor clustering. *Y. pseudotuberculosis* displays invasin, capable of interacting with  $\beta_1$  integrins, while *L. monocytogenes* uses internalin A, a protein that binds E-cadherin. Cadherin and integrins are usually involved in anchoring cell junctions and the invading bacteria mimic initiation of junction formation by a neighboring cell.
- (b) Phagocytic cup formation. Responding to the mistaken signal, the target cell extends its surface towards the signal origin via Arp2/3 (actin-related-protein 2 and 3) and Rac1 (Ras-related C3 botulinum toxin substrate 1) mediated actin polymerization, attempting to cover the adhesive surface. This leads to engulfment of the invading bacterium.
- (c) Phagocytic cup closure. In a process probably involving the phosphotransferase phosphatidylinositol-4-phosphate 5-kinase, catalyzing the conversion of PI(4)P (phosphatidylinositol 4-phosphate) to PI(4,5)P<sub>2</sub> (phosphatidylinositol 4,5-bisphosphate), the protruding membrane sections fuse and actin depolymerization returns the plasma membrane to its original state.

**Trigger mechanism.** In trigger schemes, bacterial T3SS weakly adhere to target cell receptors and inject effector molecules into the host cytosol through a pore formed by the syringe like delivery mechanism. These proteins directly interact with actin regulatory cellular components, causing major actin rearrangement characterized by membrane ruffling. This in turn leads to non-specific engulfment of bacteria and surrounding particles. Four steps have been identified by [Cossart and Sansonetti](#):

- (a) Pre-interaction stage. Effector molecules, stored in bacterial cytosol, are associated with dedicated chaperones in order to prevent aggregation, degradation and once secretion is initiated guide them towards T3SS. T3SS needle complexes (several hundred per bacterium are possible) are fully assembled and integrated into the two membranes of gram-negative bacteria.
- (b) Interaction stage. The tip of T3SS recognizes the host cell membrane and activates secretion via a process not well understood. Originally it was believed that the needle complex was able to puncture the target membrane but recent evidence suggests that translocator proteins are ejected which subsequently form a pore and thusly facilitate insertion of the secretion

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**Table 2.2:** A selection of T3SS genes in *Salmonella* and *Shigella*. The *Salmonella* genome encodes two separate secretion systems which are deployed at distinct phases of infection. The data was obtained from [Wang et al. \(2012\)](#).

T3SS component	<i>Salmonella</i> SPI-1	<i>Salmonella</i> SPI-2	<i>Shigella</i>
Outer membrane ring	InvG	SsaC	MxiD
Inner membrane ring	PrgK, PrgH	SsaJ, SsaD	MxiJ, MxiG
Cytoplasmic ring	SpaO	SsaQ	Spa33
Export apparatus	SpaP, SpaQ, InvA	SsaR, SsaS, SsaT	Spa24, Spa9, MxiA
Needle assembly	InvI, InvJ, OrgA	SsaO, SsaP, SsaK	Spa13, Spa32, MxiK
Needle major subunit	PrgI	SsaG	MxiH
Needle minor subunit	PrgJ	SsaI	MxiI
Translocon	SipB, SipC, SipD	SseB, SseC, SseD	IpaB, IpaC, IpaD
ATPase	InvC	SsaN	Spa47
Effector export	InvE		MxiC
Chaperone	SicA, InvB, SigE	SseA, SscA, SscB	IpgC, IpgE, IpgA
Secreted effector	SipA, SptP, SopE, SopE2, SopB, SopA	SifA, SifB, SseE, SpvC, SseG, SseI	IpaA, IcsB, IpgD, IpgB1, IpgB2, VirA
Transcription regulator	InvF, HilA, HilC, SirA, SirB, PhoP	SsrA, SsrB, YdgT, OmpR, H-NS, Hha	MxiE, VirB

system. Once bacterial and cellular cytoplasms are connected, effector chaperones dissociate and unfolded proteins (the needle passage is only 3 nm wide) enter the host in a probably adenosine triphosphate (ATP) dependent manner. Chaperones serve a double purpose as transcription factors, encouraging synthesis of new effector protein when not attached to their substrate.

- (c) Formation of macropinocytic pocket. Massive, but localized membrane protrusions emerge due to action of bacterial effectors. In *Shigella* entry, VirA causes local destabilization of microtubules by binding to  $\alpha/\beta$ -tubulin heterodimers. This in turn stimulates Rac1 activity, Cdc42 (cell division control protein 42 homolog) recruitment and subsequent Arp2/3 activation leading to actin polymerization. IpaC recruits the Src tyrosine kinase further enhancing actin dynamics. *Salmonella* inject the proteins SopE and SptP, an activator/inhibitor pair for the GTPase complex Rac1/Cdc42. Guanine nucleotide exchange factor (GEF) activity of SopE induces actin rearrangements leading to formation of the macropinocytic

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cup.

- (d) Closing of macropinocytic pocket. In *Salmonella* invasion, GTPase activating protein (GAP) activity of secreted effector protein (SptP), restores the inactive guanosine diphosphate (GDP) state of Rac1/Cdc42 and leads to actin depolymerization. Its GEF partner SopE is degraded more rapidly than SptP, enabling reversible control over the pathways exploited for entering. In case of *Shigella*, actin depolymerization is initiated by binding of IpaA to vinculin, a key protein of focal adhesion plaques.

**Other entry pathways.** In addition to trigger and zipper type uptake, other atypical mechanisms exist. Host cell entry of *Brucella abortus*, for example, has been described as invasome mediated ([Dehio, 2005](#)). In this actin dependent process, bacteria aggregate on the cell surface and trigger their engulfment by injecting bacterial effectors into the host via T4SS. The internalized structure is called an invasome.

Actin-independent uptake albeit rare, is possible as evidenced by *Campylobacter jejuni* and *Citrobacter freundii*, which have evolved a microtubule dependent invasion strategy ([Kopecko et al., 2001](#)). A further example of microtubule involvement is presented by *Clostridium* spp. (a genus that includes the etiological agent of tetanus and several species capable of botulinum toxin synthesis). These intercellular pathogens induce formation of long protrusions formed by microtubule filaments that wrap around the bacteria and fix them in close proximity. It has been speculated that such a mechanism could be exploited by intracellular pathogens to promote adherence ([Haglund and Welch, 2011](#)).

### 2.1.3 Intracellular Survival

While bacteria that successfully subvert endocytic pathways and trigger their uptake have defied innate and evaded adaptive immune responses, they are still faced with defensive mechanisms by their new hosts. A multitude of strategies has evolved in order to undermine hostile actions directed at intruding bacteria and for establishing a replicatory niche within this initially adverse but nevertheless potentially propitious environment.

Phagocytic vacuoles, containing internalized microorganisms are destined for endocytic maturation. These compartments undergo successive acidification and finally develop into mature degradative phagolysosomes. Most intracellular pathogens either escape the endocytic vacuole before fusion with lysosomes occurs or manipulate cellular pathways that control its maturation, thereby creating a niche permissive to their survival. Two important themes for cytosolic

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bacteria are efficient means of locomotion and evasion of cellular responses such as autophagy.

**The phagocytic vacuole.** In order to survive an environment characterized by constantly decreasing pH, poor nutrient content and an increasing concentration of antibacterial and lysosomal enzymes, most successful pathogens incapable of escaping their internalization compartment alter biogenesis and dynamics of their surroundings. Examples include *Salmonella*, *Mycobacterium tuberculosis* and *Legionella pneumophila* (Ham et al., 2011).

*Salmonella* replicates in plasma membrane derived perinuclear *Salmonella*-containing vacuoles (SCVs) that are neither early, nor late endosomes. While some fusion events with vesicles of the endocytic pathway take place, maturation into lysosomes is inhibited. SopB, an SPI-1 encoded T3SS effector, hydrolyzes PI(4,5)P<sub>2</sub> and reduced concentration of PI(4,5)P<sub>2</sub> inhibits recruitment of cellular RAB GTPases required for phagosome-lysosome fusion. A key role in manipulating membrane trafficking is played by the SPI-2 encoded T3SS effector SifA. Endosomal maturation is associated with microtubule dependent vacuole relocation and SifA has been shown to regulate kinesin activity, thereby contributing to SCV integrity. Furthermore, SifA activity is essential in increasing the SCV volume and creating the necessary space for replication.

Similarly to SopB in *Salmonella*, the bacterial phosphatase SapM expressed by *M. tuberculosis*, dephosphorylates phosphatidylinositol 3-phosphate (PI(3)P) which blocks phagosomes from fusing with late endosomes and consequently arrests maturation. A different approach is employed by *L. pneumophila* which secrete SidC via T4SS, an effector protein capable of binding and displaying PI(4)P on the vacuolar surface. This leads to recruitment of ER-derived vesicles which turns the phagosome into an LCV (*Legionella*-containing vacuole) which is removed from the endocytic pathway and becomes sufficiently spacious for replication.

**Cytosolic replication.** Pathogens that evolved to replicate in the cytosol must quickly escape the internalization vacuole. Most species are capable of triggering their release within 30 minutes of infection, highlighting the importance of quick action in order to prevent damage incurred by acidification of the phagosome. In all known cytosolic bacteria vacuolar egress is a pathogen driven process and most species employ lytic enzymes capable of forming trans-membrane pores. Examples of bacteria that enter the cytosol as part of their life-cycle include *Listeria monocytogenes*, *Shigella flexneri*, *Burkholderia pseudomallei*, *Francisella tularensis* and species of the *Rickettsia* genus.

## 2.1. Microbial Host-Cell Infection

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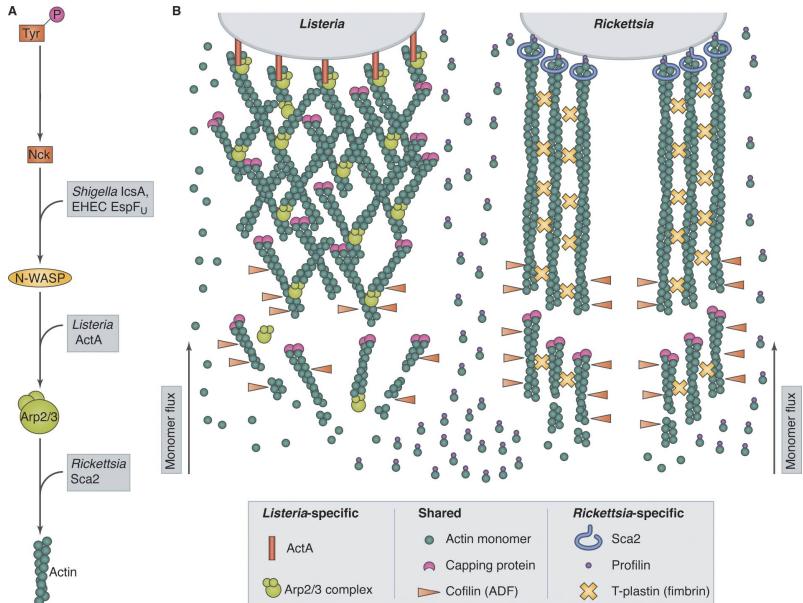
The best studied organism, *L. monocytogenes*, secretes listeriolysin O (LLO) a hemolytic enzyme, capable of inserting into the target membrane, oligomerize and through a conformational change form a pore. This delays endosomal maturation, prevents fusion with lysosomes and finally releases the bacteria into the cytosol. Regulation of LLO is pH mediated and involves the host factor  $\gamma$ -interferon-inducible lysosomal thiol reductase (GILT), specific to endosomes, phagosomes and lysosomes. Vacuole escape by *S. flexneri* is dependent on the three T3SS translocator proteins *Shigella* invasion protein (Ipa)B, IpaC and IpaD with IpaB and IpaC forming a pore complex and IpaD facilitating insertion into the membrane. For both *Rickettsia* spp. and *B. pseudomallei*, little information on vacuole-lytic mechanisms exist. Hemolysin C and phospholipases have been implicated of playing important roles but more work is necessary to uncover their exact mechanistic relevance. *F. tularensis* stands out among the previously mentioned organisms in that lysis of the internalization vacuole is followed by re-entry of another membrane bound compartment, the *Francisella*-containing vacuole (FCV).

Interestingly, it is not known to what extent, the nutritional content of mammalian cytosol is permissive to bacterial growth. As evidenced by *S. flexneri* which can grow with a doubling time of only 40 minutes (growth rates in laboratory medium are comparable) it is possible for microorganisms to adapt to this environment. While it seems natural to assume that cellular cytosol provides ideal conditions for bacterial growth and replication, this raises the question of why only comparably few pathogenic organisms exploit this ecological niche. Indeed, nutritional arguments are not the only reasons to consider, as it is clear that reaching this habitat is no easy feat and further defense against foreign particles in the cytosol, for example autophagocytosis, exist ([Ray et al., 2009](#)).

**Actin-based motility.** As for large viruses, free diffusion within the cytoplasm is not readily possible for bacteria and a feature common to most cytosolic bacterial is ABM. Actin monomers exist in two forms, G-actin (globular) and F-actin (filamentous). ATP binds G-actin, and upon formation of a trimeric nucleus the nascent chain grows in both directions while ATP is hydrolyzed. When the chain reaches a certain length, growth becomes directional and association of monomers at the plus end compensates for dissociation of monomers at the minus end. This leads to a self-sustaining scheme described as treadmilling. Limiting to actin polymerization is the kinetically unfavorable nucleation step and in vivo, this is stimulated by cellular factors such as Arp2/3 which require activation by NPFs, including members of the Wiskott-Aldrich syndrome protein (WASP) family.

ABM exhibiting pathogens can be divided into two groups depending on

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**Figure 2.3:** The capability for actin based motility evolved independently in several intracellular bacterial pathogens. (A) The entry points into the cellular actin assembly pathway vary, with some organisms providing their own nucleation promoting factor (*Listeria* and *Rickettsia*), while others rely on cellular NPF (*Shigella* and enterohemorrhagic *E. coli*). The branching pattern of actin tails differs between organisms relying on cellular Arp2/3 from those capable of Arp2/3 independent ABM (B). Cofilin is responsible for actin depolymerization, profilin is a regulatory protein catalyzing the exchange of ADP to ATP and fimbrin crosslinks actin strands into bundles. (Haglund and Welch, 2011).

whether they provide NPFs of their own or recruit host cell NPFs. Expression of ActA by *L. monocytogenes* is both necessary and sufficient for ABM as shown in a cell free system using ActA coated beads, actin, Arp2/3, CapZ, cofilin and ATP. No cellular NPF is required as ActA mimics WASP and directly activates Arp2/3. *Rickettsia* are among the few pathogens capable of ABM without relying on Arp2/3, leading to formation of actin tails with a distinct morphology. The formin-like protein Sca2 directly interacts with actin and RickA, exhibiting some homology to WASP can act as NPF. A further organism that mimics cellular NPF is *B. pseudomallei*, which is capable of BimA-mediated, Arp2/3 independent actin polymerization.

Bacterial pathogens that rely on cellular NPF instead of supplying their own include *S. flexneri*, *Mycobacterium marinum* and enterohemorrhagic *E. coli* (EHEC).

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In *S. flexneri* infection, bacterial IcsA, capable of recruiting neural Wiskott-Aldrich syndrome protein (N-WASP), is the only required bacterial factor for triggering ABM. By inducing conformational changes in N-WASP, IcsA is thought to mimic activation by Cdc42, allowing for association of N-WASP with Arp2/3. Details of ABM in *M. marinum* are less well known but as it is both WASP and Arp2/3 dependent, it is assumed that the employed mechanism is similar to that of *S. flexneri*.

Both EHEC and EPEC, while not intracellular pathogens, are able to induce cytosolic actin polymerization. By inserting the transmembrane protein Tir into the plasma membrane they are able to initiate actin polymerization via the same pathway as ABM, leading to the formation of pedestals protruding outwards. This is thought to provide means of extracellular motility. Intercellular motility by ABM capable bacteria is achieved by pushing against the host cell membrane into the neighboring cell body until being engulfed in a double membrane vacuole that is subsequently lysed (Stevens et al., 2006; Haglund and Welch, 2011).

**Autophagy.** The basic catabolic cellular mechanism of autophagy is initiated as response to stress signals such as nutrient starvation, damaged cellular compartments and foreign particles. Cytoplasmic components marked for degradation are captured by the double-membrane autophagosome, intended for fusion with lysosomes. As cellular defense mechanism, cytosolic pathogens are targeted by selective autophagy in a process also related to xenophagy. Bacterial response can be categorized as interference with the autophagy pathway, evasion of autophagy recognition or escape from the autophagosome.

*S. typhimurium*, while not usually considered a cytosolic pathogen, has been shown to occasionally escape the phagocytic vacuole and replicate in the cytosol. Membrane damage triggers an amino acid starvation response that activates autophagy signaling by relocating mechanistic target of rapamycin (mTOR) from the late endosome to the cytosol. The bacteria respond via unknown action probably involving SPI-2 T3SS secretions that are able to restore both the cytosolic amino acid pool and mTOR localization, successfully inhibiting autophagy. Further examples of autophagy-initiation signaling inhibition are provided by *M. tuberculosis* which is able to produce enhanced intracellular survival protein (Eis), an inhibitor to production of reactive oxygen species (ROS) needed as an autophagy signal. Furthermore, bacterial toxins apt for interfering with cyclic adenosine monophosphate (cAMP) regulation can block autophagy in mammalian cells.

After autophagy has been initiated, the procedure can still be modified as shown by *L. pneumophila* and *S. flexneri*. The former organism secretes RavZ

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via T4SS which decouples the autophagy marker LC3 from the phagosomal membrane while the latter expresses VirA via T3SS, a GAPcapable of inactivating the autophagy regulator RAB1. Another group of bacteria prevent or delay lysosome fusion and accumulate in non-degradative vacuoles at neutral pH. Examples include *M. marinum*, *Chlamydia trachomatis*, *Yersinia pestis* and *Helicobacter pylori*.

Evasive measures have been suggested to be employed for *S. flexneri* and *L. monocytogenes*. IcsB of *S. flexneri* and ActA of *L. monocytogenes* have been implicated of masking the bacteria from autophagy targeting mechanisms and it has been suggested that ABM might somehow facilitate hiding from host cell detection. By also currently unknown means, *B. pseudomallei* can escape the phagosome of an autophagy-related process, possibly involving BopA secretion via T3SS ([Huang and Brumell, 2014](#)).

## **2.2 Select Bacterial Pathogens**

A total of 5 bacterial pathogens were selected for study within the InfectX research, technology and development (RTD) project by SystemsX. This section shortly describes each organism in terms of microbiological features, pathogenesis, epidemiology and diseases caused in humans. For each organism, a chapter of [Rolain and Raoult \(2006\)](#) serves as basis and is enriched by one or two review articles referenced in the first paragraph of each section.

### **2.2.1 *Bartonella Henselae***

*Bartonella henselae* is a short, rod shaped, unflagellated proteobacterium, phylogenetically closely related to the genus *Brucella*, presenting 94.4% 16S rRNA gene sequence homology, compared with *Brucella abortus*. The Gram-negative bacillus is a facultative anaerobic, intracellular parasite and was first described in 1992. Relatively harmless for healthy humans, infections can become life threatening in immunocompromised patients, making the species an important opportunistic pathogen ([Anderson and Neuman, 1997](#); [Harms and Dehio, 2012](#)).

**Diseases.** In immunocompetent humans, infection with *B. henselae* can lead to a condition known as Cat Scratch Disease (CSD). As the name suggests, most patients report being in contact with a cat and transmission often occurs through scratches and bites. Affecting primarily children and young adults (80% are 21 or younger), the self limiting infection typically presents itself with lymphadenopathy. Most patients remain afebrile and do not report feeling ill, with low-grade fever and malaise shown in roughly 30% of the cases. Recovery

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from uncomplicated CSD usually takes 2 to 6 months and requires no specific treatment.

Possible complications include Parinaud's oculoglandular syndrome (granulomatous conjunctivitis in one eye and parotid lymphadenitis on the same side), splenomegaly and hepatic or splenic abscesses, accompanied by fever, weight loss, fatigue and malaise. In 1 to 7% of the cases, the disease spreads to the central nervous system, leading to encephalopathy, but recovery is usually rapid (within several weeks).

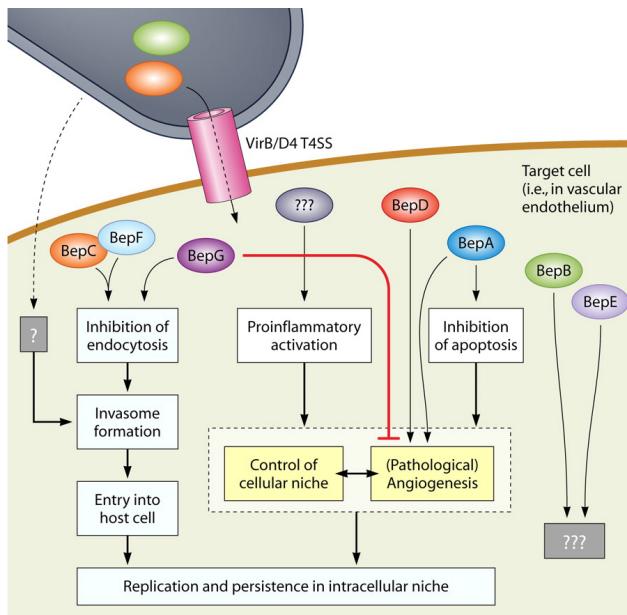
Infections with *B. henselae* tend to have more severe consequences for immunocompromised patients, such as bacillary angiomatosis, bacteremia and endocarditis. Acquired immune deficiency syndrome (AIDS) patients suffering from CSD usually experience severe, progressive disease with infection spreading systematically and without appropriate treatment, fatal outcome. *Bartonella* spp. are the only prokaryotes known to be able to induce angiogenic tumors such as bacillary angiomas, which may involve skin, respiratory or gastrointestinal epithelia, heart, liver, spleen, bone marrow, muscles, or lymph nodes. Bacteremia may lead to inflamed heart valves, usually requiring endocarditic patients to have heart valve replacement surgery.

**Pathogenesis.** *B. henselae* are capable of intracellular growth in both epithelial cells and erythrocytes but the focus of this section lies on infection of the former cell type. Initial attachment is mediated by the trimeric autotransporter adhesin (TAA) BadA which is capable of both interacting with the extracellular matrix (ECM) and  $\beta_1$ -integrin, followed by effector secretion via the bacterial T4SS VirB/D4. For host cell entry, two mutually exclusive mechanisms have been described. Either single bacteria or small groups are phagocytosed via a zipper-like mechanism or large clusters are internalized in a unique cellular structure termed an invasome. While invasome formation is a slow process, taking 16 to 24 hours, *Bartonella*-containing vacuoles (BaCVs) resulting from endocytosis are visible within minutes. It has been suggested that inhibition of endocytosis by either a combination of effector proteins BepC and BepF or the exclusive action of BepG is crucial to invasome formation as it allows for aggregation of bacteria on the cell surface. Not the activity of effector proteins but the clustering of cellular receptors may trigger large-scale internalization (c.f. figure 2.4).

Pathological angiogenesis can be induced as the net result of a set of agonistic and antagonistic effectors. While BepG is a strong inhibitor to angiogenesis, both BepD (weakly) and BepA (strongly) promote sprouting of new blood vessels. Although the exact mechanisms of induction and regulation remain to be uncovered, the secretion of vascular endothelial growth factor (VEGF) has been

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**Figure 2.4:** Several effectors secreted by T3SS in *B. henselae* infection serve as virulence factors for colonization of the intracellular replicatory niche. The ones best characterized alongside their phenotype are schematically summarized. ([Harms and Dehio, 2012](#)).

demonstrated in vasoproliferative tumors caused by infection with *B. henselae*. Furthermore, transcriptional promotion of various factors supporting angiogenesis, including interleukin-8 (ICAM-1), intercellular adhesion molecule 1 (IL-8) and angiopoietin-2 by bacterial effectors are unanimously agreed upon. Curiously, secretion of VEGF by infected endothelial cells has so far not been possible to show.

Inhibition of apoptosis is decisive to intracellular survival and it is assumed that BepA is capable of cAMP mediated antiapoptotic action. The role of two further effectors that have been identified as part of the T4SS in *B. henselae*, BepB and BepE, are unknown, as are the mechanisms leading to activation of proinflammatory signalling via NF- $\kappa$ B (nuclear factor  $\kappa$ -light-chain-enhancer of activated B cells).

**Epidemiology.** The role of cats and in particular kittens, as reservoirs to *B. henselae* has been firmly established. Infected felines are asymptomatic and

## 2.2. Select Bacterial Pathogens

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show no signs of illness. Cat fleas (*Ctenocephalides felis*) serve as vectors to spread the bacteria among cats and have also been suspected of infecting humans. The main path of transmission to humans however, is through scratches and bites by infected cats. *B. henselae* has also been found in ticks and tick bites prior to contraction of CSD have been reported.

In the United States, 24000 cases of CSD are reported yearly, yielding 2000 hospital admissions with an estimated health care cost of \$12 million. Children are more likely to be affected (80%) and incidence is higher in males (60%). The seasonal pattern (occurrences higher in fall/winter) is attributed to cat mating patterns, as well as pet acquisition fluctuations.

### 2.2.2 *Brucella Abortus*

The Danish physician David Bang first isolated *Brucella abortus* in 1895 from cystic cattle tissue, investigating a contagious disease causing abortions in cows. *B. abortus* are small, unflagellated proteobacteria with a cell wall consisting of an outer layer of lipopolysaccharide (LPS) (9 nm) and an inner layer of muramyl mucopeptide complexes (3–5 nm). The Gram-negative cocobacilli appear to have evolved from free-living, soil-dwelling species and are closely related to other human pathogens such as *Bartonella* spp., based on 16S rRNA sequences. *Brucella* species were investigated for possible use as warfare agents in the mid 20th century by several armed forces. ([Atluri et al., 2011](#); [von Bargen et al., 2012](#))

**Diseases.** Brucellosis is a human disease caused by several pathogenic *Brucella* species, most importantly *B. abortus*, *B. melitensis*, *B. canis* and *B. suis*. Onset may be acute or insidious and due to protean symptoms, diagnosis based on clinical presentation alone is difficult. The febrile disease is generalized and may involve many parts of the body, including nervous, skeletal, gastrointestinal, cardiovascular, respiratory and genitourinary systems. Furthermore, as the bacteria spread to other reservoir hosts via their reproductive systems, persistence of infection is crucial to the pathogen and it comes as no surprise that brucellosis can manifest as a chronic disease in humans too.

Fever is the most consistent sign of *Brucella* infection and depending on what specific organs are affected, further symptoms include asthenia, anorexia, nausea, malaise, arthritis, hepatomegaly, splenomegaly, epididymo-orchitis in males, and pulmonary manifestations such as bronchitis or pneumonia. A rare complication (less than 2%), albeit the most lethal, is infective endocarditis. Invasion of the nervous system develops in less than 5% of cases and often results in meningitis or meningoencephalitis with good prognosis under antimicrobial treatment.

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**Pathogenesis.** Host entry occurs primarily via the digestive system but is also possible through the respiratory tract or skin lesions. Via the gastrointestinal route, *Brucella* spp. target Peyer's patches (lymphoid nodules localized towards the end of the small intestine) and must therefore pass through acidic conditions in the stomach. This is facilitated by expression of two ureases capable of hydrolyzing urea and producing a protective bicarbonate buffering system. When entering through the respiratory system, *B. abortus* target alveolar macrophages which serve as access point to the lymphatic system therefore facilitating systematic spread.

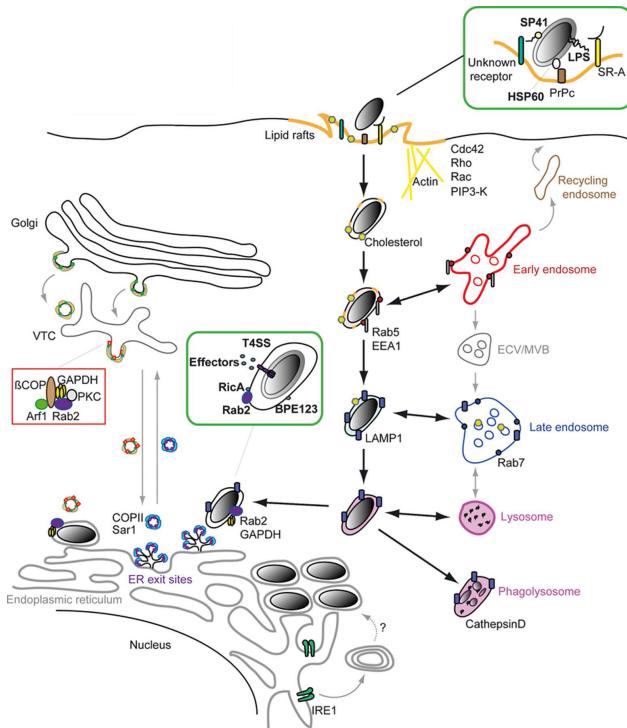
In order to persist at systemic sites, both active and passive mechanisms for evading the immune system are in place. LPS of the outer cell wall disguises the bacteria from toll-like receptors (TLRs) and expression of two proteins containing toll-interleukin-1 receptor (TIR) domains actively interferes with TLR signaling.

Uptake by macrophages happens via phagocytosis, which is either triggered by nonopsonized bacteria through a lipid raft mediated mechanism or by opsonization. Although opsonin marked bacteria are 10-fold more likely to be ingested, the number of pathogens reaching their destination within the host cell is higher for nonopsonized bacteria. Still, most bacteria (up to 90%) are unsuccessful in evading their digestion and only very few are able to establish a replicative niche. Apart from professional phagocytes, epithelial cells may also be infected and the following paragraphs focus on this particular cell type.

Initial attachment is mediated by unknown eukaryotic receptors containing sialic acid residues that interact with *Brucella* surface protein (SP41). While involvement of bacterial HSP60 and cellular prion protein (PrPC) has been proposed, this remains controversial. Maturation of early *Brucella*-containing vacuoles (BrCVs) is important for successful infection as preventing acidification (through addition of baflomycin A) or fusion with lysosomes (through suppression of the late-endosomal GTPase Rab7) interferes with bacterial replication. This observation can be explained with acid serving as a trigger for expression of the T4SS VirB. However fusion events with late endosomes and lysosomes are only limited and under bacterial control.

Upon acquisition of late endosomal markers and acid initiated expression of T4SS, fusion with autophagic vacuoles occurs leading to formation of an autophagosome-like compartment. Subsequent interactions with ER exit sites, mediated by secreted effectors, further modify the BrCV into an ER-derived vacuole, coated with ribosomal particles. At this stage, located within the ER-Golgi intermediate compartment (ERGIC), the replicatory niche is reached. Blocking the small GTPase Sar1 inhibits intracellular replication by preventing acquisition of coat protein complex II, involved in anterograde ER-Golgi

## 2.2. Select Bacterial Pathogens



**Figure 2.5:** Schematic representation of the *B. abortus* intracellular life cycle, from cell entry via maturation of the *Brucella* containing vacuole to establishing an intracellular replicatory niche. Interaction with many different cellular compartments of the endocytic pathway and the *cis* Golgi network are required for successful infection (von Bargen et al., 2012).

transport (COPII) by ER-exit vesicles and the small GTPase Rab2, involved in ER-cis-Golgi traffic, is required for maximal proliferation, illustrating the dependence of *Brucella* on intercepting vesicular traffic.

Despite multiplying intra-cellularly in high numbers, host cells are kept alive and are even able to replicate despite infection. Furthermore *Brucella* species are able to interfere with apoptosis, maintaining their replicatory niche, protected from immune response. It remains unknown what happens when the hosts capacity for freeloaders is reached, as well as how bacteria exit the host cell and spread.

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**Epidemiology.** Preferred natural reservoir species for *B. abortus* are cattle (*Bos taurus* and *Bos indicus*) and almost all parts of the world are affected. The disease exists in both domestic and wild animals and is most prevalent in Mediterranean countries, North Africa, throughout the Middle East, India, Central Asia, as well as South and Central America. Zoonosis most often occurs through ingestion of unpasteurized milk products but airborne transmission is also possible, putting professionals involved in animal husbandry at risk. Vertical transmission among reservoir hosts can occur through lactation and horizontal transmission is facilitated by mating and placental discharge associated with aborted gestation. Human-to-human transmission is rare (but has been suspected to be possible via sexual intercourse), making humans dead-end hosts. As opposed to *Bartonella*, immunodeficient patients do not seem to be especially susceptible towards *Brucella* infections.

Worldwide, an estimated 500000 new cases of brucellosis occur annually, making it one of the most prevalent zoonoses. Although usually susceptible to combined antibiotic therapies of at least two agents (usually a tetracycline antibiotic combined with an aminoglycoside or rifampin), untreated brucellosis leads to a high degree of morbidity, leading to being classified a neglected zoonosis by the World Health Organization (WHO).

### 2.2.3 *Listeria Monocytogenes*

The short, Gram-positive bacilli are non-sporeforming facultative anaerobes, capable of growing in a wide temperature range (0–50°C) and in many different environments. Flagellation is temperature dependent with flagellin being expressed and assembled into peritrichous flagella around 20–25°C but not at 37°C. First described in 1924 by Murray after isolation from lymph glands of diseased laboratory animals, the pathogen was found to also infect humans four years later. For much of the time since, listeriosis was considered a rare zoonotic disease and did not receive much attention. It was not until the 1980s, when several food-borne listeria outbreaks caused a shift in interest towards the pathogen, which has since become a well studied facultatively intracellular parasite (Farber and Peterkin, 1991; Cossart and Lebreton, 2014).

**Diseases.** Maternal and neonatal listeriosis accounts for almost half of all infections. Listeriosis in pregnancy typically manifests in bacteraemia and presents as a self-limiting febrile disease with flu-like symptoms. Many cases, however are asymptomatic and the first sign of infection is abortion or neonatal listeriosis. Maternal infection does not necessarily carry over to the fetus, especially if proper chemotherapy is administered. Perinatal incidences are divided into early and late onset (>5 days after parturition), with former cases

## 2.2. Select Bacterial Pathogens

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typically resulting in septicemia and latter cases in meningitis. While in early onset cases the predominant route of transmission is transplacental, the situation is less clear in late onset cases. Both the maternal genital tract during child birth and environmental sources have been implicated. Despite antibiotic treatment, overall mortality rates of 30–40% are typical and prognosis for early onset disease is worse, as it is often associated with preterm birth and advanced stage of infection.

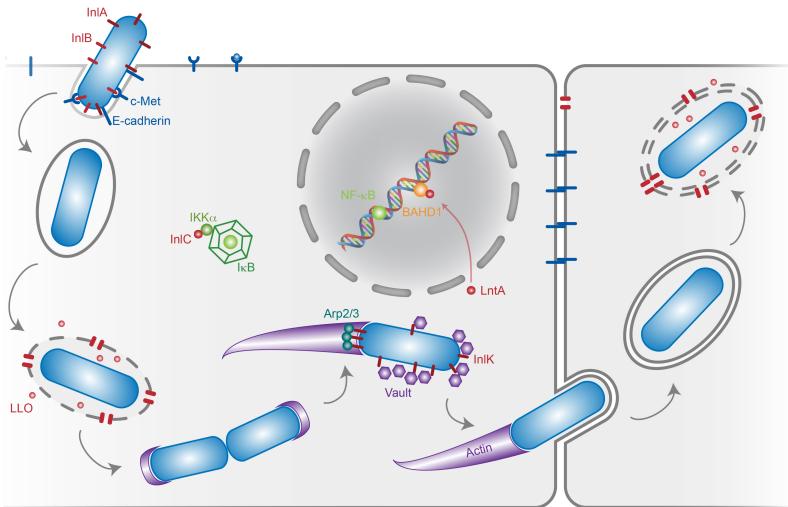
Among adults, most cases of listeriosis occur in T-cell deficient individuals. HIV infection, for example, increases incidence 150–300 fold compared to general population control groups. Predisposing conditions include lymphoreticular neoplasms, deliberate immunosuppression (e.g. antirejection treatment after organ transplants), alcoholism and diabetes mellitus. Despite increased susceptibility caused by immunodeficiency, roughly 30% of all infections affect immunocompetent individuals. In healthy adults, consumption of food contaminated with *L. Monocytogenes* can either lead to self-limiting febrile gastroenteritis with short incubation time (<24 h) or invasive listeriosis with much longer incubation periods (3–4 weeks). The systemic form of infection often manifests as bacteraemia or as a neurological infection, but can also involve endocarditis and spread to other parts of the body. Central nervous system involvement occurs in as much as 75% of cases and either presents as meningitis or encephalitis. Mortality rates of 35–45% have been reported for listeriosis in adults.

**Pathogenesis.** The predominant entry path for *L. Monocytogenes* into the human body is via the gastrointestinal tract, where Peyer's patches are targeted. The bacteria can induce cellular uptake, by non-phagocytic host cells via expression of cell-surface associated internalins through a zipper-type entry program (see section 2.1.2). The cellular receptor for bacterial internalin A is E-cadherin and internalin B interacts with the receptor tyrosine kinase c-Met. Upon internalization, the phagosomal membrane is lysed, mediated by secretion of listerial haemolysin LLO in a cholesterol dependent mechanism whereby LLO monomers associate, oligomerize and form 35 nm pores. LLO is acid activated with an optimum around pH 5.5, which is reached in late endosomes. Moreover, LLO is required for autophagy modulation (see section 2.1.3) and has been implicated in regulating inflammatory response.

Growth and replication occur in the cytoplasm and ActA mediated ABM (see section 2.1.3) provides means of intracellular and intercellular movement. Adjacent cells can be entered by pushing against the plasma membrane and forming a pseudopod-like structure which in turn is taken up the neighbor. The resulting double-membrane vacuole is escaped by cytolysis, again dependent on LLO.

## 2. BIOLOGICAL BACKGROUND

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**Figure 2.6:** *Listeria* enter host epithelial cells via a zipper-type entry mechanism mediated by the bacterial internalins InlA and InlB. In order to reach the cytosolic replicatory niche, lytic LLO is secreted and ABM provides means of locomotion. Evading host detection and maintaining a permissive environment is controlled by several bacterial effector molecules including InlC, InlK and LntA. ([Cossart and Lebreton, 2014](#)).

Several mechanisms for persistence within the replicatory niche have recently been uncovered. Down-regulation of the host pro-inflammatory response is dependent on secretion of InlC, a virulence factor of the internalin family which prevents phosphorylation of NF- $\kappa$ B and thus prevents nuclear translocation. Both InlK and ActA are associated with ABM and are simultaneously involved in autophagy evasion. Finally, mechanisms for several epigenetic host modifications have been demonstrated. Both LLO action and Met-binding during invasion, trigger histone modifications and upon entry, the virulence factor LntA localizes to the nucleus where it interacts with the newly characterized chromatin component BAHD1. While illustrating that *L. monocytogenes* is capable of reprogramming host gene expression, the exact implications of this capability have yet to be elucidated.

In an extracellular setting, haemolysins serve to rupture erythrocytes in order to generate free iron, a limiting growth factor. Furthermore the nonspecific immune system has to be evaded and haemolysins have also been shown to be cytotoxic towards leukocytes. Additionally, expression of bacterial superoxide dismutase mitigates the effect of free superoxide radicals which play an important role in killing phagocytized bacteria.

## 2.2. Select Bacterial Pathogens

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**Epidemiology.** Incidence of listeriosis has initially been increasing since its recognition as food-borne disease but effects of awareness and diagnostic methods are unclear. While typically long incubation periods do pose difficulties for clinical diagnosis, the number of susceptible individuals is on the rise and certain aspects modern processing and handling of foods may be beneficial for growth of *L. Monocytogenes*. Disease rates of 2–15 cases per million population per year have been reported and listeriosis is among the leading case of lethal food-borne pathogen infections. Most recent data, however suggests that incidence is decreasing again.

Due to its non-fastidious lifestyle, *L. Monocytogenes* has been isolated from a wide array of ecological niches, including soil, sewage and water (both fresh water bodies and estuaries). High persistence (up to 4 years) in soil samples is problematic when contaminated manure is used as fertilizer and biofilm formation poses challenges for eradication from food processing plants. Additionally, the ability to grow in refrigerated foods and resistance towards heat treatment such as pasteurization warrant alertness and special preventive care. Many different foods have been implicated in listeriosis outbreaks, including vegetables (potatoes, radishes and celery), seafood (shrimp, crabmeat and smoked fish), dairy products (soft cheese, pasteurized and unpasteurized milk) and meats (poultry, various types of sausages and pâté).

Despite high prevalence in food (studies have found 20–80% of meat product samples and 1–10% of dairy product samples contaminated with *L. Monocytogenes*), comparatively few successful infections occur. The bacteria are ingested frequently in small doses and stool sample examinations suggest that 10–70% of investigated populations could be intestinal carriers. While the minimum infectious dose has not been settled definitively, approximations range from  $10^6$  to  $10^9$  bacteria.

### 2.2.4 *Salmonella* Typhimurium

*Salmonella* are non-sporulating, Gram-negative bacilli, belonging to the family Enterobacteriaceae. The motile bacteria are able to produce peritrichous flagella and diameters span 0.7 to 1.5  $\mu\text{m}$  while typical lengths range from 2 to 5  $\mu\text{m}$ . They are closely related to the genus *Escherichia*, showing only 15% chromosomal sequence disparity. Currently, two distinct species, *S. bongori* and *S. enterica*, within the genus *Salmonella* are recognized, both of which are pathogenic towards a wide array of hosts. *S. enterica* is further divided into 6 subspecies, the most relevant of which for human and domestic animal hosts being *enterica*. A large number of serovars (more than 2500) for *S. enterica* subsp. *enterica* have been characterized and due to an originally mistaken classification as separate species, some serovars are designated with shortened

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names. *S. Typhimurium*, therefore is shorthand for *S. enterica* subsp. *enterica*, serovar *Typhimurium* and to emphasize that *Typhimurium* is not a species description it is not italicized.

The first description of the genus *Salmonella* dates back to an investigation into swine fever led by Salmon and Smith in 1885. The newly isolated bacterium was wrongly proposed as the etiological agent, as the disease later turned out to be caused by a virus (Fàbrega and Vila, 2013; Haraga et al., 2008).

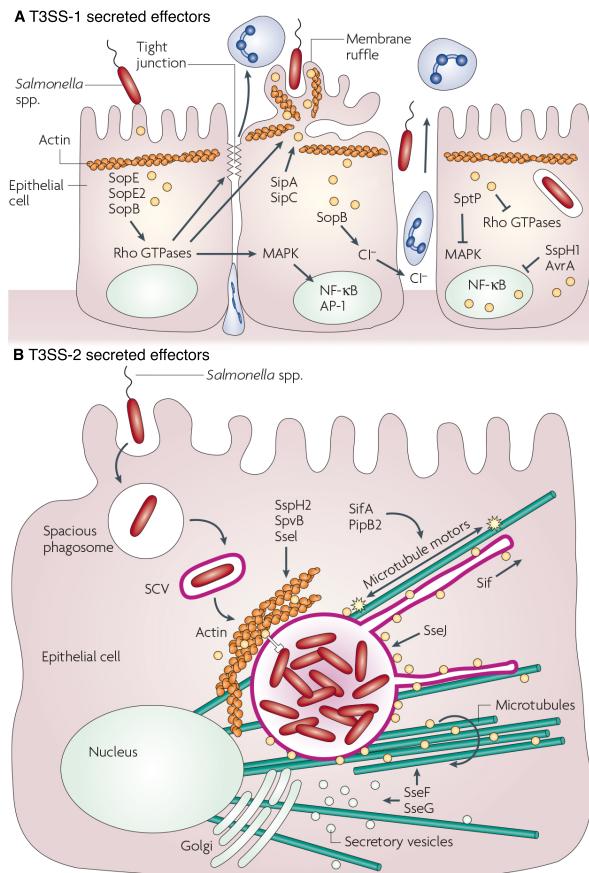
**Diseases.** Two distinct disease patterns are associated with *Salmonella* spp. infections, typhoid fever and salmonellosis. While the former is exclusively caused by the serovars *Typhi* and *Paratyphi*, the latter is associated with several serovars, the most frequent being *Enteritidis* and *Typhimurium*, accounting for 65% and 12% of cases worldwide. The current and following sections will not discuss typhoid fever.

Salmonellosis is a diarrheal disease with a short incubation period of 6–24 h, followed by nausea, vomiting, loose or liquid bowel movements, abdominal cramps and fever. Clinical features are similar to those of dysentery and other gastroenteric disease and can include bloody and mucosal stool. In most cases, the infection is self-limiting and symptoms fade away within 4 to 7 days. The most common complication is bacteraemia, which presents in 5% of cases and is more likely to develop in children, especially if malnourished, and immunocompromised individuals. Further manifestations of invasive infections include meningitis, osteomyelitis, cholangitis, pneumonia and endocarditis. While mortality in immunocompetent hosts in developed countries is as low as 0.1%, it can increase to 77% for HIV positive patients in undeveloped countries.

**Pathogenesis.** In order to reach the small intestine, ingested *S. Typhimurium* first have to defy the hostile environment of the stomach. A set of proteins, summarized as acid tolerance response (ATR) helps mediate the acidic conditions and improves survival rates. The remaining bacteria subsequently end up in the small bowel and target epithelial cells, with preference towards micro-fold cells (M cells). Flagellar motility enables penetration of intestinal mucus secretions and improves the chance of reaching the intestinal walls where adhesion can be established. Fimbriae are important attachment factors, capable of interaction with host-cell laminin and fibronectin and provide an initial platform for pathogen induced phagocytosis via trigger mechanism (see section 2.1.2).

*S. Typhimurium* utilize two separate T3SS systems for host-cell colonization, the first of which (T3SS of SPI-1) mediates invasion. In addition to strength-

## 2.2. Select Bacterial Pathogens



**Figure 2.7:** Two separate T3SS are encoded in the *Salmonella* genome which serve as central virulence factors at different stages of infection. T3SS of SPI-1 secretes effectors required for triggering internalization (A), while T3SS of SPI-2 is critical for maturation of the phagosome into a replication permissive SCV. Figures adapted from [Haraga et al. \(2008\)](#).

ening initial interactions attaching the pathogen to its target, the needle like structure serves as delivery mechanisms, capable of injecting effector proteins, including SopE, SopE2, SopB (also known as SigD) and *Salmonella* invasion protein (Sip)A. SopE serves as GEF and activates Rho GTPases Rac1 and Cdc42 via GDP/GTP exchange, which in turn initiate cytoskeletal rearrangements. SopE2 is a further GEF, highly homologous to SopE and it is assumed to provide some

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level of redundancy for SopE as shown by *Salmonella* strains missing the SopE gene. SopB is a phosphatase, capable of hydrolyzing various substrates, including inositol 1,3,4,5,6-pentakisphosphate (I(1,3,4,5,6)P<sub>5</sub>), which has been linked to intestinal fluid secretion causing diarrhea and PI(4,5)P<sub>2</sub>, involved with membrane rigidity.

The actin binding proteins SipA and SipC stimulates actin polymerization and promotes membrane ruffling, yielding a macropinocytic pocket. Mitogen-activated protein kinases (MAPKs) initiate pro-inflammatory response via NF-κB signaling, leading to the expression of IL-8. Furthermore, damage the tight junctions make the epithelial barrier permissive to bi-directional leakage. After internalization, actin structure is restored and MAPK signaling down regulated by SptP, SspH1 and AvrA.

Upon engulfment, maturation of the phagosome and fusion with lysosomes have to be prevented. Unlike other pathogens that escape the digestive mechanisms of phagocytic vesicles by moving to the cytosol, *Salmonella* replicate inside phagosome-derived SCVs. In this setting, the second translocation complex (T3SS of SPI-2) is activated, and used to secrete effectors, including SsaB and SifA, capable of interacting with vesicular trafficking mechanisms and guiding the SCV away from its original degradation pathway. Furthermore T3SS-2 translocated effectors such as SspH2, SpvB and Ssel induce actin polymerization events, which relocate the SCV toward a perinuclear position. A last step in creating the intracellular niche needed for replication, is formation of *Salmonella*-induced filaments (SIFs), extending outwards from the SCV. T3SS-2 secretes effectors such as SifA, PipB2, SseF and SseG, which mediate the microtubule dependent processes by bundling and accumulating filaments and regulating microtubule motor function.

**Epidemiology.** The global disease burden caused by nontyphoidal salmonellosis is estimated at 90 million cases per year and 150000 deaths. Incidence rates are highest in East and Southeast Asia (up to 4000 cases per 100000 population per year) and both developed and undeveloped countries are affected (incidence rates in Africa are estimated at 320 while estimates for Europe are around 690 cases per 100000 population per year). An estimated 80.3 million or 86% of reported cases are food borne ([Majowicz et al., 2010](#)).

In order to control *Salmonella* outbreaks, preventive measures in food production and processing is of major importance. This starts with disease containment in domestic animals, such as vaccination of chickens, enforcing hygiene standards in manufacturing and distribution facilities and ends with proper preparation, exploiting heat sensitivity of the organisms. As the main route of transmission is fecal-oral, good sanitary infrastructure, treatment of

## 2.2. Select Bacterial Pathogens

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sewage and water processing are crucial prerequisites in combating outbreaks of salmonellosis.

### 2.2.5 *Shigella Flexneri*

*Shigellae* are small, non-sporeforming, Gram-negative bacilli and belong to the family *Enterobacteriaceae*, along with *Escherichia*, *Salmonella* and *Yersinia*. While flagellar genes are present and their expression is observed under certain conditions, the bacteria are usually described as non-motile and unflagellated. The facultative intracellular parasite shows strong specificity towards human hosts where it typically infects the lower gastrointestinal tract.

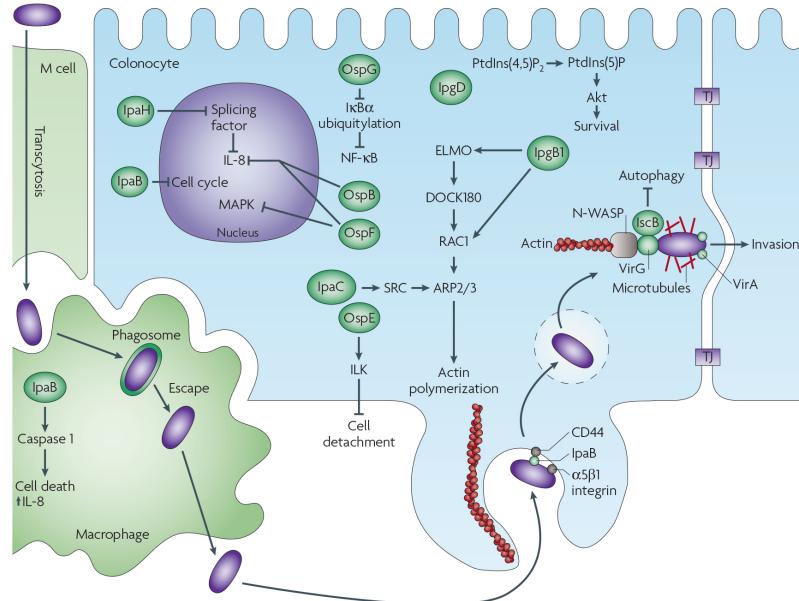
*Shigella dysenteriae* was identified as the etiological agent of dysentery by Shiga in 1897 during an epidemic in Japan with 91000 reported cases and a >20% mortality rate. *S. Flexneri* was first described by Flexner in 1900, while investigating diseases endemic to the Philippines. Recent genetic studies suggest, that *Shigella* spp. belong to the species *Escherichia coli*, rather than forming a distinct genus, as only marginal sequence divergence (1.5%) between *E. coli* and *S. Flexneri* was found ([Schroeder and Hilbi, 2008](#); [Croxen and Finlay, 2010](#)).

**Diseases.** Bacillary dysentery is an acute infection of the intestine. Mild cases of the disease are self-limiting and afebrile with diarrhea and possibly vomiting as the only symptoms. In as little as 24 hours after onset, bowel movements usually begin to normalize and the condition is resolved within a few days. More severe cases are accompanied by strong abdominal cramps, fever and watery diarrhea containing blood and mucus, indicative of injury to the intestinal epithelium. While still usually self limiting in healthy individuals, recovery takes 10–14 days and relapses are possible. In immunocompromised patients, young children, especially if malnourished, and elderly individuals, life threatening complications including bacteraemia, renal failure, intestinal perforation and toxic megacolon are more frequent. Involvement of the central nervous system and respiratory tract is rare.

Administration of antibiotics is not recommended in mild to moderate cases as the disease can usually be overcome by the immune system and AMR in *Shigellae* is becoming an increasing concern. Oral rehydration therapy is the most effective treatment, helping the body replenish liquids and salts lost due to diarrhea. For severe infections, the use of antibiotics can become necessary and testing for resistance patterns, if possible, is advised.

**Pathogenesis.** Main targets of *S. Flexneri* are mucosa of the distal ileum and colon where they enter epithelial cells from the basolateral side. For initial

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**Figure 2.8:** *S. flexneri* cross the epithelial barrier of the distal ileum and colon by transcytosis via M cells, followed by escape from macrophage digestion. Internalization into epithelial cells from the basolateral side is mediated by trigger mechanism endocytosis. Several bacterial effectors are instrumental in reaching the replicatory niche and ensuring host survival [Croxen and Finlay \(2010\)](#).

crossing over from the apical side, action of M cells at Peyer's patches is exploited. These specialized enterocytes constantly sample antigens from the gut lumen and pass them to intraepithelially located dendritic cells and lymphocytes. Upon uptake by basolaterally located macrophages, *S. Flexneri* survive digestive action of the phagosomal vacuole by disrupting the surrounding membrane and initiating host-cell apoptosis, mediated by the bacterial effector protein IpaB, capable of acting on a caspase 1 regulated apoptotic pathway (c.f. figure 2.8). The bacteria are subsequently released into the sub-mucosa, where they induce phagocytosis by normal epithelial cells via trigger mechanism (see section 2.1.2).

Initial contact between pathogen and target host cell is mediated by cellular  $\alpha_5\beta_1$  integrins and binding of IpaB to CD44 receptors may initiate first actin rearrangements, preparing the cell for uptake. Subsequent injection of at least 6 effector proteins into the epithelial cytosol through the T3SS triggers engulfment of the bacterium in an actin dependent process, involving the small GT-

## 2.2. Select Bacterial Pathogens

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Pases Rac1 and Cdc42, which recruit the actin nucleation complex Arp2/3. IpaC, a component of the translocation complex, is involved in stimulating Rac1 and Cdc42 GTPase activity through an unknown mechanism and the secreted effector proteins IpgB1, IpgB2, IpgD and IpaA facilitate actin polymerization. IpgD, an inositol phosphate phosphatase, catalyzes the hydrolysis of PI(4,5)P<sub>2</sub> to PI(5)P which promotes disassociation of cytoskeleton and membrane, increasing actin availability and making the plasma membrane more susceptibility to manipulation. The mechanism of action of IpgB2 remains to be resolved, while IpgB1 is assumed to mimic activated RhoG (Ras homology growth-related), a small GTPase, located upstream in the signaling pathway of Rac1. Finally, binding of IpaA to vinculin induces depolymerization of actin, which is assumed to be important for closing of the phagocytic cup.

The resulting phagocytic vacuole has to be escaped before maturation progresses, which is accomplished in a non-acid dependent fashion, by the translocator proteins IpaB, IpaC and IpaD via membrane lysis. With release into the epithelial cytosol, the replicatory niche of *S. Flexneri* is reached. Actin mediated intracellular motility enables intercellular dissemination (see section 2.1.3) and targeting of epithelial tight junctions initiates break-down of the epithelial barrier, providing more pathogens with access to the basolateral side of the gut lining.

ABM, is driven by activity of two bacterial proteins. VirA is secreted at the forward facing end of the rod shaped bacilli, which promotes degradation of tubulin structures and therefore clearing a path through the dense network of microtubules and surface bound Ics (*Shigella* intracellular spread protein)A (VirG) facilitates actin polymerization at the opposite end. Both Arp2/3 and N-WASP are involved in actin nucleation, the directed nature of which provides the driving force for locomotion.

In order to maintain their intracellular niche, several strategies have evolved within the *Shigella* infectome. Autophagy is inhibited by IscB (see section 2.1.3) and via the previously mentioned phosphatase action of IpgD, cellular Akt proteins are activated which regulate cell survival and inhibit apoptosis. Furthermore, IpaD interacts with cell cycle regulatory protein MAD2L2, mediating cell cycle arrest. Together with OspE action on integrin linked kinase (ILK), downregulating cell detachment, this prevents turnover of epithelial cells. Inflammatory response is muddled by a combination of at least four bacterial effectors. Cytoplasmically acting OspG inhibits NF-κB, while nuclearly located IpaH and OspB reduce IL-8. Adding to that, OspF dephosphorylates MAPKs that are required for transcription of genes of the NF-κB pathway.

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**Epidemiology.** Estimates by the WHO peg the disease burden caused by *Shigella* spp. at 80 million annual cases worldwide, leading to 700000 deaths. Developing countries are disproportionately affected, representing 99% of all cases, as are children less than 5 years old, accounting for 70% of cases and 60% of deaths. In developed countries, incidence rates of 1–2 per 100000 population are typical and *Shigellae* are common causes of Traveler's diarrhea.

The predominant route of transmission is fecal–oral, highlighting the importance of sanitary precautions for infection control. Proper treatment of fecal matter is important for preventing contamination of drinking water and inhibiting spreading by disease vectors such as house flies. During acute phases, diseased individuals excrete pathogens in large numbers and as few as 100–200 organisms are sufficient of causing a new infection.

### **2.3 Select Viral Pathogens**

In addition to the previous 5 bacterial pathogens, 3 viruses were selected for study within the InfectX RTD project by SystemsX. This section shortly describes each pathogen in terms of physical features, pathogenesis, epidemiology and diseases caused in humans. For each section, a chapter of [Craighead \(2000\)](#) serves as basis.

#### **2.3.1 Adenoviruses**

The family *Adenoviridae* encompasses 5 genera of non-enveloped, medium sized (90 nm diameter) viruses, capable of infecting a broad range of vertebrate hosts. The capsid is of  $T = 25$  icosahedral symmetry, composed of 720 hexons arranged as 240 trimers which form the triangular facets and 12 penton capsomeres located at the vertices. A homotrimeric fiber glycoprotein protrudes from each vertex, attached to the penton base via interactions of its N-terminal domains and ending in a globular, C-terminal knob. The genome is present as double stranded DNA (Baltimore group I), is non-segmented, linear, 35–35 kb long and encodes 40 proteins.

Adenoviruses were first isolated from human adenoid tissue cultures by Rowe in 1953 and their study led to the discovery of alternative splicing in 1977, a commonly encountered phenomenon among eukaryotes. Currently, 57 serovars are recognized as pathogenic towards humans, all belong to the genus *Mastadenovirus* and are classified into 6 species, labeled A through G. The following sections are mostly concerned with *Human adenovirus C* ([Lenaerts et al., 2008](#)).

## 2.3. Select Viral Pathogens

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**Diseases.** In immunocompetent individuals, adenoviruses seldom cause more than transient disease with many infections even occurring subclinically and fatal outcome being very uncommon. Symptomatic cases usually manifest as respiratory tract infections or conjunctivitis and less frequently as hemorrhagic cystitis, nephritis or gastroenteritis. Infections of the oropharynx can spread to the lower respiratory tract, causing bronchitis, bronchiolitis or pneumonia, which can become chronic, leading to desquamated epithelial tissue and long-term damage to the respiratory mucosa. Heart failure and central nervous system involvement can occur in severe cases. Ocular infections range from mild, short-term follicular conjunctivitis to highly contagious keratoconjunctivitis with possible long-term damage to the cornea. *Human adenovirus C* serotypes are mostly associated with respiratory diseases but have also been implicated in eye infections.

Invasive forms of disease are mostly limited to immunocompromised individuals, including transplant recipients, HIV infected, hereditary immunodeficient and cancer patients subject to chemotherapy. In addition to the lungs, a wide variety of organs has been reported to be affected, such as parotid glands, liver, gall bladder, colon, brain and kidney and pathological changes range from perivascular cuffing to parenchymal necrosis.

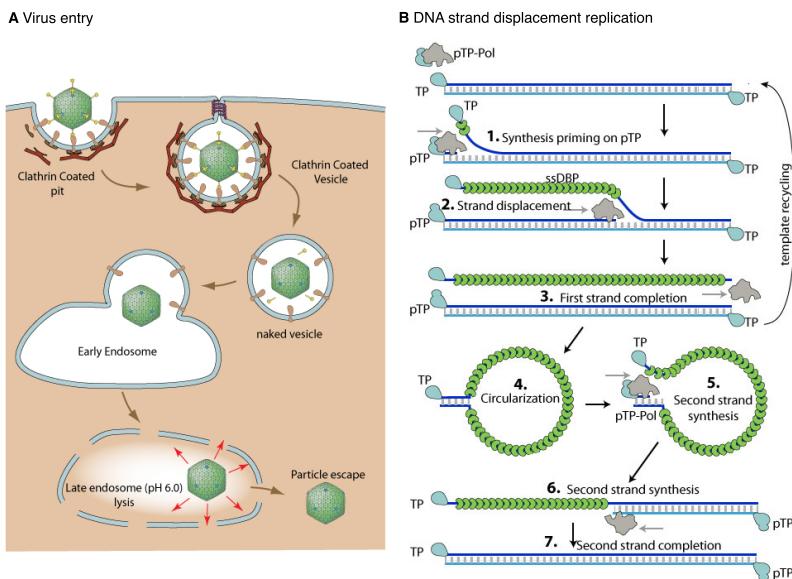
**Pathogenesis.** Initial attachment of virions is mediated by interactions between the globular knobs of viral fiber proteins and target cell CARs (coxsackievirus and adenovirus receptors). In *Human adenovirus C* infections, cellular heparan sulfate proteoglycans serve as additional attachment factors, reinforcing adhesion. Subsequent binding of penton bases to  $\alpha,\beta$ -integrin receptors induces clathrin-mediated endocytosis and leads to loss of viral fiber proteins (see figure 2.9, A).

The adenoviral replication cycle is divided into early (E) and late stages (L), with each seeing expression of a typical set of genes. Upon engulfment by the host cell and triggered by endosomal acidification, hexon bound protein VI disassociates from the capsid structure and induces lysis of the endosomal membrane. The remainder of the now cytosolic virion is shuttled to the nucleus by microtubular transport where viral protein IX recruits kinesin thereby procuring capsid disruption.

Nuclear penetration is mediated by core protein VII and occurs at nuclear pore complexes (NPCs), leading to transcription of early viral genes by host RNA polymerase II. The resulting proteins manipulate various cellular processes, such as evasion of host immune response (E3, E19), cell cycle (E1A), apoptosis (E1B and E4), autophagy (E1B) and messenger RNA (mRNA) transport (E4), while also promoting viral DNA replication (E2). Modulation of the adaptive

## 2. BIOLOGICAL BACKGROUND

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**Figure 2.9:** Schematic representation of molecular mechanisms for cell entry and genome replication in adenovirus infection. Formation of the endocytic vesicle is mediated by viral fiber proteins interacting with cellular receptors that in turn recruit clathrin. The CCP is pinched off by membrane scission proteins dynamin-1 and dynamin-2, leading to release of a cytosolic CCV (A). The adenoviral genome is replicated via a mechanism described as DNA strand displacement replication whereby single stranded DNA is synthesized from the viral template in a protein primed fashion, which in turn is copied into double strand DNA. Figures adapted from [Hulo et al. \(2011\)](#).

immune response is mediated by the tapasin (TAP) inhibitor E19, as binding of TAP prevents loading of peptides onto major histocompatibility complex (MHC) class I molecules for display on the cell surface. In order to create an optimal environment for replication, viral protein E1A induces a G1/S cell cycle transition which increases the concentration of cellular enzymes involved in DNA replication. This, however also leads to accumulation of cellular p53, participating in apoptosis signaling. Several viral proteins, including E1B 55K and E4 orf6 have been shown to inhibit the apoptotic pathway. E1B 19K is involved in activation of autophagy thereby contributing to the delay or inhibition of apoptosis.

A virally encoded DNA polymerase replicates the genome by DNA strand displacement in an unusual protein primed fashion involving precursor terminal protein (pTP) acting as primer and DNA-binding protein (DBP), stabilizing single stranded DNA, as well as several host proteins (figure 2.9, B). With onset

## 2.3. Select Viral Pathogens

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of replication, translation of late genes by host RNA polymerase II is initiated, leading to the production of structural proteins and proteins required for virion assembly. Encapsidation occurs in the nucleus and progeny virions are released by lysis of the host cell.

**Epidemiology.** Adenoviruses are endemic and ubiquitous, causing an estimated 2–5% of all respiratory infections. Distribution is worldwide and incidence higher in the first half of the year. Children are frequently infected as serological studies show that by the age of 5 years some 50% present antibodies towards the most common species, including *Human adenovirus C*. On the order of 1 in every  $10^7$  lymphoid cells in the oropharynx harbor fully assembled latent state virions, making most humans latent carriers. Transmission can both occur via respiratory droplet or fecal-oral routes and virions are very stable towards chemical and physical agents, surviving for long periods outside their host.

Adenovirus outbreaks are a common cause of pneumonia in militaries, leading to the development of live, oral vaccines in the 1960's by the U.S. Army. The only supplier however ceased production and as of 1999, vaccinations could no longer be administered, resulting in reemerging incidence. In the first 5 years after loss of the vaccine, 110000 cases of febrile respiratory illness were reported, of which an estimated 90% are considered preventable. By October 2011, new vaccine again was available and has been administered to new recruits since.

### 2.3.2 Rhinoviruses

Investigations into the etiological agent of the common cold within the British Common Cold Research Unit led to the discovery of rhinoviruses in 1953. Initially classified as a separate genus of the family *Picornaviridae*, recent genomic evidence led to a revised taxonomy, recognizing three species of rhinoviruses (A through C) as members of the genus *Enterovirus*. Over 100 distinct serotypes have been isolated from humans, 74 belong to species A, 25 to B and the newly identified species C, currently under active study, may encompass an additional 55 types.

Rhinovirus virions are small (30 nm in diameter), non-enveloped, with a pseudo  $T = 3$  icosahedral capsid, consisting of 4 different polypeptides (VP1, VP2, VP3 and VP4) surrounding the RNA genome. There are 60 copies of each structural protein and VP1-3 face towards the outside and are responsible for antigenic diversity, while VP4 faces inwards and anchors the RNA core to the capsid. A canyon formed by VP1 and VP3 serves as receptor binding site. The viral genome consists of monopartite, linear, single stranded, positive sense RNA,

## 2. BIOLOGICAL BACKGROUND

roughly 7.2 kb in length and encodes a single polyprotein, which cleaved by viral proteases yields 11 proteins. Instead of a methylated 5' cap, the RNA genome is terminated by a viral protein (VPg) at its 5' end ([Jacobs et al., 2013](#)).

**Diseases.** Over half and up to two thirds of all cold-like illnesses are thought to be caused by rhinoviruses. In addition, asymptomatic infections, especially in children are very common with rates among children under 4 years ranging from 12 to 32%. These surprisingly high numbers may to some extent result from virus persistence in hosts that have recovered in addition to disease that has not broken out. In adults, rates of asymptomatic carriage are significantly lower, reported at 0–2%.

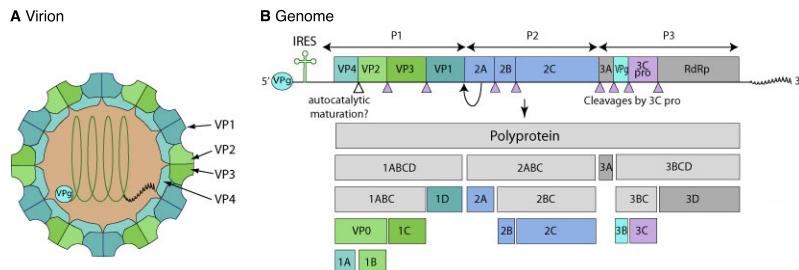
In immunocompetent individuals, symptomatic disease typically manifests as upper respiratory infection with clinical syndromes associated with common cold, including rhinorrhea, nasal congestion, sore throat, headache and possibly fever and general malaise. Disease is self-limited, incubation periods are between 12 and 72 h and within 7 to 14 days, symptoms wear off. A common complication is acute otitis media, which has been reported to happen in up to 30% of cases in early childhood. In 41–45% of investigated middle ear infections, rhinoviruses were detected. Further cavities that are frequently infected are the paranasal sinuses. Nose blowing has been suggested as mechanism for spreading the virus and causing rhinosinusitis.

Initially thought to primarily cause benign upper respiratory infections, recent data clearly implicates rhinoviruses in more severe lower respiratory infections. Croup, bronchiolitis and community-acquired pneumonia (CAP) have been associated with rhinovirus infections and studies have shown that in 12, 14 and 18–26% of respective cases, rhinoviruses were present. Furthermore, a study of children admitted to intensive care units (ICUs) with lower respiratory tract infections detected no other pathogens in 49% of the patients. Immunocompromised individuals are predisposed to contracting more serious forms of disease, with morbidity and mortality comparable to that of pandemic H1N1 influenza.

While not typically associated with cytopathogenic effects on epithelia of the upper respiratory tract, cell damage to lung tissue, especially among children, has been documented. Thus, rhinoviruses are linked to exacerbations of chronic pulmonary diseases such as asthma, chronic obstructive pulmonary disease and cystic fibrosis.

**Pathogenesis.** Members of rhinovirus species A and B are divided into two group according to their host cell receptors. Members of the major group form interactions with ICAM-1 while minor group types (including serotype 1a) as-

## 2.3. Select Viral Pathogens



**Figure 2.10:** Capsid proteins VP1 through VP4 form a pseudo  $T = 3$  icosahedral coat, roughly 30 nm in diameter around the RNA genome (A), which is monopartite, linear, 7.2 kp long and encodes 11 proteins (B). Figures adapted from [Hulo et al. \(2011\)](#).

sociate with very low-density lipoprotein receptors (VLDLRs). Attachment of species C have very recently been identified as induced by cadherin-related family member 3 (CDHR3). Upon receptor mediated endocytosis, pH dependent conformational changes in capsid structure exposes VP4 which has pore-forming properties and facilitates release of viral genomic material into the cytosol.

Host cell ribosomes readily translate the released positive-sense RNA into polyprotein, which is cleaved in *cis* by 2A and 3Cpro, yielding preproteins P1, P2 and P3 (see figure 2.10). P1 is digested into structural capsid proteins while P2 and P3 are further processed to produce the replication machinery. Viral RNA-dependent RNA polymerase synthesizes a complementary, negative-sense RNA strand, primed by VPg, which in turn serves as template for many copies of the viral genome. These can be both translated into more protein and in a later stage of replication also be packaged into progeny virions. Final cell export is mediated by host-cell membrane lysis.

**Epidemiology.** Despite most infections with rhinoviruses only resulting in mild disease, economic impact both due to health care costs and loss of productivity is considerable. This is primarily owed to the overwhelming prevalence of these pathogens. Respiratory illnesses attributed to rhinoviral infections occur throughout the world and all year round with peak incidences in early fall and spring. Vaccination efforts so far have been futile, mainly because of the large number of serotypes and lack of individual epidemiological data.

Due to acid-sensitivity, fecal-oral infection is unlikely most person-to-person transmission occurs through aerosols and contact spread either direct or via fomites. Extra-host survival times of hours to days have been reported for indoor environments and up to 2 h on undisturbed skin.

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### **2.3.3 Vaccinia**

*Vaccinia virus* is a species within the genus *Orthopoxvirus*, alongside the exceptionally virulent *Variola virus*, the etiological agent of smallpox. Immunological similarities between the two species allows for cross-protection, which coupled with the large discrepancy in pathogenicity presents a fortunate opportunity for artificially inducing acquired immunity. This was recognized by Jenner in 1798, while investigating the zoonosis of *Cowpox virus* and subsequent susceptibility towards contraction of smallpox. The origins of vaccinia are unknown. It has been speculated to have derived from cowpox or smallpox, to be a hybrid of both and of being the prototype orthopoxvirus, as well as descending from an extinct ancestor.

The virions are large and brick shaped, measuring 200 by 250 by 340 nm and exist as both mature virion (MV) and extracellular virion (EV). Structurally they are unusually complex. The linear, double-stranded DNA genome, roughly 200 kb long, is encased in an S-shaped, tube-like nucleocapsid with an outside diameter of 50 nm, a cavity of 10 nm and an overall length of 250 nm. Additionally, 47 viral proteins, of which 16 are involved in early mRNA synthesis and 28 have no known enzymatic function, are packed into a core structure. The core wall consists of two layers, the palisade (outer) layer which is 17 nm thick and an inner smooth layer, measuring 8 nm across. Centered on the top and bottom faces, two proteinaceous lateral bodies separate core from the surface membrane, forming the virion core into a biconcave disc with dumbbell shaped vertical cross sections. The lipidic surface membrane also consist of two layers, the outermost measuring 9 nm and the innermost domain is 5 nm thick. EV virions are additionally wrapped in a membrane derived from Golgi cisternae ([Marennikova et al., 2005](#); [Condit et al., 2006](#)).

**Diseases.** While infection with variola causes severe disease manifesting in skin lesions covering the whole body, accompanied by 20–40% mortality rates, the closely related *Vaccinia virus* is far less invasive. Virulence of the latter pathogen is so low that it has been routinely used as live vaccine against the former. Owing to the massive effort put into the worldwide fight against smallpox led by the WHO in the late 1960's and throughout the 1970's, the disease was found to be eliminated by 1980. At the center of the smallpox eradication program was the administration of freeze-dried, calf lymph derived vaccinia with a bifurcated needle, by multiple pricking of the skin. Towards the end of the initiative, 200 million vaccinations were performed annually.

The predominant reaction to vaccinia inoculation is localized, self-limited disease. After an incubation period of 3–4 days, a papule with a sunken center develops at the site of infection, accompanied by erythema and swelling. Over

## 2.3. Select Viral Pathogens

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the following days the papule increases in size and liquid begins to accumulate within. Fever may develop around days 7–10, possibly followed by malaise, headache and anorexia. Local lymphadenopathy is frequently encountered and days 8–10 typically mark the beginning of involution of the papule, which subsequently dries out and forms a scab.

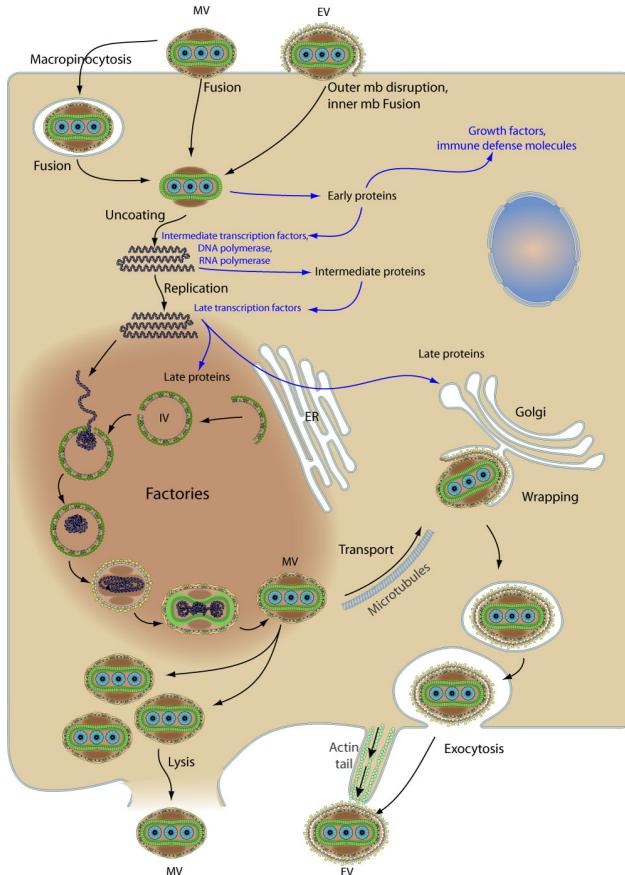
Of great concern for routine vaccination procedures are complications which inevitably occur in a small numbers. Atypical side effects develop in roughly 1 per mill of cases and potentially life threatening reactions usually manifest as neurological (477.4 cases and 47.0 fatalities per 1 million) or cutaneous disease (278.4 cases and 0.2 fatalities per 1 million). Predisposing conditions for central nervous system involvement are not known and despite decades of inquiry into this pathology, it remains poorly understood. Symptoms range from febrile seizures to severe encephalitis, but so far no neuropathological characteristics have been identified. Complications affecting the skin and mucous membranes are classified as progressive vaccinia, eczema vaccinatum and generalized vaccinia and disease severity decreases in this order. Predisposing conditions for progressive and generalized vaccinia are immunodeficiencies while a history of eczema is a risk factor for eczema vaccinatum.

**Pathogenesis.** Initial attachment is mediated by interaction between viral proteins and cellular heparan sulfate chains. For cell entry, various strategies have been reported, depending on the strain under investigation. WR strains induce macropinocytosis and proteins A25/A26 act as fusion suppressors that only lift their embargo under acidic conditions encountered in a maturing endosome, while other strains, such as Copenhagen, present no A25/A26 on their outer membrane and fuse directly with the target plasma membrane. Due to the additional membrane of EVs, a differing entry mechanism needs to be employed. In a currently not well understood fashion, the outer membrane is lost by non-fusogenic disruption, followed by fusion of the inner virion membrane. All pathways lead to cytosolic localization of virions devoid of their envelope (c.f. figure 2.11).

Members of the *Poxviridae* family are special among Baltimore group I viruses in that their genome encodes all necessary replicatory proteins, allowing for cytosolic localization. Replication is temporally tightly regulated and consists of distinct phases of early, intermediate and late gene expression. Each class of genes encodes factors capable of initiating the succeeding stage, providing transcription level regulation. Uncoating of the core structure releases early proteins, including RNA polymerases and enzymes for mRNA processing which start transcribing early genes. At least 50 different products, such as DNA replicatory enzymes, additional RNA polymerase, mRNA processing machinery, host defense molecules and intermediate gene transcription factors, have

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**Figure 2.11:** Replication cycle of *Vaccinia* viruses for both mature virion (MV) and extracellular virion (EV). Cell entry is either macropinocytic or proceeds via membrane fusion, followed by uncoating and replication. Host exit proceeds by either lytic or exocytic mechanisms. ([Hulo et al., 2011](#)).

been identified and account for 25% of the viral coding capacity. Early gene transcripts are detectable 20 minutes after cell entry and reach their productive peak within 100 minutes of infection.

Expression of intermediate genes is initiated by accumulation of intermediate transcription factors and the onset of DNA replication. Only 7 products of this

## 2.4. RNA Interference

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phase are known, which functionally are mostly concerned with host defense, DNA/RNA metabolism and commencement of the final phase. Beginning 100 minutes after infection, intermediate gene transcription reaches its peak at 120 minutes and is thereafter superseded by late gene transcription, beginning 140 minutes after cell entry. Products of the final phase comprise a large number of genes (up to 75% of the vaccinia genome) and include enzymes needed for initiating replication (RNA polymerase and modification proteins), early transcription factors and structural proteins, as well as virion assembly machinery.

DNA replication not only serves for progeny virions, but also to increase the concentration of templates used for gene expression. Both DNA synthesis and virion assembly occur within factories, readily visualized by electron microscopy as electron dense cytoplasmic inclusion bodies. Owing to the complex virion structure DNA packaging and virion assembly is an involved procedure with is incompletely understood.

**Epidemiology.** It is unknown if a natural reservoir of vaccinia exists. It has been speculated that the virus is maintained only within research laboratories and vaccination production facilities, while others have implicated some rodent species as possible reservoir hosts. Small scale zoonotic outbreaks of vaccinia have been documented in Brazil and it was initially suspected that these were linked to vaccine that had escaped into the environment. Recent phylogenetic studies however were able to rule out this explanation but were unable to shed further light into underlying epidemiological mechanisms.

While transmission from vaccinees to unvaccinated individuals is rare, direct contact transmission is possible and such occurrences have been documented. Special care has to be taken to avoid direct contact between recently vaccinated and individuals predisposed towards developing complications.

## 2.4 RNA Interference

First described only two decades ago, regulation of gene expression by short strands of RNA has become an indispensable tool to both experimental biology and bioinformatics. Recognizing the importance of applications made possible by this discovery, the 2006 Nobel prize in Physiology or Medicine was awarded to Fire and Mello who studied RNA interference in the nematode worm *Caenorhabditis elegans* and published their findings in 1998. Building on studies by Guo and Kemphues, who showed that sense RNA, as well as antisense RNA was capable of suppressing gene expression, Fire, Mello and coworkers found that double stranded RNA was at least ten-fold more effective as silencing agent than individual single stranded fragments. Further investi-

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gations showed that several gene regulatory processes, previously thought to be unrelated, were in fact manifestations of RNA interference and that the underlying mechanism was conserved in many if not most eukaryotic organisms ([Hannon, 2002](#)).

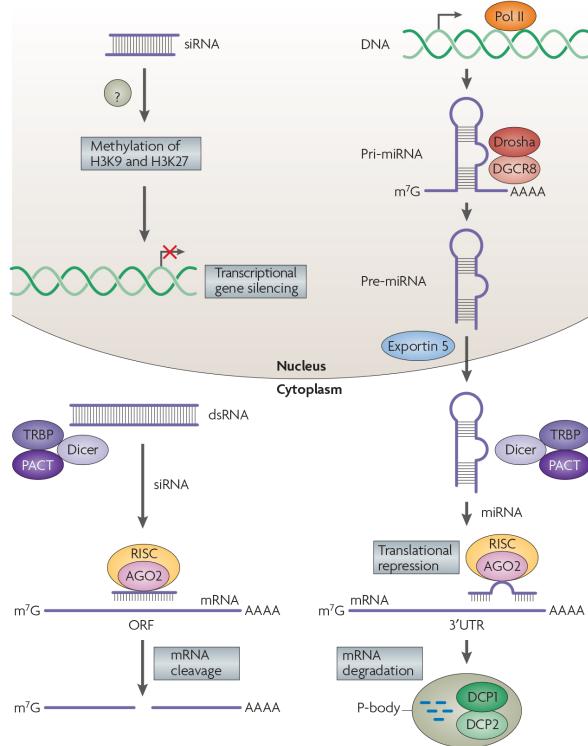
The RNAi pathway can take as input two separate types of RNA molecules, miRNA and siRNA, of differing origins. While miRNAs are endogenous and purposively employed in post-transcriptional regulation of gene expression, siRNAs are exogenous synthetic or viral inducers of gene suppression, in which case, RNA interference can be viewed as an immune response to foreign genetic material. Parsimony-based phylogenetic analysis of involved genes suggests that the key components to an RNAi system were already present in the last common ancestor of eukaryotes and were subsequently lost or extensively simplified in some protists. The original function of RNAi is hypothesized to be that of a defense mechanism against genomic parasites as indicated by the extent of its conservation, whereas miRNA-directed silencing most probably was introduced at a later point in evolution ([Cerutti and Casas-Mollano, 2006](#)).

### **2.4.1 Molecular Mechanism**

RNA interference refers to three separate mechanisms for regulation of gene expression by small RNAs, as visualized by figure [2.12](#). While siRNAs are involved both in transcriptional and post-transcriptional gene silencing, the miRNA pathway is focused on translational repression. The severity of action on the targeted genes once again highlights the differing purposes of the siRNA and miRNA pathways, one tasked with inhibitory and the other with regulatory measures ([Wilson and Doudna, 2013](#); [Kim and Rossi, 2007](#); [Carthew and Sontheimer, 2009](#)).

**Translational repression by miRNA.** The biogenesis of miRNA occurs in the nucleus and is initiated by RNA polymerase II transcription of long (>1000 nt) primary miRNA (pri-miRNA) segments, characterized by double-stranded hairpin loops with single-stranded 5'- and 3'-terminal overhangs which are polyadenylated and capped. Subsequent processing by the microprocessor complex consisting of the RNase III family enzyme Drosha and DGCR8 (DiGeorge syndrome critical region gene 8) yields ~60–70 nt stem-loop structured precursor miRNA (pre-miRNA) fragments. DGCR8 recognizes pri-miRNAs by the junction of stem and single-stranded overhang and helps positioning the substrate for endonucleolytic cleavage by Drosha at a site ~11 nt from the junction. Nuclear export is mediated by the transport facilitator exportin-5 and is Ran-GTP dependent.

## 2.4. RNA Interference



**Figure 2.12:** RNA interference comprises of three distinct mechanisms that yield control over gene expression. Exogenous double-stranded RNA are processed into siRNA fragments that both act inside the nucleus as transcriptional silencing agents and in the cytoplasm, post-transcriptionally cleaving mRNA strands. Endogenous miRNA is synthesized by RNA polymerase, originates from the nucleus in processed form and mediates milder translational repression (Kim and Rossi, 2007).

In the cytoplasm, pri-miRNAs are targeted by Dicer and the associated dsRNA binding proteins TRBP (TRA RNA-binding protein) and PACT (protein activator of protein kinase PKR) for further processing into 21–25 nt dsRNA strands with 2 nt overhangs at the 3' termini and phosphate groups at each of the recessed 5' ends. The mature miRNAs are loaded onto Argonaute (Ago) by Dicer, which leads to the formation of RNA-induced silencing complex (RISC). Concomitantly with RISC-loading, one of the two RNA stands is selected as guide strand whereas its complement (the passenger strand) is ejected and degraded. Thermodynamic asymmetry between the two strands is exploited in this step and the strand with the less stable 5' end is preferred. As opposed to strand

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separation in siRNAs, the passenger strand is not cleaved but rather unwound by helicase activity, facilitated by imperfect sequence alignment.

Finally, active RISC, exposing the Argonaute-bound guide strand, interacts with the 3' untranslated region of mRNA targets and directs translational repression and mRNA degradation. Sequence homology between mRNA and the miRNA seed sequence (the first 2–6 or 2–8 nt from the 5' end) is critical while mismatched nucleotides towards the 3' end of the miRNA are readily tolerated. The extent of base-pairing influences the subsequent mechanism of silencing, ranging from direct target cleavage (perfect match) over deadenylation (followed by degradation), to nonendonucleolytic translational repression (imperfect match).

**Post-transcriptional gene silencing by siRNA.** Precursors to siRNA are long, linear, perfectly base-paired double stranded sequences of RNA, typically of exogenous origin either introduced directly into the cytoplasm, or taken up from the environment. A complex consisting of Dicer and several RNA-binding proteins are responsible for trimming down dsRNA fragments to the appropriate size for loading onto Ago2. Of the four Argonaute family members in humans, capable of associating with miRNA, only Ago2 seems to be involved with siRNA. Furthermore, Ago2 is the only mammalian Argonaute bearing endonucleolytic functionality and therefore capable of directly cleaving targeted mRNA.

Strand selection is again based on differences in stability of base-pairing at the 5' termini with the weaker interacting end being favored as guide strand. Accuracy of discrimination can be low, leading to incorporation of both strands with equal frequency. In contrast to miRNA loading, the passenger strand is not merely separated but directly cleaved by Ago2 and the differing treatment seems to only depend on perfect strand complementarity given in siRNA and absent in miRNA. Upon RNA incorporation, RISC is formed and activated by cleavage of the passenger strand. The 3' guide strand end is bound by the Argonaute's PIWI-Argonaute-Zwille domain (PAZ), while the 5' end interacts with Argonaute middle domain (MID), closely located to the catalytic RNase H-like P-element-induced whimpy testes domain (PIWI).

Post-transcriptional gene silencing is accomplished by endonucleolysis of perfectly matching mRNA precisely at the phosphodiester linkage between bases 10 and 11 relative to the 5' terminus of the siRNA guide strand. Following cleavage, the target dissociates, freeing RISC for further catalysis, and the mRNA fragments are degraded by cellular exonucleases. Imperfectly matched mRNA may be targeted, much like it is the case with miRNA, leading to siRNA off-target effects which are of great practical importance.

**Transcriptional gene silencing by siRNA.** In addition to post transcriptional action of siRNA, nuclear inhibition of gene transcription has been described in many eukaryotes. Diced siRNA fragments are transported into the nucleus where they are assembled with a group of proteins, including Ago1, to form RNA-induced transcriptional silencing complex (RITS). Currently only incompletely understood, the siRNA guide strand is thought to recognize RNA transcripts as they emerge from RNA polymerase II, followed by recruitment of factors that enable covalent modifications of nearby histones. Methylation of lysines 9 and 27 in H3 by histone methyltransferases leads to chromatin compaction and heterochromatin formation. RITS has also been shown to induce direct methylation of DNA, repressing gene expression even further.

Contributing to the potency of RNA interference, engagement of RITS with nascent transcripts activates the RNA-dependent RNA polymerase complex (RDRC), capable of generating secondary siRNA fragments and therefore amplifying silencing capabilities. The role of this reinforcement mechanism has been firmly established in many eukaryotic RNAi systems with the notable exceptions of vertebrates and insects. Whether a similar system exists in these organisms remains an open question.

#### 2.4.2 Biological Function

The mechanisms of RNA interference have most probably evolved in order to protect against foreign genetic material such as parasitic DNA sequences or viral RNA. Transposable elements (transposons) are DNA sequences that are mobile within the genome, can make up a significant fraction of eukaryotic genomes and are typically considered non-coding. Transposition is mediated by transposases, enzymes often encoded within the transposons themselves, that act on specific sequences at the transposon ends and cause unspecific insertion into new target sites. Retrotransposons move via an RNA intermediate which is reverse transcribed to DNA and inserted, while DNA transposons employ a cut and paste mechanism. Retroviruses therefore can be viewed as transposons and in general, transposable elements are a form of selfish DNA that often incur deleterious effects.

RNAi is an important regulatory force to transposon activity, both by processing transcripts of retrotransposons, thereby reducing their concentration and eliciting epigenetic modifications, as well as transcriptional inhibition via heterochromatin formation. The importance of keeping transposable elements in check is highlighted by their prevalence, with roughly half of the human genome being thought to derive thereof.

Antiviral mechanisms are particularly important in organisms lacking an adap-

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tive immune system as found in vertebrates and exploiting the orthogonality of most genomic systems to double-stranded RNA puts RNA interference in a powerful position. Corroborating this notion is the observation that, in *Drosophila melanogaster*, three key proteins of the RNAi pathway (Dicer-2, Ago2 and R2D2, a protein involved in RISC loading) are among the top 3% in terms of genetic instability. Furthermore, miRNA pathway paralogs to these three genes (Dicer-1, Ago1 and R3D1), not being involved in immune response, evolve at a much slower rate ([Obbard et al., 2009](#)).

Although small RNA-guided, Ago-dependent up-regulation of gene expression (termed RNA activation or RNAa) has been described, most regulation of gene expression by miRNA is of inhibitory nature. This widespread mechanism, consisting of >1000 miRNA sequences (as much as 5% of the human genome) controls at least 30% of human genes and is responsible for vital processes including cell growth, tissue differentiation and cell proliferation.

### **2.4.3 Applications**

In *C. elegans*, RNA interference is especially powerful, making it a popular model organism for RNAi. Not only is delivery efficiently possible simply by feeding the nematodes with bacteria such as *E. coli* that carry the desired dsRNA, but the resulting gene silencing effects are hereditary. Moreover, RNAi response is not stoichiometric but catalytic, is amplified in a feedback loop and in many organisms, systemic spread has been documented.

The initial burst of excitement surrounding possible applications was somewhat moderated by difficulties in applying RNAi to mammalian systems. At first it seemed impossible to use this technology in somatic cells as the introduction of dsRNA is typically met with overwhelming non-specific responses, including RNA-dependent protein kinase pathway (PKR), which leads to arrest of translation and apoptosis. This issue was shown to be overcome by exclusively using siRNAs duplexes of 21–23 nt with 2-nt 3' overhangs that mimic Dicer products and are too short for inducing PKR. Mammalian RNAi response, however is transient, lacking amplification and spreading mechanisms documented in other organisms (mainly plants and worms) and delivery, especially *in vivo* remains problematic. A further issue that continues to be an actively researched area of interest is that of off-target effects (OTE), which considerably complicate the interpretation of phenotypic data.

**Gene knockdown studies.** Large-scale loss-of-function (LOF) and modifier or synthetic lethality screens are readily possible by means of RNAi based high throughput screening (HTS). Such experiments usually proceed by arraying libraries of gene specific siRNAs onto microtiter plates (96 and 384 well for-

## 2.4. RNA Interference

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mats are common), followed by the addition of liquid cell cultures. After an appropriate transfection time, the cells may be subjected to an additional treatment, such as exposure to drugs or pathogens (modifier screen) or LOF phenotypes can be investigated directly. Assay readout is performed via optical measurements such as detection of fluorescence or luminescence signals or by microscopic imaging (confocal or wide-field).

Transcriptional reporters, fluorescent dyes that detect enzymatic activity and protein-modification specific antibodies have been employed in plate reader-based investigations which yield a single numerical readout per well. This quantitative approach is contrasted with microscopy based assay read-outs that are able to capture spatial information on antibody stained proteins, fluorescently labeled cellular structures and green fluorescent protein (GFP) expression, yielding much more data per well. Significant challenges incurred by automated high-content imaging have successfully been addressed by computational image analysis software.

A multitude of sources of technical and biological noise contribute to serious problems in interpretability and comparability of observed data. Common to all high throughput approaches, errors arise from difficulties guaranteeing equal conditions in a large number of parallel experiments. Liquid dispensing errors, temporal disparities caused by bottleneck resources such as imaging equipment and spatial discrepancies, for example inhomogeneous temperature distribution over the plate or liquid evaporation in border wells, are only a few issues that come to mind. Biological sources of error include OTEs, varying potency of reagents (both the knockdown strength and time required to achieve optimal knockdown may be affected) and obscuration of assay phenotype by the knockdown phenotype (e.g. cell death). Furthermore, incorrect gene models lead to errors in library design and detection may be hampered by weak phenotypes. Replicates, although expensive in large-scale experiments and control wells embedded in every assay plate are indispensable measures in order to assess reproducibility of the data ([Echeverri and Perrimon, 2006](#); [Perrimon and Mathey-Prevot, 2007](#)).

Despite being a young technology, RNA interference has already proven itself as an invaluable tool and has yielded many insights with significant impact on various fields. A review by [Mohr et al. \(2010\)](#) lists some of these findings which lead to refined understanding of cell proliferations, cancer biology, cell cycle regulation, mitochondrial diseases, signal transduction, RNA biology and pathogen response.

**Biotechnological applications.** Intercellular, systemic spread of RNAi response in plants and even its heredity over several generations have been documented

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**Table 2.3:** A non exhaustive list of RNAi based drugs that currently are in clinical trials. The data was obtained from the clinicaltrials.gov database [McCray and Ide \(2000\)](#)

Company	Disease	Delivery system	Status
Alnylam	(TTR)-Mediated Amyloidosis	siRNA-GalNAc conjugate	Phase III recruiting
Alnylam	Antitrypsin Deficiency Liver Disease	Liposome	Phase II recruiting
Alnylam	Acute Intermittent Porphyria	siRNA-GalNAc conjugate	Phase I recruiting
Silenseed	Advanced pancreatic cancer	Polymer (LODER)	Phase III planned
Sylentis	Dry eye syndrome	Naked siRNA	Phase II completed
Sylentis	Open angle glaucoma	Naked siRNA	Phase II recruiting
Tacere	Chronic hepatitis C	adeno-associated virus (AAV) vector	Phase II recruiting
Tekmira	Advanced Hepatocellular Carcinoma	Liposome	Phase II recruiting

and it comes as no surprise that the technology is investigated for possible utilization in crop improvement. Removal of plant endotoxins by targeting genes of toxin biosynthesis has been accomplished, leading to the production of decaffeinated coffee plants (knockdown of theobromine synthase), tobacco with reduced concentration of carcinogenic compounds (inhibition of nicotine demethylase activity) and edible cotton seeds (reduction of delta-cadinene synthase leads to low levels of gossypol, a toxic terpenoid), which are naturally rich in dietary protein.

In addition to investigations with consumer health in mind, improvements in environmental resistance have been studied in many organisms. Susceptibility to bacterial and viral pathogen infection has been reduced in rice, bean, barley and lemon, while fungal resistance has been increased in potato, tobacco and wheat. Successful RNAi application as insecticide has been shown in cotton and maize and even improved resistance to parasitic weeds could be demonstrated in transgenic tomato plants. Despite these achievements, concerns over environmental issues and adverse health effects have so far prevented RNAi based genetic modifications from exiting experimental stages ([Saurabh et al., 2014](#)).

**Therapeutic potential.** Great promise lies in therapeutic application of RNAi as theoretically, any gene can be targeted, yielding unparalleled flexibility not

## 2.4. RNA Interference

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encountered with typical small molecule drugs. An initial obstacle to harnessing this power in humans is the issue of delivery. Systemic spread of naked siRNAs is hampered by kidney filtration, phagocytic uptake and degradation by serum RNases. Movement across capillary walls is not readily possible for molecules larger than 5 nm and phagocytes patrol the extracellular matrix, ingesting foreign genetic material. Furthermore, polyanionic macromolecules do not easily penetrate hydrophobic cellular membranes.

Topical or local administration offers advantages, including increased bioavailability and reduced side effects and is therefore preferred for treatment of eye, skin and mucosal diseases, as well as localized tumors. If targets are not localized or difficult to reach, injection into the bloodstream may provide a mode of systemic application. Chemical modification of the RNA backbone (2'-O-methylation or 2'-fluorination of ribose) has been shown to provide resistance to RNase and covalent attachment to cholesterol promotes cellular uptake. Encapsulation of siRNA in liposomes and cationic polymers are further proven techniques for improving extracellular stability, stimulate endocytosis and facilitate endosomal escape.

Some sort of target selectivity is desirable on order to avoid high dosages, associated concerns of toxicity and potential OTEs in non-target tissue. Coupling of siRNA reagents to antibodies specific for HIV envelope glycoproteins has been successfully employed for selectively entering infected cells, while aptamers (oligonucleotides specifically engineered for binding a given target), carrying siRNAs have also been shown to be capable homing mechanisms.

Viral delivery of of RNAi inducing agents presents an alternative technique to the above and in case of retroviral transport vessels, short hairpin RNAs (shRNAs) that are reverse transcribed and integrated into the host genome, provide stable expression and prolonged RNAi activity. Adenoviral and AAV vectors have also been employed, resulting in a more transient response. While health concerns associated with perpetuity of gene therapy no longer apply, repeated administrations may prove problematic, triggering strong immune response and thereby limiting therapeutic potential.

Currently, multiple siRNA based therapeutics are in clinical trials, including stage III studies (see table 2.3). Due to the unprecedented pace at which RNAi technology went from discovery to development of applications, much uncertainty remains surrounding long term effects of exposure to such drugs. Chronic diseases including hepatitis C or HIV infections require life-long treatments and consequences of repetitively triggering RNAi response has not been thoroughly studied (unwanted changes to chromatin structure for example, are one area of concern). Apart from safety issues, several implementation aspects require further study. While neurodegenerative diseases have successfully been

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treated in mouse models via direct injection into the brain, this is not easily feasible in human patients and currently no delivery vehicle capable of crossing the blood-brain barrier. ([Kim and Rossi, 2007](#); [Whitehead et al., 2009](#)),

## Chapter 3

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# Mathematical Background

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Modeling the relationship among variables is one of the most important applications of statistical theory. The study of regression analysis (and the closely related notion of correlation) started to form towards the end of the 19th century with Sir Francis Galton's study of height heredity in humans and his observation of regression towards the mean. Over the next few years, Udny Yule and Karl Pearson cast the developed concepts into precise mathematical formulation, in turn building on work performed by Adrien-Marie Legendre and Carl Friedrich Gauss who developed the method of least squares almost a century earlier ([Allen, 1997](#)).

A multiple linear regression model can be written in matrix-vector form as

$$y = X\beta + \varepsilon \quad (3.1)$$

where  $y \in \mathbb{R}^n$  is the vector of observations on the dependent variable, the design matrix  $X \in \mathbb{R}^{n \times p}$  contains data on the independent variables,  $\beta \in \mathbb{R}^p$  is the  $p$ -dimensional parameter vector and the error term  $\varepsilon \in \mathbb{R}^n$  captures effects not modeled by the regressors. Without loss of generality, all variables are assumed to be expressed as deviations from their means and measured on the same scale.

In order to find unknown coefficients  $\beta_i$ , the ordinary least squares estimator minimizes the residual sum of squares, the squared differences between observed responses and their predictions according to the linear model.

$$\hat{\beta} = \arg \min_{\beta} \|y - X\beta\|^2 \quad (3.2a)$$

$$= (X^T X)^{-1} X^T y \quad (3.2b)$$

### 3. MATHEMATICAL BACKGROUND

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Some assumptions are typically associated with linear regression models that yield desirable attributes for the estimates. None of these restrictions are imposed on the explanatory variables; they can be continuous or discrete and combined as well as transformed arbitrarily. Furthermore, in practice, it is irrelevant whether the covariates are treated as random variables or as deterministic constants. With exception of the field of econometrics it appears that the majority of literature adheres to the latter interpretation and therefore, statements will not explicitly be conditional on covariate values.

**Linearity.** The relationship between dependent and independent variables should be linear (after suitable transformations) and individual effects additive. If this cannot be satisfied, a linear model is not suitable.

**Full rank.** For the matrix  $X^T X$  to be invertible, it has to have full rank  $p$ . Therefore  $n \leq p$  and all covariates must be linearly independent.

**Exogeneity.** All independent variables should be known exactly i.e. contain no measurement or observation errors as only the mean squared error of the dependent variable is minimized. Additionally, all important causal factors have to be included in the model. Exogeneity implies  $E[\varepsilon_i] = 0 \ \forall i$ , as well no correlation between regressors and error terms (Hayashi, 2000).

**Spherical errors.** This includes both homoscedasticity or constant error variance:  $E[\varepsilon_i^2] = \sigma^2 \ \forall i$  and uncorrelated errors  $E[\varepsilon_i \varepsilon_j] = 0 \ \forall i \neq j$ . These two conditions can be written more compactly as  $\text{Var}[\varepsilon] = \sigma^2 I_{n \times n}$ .

**Normality.** For the estimated coefficients to gain some additional desirable characteristics, it can be required that the errors  $\varepsilon_i$  be jointly normally distributed. Together with the above restrictions on expectation and variance, this yields  $\varepsilon \sim \mathcal{N}_n(0, \sigma^2 I_{n \times n})$ .

Violations of these assumptions have varying consequences. In case of perfect multicollinearity, the ordinary least squares estimator  $\hat{\beta}$  as defined in (3.2b) does not exist. Recovering such a situation is possible by using a generalized matrix inverse (for example the Moore–Penrose pseudoinverse) or employing a regularization scheme such as ridge regression.

Omitting a variable that is both correlated with dependent variables and has an effect on the response (a nonzero true coefficient) will introduce bias in the parameters. The method of instrumental variables can help to produce an unbiased estimator.

The assumption of spherical errors ensures that the least squares estimator is the best linear unbiased estimator in the sense that it has minimal variance among all linear unbiased estimators. Heteroscedasticity and autocorrelation do not cause coefficient estimates to be biased but can introduce bias in OLS

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### **3.1. Generalized Linear Models**

estimates of variance, causing inaccurate standard errors. A generalized least squares estimator (for example weighted least squares)

#### **3.1 Generalized Linear Models**

#### **3.2 Multivariate Adaptive Regression Splines**

#### **3.3 B-Splines**



## Chapter 4

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# Data

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InfectX is an RTD project funded by SystemsX, the Swiss initiative for systems biology with the goal of studying and identifying components of the human infectome for a set of bacterial and viral pathogens. A central effort consists of generating kinase- and genome-wide siRNA screens for each of the investigated pathogens, capturing image data and performing image analysis. The acquired data is publicly available on the internet at [the InfectX website](#).

### 4.1 InfectX Workflow

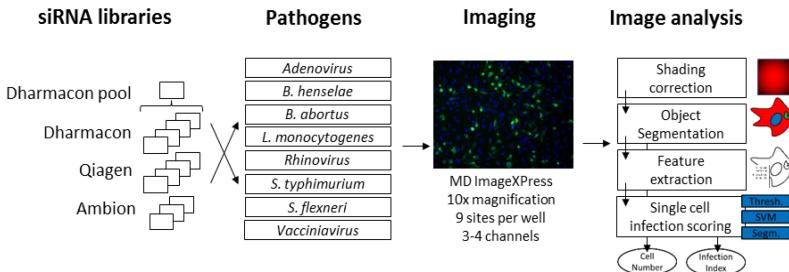
Due to the large scale accompanied by screening multiple libraries, using numerous pathogens and desiring replicates, several labs were involved in carrying out wet-lab experimentation. In order to obtain reproducible results, a strong emphasis was put on developing unified procedures for cell culture, siRNA transfection, pathogen infection and imaging. Figure 4.1 summarizes the central steps, beginning with siRNA libraries arrayed onto 384-well plates that are used for transfection of added cells, carrying on with pathogen infection, subsequent microscopic imaging and concluding with computational image analysis. Much of the information contained in this chapter has been published by [Rämö et al. \(2014\)](#) and is summarized for the reader's convenience.

The model system chosen for investigation is HeLa (ATCC CCL-2), the oldest and most wide spread human cell line and a proven system for studying infelicitous disease<sup>1</sup>. Culturing was performed at 37 °C, under 5% CO<sub>2</sub> atmosphere

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<sup>1</sup>Collected 60 years ago from a cervical adenocarcinoma, these epithelial cells have led to much

## 4. DATA



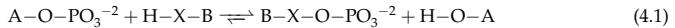
**Figure 4.1:** A total of 11 single siRNA libraries, produced by 3 separate manufacturers, (4 from Dhamacon, 4 from Qiagen and 3 from Ambion), as well as one pooled library (Dhamacon) were screened with 8 pathogens. Plates were imaged under wide-field microscopes and the resulting images run through an image analysis pipeline. ([Rämö et al., 2014](#)).

for maintaining optimal pH, using Dulbecco Modified Eagle Medium (DMEM) supplemented with 10% inactivated fetal bovine serum (FBS), both supplied by Invitrogen.

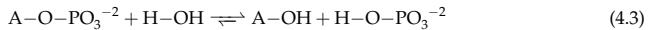
While genome-wide siRNA screens were also produced within the InfectX framework, this report focuses on kinase-wide investigations. Introduced by [Manning et al. \(2002\)](#), the term kinase refers to the subset of genes encoding protein kinases<sup>2</sup>. As phosphorylation reactions have been identified to constitute the most widespread signaling mechanism in eukaryotic cells, the set of 518 genes identified by [Manning et al.](#) are a popular target for functional genomics studies. Up to 30% of intracellular proteins may be phosphorylated,

insight into human cell biology. Prior to their discovery, attempts at growing human tissue in vitro were futile and development of protocols for sustaining cancerous human tissue was thought to hold great promise for cancer research. One of the early successes involving HeLa cells was the development a polio vaccine. For this endeavor, a large amount of human cells were needed and the installment of a production facility capable of meeting the high demand might have contributed significantly to the predominance of these cells. ([Masters, 2002](#))

<sup>2</sup>Kinases are part of the larger enzymatic family of transferases and catalyze phosphorylation reactions of the form



where A represents the donating (typically ATP) and B the accepting molecule. Kinases are further subdivided according to the type of acceptor which can be an alcohol, carboxy, nitrogenous or phosphate group, or in case of protein kinases, a tyrosine, serine, threonine or histidine residue.



Phosphorylases are a further group of transferases that involve phosphate but unlike kinases, they utilize inorganic phosphate sources (4.2). Often phosphorylases are involved in breaking down biological polymers such as polysaccharides and polynucleotides. Finally, phosphatases catalyze the reverse reaction, the removal of a phosphate group (4.3).

## 4.1. InfectX Workflow

**Table 4.1:** Number of replicates performed for each of the pathogens and siRNA libraries. The primary values indicate how many were performed in total while the value in parenthesis represents the number of screens that turned out to be unusable and had to be discarded. The effectively available number of replicates is the difference between the two. ([Rämö et al., 2014](#))

Pathogen	Dharmacon (1x pooled)	Ambion (3x single)	Quiagen (4x single)	Dharmacon (4x single)
<i>B. abortus</i>	8 (1 rem.)	2	1	2
<i>B. henselae</i>	5 (1 rem.)	4 (1 rem.)	2 (1 rem.)	1
<i>L. monocytogenes</i>	4	4 (2 rem.)	1	1
<i>S. flexneri</i>	6 (2 rem.)	2	1	1
<i>S. typhimurium</i>	7 (4 rem.)	3	1	1
adenovirus	8 (1 rem.)	2	1	1
rhinovirus	8 (2 rem.)	2	1	1
vaccinia virus	2 (1 rem.)	2	1	1

leading to 20000 phosphoprotein states, all regulated by expression of varyingly substrate specific kinases ([Johnson and Hunter, 2005](#)). Due to the importance of kinases for cell behavior, they represent a major drug target in cancer therapeutics and might therefore also be attractive for HDT.

In order to offset potential bias introduced by siRNA design paradigms employed by manufacturers, libraries from three different companies (Abion, Dharmacon and Qiagen) were obtained. To further account for the effect of OTEs, several siRNA sequences per target were used for both pooled and unpooled experiments. The Ambion Silencer Select Human Kinase siRNA Library V4 targets 710 genes of kinases and kinase-associated proteins with 3 sequences each, while the Qiagen HP OnGuard Human Kinase siRNA Set V4.1 comprises of 718 targets and 4 siRNAs per gene. The Dharmacon Human ON-TARGETplus siRNA Protein Kinase Libraries are designed with 715 genes in mind and are both available in 1 siRNA per well (unpooled) and 4 siRNAs per well (pooled) formats.

Depending on library and pathogen, screens were repeated 1–8 times (see table 4.1). The primary values denote the total number of replicated performed and the values in parenthesis indicate the number of screens that had to be removed due to issues with transfection, weak signal intensities or usage of a protocol other than the one eventually agreed upon. The Dharmacon pooled screen was used for optimizing the assay procedures which is why for almost all pathogens some replicates had to be removed. The number of available screens is the difference between primary and parenthesized values.

## 4. DATA

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**Table 4.2:** Despite putting much emphasis on using identical protocols throughout all screens, some assay parameters were fine-tuned in order to obtain phenotypes such as infection and cell counts that are similar among the investigated pathogens. ([Rämö et al., 2014](#))

Pathogen	Seeded cell number	MOI	Pathogen entry time	Total infection time <sup>c</sup>	DNA stain	Actin stain <sup>a</sup>	Pathogen stain	Additional stain <sup>b</sup>
<i>B. abortus</i>	500	10000	4 h	44 h	DAPI	DY-547	GFP	-
<i>B. henselae</i>	300	400	24 h	24 h	DAPI	DY-547	GFP	-
<i>L. monocytogenes</i>	600	25	1 h	5 h	DAPI	DY-647	GFP	Alexa546
<i>S. flexneri</i>	600	15	30 min	3.5 h	Hoechst	DY-495	DsRed	Alexa647
<i>S. typhimurium</i>	550	80	20 min	4 h	DAPI	DY-547	GFP	-
adenovirus	700	0.1	16 h	16 h	DAPI	DY-647	GFP	-
rhinovirus	1000	8	7 h	7 h	DAPI	DY-647	GFP	-
vaccinia virus	600	0.125	1 h/8 h <sup>c</sup>	24 h	Hoechst	DY-647	GFP/RFP	-

<sup>a</sup>Phalloidin-based actin stains were supplied by Dymomics and depending on absorption wavelength, different imaging channels were used: GFP for DY-495, RFP for DY-547 and Cy5 for DY-647.

<sup>b</sup>The Cy3 channel was used Alexa546, staining bacterially secreted InIC, while Cy5 was used for Alexa647, staining cellular IL-8 during imaging.

<sup>c</sup>The two values stand for primary and secondary infection times, respectively. The same goes for pathogen dyes.

## 4.2 RNA Interference Protocols

Central to the siRNA screens produced by InfectX was the effort to develop unified protocols for wet-tab experiments and subsequent analysis. While this approach was successfully implemented for many aspects, some deviations among the pathogens are inevitable, while others are intentionally developed to achieve similar phenotypes. Table 4.2 summarizes some key parameters that vary between pathogens, which include seeded cell number, multiplicity of infection (MOI) and infection times, all optimized to yield infection rates that are of comparable magnitude. The target value for cell number was 1500 per well in order to create densely populated areas interspersed with some empty spaces, leading to cells living on colony edges and infection rates of 30–50% were aimed for. Pathogen properties made it in some cases impossible to hit these goals and infection rate for *B. abortus* remained low (~5%), while being high in *B. henselae* (~90%) despite best efforts.

The usage of control wells enables diagnosis of possible problems that may occur.

## 4.2. RNA Interference Protocols

cur in RNAi screens, including cytotoxicity of siRNA, low transfection yields, failure of RNAi pathway induction, dominance of non-specific responses, and therefore should be embedded in every assay plate. Three types of control experiments are typically employed: positive, negative and mock (no siRNA treatment). Positive controls are used to confirm expected response, while negative controls help distinguish sequence specific from unspecific effects, and mock experiments help to establish a baseline ([Sittampalam et al., 2004](#)).

Positive controls ideally constitute of siRNAs with known effect on the assay under investigation and are therefore often unavailable beforehand. Instead, controls to check transfection efficiency and reporter quality are usually employed. One straightforward possibility for monitoring delivery is by targeting a gene that is vital to the cell. Kinesin family member 11 (KIF11), for example is a gene involved in cell cycle progression, the down-regulation of which induces apoptosis. Furthermore there are mixtures of siRNAs available (e.g. AllStars Hs Cell Death Control siRNA by Qiagen) optimized for killing cells by targeting several ubiquitously expressed genes essential for cell survival. The downside of assessing transfection by killing cells is that a potentially toxic effect of the delivery mechanism itself may be masked. This can be mitigated by either performing negative control experiments (which should be done anyways) or by targeting housekeeping genes that are abundantly expressed but do not affect cell viability. Dharmacon suggests three such genes, Peptidyl-prolyl cis-trans isomerase B (PPIB), glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and lamin A/C (LMNA), for which they sell specially branded control siRNAs (as does Ambion).

Fluorescent dyes are also frequently employed in positive control experiments, typically by labeling siRNA, allowing for visual inspection of reagent localization within the cell (nuclear uptake indicates efficient transfection), or by targeting reporter genes. The latter method either allows for confirming that the reporting mechanism (usually expression of GFP or luciferase) works as intended, or establishing that siRNA transfection was successful. Again, siRNA products targeting the commonly used reporter genes are readily available from manufacturers.

Negative controls should lack homology with known targets in order to separate non-specific effects from sequence specific silencing. Such siRNAs are therefore engineered to contain a passenger strand seed region (the first 2–6 nt from the 5' end) that matches no known gene and generally have poor overall sequence identity between passenger strand and any known gene. One way of generating such a sequence is taking an assay siRNA and randomizing the order of nucleotides while keeping the nucleotide composition unchanged (i.e. scrambling the sequence). Multiple proposals are usually generated and subse-

## 4. DATA

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**Table 4.3:** Depending on screen type and pathogen, different genes were targeted for control experiments. Abbreviations: AU, Ambion unpooled; DP, Dharmacon pooled; DU, Dharmacon unpooled; and QU, Qiagen unpooled.

	Adeno AU	Adeno DP	Adeno DU	Bartonella AU	Bartonella DP	Bartonella DU	Bartonella QU	Brucella AU	Brucella DP	Brucella DU	Brucella QU	Listeria AU	Listeria DP	Listeria DU	Listeria QU	Rhino AU	Rhino DP	Rhino DU	Rhino QU	Salmonella AU	Salmonella DP	Salmonella DU	Salmonella QU	Shigella AU	Shigella DP	Shigella DU	Shigella QU	Vaccinia AU	Vaccinia DP	Vaccinia DU	Vaccinia QU		
ARF1																																	
ARPC3	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓			
ATP6V1A	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓			
Abi1		✓							✓																								
CBL																																	
CDC42	✓	✓	✓	✓	✓	✓	✓		✓	✓	✓																						
CDH4		✓							✓																								
CFL1																																	
CHUK																																	
CLTC																																	
DNM2																																	
FRAP1	✓	✓																															
GFP 1 <sup>a</sup>	✓																																
GFP 2 <sup>a</sup>	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓			
ITGAV																																	
ITGB1		✓																															
Kif11	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓			
Kill <sup>b</sup>		✓																															
MAP3K7		✓																															
MET		✓																															
MOCK	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓			
NOD1																																	
PAK1																																	
PI4KA																																	
PI4KB	✓								✓																								
PIK3R1		✓	✓																														
PRKCA		✓	✓																														
PSMA6																																	
PSMC3	✓								✓																								
PXN	✓	✓							✓																								
RAB7A																																	
RAC1	✓	✓																															
Rab2																																	
SNX9	✓	✓																															
Scra 1 <sup>c</sup>	✓								✓																								
Scra 2 <sup>c</sup>	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓			
TLN1																																	
TSG101																																	

<sup>a</sup>GFP targeting siRNA sequences are provided by Ambion (Ambion Silencer eGFP; GFP 1) and Dharmacon (GFP Duplex III; GFP 2).

<sup>b</sup>A positive control cell killer is provided by Qiagen (AllStars Hs Cell Death Control)

<sup>c</sup>Scrambled siRNA sequences are provided by Ambion (Silencer Select Negative Control; Scra 1) and Dharmacon (ON-TARGETplus Non-targeting Control; Scra 2).

## 4.2. RNA Interference Protocols

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quently checked for applicability by sequence alignment to the target genome. While scrambling has the advantage that a possible effect of nucleotide composition is removed, it is infeasible for large-scale screens and often a set of predefined sequences sold by manufacturers (for example Silencer Select Negative Control from Ambion, ON-TARGETplus Non-targeting Control siRNAs from Dharamcon or AllStars Negative Control siRNA from Qiagen) is used instead (while still being called scrambled controls).

Table 4.3 lists the siRNA control experiments that were performed throughout all screens and indicates the set of controls each screen contains. Common to all pathogens and all libraries are the previously mentioned positive control Kif11, one of two GFP targeting sequences, scrambled siRNAs, as well as mock experiments. Additionally, several controls for general infection mechanisms were included in most screens, including ATP6V1A (a H<sup>+</sup> transporting ATPase, responsible for acidification of endocytic and lysosomal vesicles), ARPC3 (Arp2/3), and Cdc42, both part of actin-dependent processes surrounding pathogen uptake and ABM. Some controls however are specific to a subgroup of pathogens or single pathogens and include the following (grouped by pathogen).

**Bartonella/Brucella:** The small GTPase Rab2 is required for anterograde ER-Golgi transport, capable of interacting with RicA of *B. abortus* ([de Barsy et al., 2011](#)) and TLN1 (talin 1) has been determined to be necessary for invasome formation in *B. henselae* infection via β<sub>1</sub> signaling ([Truttmann et al., 2011](#)).

**Listeria:** During endocytosis of *L. monocytogenes*, the ubiquitin ligase Cbl is recruited to the site of entry and seems to be involved in InlB mediated, clathrin dependent (CLTC encodes the heavy chain 1 component of clathrin) bacterial uptake ([Veiga and Cossart, 2005](#)). DNM2 (dynamin-2) is a further protein involved in host entry and PIK3R1 is a phosphatidylinositol 3-kinase, downstream to Met, the cellular receptor to InlB.

**Salmonella:** The actin-modulating protein CFL1 (cofilin-1), responsible for actin depolymerization, the cellular integrin receptor ITGAV and RAB7A, a small GTPase that regulates vesicular trafficking, have all been shown to be hits in a salmonella invasion screen ([Misselwitz et al., 2011](#)).

**Shigella:** Down regulation of ARF1 (a guanosine triphosphate (GTP) binding protein involved in vesicle trafficking) and phosphatidylinositol 4-kinase PI4KA has been suspected of interfering with pathogen entry, while suppression of CHUK/NOD1 (both involved in NF-κB signaling) may inhibit IL-8 production by uninfected bystander cells, thereby possibly promoting infection ([Kasper, 2012](#)).

## 4. DATA

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**Adenovirus/rhinovirus:** SNX9 regulates dynamin assembly and is therefore crucial to viral endocytosis and PRKCA is a Ser-/Thr-specific protein kinase responsible for a wide array of regulatory signals. PRKCA might be involved in influenza virion budding and has been implicated in playing a role in intracellular proliferation of hepatitis ([Kanehisa and Goto, 2000](#)).

**Vaccinia virus:** The Ser-/Thr-specific protein kinase PRKCA is targeted by Cdc42/Rac1 and has been shown to be required for MV entry ([Mercer and Helenius, 2008](#)). Up-regulation of PSMA6 (a proteasome complex component) enables evasion of intracellular immune surveillance ([Zhou et al., 2014](#)) and TSG101, ubiquitin-conjugating enzyme, hampers EV production ([Honeychurch et al., 2007](#)).

Many of these pathogen specific positive control targets are not well established and while some have been previously been identified and validated, others represent best guesses and might not serve their purpose particularly well. Controls wells are typically located at the plate border (rows A and P; columns 1, 2, 23 and 24) and the different control experiments were replicated multiple times on each plate.

For all screens, siRNA transfection was carried out by adding 25 µl of RNAi-MAX/DMEM (0.1 µl/24.9 µl) transfection agent to 1.6 pmol siRNA diluted in 5 µl RNase-free ddH<sub>2</sub>O contained in each of the 384 wells per screening plate. After 1 h incubation at room temperature, the required number of cells were added (see table 4.2), suspended in 50 µl DMEM/16% FBS. Plates were subsequently incubated for 72 h at 37 °C and 5% CO<sub>2</sub>, followed by the pathogen specific infection procedure (see section B.1 for details). Following infection, cells were fixed in paraformaldehyde (PFA) and stained for DNA, F-actin and additional pathogen specific markers (see table 4.2). Plates were sealed prior to imaging.

### 4.3 Image Acquisition and Data Processing

Imaging was performed both at the University of Basel and the Light Microscopy and Screening Center of ETH Zürich, using ImageXpress micro (IXM) HCS wide-field microscopes from Molecular Devices, equipped with Thermo Scientific CataLyst Express robotic plate handlers capable of storing and serving up to 45 plates. Lumencor Spectra X solid-state light engines (LED light sources), 10x S Fluor objective lenses by Nikon with a numerical aperture of 0.45 and Photometrics CoolSNAP HQ 14-bit CCD cameras, resolving 1392 × 1040 pixels (individual pixel size of 6.45 µm × 6.45 µm), complete the hardware setup. Channel selection is assay specific and stain dependent Semrock

### 4.3. Image Acquisition and Data Processing

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filters (DAPI/Hoechst, GFP/FITC/Alexa488, Cy3, Cy5, Quadbard DAPI-GFP-mCherry-Cy5) are employed (see table 4.2 for details).

Wells are divided into  $3 \times 3$  grids for most plates while some  $2 \times 3$  site images exist too, with no spacing and no overlap, and Molecular Devices MetaXpress High-Content Image Acquisition and Analysis Software was used for recording images. Software parameters include no gain, well bottom as autofocus target, site-specific autofocusing, enabling of laser-based focusing and no shading correction. For each imaging channel, focus Z-offset was selected manually and exposure time was automatically calculated. In cases of poor dynamic range or overexposure, manual correction to exposure time was applied. Upon imaging, data was transferred to iBrain2/screeningBee (Rouilly et al., 2012) for further processing.

#### 4.3.1 Data Handling (iBrain2/screeningBee)

In case of microscopy based siRNA screening experiments, a complex task of data handling and processing follows the imaging stage. A wealth of data is generated by imaging devices<sup>3</sup> which has to be accessibly and redundantly stored. Furthermore, analysis of image data is a processing intensive task that quickly becomes reliant on high performance computing (HPC) resources, entailing specialized requirements due to the oftentimes shared and centralized nature of such systems. The authors of iBrain2 summarize the key steps in RNAi high content screening (HCS) data processing as follows (Rouilly et al., 2012):

1. **Data acquisition.** The raw data of siRNA screens is produced as digital images by microscopy. Acquisition times of several hours per plate are typical and a single plate yields ~20 GB of data.
2. **Permanent storage.** Due to infeasibility of re-screening plates, all raw data has to be stored in a sufficiently redundant manner. Furthermore, the permanent data store has to be able to serve portions of the dataset quickly and efficiently.
3. **Temporary data staging to HPC.** In order for the compute cluster not to be bound by network latency, it is often necessary to stage the data to be analyzed to local scratch space. This step becomes superfluous whenever the permanent storage system is directly integrated in the cluster's high-speed network.

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<sup>3</sup>A genome-wide screen (~27000 individual experiments) in 384 well format involves 70–100 plates depending on the number of controls per plate. Each plate yields 10000–14000 images (384 wells, 9 sites and 3–4 channels), leading to 700000–1400000 individual images and requiring multiple terabytes of storage.

## 4. DATA

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4. **Data analysis.** Many aspects of processing a large number of images are embarrassingly parallel and a cluster environment is ideally suited to tackle this computation intensive task.
5. **Permanent storage of results.** Some results produced by the analysis procedure will be saved back to the permanent storage system. While it may be sensible to save storage space and carry out some procedures on the fly, this is not feasible for all analysis routines.
6. **Publication and archiving.** Upon completion of the project, some data will be made available publicly and all data worth keeping is moved to an archival system capable of cheap long-term storage where quick retrieval is unimportant.

The requirements outlined above led to the development of iBrain2 as a modular workflow manager capable of setting up reproducible procedures. The software is implemented in Java and uses an XML-based format for defining workflows. Furthermore, it interfaces with other open source projects, such as openBIS ([Bauch et al., 2011](#)) which can be used as storage system and CellProfiler ([Carpenter et al., 2006; Kamentsky et al., 2011](#)), a popular tool for analyzing cell based image material. Within InfectX, not iBrain2 itself is used, but a derivative thereof called screeningBee, featuring tighter openBIS integration and a large set of CellProfiler extensions.

### 4.3.2 Image Analysis (CellProfiler)

Prior to the development and release of CellProfiler in 2006, large-scale screens were routinely evaluated through tedious and labor intensive visual inspection by expert biologists. While humans are able to perceive and contextualize image data in ways that computers still struggle to emulate, a lot of detail is lost through human interpretation. Small differences are hard to spot, leading to subtle patterns being missed and humans focus on a handful of features, not being able to take into account the plethora of information encoded in crowded images. Furthermore, human study of image data leads to qualitative evaluations, while computational analysis yields a potentially large number of quantitative measures. Lastly and perhaps most importantly, human based assessment of imagery coming from a project the size of InfectX is simply infeasible. In the words of [Carpenter et al.](#), the authors of CellProfiler, consequences of implementing computational image analysis for HTS are as follows:

With the successful application of sophisticated image analysis methods, the bottleneck of image-based genome-wide screens is now moving downstream to data visualization, exploration, and statisti-

### 4.3. Image Acquisition and Data Processing

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cal analysis in order to accommodate the number and richness of measurements that result from image-based genome-wide assays.

CellProfiler is an open-source software solution catering to the needs of HTS screening initiatives and providing a customizable, modular platform for cellular image analysis. CellProfiler processing is based on modules which are placed in sequential to form a pipeline, usually starting out with image processing, followed by object identification and concluded by calculation of feature measurements on these objects. The pipeline's modules and settings can be saved to a configuration file, in order to ensure reproducibility and transferability of workflows. The original version of CellProfiler was written in MATLAB and a more recent version 2 has been ported to python. InfectX procedures are based on version 1 ([Carpenter et al., 2006](#)).

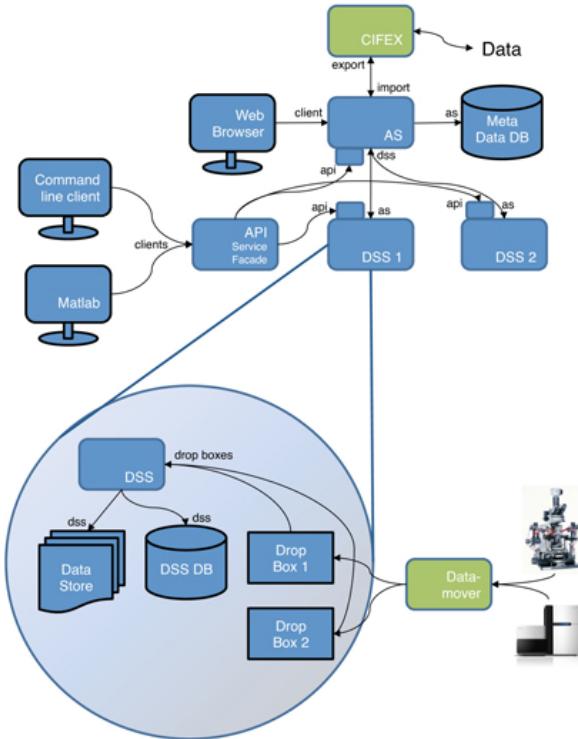
The InfectX image analysis routine starts out with metadata collection and image quality assessment modules, followed by shading correction. Detecting out of focus images, wells that show signs of experimental error or problematic plates, however is still largely carried out by humans. Position dependent illumination and sensor inhomogeneities need to be corrected for, as they lead to decreased sensitivity in darker areas and make comparison of intensity based features between objects belonging to different areas of the image problematic.

Shading correction, while offered by a default module of CellProfiler, is implemented in a module developed specifically for InfectX. A shading model is generated per microscope, lamp, channel, pathogen and assay which can simply be applied to all corresponding images. This reduces the computational complexity of shading correction significantly and avoids errors associated with an automated procedure being applied to a diverse range of instances. Shading models are produced by overlaying images and calculating histograms per pixel position, yielding robust estimates for foreground intensities and by fixed offsetting, also estimates for background intensities. Subsequently, a foreground model is calculated and applied, leading to conservatively corrected images. With only little shading remaining, reliable object segmentation is possible, the result of which are used to calculate a further, final shading model based on pixel histograms and average object intensities.

Illumination corrected images are stitched together, channel separated and forwarded to CellProfiler modules for segmentation and feature calculation. Cell identification is a hard problem, especially if clumps are present. Whenever objects are present that can easily be identified, such as nuclei if DNA was stained, these are identified first (primary objects), facilitating identification of secondary objects, e.g. cells. In case of clumped cells, a three step algorithm is employed: clumps are recognized, then the dividing lines are defined and finally, some resulting objects may be removed or merged based on measure-

#### 4. DATA

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**Figure 4.2:** An openBIS instance is deployed as a service consisting of an application server (AS) and one or more data store servers (DSSs). At both levels, clients can interact with the system to query, fetch or deposit data. ([Bauch et al., 2011](#)).

ments such as size or shape.

Several modules developed in-house are used during the feature extraction stage, extending CellProfiler functionality to specific requirements including invasome and bacterial aggregate detection, more efficient neighbor feature measurement and collection of various properties per sub-cellular object. The features calculated by the CellProfiler pipeline are explored more in-depth in section 4.4. All results produced by image analysis are saved to an openBIS instance.

#### 4.3.3 User Accessible Data Storage (OpenBIS)

The authors of openBIS (open Biology Information System) argue that the availability of a domain specific information management system plays an important role in data-driven biological research, serving as a basis for management of large amounts of experimental data and acting as both a source and a sink for analysis procedures. Furthermore, such a system should provide a user interface capable of visualizing complex dependency structures and a set of application programming interfaces (APIs) for accessing the stored data programmatically ([Bauch et al., 2011](#)). Finally, it should be possible to make a subset of the data available to the public, providing a simplified interface of the browsing and searching capabilities.

In order for openBIS to be applicable to the broad range of scenarios it is intended for, extensibility and transparent interfaces are paramount, as is scalability and performance. To address the latter aspect, separation of bulk data from metadata is a guiding principle. Both ingestion of high-throughput, high-content data as well as serving it to users and analysis routines is CPU, I/O and network intensive and therefore should not be performed on the same system as user-facing metadata queries which only deals with comparably inexpensive tasks that scale well. Additionally, scalability requires that bulk data storage may be distributed among several systems.

With these requirements in mind, openBIS is designed to consist of an application server (AS), dealing with metadata, serving user queries such as searches and visualizing dependencies via the browser based graphical user interface, as well as one or several data store servers (DSSs) that handle the bulk data. Communication between the two layers is asynchronous.

All datasets in openBIS are immutable. While this is potentially wasteful in terms of storage resources, it can be argued that storage space is cheap and such an architecture improves traceability of constant evolution of analysis procedures and corresponding results. Datasets are subject to a hierarchical structure paradigm consisting of the entities (1) *Data Space*, (2) *Project*, (3) *Experiment*, and (4) *Sample*. In the example of HTS data, a sample represents a plate and associated datasets include raw images, feature measurements and infection scores. Metadata can be provided either in unstructured form which is handled as attachments associated with project, experiment or sample entities, or as structured objects (consisting of name, label, description and value) that are linked to experiments, samples and datasets.

Data storage by a DSS follows a hybrid model consisting of a relational database, storing index information, file metadata and selected results, in addition to flat-file data store which can be distributed between several file systems. Man-

## 4. DATA

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agement of fill state of the associated storage shares and data distribution is all managed by the individual DSSs. Some data presentation task handled by DSSs include data visualizations such as plate heatmaps and assembly of multichannel images, as well as overlay of analysis results (e.g. segmentation).

Ingestion of new data can either be triggered by web import, custom software talking to the corresponding API or simply by adding to folders that are monitored by a DSS as dropboxes. Responding to write activity, the highly customizable ETL (extract, transform, load) routine extracts metadata, creates datasets annotated with metadata in the AS database, links them to the appropriate entities and adds the new datasets to the data store. Export of data is either browser based or can be handled by provided command line tools, accessing openBIS via a Java service facade. Two separately designed tools are integrated for used with openBIS: CIFEX and Datamover. The former enables browser based transfer of large files and supports interruption of transfers as well as checksumming for file integrity, while the latter can be used to automatically transfer data off of an instrument computer into a dropbox folder via SSH or rsync protocols.

### 4.4 Single Cell Feature Data

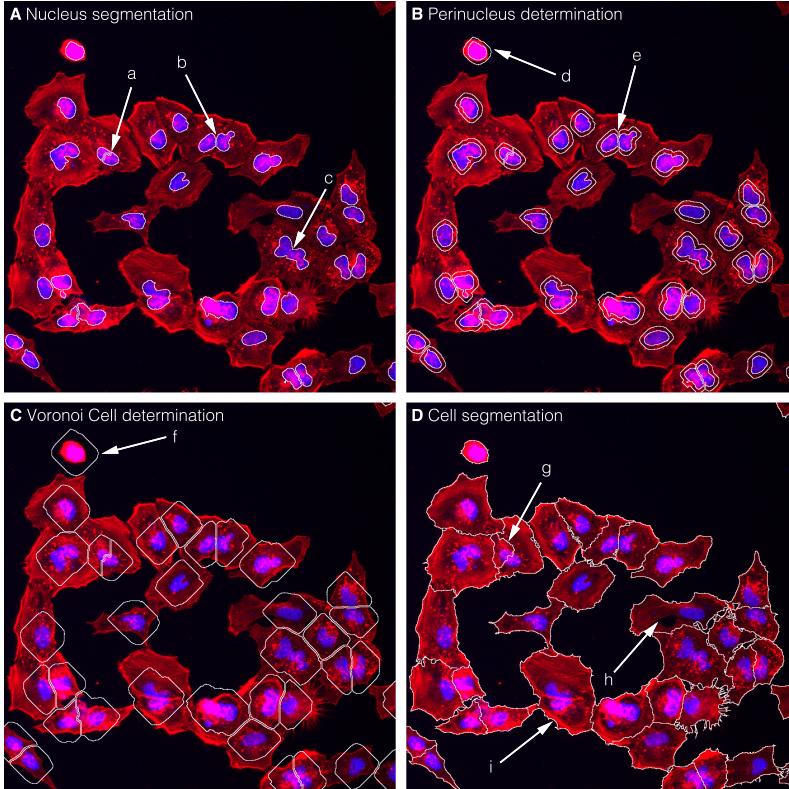
The set of accessible single cell features varies among screens and pathogens to some extent. While there are some general features, such as geometry, intensity and texture of nuclear and cellular objects available for every plate dataset, others, including invosome and IL-8 features are specific to a subset of the pathogens, while others still, such as certain neighbor features or Vononoi cell segmentation were only recently added to the feature extraction pipeline and therefore have not yet been calculated for all screens. Such discrepancies are not treated by this overview which focuses on general aspects of currently available single cell features.

Nomenclature of single cell features follows a hierarchical pattern consisting of an object specifier, a measurement group, the measurement name itself and the imaging channel it was applied to, as follows:

```
<object>.<group>_<measurement>_<channel>
Cells.Intensity_MedianUpperTenPercentIntensity_CorrDNA
```

In the above example, the measured objects are cells, the feature is of intensity type, the feature name indicates that it represents the mean of intensities above the upper decile and the channel description reveals that DNA stain values are used. Throughout the different screens, 14 single cell feature objects are available:

#### 4.4. Single Cell Feature Data



**Figure 4.3:** Object identification is started with easily recognizable nuclei, which become primary objects and serving as basis for detecting secondary objects. Problems with nuclei detection include superfluous splitting (a) and failure to separate some touching objects (c). Mostly results are acceptable and some touching nuclei are correctly separated (b). Perinuclei extend the nuclear border and therefore suffer from problems introduced in nuclear recognition. Further issues include overlap with extracellular space (d) and small perinuclear area in densely populated areas (e). Voronoi cells further extend the nuclear area and often grow beyond cell boundaries (f). Finally, cell segmentation is a hard problem due to irregular shapes, variability in size and inhomogeneities in actin distribution (h). Results suffer from falsely identified nuclei (g and i).

## 4. DATA

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**Nuclei:** Using the DNA channel, nucleus recognition is fairly accurate owing to generally uniform morphology of nuclei, uniform distribution of DNA and good contrast of DNA staining. The current implementation for segmentation is based on Otsu's method which converts a monochromatic image into a binary image by finding an appropriate threshold, effectively separating foreground from background. Objects that have an Euler characteristic  $\chi \leq 0$  are filled and apparently clumped objects are split. Finally, objects outside of an allowed size range are discarded (mostly dust particles which are too big and pathogen DNA which yields objects that are too small). Nuclei represent primary objects with serve as basis for the detection of secondary objects (see figure 4.3, A).

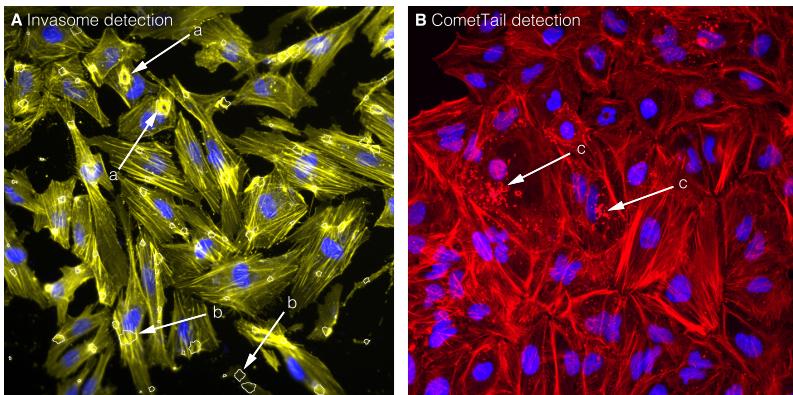
**PeriNuclei:** The perinuclear region is defined as an extension of the nuclear border by 8 pixels and removal of the nucleus itself. As some pathogens establish their replicatory niche in this region it lends itself to closer investigation. Furthermore the border is only dependent on the reliably established nuclear object. Potential pitfalls include extension of the perinuclear region into extracellular space or neighboring cell when the nucleus is located close to the plasma membrane, as well as reduced area whenever cells are clustered, as perinuclear regions may not overlap (see figure 4.3, B).

**VoronoiCells:** Simultaneously extending all nuclei by maximally 24 pixels or until a neighboring border is met leads to Voronoi cells. This type of defining cell bodies is very prone to extending into extracellular space, as the actin channel is not taken into account (see figure 4.3, C).

**Cells:** The cell body border is determined by propagation segmentation based on thresholding. Starting from the nucleus, the cytoplasmic region is extended until the intensity gradient changes abruptly, indicating either the beginning of background, or the boundary to a neighboring cell. Non-uniform actin intensity patterns, irregular shape, large variability in size and frequent touching of neighbors makes this a hard problem and results can be unreliable. Common to all secondary objects, errors in determining primary objects will propagate and lead to cells erroneously being cut up or aggregated. Furthermore, borders to neighbors are frequently not determined correctly, but significant discrepancies among human expert labeling are to be expected in such situations as well (see figure 4.3, D).

**ExpandedNuclei:** Serving as precursors to PeriNuclei objects, these objects are not further quantified and only have coordinates and number of perinuclear children (almost exclusively 1) associated.

**Bacteria/Viruses/Pathogen:** Due to the heterogeneity between pathogens, detection is assay specific. Generally, pathogens are primary objects and not derived of cell nuclei. In *Bartonella* and *Brucella* screens, large and small



**Figure 4.4:** Both CometTails and Invasomes are detected on actin channels, which poses difficulties due to noisy background. Invasomes are characterized by their ring-shaped morphology (a), while CometTails appear as actin streaks (c). Currently invasome detection yields many false-positives (b).

pathogen clusters are segmented using Otsu's method on the pathogen channel. Individual bacteria in *Listeria* screens, SCVs in *Salmonella* screens and small/medium pathogen clusters in both *Shigella* and rhinovirus screens are identified using wavelet decomposition of the pathogen channel, while for adenovirus screens, detection of large pathogen aggregates relies on propagation outwards from nuclei, much like cell body identification but using the pathogen channel. For vaccinia virus, direct pathogen segmentation is currently not possible.

**IntBacteria/ExtBacteria:** In *Bartonella* assays, different stains were used for intracellular (GFP) and extracellular (Cy5) bacteria which consequently can be distinguished during image analysis. This is helpful to prevent bacteria the are sit on top or below a cell from being considered intracellularly located.

**BlobBacteria:** Specific to *Brucella* screens, these large bacterial aggregates do not have any real measurements associated, except coordinates.

**Invasomes:** As one of two internalization structures in *Bartonella* infection, invasomes are characterized as actin surrounded large bacterial aggregates. Exploiting this morphology, the current invasome detection uses a ring template to search the actin channel for candidates and exploits the intensity difference to the cell mean for confirmation. Unfortunately, this yields many false positives. Improvements are being discussed but have yet to be implemented (see figure 4.4, A).

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**IL8:** In *Shigella* assays, cellular IL-8 was stained in order to study proinflammatory response of uninfected bystander cells. Due to these objects having their own channel, segmentation is fairly straightforward.

**CometTails:** Specific to *Listeria* screens, the telltale signs of ABM can be identified as dense strokes on the actin channel. CometTails contain only few measurements beyond their location (see figure 4.4, B).

**Neighbors:** While not representing detected objects themselves, features involving identities of adjacent objects are summarized within this category. Currently, indices of Cells and Bacteria that come closer than 2 pixels and of Voronoi cells that do not lie further apart than 8 pixels are collected. These thresholds compensate for slight segmentation errors. Furthermore, for each pair of detected neighbors, the length of common border is recorded. Taken together, features of this category allow construction of both a binary adjacency matrix and a weighted adjacency matrix corresponding to each of the objects that were considered.

Once images are segmented into objects, measurements are applied to the individual image areas. Dependent on object class, the number and types of available features differ, sometimes owing to systematic causes and at other times, only due to differences in implementations of per-object feature recognition. The most commonly available features are grouped into categories *AreaShape*, *Intensity*, *Texture*, *RadialDistribution*, *Location*, *Neighbors/IdentityOfNeighbors/PercentTouchingNeighbors* and *Parent/Children*.

Features belonging to the *AreaShape* category are not applied to a specific imaging channel but serve to summarize geometric properties of detected objects (*cf.* table 4.4). When applied to actin channel objects that cannot reliably be segmented, such as cell bodies, some caution has to be exerted, as these features are strongly affected by upstream errors. Nevertheless, this group constitutes features that are among the most frequently used for analysis procedures. Some infection phenotypes, for example membrane ruffling encountered during trigger-type host entry mechanisms are captured by some of these features (FormFactor is able to measure raggedness).

Intensity features mostly consist of summary statistics being applied to all or a portion of per channel intensity data within a given object. The functions used to describe the underlying distributions are sum (Integrated), lower quartile, maximum (Max), mean, median, minimum (Min), standard deviation (Std), upper quartile, upper vigintile (UpperFivePercent), upper decile (UpperTenPercent) and upper 50-quantile (UpperTwoPercent). Additional features are generated by using certain functions only on the object border pixels, yielding *IntensityEdge* features (e.g. IntegratedIntensityEdge) and by only including

#### 4.4. Single Cell Feature Data

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**Table 4.4:** List of AreaShape features with corresponding descriptions. Some information is taken from the CellProfiler manual ([Carpenter et al., 2006](#)).

Feature name	Feature description
Area	The number of pixels enclosed by the object border.
Eccentricity <sup>a</sup>	Ratio of the distance between the foci and major axis length of the corresponding ellipse. The value is between 0 (circle) and 1 (line segment).
EulerNumber	1 minus the number of holes within the object, assuming 8-connectivity.
Extent	Area of the object divided by the area of the bounding box.
FormFactor	Calculated as $4\pi * \text{area} / \text{perimeter}^2$ . Equals 1 for a perfectly circular object.
<Spec>AxisLength <sup>a</sup>	Spec = [Major, Minor]; The major/minor axis length (in pixels) of the corresponding ellipse.
Orientation <sup>a</sup>	The angle (ranging from $-90^\circ$ to $90^\circ$ ) between the x-axis and the major axis of the corresponding ellipse
Perimeter	The total number of pixels around the object boundary.
Solidity	Proportion of the pixels in the convex hull that are also part of the object, i.e. $\text{area}_{\text{object}} / \text{area}_{\text{conv.hull}}$ .
<Sub <sup>b</sup> >Area	The total number of pixels enclosed by all subcellular pathogen object borders.
<Sub <sup>b</sup> ><Stat <sup>c</sup> >Distance-ToNuclNorm	Distances between the nucleus and all subcellular pathogen objects are normalized and a summary statistic is applied.
<Sub <sup>b</sup> ><Stat <sup>c</sup> >Distance-ToNucl	Distances from nucleus to all subcellular pathogen objects are calculated and a summary statistic is applied.
<Sub <sup>b</sup> ><Stat <sup>c</sup> >Pairwise-Distance	A summary statistic is applied to all pairwise distances between subcellular pathogen objects.
<Sub <sup>b</sup> ><Stat <sup>c</sup> >Shortest-Distance	For each subcellular pathogen object, the shortest distance among all distances to other subcellular objects is selected and a summary statistic is applied to the collection of shortest distances.

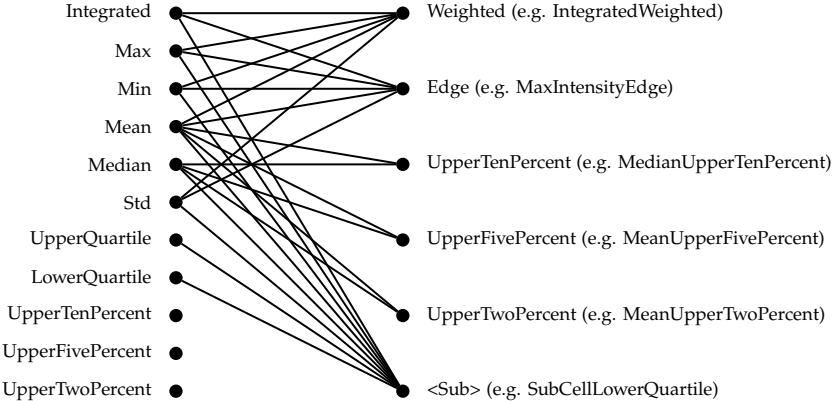
<sup>a</sup> The corresponding ellipse is defined as the best fitting ellipse in the sense that it has identical second moments compared to the original object ([Rocha et al., 2002](#)).

<sup>b</sup> Naming of these features currently is fairly heterogeneous with possible prefixes *PerObj*, *SubCellBacteria*, *SubCellViruses*, *SubCell*. All are based on the set of subcellularly located pathogens.

<sup>c</sup> The statistics applied in order to describe the underlying distributions are mean, median, lower and upper quartiles, standard deviation and sum.

## 4. DATA

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**Figure 4.5:** Common summary statistics that are applied to intensity data, along with their possible modifiers. The subcellular intensity features have different prefixes, including SubCellBacteria, SubCellCometTails, SubCellViruses, PerObject and SubCell.

the upper 10%, 5% and 2% of the data. Weighted features are calculated by extracting weights from a corresponding grayscale image and multiplying individual intensity values before applying the summary statistic. Restricting the data to subcellular pathogen objects yields features such as SubCellLowerQuartile. For a complete list of modifiers and implemented combinations, refer to figure 4.5. The only currently available intensity feature that is not shown in 4.5 is MassDisplacement, which quantifies the distance between the centers of gravity in the gray-level representation of the object and the binary representation of the object.

All texture measurements are either based on Haralick features (Haralick et al., 1973) or Gabor wavelet features (Gabor, 1946), as implemented in CellProfiler and are calculated per image channel. The gray-level co-occurrence matrix  $G_k$  with dimensions  $n \times n$  where  $n$  is the number of gray levels serves as basis for Haralick's texture features (Carpenter et al., 2006).

$$G_k = \begin{bmatrix} p(1,1) & p(1,2) & \cdots & p(1,n) \\ p(2,1) & p(2,2) & \cdots & p(2,n) \\ \vdots & \vdots & \ddots & \vdots \\ p(n,1) & p(n,2) & \cdots & p(n,n) \end{bmatrix} \quad (4.4)$$

#### 4.4. Single Cell Feature Data

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Element  $p(i, j)$  is calculated by counting the number of times, a pixel with value  $i$  is adjacent to a pixel with value  $j$ , divided by the total number of comparisons. Consequently,  $p(i, j)$  can be thought of as representing a probability. Moreover, assuming 8-connectivity, 4 different types of adjacency are possible: horizontal, vertical, top left to bottom right diagonal and top right to bottom left antidiagonal, yielding 4 co-occurrence matrices  $G_k, k \in \{1, 2, 3, 4\}$ . 14 statistics were developed by [Haralick et al.](#), 13 of which are included with CellProfiler:

$$\begin{aligned}
p_x(i) &= \sum_{j=1}^n p(i, j), & p_y(j) &= \sum_{i=1}^n p(i, j), \\
p_{x+y}(k) &= \sum_{i=1}^n \sum_{\substack{j=1 \\ i+j=k}}^n p(i, j), & p_{x-y}(k) &= \sum_{i=1}^n \sum_{\substack{j=1 \\ |i-j|=k}}^n p(i, j), \\
\mu_x &= \mathbf{E}[p_x], & \mu_y &= \mathbf{E}[p_y], \\
\sigma_x &= \sqrt{\text{Var}(p_x)}, & \sigma_y &= \sqrt{\text{Var}(p_y)}, \\
HX &= - \sum_{i=1}^n p_x(i) \log(p_x(i)), & HY &= - \sum_{i=1}^n p_y(i) \log(p_y(i)), \\
HXY1 &= - \sum_{i=1}^n \sum_{j=1}^n p(i, j) \log(p_x(i)p_y(j)), \\
HXY2 &= - \sum_{i=1}^n \sum_{j=1}^n p_x(i)p_y(j) \log(p_x(i)p_y(j)) \\
\text{AngularSecond-} & f_1 = \sum_{i=1}^n \sum_{j=1}^n p(i, j)^2 & (4.5) \\
\text{Moment} & & \\
\text{Contrast} & f_2 = \sum_{k=0}^{n-1} k^2 \left( \sum_{\substack{i=1 \\ |i-j|=k}}^n \sum_{j=1}^n p(i, j) \right) & (4.6) \\
\text{Correlation} & f_3 = \frac{\sum_{i=1}^n \sum_{j=1}^n (ij)p(i, j) - \mu_x \mu_y}{\sigma_x \sigma_y} & (4.7) \\
\text{Variance} & f_4 = \sum_{i=1}^n \sum_{j=1}^n (i - \mu)^2 p(i, j) & (4.8)
\end{aligned}$$

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#### 4. DATA

$$\text{InverseDifferenceMoment} \quad f_5 = \sum_{i=1}^n \sum_{j=1}^n \frac{1}{1 + (i - j)^2} p(i, j) \quad (4.9)$$

$$\text{SumAverage} \quad f_6 = \sum_{i=2}^{2n} i p_{x+y}(i) \quad (4.10)$$

$$\text{SumVariance} \quad f_7 = \sum_{i=2}^{2n} (i - f_8)^2 p_{x+y}(i) \quad (4.11)$$

$$\text{SumEntropy} \quad f_8 = - \sum_{i=2}^{2n} p_{x+y}(i) \log(p_{x+y}(i)) \quad (4.12)$$

$$\text{Entropy} \quad f_9 = - \sum_{i=1}^n \sum_{j=1}^n p(i, j) \log(p(i, j)) \quad (4.13)$$

$$\text{DifferenceVariance} \quad f_{10} = \sum_{i=0}^{n-1} i^2 p_{x-y}(i) \quad (4.14)$$

$$\text{DifferenceEntropy} \quad f_{11} = - \sum_{i=0}^{n-1} p_{x-y}(i) \log(p_{x-y}(i)) \quad (4.15)$$

$$\text{InfoMeas1} \quad f_{12} = \frac{f_9 - HXY1}{\max\{HX, HY\}} \quad (4.16)$$

$$\text{InfoMeas2} \quad f_{13} = (1 - \exp[-2(HXY2 - f_9)])^{\frac{1}{2}} \quad (4.17)$$

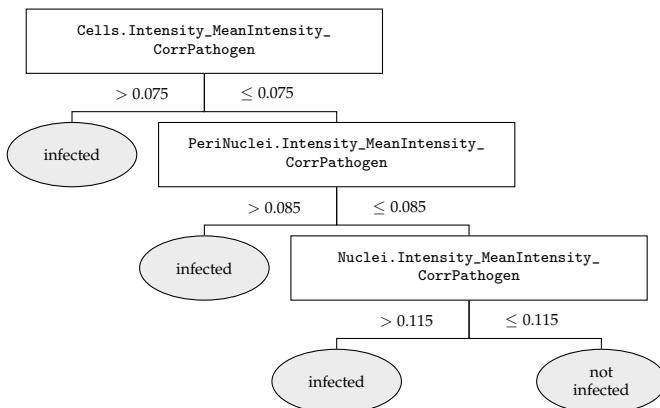
The individual statistics are averaged over the four co-occurrence matrices to yield a single value per object per imaging channel. Computation of Haralick features is comparably inexpensive and the resulting texture characterization has successfully been used in image-based classification scenarios. Gabor filters on the other hand are more complicated and their application can incur significant computational cost. They quantify striped texture that is parallel to a given angle and sets of Gabor filters at certain angles are frequently used for pattern recognition tasks such as optical character recognition or fingerprint recognition. The directions for which Gabor filters are applied to segmented objects are the x- and y-axes, yielding the features GaborX and GaborY.

RadialDistribution features contain 3 measurements that are applied to each image channel. Given an object, its area is divided into bins formed by concentric rings, where the center is the point that is the farthest from any edge. FracAtD records the fraction of total stain in the object at a given radius.

#### 4.5 Infection Scoring

#### 4.5. Infection Scoring

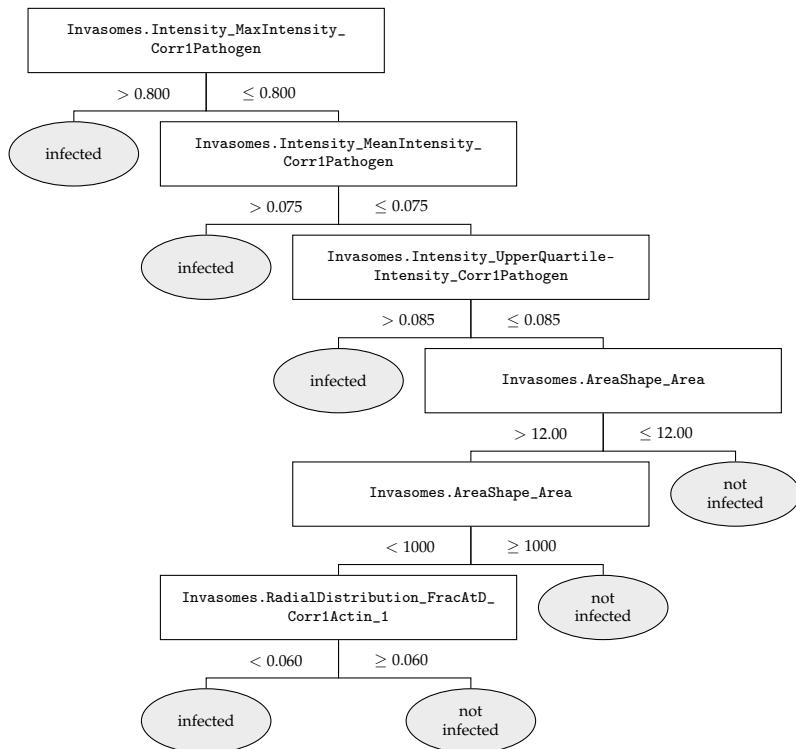
---



**Figure 4.6:** For adenovirus infection scoring, the decision tree classifier checks if enough pathogen is detected within the cell body, the perinuclear region or the nucleus. The threshold decreases as the region of interest concentrates on areas associated with progressively involved in infection.

## 4. DATA

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**Figure 4.7:** Decision tree for *Bartonella* infection scoring. In order to detect bona-fide invasomes, the first three or-linked decisions assemble a list of candidates while the following three and-linked decisions discard some erroneously included instances. In order to obtain the desired cell-based infection score, invasomes are mapped to cellular objects in a subsequent step.

## Appendix A

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### R Package **singleCellFeatures**

---

Additional material. For example long mathematical derivations could be given in the appendix. Or you could include part of your code that is needed in printed form. You can add several Appendices to your thesis (as you can include several chapters in the main part of your work).

```
facetBorder <- function(x, y, img, facet) {  
  facet.size <- img / facet  
  # calculate facets (2d binning)  
  x.bin <- ceiling(x / facet.size[1])  
  y.bin <- ceiling(y / facet.size[2])  
  # initialize empty grid/border matrices  
  grid <- matrix(0, facet[2], facet[1])  
  # calculate col-major grid index for each object  
  index <- y.bin + (x.bin - 1) * facet[2]  
  # summarize as counts  
  counts <- table(index)  
  # fill grid with counts  
  grid[as.numeric(names(counts))] <- counts  
  grid.res <- grid  
  grid <- grid > 0  
  # extend grid with a frame of ones  
  grid.ext <- rbind(rep(1, (facet[1] + 2)),  
                    cbind(rep(1, facet[2]), grid, rep(1, facet[2])),  
                    rep(1, (facet[1] + 2)))  
  # set up stencil  
  row <- rep(rep(1:facet[2], facet[1]))  
  col <- rep(1:facet[1], each=facet[2])  
  colP1 <- col + 1  
  colM1 <- col - 1  
  rowP1 <- row + 1  
  rowP2 <- row + 2
```

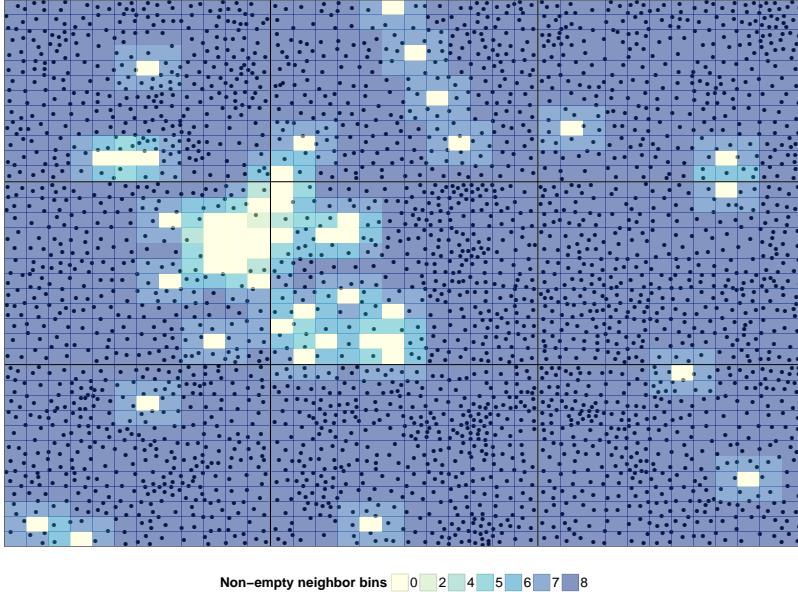
## A. R PACKAGE SINGLECELLFEATURES

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```
nrowP <- facet[2] + 2
stencil <- cbind(row  + colM1 * nrowP, # northwest neighbor
                  row  + col   * nrowP, # north neighbor
                  row  + colP1 * nrowP, # northeast neighbor
                  rowP1 + colP1 * nrowP, # west neighbor
                  rowP2 + colP1 * nrowP, # east neighbor
                  rowP2 + col   * nrowP, # southwest neighbor
                  rowP2 + colM1 * nrowP, # south neighbor
                  rowP1 + colM1 * nrowP) # southeast neighbor
# apply stencil row-wise to grid
border <- apply(stencil, 1, function(ind, mat) {
  return(sum(mat[ind]))
}, as.numeric(grid.ext))
# map col-major object index to border array
return(border[index])
}

edgepos <- function(x, y, img, n) {
  empty <- logical()
  xst <- img[1] / n
  yst <- img[2] / n
  sgrid <- matrix(0, nrow=n, ncol=n)
  for (i in 1:n) {
    for (j in 1:n) {
      ispos <- (x > (i - 1) * xst) & (x <= (i * xst)) &
        (y > (j - 1) * yst) & (y <= (j * yst))
      sgrid[i, j] <- sum(ispos)
    }
  }

  for (i in 1:n) {
    for (j in 1:n) {
      ispos <- (x > (i - 1) * xst) & (x <= (i * xst)) &
        (y > (j - 1) * yst) & (y <= (j * yst))
      isempty <- F
      if ((i > 1) && (j > 1) && (sgrid[i - 1, j - 1] == 0))
        isempty <- T
      else if ((i > 1) && (sgrid[i - 1, j] == 0))
        isempty <- T
      else if ((i > 1) && (j < n) && (sgrid[i - 1, j + 1] == 0))
        isempty <- T
      else if ((j > 1) && (sgrid[i, j - 1] == 0))
        isempty <- T
      else if ((j < n) && (sgrid[i, j + 1] == 0))
        isempty <- T
      else if ((i < n) && (j > 1) && (sgrid[i + 1, j - 1] == 0))
        isempty <- T
      else if ((i < n) && (sgrid[i + 1, j] == 0))
        isempty <- T
      else if ((i < n) && (j < n) && (sgrid[i + 1, j + 1] == 0))
        isempty <- T
    }
  }
}
```



**Figure A.1:** Cell colony edges are detected by 2D binning of cell center locations. Dots represent cell centers within the well H6 of plate J107-2C. Each of the nine images is segmented into 12 horizontal and 12 vertical sections yielding 144 tiles (1296 bins for the entire well). The tiles are colored according to the number of non-empty neighboring bins.

```

        isempty <- T
        empty[ispos] <- isempty
    }
}
return(empty)
}

```

Times for my version are mean 7.902 ms (with sd 2.744 ms) and total runtime for a plate is 3.034 s, while theirs runs with mean 529.369 ms (sd 22.81 ms) and for a plate 203.278 s.



## Appendix B

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# InfectX Protocols

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### B.1 Materials and Methods for Wet-Lab Procedures

The following sections describe materials and methods employed in pathogen specific protocols. This information has been published in [Rämö et al. \(2014\)](#) and is only reproduced for the reader's convenience.

**B. *henselae*-specific protocol.** *Bartonella henselae* ATCC49882<sup>T</sup>  $\Delta$ bepG containing plasmid pCD353 ([Dehio et al., 1998](#)) for IPTG-inducible expression of GFP were grown on Columbia base agar (CBA) plates supplemented with 5% defibrinated sheep blood (Oxoid) and 50  $\mu$ g ml<sup>-1</sup> kanamycin. Bacteria were incubated at 35 °C in 5% CO<sub>2</sub> for 72 h before re-streaking them on fresh CBA and further growth for 48 h. Cells were washed once after siRNA-transfection with M199 (Invitrogen)/10% FBS using a plate washer (ELx50-16, BioTek). Cells were infected with *B. henselae* at an MOI of 400 in 50  $\mu$ l of M199/10% FBS and 0.5 mM IPTG (Applichem) and were incubated at 35 °C in 5% CO<sub>2</sub> for 30 h. Fixation at RT was performed using a Multidrop 384 (Thermo Scientific) to wash cells with 50  $\mu$ l of PBS, fixed in 20  $\mu$ l of 3.7% PFA for 10 min, and washed once more with 50  $\mu$ l of PBS. Staining was performed on a Biomek liquid handling platform. Fixed cells were washed twice with 25  $\mu$ l of PBS and blocked in PBS/0.2% BSA for 10 min. Extracellular bacteria were labeled with a rabbit serum 2037 against *B. henselae* ([Dehio et al., 1997](#)) and a secondary antibody goat anti rabbit A647 (Jackson Immuno) in PBS/0.2% BSA. Antibodies were incubated for 30 min each and both incubations were followed by two washings with 25  $\mu$ l of PBS. Cells were then permeabilized with 20  $\mu$ l of 0.1% Triton X-100 (Sigma) for 10 min and afterwards washed twice with 25  $\mu$ l of PBS, followed by the addition of 20  $\mu$ l of staining solution (PBS containing 1.5  $\mu$ g ml<sup>-1</sup> DY-547-

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Phalloidin, Dyomics and  $1\text{ }\mu\text{g ml}^{-1}$  DAPI, Roche). After 30 min of incubation in the staining solution, cells were washed twice with  $25\text{ }\mu\text{l PBS}$ , followed by a final addition of  $50\text{ }\mu\text{l}$  of PBS.

**B. *abortus*-specific protocol.** *Brucella abortus* 2308 pJC43 (*aphT::GFP*) (Celli et al., 2005) were grown in tryptic soy broth (TSB) medium containing  $50\text{ }\mu\text{g ml}^{-1}$  kanamycin for 20 h at  $37^\circ\text{C}$  and shaking (100 rpm) to an OD of 0.8–1.1.  $50\text{ }\mu\text{l}$  of DMEM/10% containing bacteria was added per well to obtain a final MOI of 10000 using a cell plate washer (ELx50-16, BioTek). Plates were then centrifuged at  $400\text{ g}$  for 20 min at  $4^\circ\text{C}$  to synchronize bacterial entry. After 4 h incubation at  $37^\circ\text{C}$  and 5%  $\text{CO}_2$ , extracellular bacteria were killed by exchanging the infection medium by  $50\text{ }\mu\text{l}$  medium supplemented with 10% FBS and  $100\text{ }\mu\text{g ml}^{-1}$  gentamicin (Sigma). After a total infection time of 44 h cells were fixed with 3.7% PFA for 20 min at RT with the cell plate washer. Staining was performed using a Biomek liquid handling platform. Cells were washed twice with PBS and permeabilized with 0.1% Triton X (Sigma) for 10 minute. Then, cells were washed twice with PBS, followed by addition of  $20\text{ }\mu\text{l}$  of staining solution which includes DAPI ( $1\text{ }\mu\text{g ml}^{-1}$ , Roche) and DY-547-phalloidin ( $1.5\text{ }\mu\text{g ml}^{-1}$ , Dyomics) in 0.5% BSA in PBS. Cells were incubated with staining solution for 30 min at RT, washed twice with PBS, followed by final addition of  $50\text{ }\mu\text{l}$  PBS.

**L. *monocytogenes*-specific protocol.** After washing an overnight culture of *L. monocytogenes* EGDe.Prfa\*GFP three times with PBS, bacteria were diluted in DMEM supplemented with 1% FBS. Cells were infected at a MOI of 25 in  $30\text{ }\mu\text{l}$  infection medium per well. After centrifugation at 1000 rpm for 5 min and incubation for 1 h at  $37^\circ\text{C}$  in 5%  $\text{CO}_2$  to allow the bacteria to enter, extracellular bacteria were killed by exchanging the infection medium by  $30\text{ }\mu\text{l}$  DMEM supplemented with 10% FBS and  $40\text{ }\mu\text{g ml}^{-1}$  gentamicin (Gibco). Both medium exchange steps were carried out with a plate washer (ELx50-16, BioTek). After additional 4 h at  $37^\circ\text{C}$  in a 5%  $\text{CO}_2$  atmosphere, cells were fixed for 15 min at RT by adding  $30\text{ }\mu\text{l}$  of 8% PFA in PBS to each well using a multidrop 384 device (Thermo Electron Corporation). PFA was removed by four washes with  $500\text{ }\mu\text{l}$  PBS per well using the Power Washer 384 (Tecan). Fixed cells were stained for nuclei, actin and bacterially secreted InIC. First, cells were incubated for 30 min with  $10\text{ }\mu\text{l}$  per well of primary staining solution (0.2% saponin, PBS) containing rabbit derived anti-InIC serum (1:250). After four washes with  $40\text{ }\mu\text{l}$  PBS per well cells were stained with  $10\text{ }\mu\text{l}$  per well of the secondary staining solution (0.2% saponin, PBS) containing Alexa Fluor-546 coupled anti-rabbit antibody (1:250, Invitrogen), DAPI ( $0.7\text{ }\mu\text{g ml}^{-1}$ , Roche), and DY-647-Phalloidin ( $2\text{ }\mu\text{g ml}^{-1}$ , Dyomics). After four washes with  $40\text{ }\mu\text{l}$  PBS per well, the cells were

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## B.1. Materials and Methods for Wet-Lab Procedures

kept in 40 µl PBS per well. The staining procedure was carried out with a Tecan freedom evo robot.

**S. typhimurium-specific protocol.** All liquid handing stages of infection, fixation, and immunofluorescence staining were performed on a liquid handling robot (BioTek; EL406). For infection the *S. typhimurium* strain S.Tm<sup>SopE\_pM975</sup> was used. This strain is a single effector strain, only expressing SopE out of the main four SPI-1 encoded effectors (SipA, SopB, SopE2 and SopE). Additionally this strain harbors a plasmid (pM975) that expresses GFP under the control of a SPI2 (ssaG)-dependent promotor. The bacterial solution was prepared by cultivating a 12 h culture in 0.3 M LB medium containing 50 µg ml<sup>-1</sup> streptomycin and 50 µg ml<sup>-1</sup> ampicillin. Afterwards a 4 h subculture (1:20 diluted from the 12 h culture) was cultivated in 0.3 M LB medium containing 50 µg ml<sup>-1</sup> streptomycin, which reached an OD<sub>600 nm</sub> ≈ 1.0 after the respective 4 h of incubation time. To perform the infection, 16 µl of diluted *S. typhimurium* (MOI of 80) were added to the HeLa cells. After 20 min of incubation at 37 °C and 5% CO<sub>2</sub>, the *S. typhimurium*-containing media was replaced by 60 µl DMEM/10% FBS containing 50 µg µl<sup>-1</sup> streptomycin and 400 µg µl<sup>-1</sup> gentamicin to kill all remaining extracellular bacteria. After additional 3 h 40 min incubation at 37 °C and 5% CO<sub>2</sub>, cells were fixed by adding 35 µl 4% PFA, 4% sucrose in PBS for 20 min at RT. The fixation solution was removed by adding 60 µl PBS containing 400 µg ml<sup>-1</sup> gentamicin. Cells were permeabilized for 5 min with 40 µl 0.1% Triton X-100 (Sigma-Aldrich). Afterwards 24 µl of staining solution containing DAPI (1:1000, Sigma-Aldrich) and DY-547-phalloidin (1.2 µg ml<sup>-1</sup>, Dyomics) was added (prepared in blocking buffer consisting of 4% BSA and 4% Sucrose in PBS). After 1 h of incubation at RT, cells were washed three times with PBS followed by the addition of 60 µl PBS containing 400 µg ml<sup>-1</sup> gentamicin.

**S. flexneri-specific protocol.** *S. flexneri* M90T  $\Delta virG$  pCK100 (PuhpT::dsRed) were harvested in exponential growth phase and coated with 0.005% poly-L-lysine (Sigma-Aldrich). Afterwards, bacteria were washed with PBS and resuspended in assay medium (DMEM, 2 mM L-Glutamine, 10 mM HEPES). 20 µl of bacterial suspension was added to each well with a final MOI of 15. Plates were then centrifuged for 1 min at 37 °C and incubated at 37 °C and 5% CO<sub>2</sub>. After 30 min of infection, 75 µl were aspirated from each well and monensin (Sigma) and gentamicin (Gibco) were added to a final concentration of 66.7 µM and 66.7 µg ml<sup>-1</sup>, respectively. After a total infection time of 3.5 h, cells were fixed in 4% PFA for 10 min. Liquid handling was performed using the Multidrop 384 (Thermo Scientific) for dispensing steps and a plate washer (ELx50-16, BioTek) for aspiration steps. For immunofluorescent staining, cells were washed with PBS using the Power Washer 384 (Tecan). Subsequently, cells were incubated

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with a mouse anti-human IL-8 antibody (1:300, BD Biosciences) in staining solution (0.2% saponin in PBS) for 2 h at RT. After washing the cells with PBS, Hoechst (5 µg ml<sup>-1</sup>, Invitrogen), DY-495-phalloidin (1.2 µg ml<sup>-1</sup>, Dyomics) and Alexa Fluor 647-coupled goat anti-mouse IgG (1:400, Invitrogen) were added and incubated for 1 h at RT. The staining procedure was performed using the Biomek NXP Laboratory Automation Workstation (Beckman Coulter).

**Adenovirus-specific protocol.** All liquid handling stages of infection, fixation, and immunofluorescence staining were performed on the automated pipetting system Well Mate (Thermo Scientific Matrix) and washer Hydrospeed (Tecan). For infection screens recombinant Ad2\_ΔE3B-eGFP (short Adenovirus) was utilized as described before ([Suomalainen et al., 2013](#); [Yakimovich et al., 2012](#)). Adenovirus was added to cells at an MOI of 0.1 in 10 µl of an infection media/FBS (DMEM supplemented with L-glutamine, 10% FBS, 1% Pen/Strep, Invitrogen). Screening plates were incubated at 37 °C for 16 h, and cells were fixed by adding 21 µl of 16% PFA directly to the cells in culture media for 45 min at RT or long-term storage at 4 °C. Cells were washed 2 times with PBS/25 mM NH<sub>4</sub>Cl, permeabilized with 25 µl 0.1% Triton X-100 (Pharmaciebiotheek). After 2 washes with PBS the samples were incubated at RT for 1 h with 25 µl staining solution (PBS) containing DAPI (1 µg ml<sup>-1</sup>, Sigma-Aldrich) and DY-647-phalloidin (1 µg ml<sup>-1</sup>, Dyomics), washed 2 times with PBS and stored until imaging in 50 µl PBS/NaN<sub>3</sub>.

**Rhinovirus-specific protocol.** All liquid handling stages of infection, fixation, and immunofluorescence staining were performed on the automated pipetting system Well Mate (Thermo Scientific Matrix) and washer Hydrospeed (Tecan). For infection assays with human Rhinovirus serotype 1a (HRV1a) were carried out as described, except that the anti-VP2 antibody Mab 16/7 was used for staining of the infected cells as described earlier ([Jurgeit et al., 2012, 2010](#); [Mosser et al., 2002](#)). Rhinovirus at an MOI of 8 was added to cells in 20 µl of an infection media/BSA (DMEM supplemented with GlutaMAX, 30 mM MgCl<sub>2</sub> and 0.2% BSA, Invitrogen). Screening plates were incubated for 7 h at 37 °C, and cells were fixed by adding 33 µl of 16% PFA directly to the culture medium. Fixation was either for 30 min at RT or long term storage at 4 °C. Cells were washed twice with PBS/25 mM H<sub>2</sub>O, permeabilized with 50 µl 0.2% Triton X-100 (Sigma- Aldrich) followed by 3 PBS washes and blocking with PBS containing 1% BSA (Fraction V, Sigma-Aldrich). Fixed and permeabilized cells were incubated at RT for 1 h with diluted mabR16-7 antibody (0.45 µg ml<sup>-1</sup>) in PBS/1% BSA. Cells were washed 3 times with PBS and incubated with 25 µl secondary staining solution (PBS/1% BSA) containing Alexa Fluor 488 secondary antibody (1 µg ml<sup>-1</sup>, Invitrogen), DAPI (1 µg ml<sup>-1</sup>, Sigma-Aldrich),

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## B.2. Decision Trees for Infection Scoring

and DY-647-phalloidin ( $0.2 \mu\text{g ml}^{-1}$ , Dyomics). Cells were washed twice with PBS after 2 h of incubation in secondary staining solution and stored in  $50 \mu\text{l}$  PBS/ $\text{NaN}_3$ .

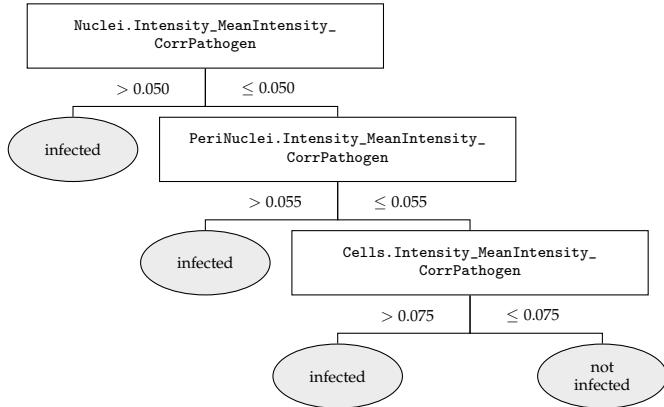
**Vaccinia virus-specific protocol.** All liquid handing stages of infection, fixation, and immunofluorescence staining were performed on a liquid handling robot (BioTek, EL406). For infection assays a recombinant WR VACV, WR E EGFP/L mCherry, was utilized. For infection, media was aspirated from the RNAi-transfected cell plates and replaced with  $40 \mu\text{l}$  of virus solution per well (MOI of 0.125). Screening plates were incubated for 1 h at  $37^\circ\text{C}$  to allow for infection, after which virus-containing media was removed and replaced with  $40 \mu\text{l}$  DMEM/10% FBS. 8 h after infection  $40 \mu\text{l}$  of DMEM/10% FBS containing  $20 \mu\text{M}$  cytosine arabinoside (AraC) was added to all wells to prevent virus DNA replication in secondary infected cells. 24 h after infection cells were fixed by the addition of  $20 \mu\text{l}$  18% PFA for 30 min followed by two PBS washes of  $80 \mu\text{l}$ . For immunofluorescence staining of EGFP, cells were incubated for 2 h in  $30 \mu\text{l}$  primary staining solution (0.5% Triton X-100, 0.5% BSA, PBS) per well, containing anti-GFP antibody (1:1000). Cells were washed twice in  $80 \mu\text{l}$  PBS, followed by the addition of  $30 \mu\text{l}$  secondary staining solution (0.5% BSA, PBS) containing Alexa Fluor 488 secondary antibody (1:1000), Hoechst (1:10000), and DY-647-phalloidin (1:1200, Dyomics). Cells were washed twice with  $80 \mu\text{l}$  PBS after 1 h incubation in secondary staining solution followed by the addition of  $80 \mu\text{l}$   $\text{H}_2\text{O}$ .

## B.2 Decision Trees for Infection Scoring

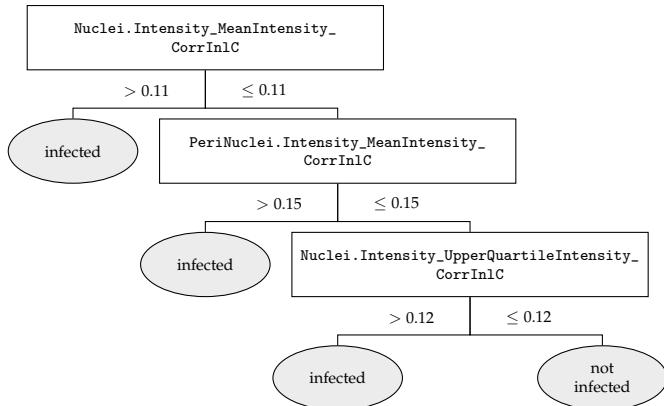
The decision trees for adenovirus and *Bartonella* are shown in section 4.5 while the ones corresponding to the remaining pathogens (*Brucella*, *Listeria*, rhinovirus, *Salmonella* and vaccinia virus) follow. Please refer to section 4.5 for more information of infection scoring.

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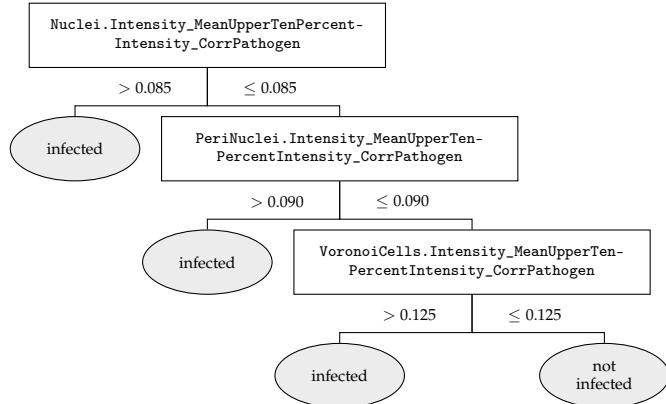
**Figure B.1:** Decision tree for *Brucella* infection scoring. While the first two decisions are modeled to capture what is considered a normal infection pattern, the last split imposes a high threshold for cells that have failed the first two steps to still be considered infected.



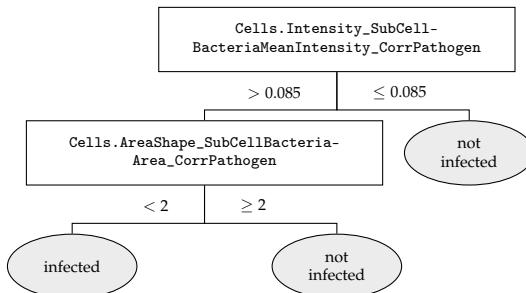
**Figure B.2:** The decision tree for *Listeria* infection scoring is based on a channel recording InIC localization and intensity instead of targeting the bacteria themselves.

## B.2. Decision Trees for Infection Scoring

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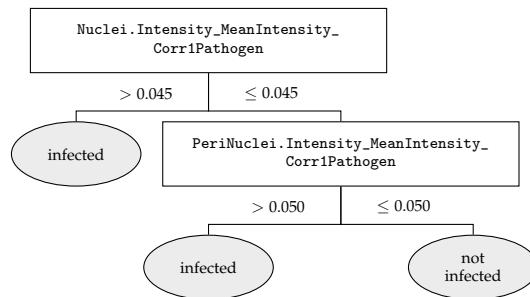
**Figure B.3:** Decision tree for rhinovirus infection scoring. Using the mean of the uppermost decile of pathogen channel intensity data yields the most stable results.



**Figure B.4:** Decision tree for *Salmonella* infection scoring. For a cell being considered infected, not only does the threshold for pathogen intensity throughout the cell need be exceeded but the bacteria also have to be sufficiently aggregated.

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**Figure B.5:** Decision tree for vaccinia virus infection scoring. A separate decision tree for distinguishing primary from secondary infections has been developed but is not shown.

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## Bibliography

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- Alberts, B., J. Wilson, and T. Hunt (2008). *Molecular biology of the cell*. New York: Garland Science.
- Allen, M. P. (1997). The origins and uses of regression analysis. In *Understanding Regression Analysis*, pp. 1–5. Springer US.
- Anderson, B. and M. Neuman (1997, apr.). *Bartonella* spp. as emerging human pathogens. *Clin. Microbiol. Rev.* 10(2), 203–219.
- Atluri, V. L., M. N. Xavier, M. F. de Jong, A. B. den Hartigh, and R. M. Tsolis (2011, sep.). Interactions of the Human Pathogenic Brucella Species with Their Hosts. *Annual Review of Microbiology* 65(1), 523–541.
- Baltimore, D. (1971, sep.). Expression of animal virus genomes. *Microbiol. Mol. Biol. Rev.* 35(3), 235–241.
- Bauch, A., I. Adamczyk, P. Buczek, F.-J. Elmer, K. Enimanev, P. Glyzewski, M. Kohler, T. Pylak, A. Quandt, C. Ramakrishnan, C. Beisel, L. Malmström, R. Aebersold, and B. Rinn (2011, jan.). openBIS: a flexible framework for managing and analyzing complex data in biology research. *BMC bioinformatics* 12(1), 468.
- Carpenter, A. E., T. R. Jones, M. R. Lamprecht, C. Clarke, I. H. Kang, O. Friman, D. A. Guertin, J. H. Chang, R. A. Lindquist, J. Moffat, P. Golland, and D. M. Sabatini (2006, jan.). CellProfiler: image analysis software for identifying and quantifying cell phenotypes. *Genome biology* 7(10), R100.
- Carthew, R. W. and E. J. Sontheimer (2009, feb.). Origins and Mechanisms of miRNAs and siRNAs. *Cell* 136(4), 642–655.

## BIBLIOGRAPHY

---

- Celli, J., S. P. Salcedo, and J.-P. Gorvel (2005, feb.). Brucella coopts the small GTPase Sar1 for intracellular replication. *Proceedings of the National Academy of Sciences of the United States of America* 102(5), 1673–8.
- Cerutti, H. and J. A. Casas-Mollano (2006, aug.). On the origin and functions of RNA-mediated silencing: from protists to man. *Current genetics* 50(2), 81–99.
- Condit, R. C., N. Moussatche, and P. Traktman (2006, jan.). In a nutshell: structure and assembly of the vaccinia virion. *Advances in virus research* 66, 31–124.
- Cossart, P. and A. Lebreton (2014, aug.). A trip in the "New Microbiology" with the bacterial pathogen Listeria monocytogenes. *FEBS letters* 588(15), 2437–45.
- Cossart, P. and P. J. Sansonetti (2004). Bacterial Invasion: The Paradigms of Enteroinvasive Pathogens. *Science* 304(5668), 242–248.
- Craighead, J. E. (2000). *Pathology and Pathogenesis of Human Viral Disease*. San Diego: Academic Press.
- Croxen, M. A. and B. B. Finlay (2010, jan.). Molecular mechanisms of Escherichia coli pathogenicity. *Nat Rev Micro* 8(1), 26–38.
- Czyż, D. M., L.-P. Potluri, N. Jain-Gupta, S. P. Riley, J. J. Martinez, T. L. Steck, S. Crosson, H. A. Shuman, and J. E. Gabay (2014, jan.). Host-directed antimicrobial drugs with broad-spectrum efficacy against intracellular bacterial pathogens. *mBio* 5(4), e01534–14.
- de Barsy, M., A. Jamet, D. Filopon, C. Nicolas, G. Laloux, J.-F. Rual, A. Muller, J.-C. Twizere, B. Nkengfac, J. Vandenhautte, D. E. Hill, S. P. Salcedo, J.-P. Gorvel, J.-J. Letesson, and X. De Bolle (2011, jul.). Identification of a Brucella spp. secreted effector specifically interacting with human small GTPase Rab2. *Cellular Microbiology* 13(7), 1044–1058.
- Dehio, C. (2005, aug.). Bartonella-host-cell interactions and vascular tumour formation. *Nat Rev Micro* 3(8), 621–631.
- Dehio, C., M. Meyer, J. Berger, H. Schwarz, and C. Lanz (1997, sep.). Interaction of Bartonella henselae with endothelial cells results in bacterial aggregation on the cell surface and the subsequent engulfment and internalisation of the bacterial aggregate by a unique structure, the invasome. *Journal of cell science* 110 ( Pt 1), 2141–2154.
- Dehio, M., A. Knorre, C. Lanz, and C. Dehio (1998, jul.). Construction of versatile high-level expression vectors for Bartonella henselae and the use of green fluorescent protein as a new expression marker. *Gene* 215(2), 223–229.

---

## Bibliography

- Dobson, A. P. and E. R. Carper (1996, feb.). Infectious Diseases and Human Population History. *BioScience* 46(2), 115–126 CR – Copyright © 1996 Oxford Universi.
- Echeverri, C. J. and N. Perrimon (2006, may.). High-throughput RNAi screening in cultured cells: a user's guide. *Nat Rev Genet* 7(5), 373–384.
- Fàbrega, A. and J. Vila (2013, apr.). Salmonella enterica serovar Typhimurium skills to succeed in the host: virulence and regulation. *Clinical microbiology reviews* 26(2), 308–41.
- Farber, J. M. and P. I. Peterkin (1991, sep.). Listeria monocytogenes, a food-borne pathogen. *Microbiological Reviews* 55(3), 476–511.
- Gabor, D. (1946). Theory of communication. Part 1: The analysis of information.
- Haglund, C. M. and M. D. Welch (2011, oct.). Pathogens and polymers: Microbe–host interactions illuminate the cytoskeleton. *The Journal of Cell Biology* 195(1), 7–17.
- Ham, H., A. Sreelatha, and K. Orth (2011, sep.). Manipulation of host membranes by bacterial effectors. *Nat Rev Micro* 9(9).
- Hannon, G. J. (2002, jul.). RNA interference. *Nature* 418(6894), 244–251.
- Haraga, A., M. B. Ohlson, and S. I. Miller (2008, jan.). Salmonellae interplay with host cells. *Nat Rev Micro* 6(1), 53–66.
- Haralick, R. M., K. Shanmugam, and I. Dinstein (1973). Textural Features for Image Classification.
- Harms, A. and C. Dehio (2012, jan.). Intruders below the radar: molecular pathogenesis of Bartonella spp. *Clinical microbiology reviews* 25(1), 42–78.
- Hawn, T. R., J. A. Shah, and D. Kalman (2015, mar.). New tricks for old dogs: countering antibiotic resistance in tuberculosis with host-directed therapeutics. *Immunological Reviews* 264(1), 344–362.
- Hayashi, F. (2000). *Econometrics*. Princeton, NJ [u.a.]: Princeton Univ. Press.
- Honeychurch, K. M., G. Yang, R. Jordan, and D. E. Hruby (2007, jul.). The vaccinia virus F13L YPPL motif is required for efficient release of extracellular enveloped virus. *Journal of virology* 81(13), 7310–5.
- Huang, J. and J. H. Brumell (2014, feb.). Bacteria-autophagy interplay: a battle for survival. *Nat Rev Micro* 12(2), 101–114.

## BIBLIOGRAPHY

---

- Hulo, C., E. de Castro, P. Masson, L. Bougueret, A. Bairoch, I. Xenarios, and P. Le Mercier (2011, jan.). ViralZone: a knowledge resource to understand virus diversity. *Nucleic acids research* 39(Database issue), D576–82.
- Jacobs, S. E., D. M. Lamson, K. St George, and T. J. Walsh (2013, jan.). Human rhinoviruses. *Clinical microbiology reviews* 26(1), 135–62.
- Johnson, S. A. and T. Hunter (2005, jan.). Kinomics: methods for deciphering the kinome. *Nat Meth* 2(1), 17–25.
- Jurgeit, A., R. McDowell, S. Moese, E. Meldrum, R. Schwendener, and U. F. Greber (2012, jan.). Niclosamide is a proton carrier and targets acidic endosomes with broad antiviral effects. *PLoS pathogens* 8(10), e1002976.
- Jurgeit, A., S. Moese, P. Roulin, A. Dorsch, M. Lötzerich, W.-M. Lee, and U. F. Greber (2010, jan.). An RNA replication-center assay for high content image-based quantifications of human rhinovirus and coxsackievirus infections. *Virology journal* 7(1), 264.
- Kahneman, D. (2012, mar.). Quotation: Kahneman on Contingencies. *Journal of the Experimental Analysis of Behavior* 97(2), 182.
- Kamentsky, L., T. R. Jones, A. Fraser, M.-A. Bray, D. J. Logan, K. L. Madden, V. Ljosa, C. Rueden, K. W. Eliceiri, and A. E. Carpenter (2011, apr.). Improved structure, function and compatibility for CellProfiler: modular high-throughput image analysis software. *Bioinformatics (Oxford, England)* 27(8), 1179–80.
- Kanehisa, M. and S. Goto (2000, jan.). KEGG: Kyoto Encyclopedia of Genes and Genomes. *Nucleic Acids Research* 28(1), 27–30.
- Kasper, C. A. (2012). *Amplifying the innate immune response : cell-cell propagation of proinflammatory signals during bacterial infection*. Phd thesis, University of Basel.
- Kim, D. H. and J. J. Rossi (2007, mar.). Strategies for silencing human disease using RNA interference. *Nat Rev Genet* 8(3), 173–184.
- Kopecko, D. J., L. Hu, and K. J. M. Zaal (2001, aug.). Campylobacter jejuni – microtubule-dependent invasion. *Trends in Microbiology* 9(8), 389–396.
- Leirião, P., C. D. Rodrigues, S. S. Albuquerque, and M. M. Mota (2004, dec.). Survival of protozoan intracellular parasites in host cells. *EMBO Reports* 5(12), 1142–1147.
- Lenaerts, L., E. De Clercq, and L. Naesens (2008, nov.). Clinical features and treatment of adenovirus infections. *Reviews in Medical Virology* 18(6), 357–374.

## Bibliography

---

- Littman, R. J. (2009, oct.). The Plague of Athens: Epidemiology and Paleopathology. *Mount Sinai Journal of Medicine: A Journal of Translational and Personalized Medicine* 76(5), 456–467.
- Majowicz, S. E., J. Musto, E. Scallan, F. J. Angulo, M. Kirk, S. J. O'Brien, T. F. Jones, A. Fazil, and R. M. Hoekstra (2010, mar.). The global burden of non-typhoidal Salmonella gastroenteritis. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* 50(6), 882–9.
- Manning, G., D. B. Whyte, R. Martinez, T. Hunter, and S. Sudarsanam (2002, dec.). The Protein Kinase Complement of the Human Genome. *Science* 298(5600), 1912–1934.
- Marennikova, S. S., R. C. Condit, and R. W. Moyer (2005). Vaccinia Virus. In *Orthopoxviruses Pathogenic for Humans SE - 3*, pp. 19–87. Springer US.
- Masters, J. R. (2002, apr.). HeLa cells 50 years on: the good, the bad and the ugly. *Nat Rev Cancer* 2(4), 315–319.
- McCray, A. T. and N. C. Ide (2000, jan.). Design and Implementation of a National Clinical Trials Registry. *Journal of the American Medical Informatics Association : JAMIA* 7(3), 313–323.
- Mercer, J. and A. Helenius (2008, apr.). Vaccinia Virus Uses Macropinocytosis and Apoptotic Mimicry to Enter Host Cells. *Science* 320(5875), 531–535.
- Misselwitz, B., S. Dilling, P. Vonaesch, R. Sacher, B. Snijder, M. Schlumberger, S. Rout, M. Stark, C. von Mering, L. Pelkmans, and W.-D. Hardt (2011, mar.). RNAi screen of Salmonella invasion shows role of COPI in membrane targeting of cholesterol and Cdc42. *Molecular Systems Biology* 7, 474.
- Mohr, S., C. Bakal, and N. Perrimon (2010). Genomic Screening with RNAi: Results and Challenges. *Annual review of biochemistry* 79, 37–64.
- Mosser, A. G., R. Brockman-Schneider, S. Amineva, L. Burchell, J. B. Sedgwick, W. W. Busse, and J. E. Gern (2002, mar.). Similar frequency of rhinovirus-infectible cells in upper and lower airway epithelium. *The Journal of infectious diseases* 185(6), 734–43.
- Obbard, D. J., K. H. J. Gordon, A. H. Buck, and F. M. Jiggins (2009, jan.). The evolution of RNAi as a defence against viruses and transposable elements. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences* 364(1513), 99–115.
- Perrimon, N. and B. Mathey-Prevot (2007, jan.). Applications of High-Throughput RNA Interference Screens to Problems in Cell and Developmental Biology. *Genetics* 175(1), 7–16.

## BIBLIOGRAPHY

---

- Prussia, A., P. Thepchatri, J. P. Snyder, and R. K. Plemper (2011). Systematic Approaches towards the Development of Host-Directed Antiviral Therapeutics. *International Journal of Molecular Sciences* 12(6), 4027.
- Rämö, P., A. Drewek, C. Arriumerlou, N. Beerewinkel, H. Ben-Tekaya, B. Cardel, A. Casanova, R. Conde-Alvarez, P. Cossart, G. Csúcs, S. Eicher, M. Emmenlauer, U. Greber, W.-D. Hardt, A. Helenius, C. Kasper, A. Kauffmann, S. Kreibich, A. Kühbacher, P. Kunszt, S. H. Low, J. Mercer, D. Mudrak, S. Muntwiler, L. Pelkmans, J. Pizarro-Cerdá, M. Podvinec, E. Pujadas, B. Rinn, V. Rouilly, F. Schmich, J. Siebourg-Polster, B. Snijder, M. Stebler, G. Studer, E. Szczurek, M. Truttmann, C. von Mering, A. Vonderheide, A. Yakimovich, P. Bühlmann, and C. Dehio (2014, jan.). Simultaneous analysis of large-scale RNAi screens for pathogen entry. *BMC genomics* 15(1), 1162.
- Ray, K., B. Marteyn, P. J. Sansonetti, and C. M. Tang (2009, may.). Life on the inside: the intracellular lifestyle of cytosolic bacteria. *Nat Rev Micro* 7(5), 333–340.
- Rocha, L., L. Velho, and P. C. P. Carvalho (2002). Image moments-based structuring and tracking of objects.
- Rolain, J. M. and D. Raoult (2006). *Bartonella* spp. In *Principles and Practice of Clinical Bacteriology*, pp. 295–304. John Wiley & Sons, Ltd.
- Rouilly, V., E. Pujadas, B. Hullar, C. Balazs, P. Kunszt, and M. Podvinec (2012). iBRAIN2: automated analysis and data handling for RNAi screens. *Studies in health technology and informatics* 175, 205–213.
- Saurabh, S., A. Vidyarthi, and D. Prasad (2014). RNA interference: concept to reality in crop improvement. *Planta* 239(3), 543–564.
- Schroeder, G. N. and H. Hilbi (2008, jan.). Molecular pathogenesis of *Shigella* spp.: controlling host cell signaling, invasion, and death by type III secretion. *Clinical microbiology reviews* 21(1), 134–56.
- Silver, L. L. (2011, jan.). Challenges of Antibacterial Discovery. *Clinical Microbiology Reviews* 24(1), 71–109.
- Sittampalam, G. S., N. P. Coussens, H. Nelson, M. Arkin, D. Auld, C. Austin, B. Bejcek, M. Glicksman, J. Inglese, P. W. Iversen, Z. Li, J. McGee, O. McManus, L. Minor, A. Napper, J. M. Peltier, T. Riss, O. J. O. J. Trask, and J. Weidner (Eds.) (2004). *Assay Guidance Manual*.
- Smith, A. (2012). Cellular Biochemistry II: Cellular Infection.
- Stevens, J. M., E. E. Galyov, and M. P. Stevens (2006, feb.). Actin-dependent movement of bacterial pathogens. *Nat Rev Micro* 4(2), 91–101.

---

## Bibliography

- Suomalainen, M., S. Luisoni, K. Boucke, S. Bianchi, D. A. Engel, and U. F. Greber (2013, nov.). A direct and versatile assay measuring membrane penetration of adenovirus in single cells. *Journal of virology* 87(22), 12367–79.
- Taubenberger, J. and D. M. Morens (2006). 1918 Influenza: the Mother of All Pandemics. *Emerging Infectious Diseases Volume* 12(1).
- Treadgold, W. (1997, jun.). *A History of the Byzantine State and Society*. Stanford: Stanford University Press.
- Truttmann, M. C., B. Misselwitz, S. Huser, W.-D. Hardt, D. R. Critchley, and C. Dehio (2011, nov.). Bartonella henselae engages inside-out and outside-in signaling by integrin beta1 and talin1 during invasome-mediated bacterial uptake. *Journal of cell science* 124(Pt 21), 3591–3602.
- Veiga, E. and P. Cossart (2005, sep.). Listeria hijacks the clathrin-dependent endocytic machinery to invade mammalian cells. *Nature cell biology* 7(9), 894–900.
- von Bargen, K., J.-P. Gorvel, and S. P. Salcedo (2012, may.). Internal affairs: investigating the Brucella intracellular lifestyle. *FEMS Microbiology Reviews* 36(3), 533–562.
- Wang, Y., H. Huang, M. Sun, Q. Zhang, and D. Guo (2012, jan.). T3DB: an integrated database for bacterial type III secretion system. *BMC bioinformatics* 13(1), 66.
- Whitehead, K. A., R. Langer, and D. G. Anderson (2009, feb.). Knocking down barriers: advances in siRNA delivery. *Nat Rev Drug Discov* 8(2), 129–138.
- Wilson, R. C. and J. A. Doudna (2013, may.). Molecular Mechanisms of RNA Interference. *Annual Review of Biophysics* 42(1), 217–239.
- World Health Organization (2003). Fact sheet No. 211: Influenza.
- World Health Organization (2012). Mortality and global health estimates: Causes of death.
- World Health Organization (2014). Antimicrobial resistance: global report on surveillance 2014. Technical report, World Health Organization.
- Yakimovich, A., H. Gumpert, C. J. Burckhardt, V. A. Lütschg, A. Jurgeit, I. F. Sbalzarini, and U. F. Greber (2012, sep.). Cell-free transmission of human adenovirus by passive mass transfer in cell culture simulated in a computer model. *Journal of virology* 86(18), 10123–37.

## BIBLIOGRAPHY

---

- Zhou, Y.-J., J.-P. Zhu, T. Zhou, Q. Cheng, L.-X. Yu, Y.-X. Wang, S. Yang, Y.-F. Jiang, W. Tong, F. Gao, H. Yu, G.-X. Li, and G.-Z. Tong (2014, jan.). Identification of differentially expressed proteins in porcine alveolar macrophages infected with virulent/attenuated strains of porcine reproductive and respiratory syndrome virus. *PloS one* 9(1), e85767.
- Zietz, B. P. and H. Dunkelberg (2004). The history of the plague and the research on the causative agent *Yersinia pestis*. *International Journal of Hygiene and Environmental Health* 207(2), 165–178.

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## **Epilogue**

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A few final words. Test 2



## Declaration of originality

The signed declaration of originality is a component of every semester paper, Bachelor's thesis, Master's thesis and any other degree paper undertaken during the course of studies, including the respective electronic versions.

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