

MATHEMATICS 412: TOPOLOGY  
TOPOLOGICAL DATA ANALYSIS PROJECT  
PART 1: BIBLIOGRAPHY

TopoCAT : Exploring the Potential Role of TDA in Detecting  
Lung Cancer

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# Part I

## Background

# Chapter 1

## The Problem

1.1 Idea and Inspiration

1.2 Hypothesis

1.3 Data

# Chapter 2

## The Approach

### 2.1 Progress So Far

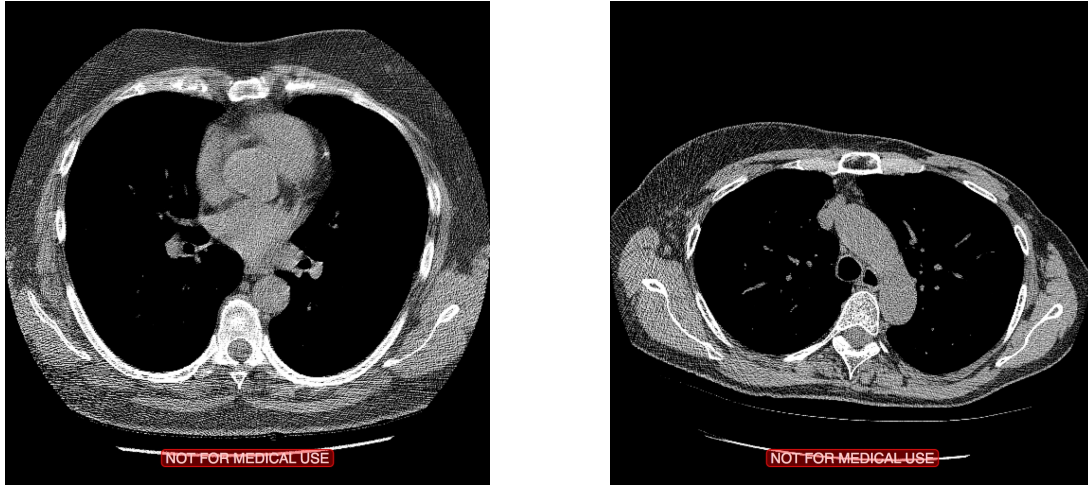
Some preliminary data processing and feature extraction using the TDA toolkit has yielded some interesting, early results. At the same time, it has uncovered some dilemmas including: how to best down-sample an image to preserve important information while making processing the image practical, what is the best way to generate a point cloud from a down-sampled image, what is the best way to summarize the persistence diagrams of each individual slide in a CT scan and the CT scan as a whole.

#### 2.1.1 Visualizing CT Scans in Osirix Lite

Osirix Lite is the free version of a popular software that is used to visualize and keep track of medical imagery, specifically scans composed of individual images taken over some height function (like CT scans).

Referring to the annotations spreadsheet that accompanies the LIDC-IDRI, a couple subjects were used to get a sense of the visual differences between patients with a large number of nodules and those with relatively "clear" lungs. The two CT scans selected were ID's 321 (25 nodules) and 306 (0 nodules).

Below are cross sections of the lung that illustrate the difference between images of lungs with virtually no growths or nodules and lungs with a large amount of nodules:



(a) A Slide from the CT Scan of Patient 306    (b) A Slide from the CT Scan of Patient 321

Figure 2.1: A figure with two subfigures

While these cross sections only illustrate the presence of nodules for a particular "height" within the lungs of the two patients, visualizing the images of these cross sections helped to foster an intuition about the topological features of the lungs or images of the lungs. Just from these images, one can see that the number of unconnected components in the image of the patient with a large number of nodules is greater than that of the patient with clear lungs.

### 2.1.2 Filtering Individual Slides of CT Scans by Pixel Intensity

The first approach to generating persistence diagrams was to triangulate an image based on pixel intensity (see `generateS.m` in Appendix) values and use that distance matrix to filter the image (`testScript.m` in Appendix). The findings did not seem to be point to any strong difference between cloudy and clear lungs using this filtration (more work needs to be done on this route to see if it yields anything interesting).

Below are the pre-processed images of the lungs before analysis with `rcalmfscm.m`.

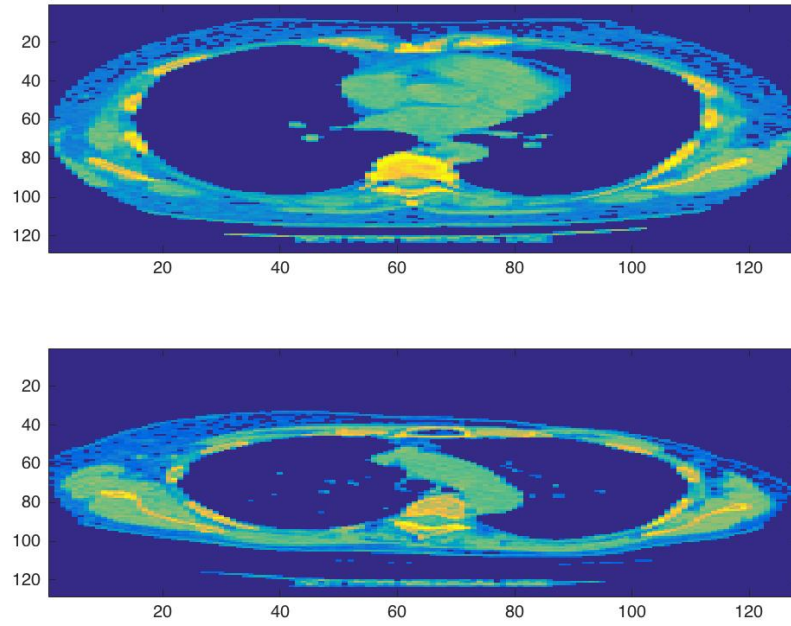


Figure 2.2: Post Image Processing: Clear Lung above Cloudy Lung

Below are the 0-dimensional and 1-dimensional persistence diagrams for the filtration of the image.



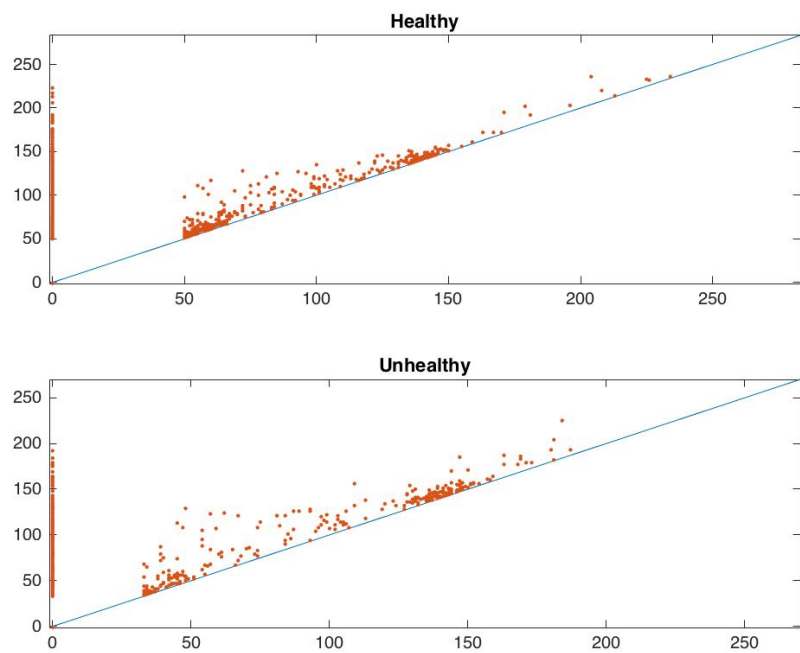


Figure 2.3:  $Dgm_0$

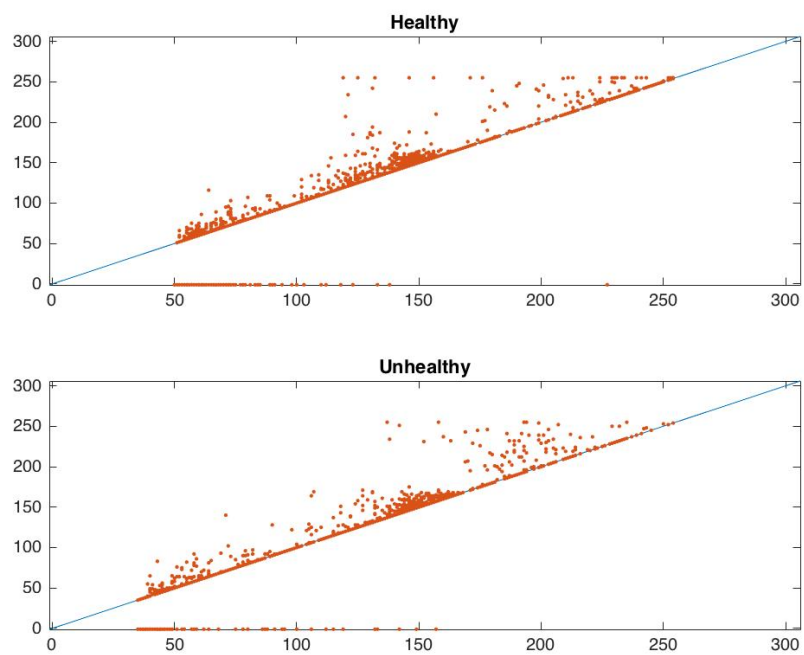


Figure 2.4:  $Dgm_1$

### 2.1.3 Filtering Individual Slides of CT Scans by Generating Point Cloud of Image and Embedding Cloud in $\mathbb{R}^2$

This approach involved flooring the background of the image and generating points out of the significant pixels (i.e. making the bright pixels in the image vertices). This point cloud was embedded in  $\mathbb{R}^2$  and `rca1dm.m` was used to compute the 0-dimensional and 1-dimensional persistence diagrams (see `testScript2.m` in Appendix). Below are images detailing the steps of the process:

Below are the original images in grayscale:

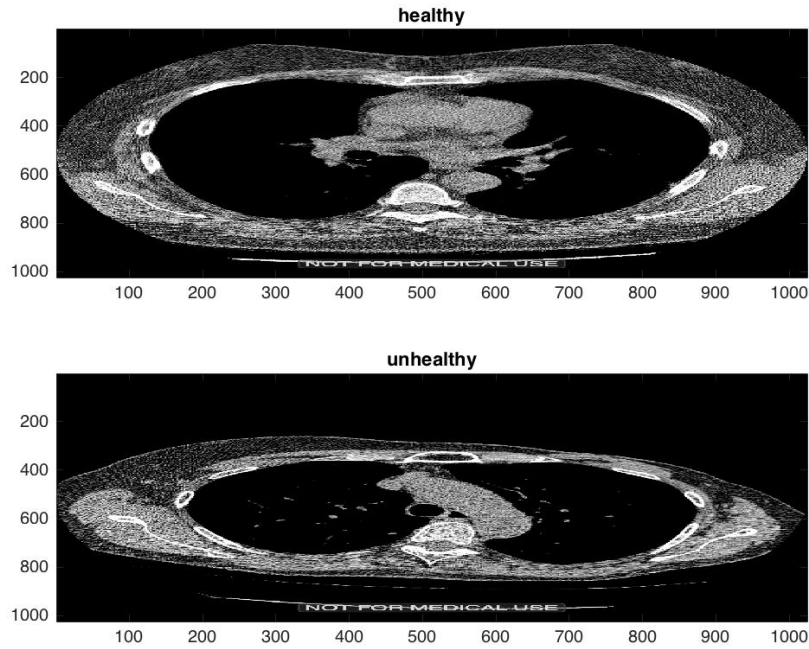


Figure 2.5: Grayscale Images of the Lungs at a Cross-section for Patients 306 and 321

Below are the same images but have been down-sampled by a factor of 0.0625 to make the size of the point cloud manageable:

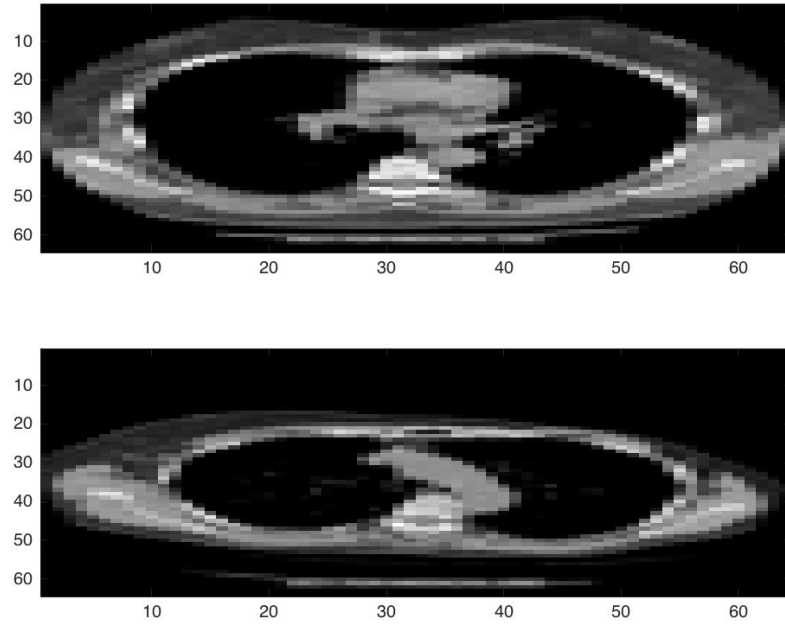


Figure 2.6: Grayscale, Down-sampled Images

Below are the down-sampled images, where the pixels with low intensity are floored to exclude insignificant points from the point cloud:

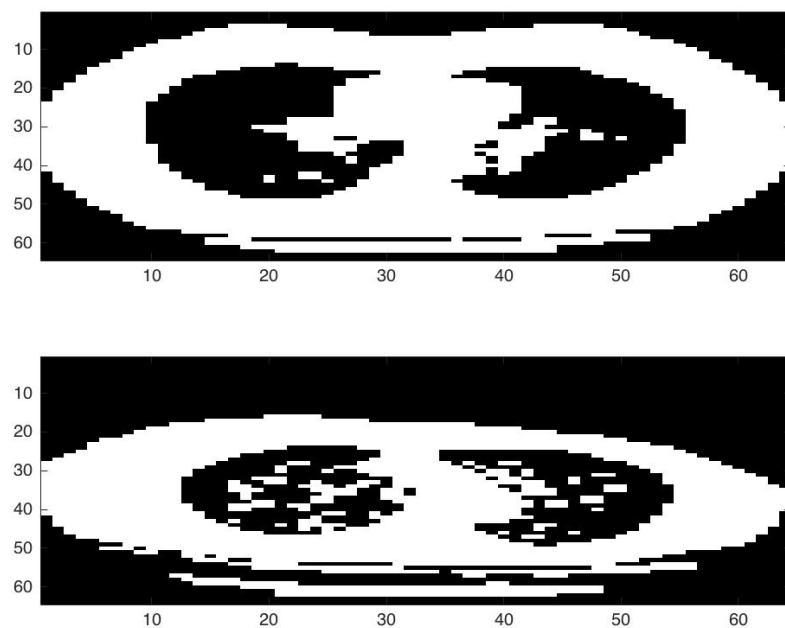


Figure 2.7: Grayscale, Down-sampled Images with Pixel Values on a Binary Scale

Below are the persistence diagrams after running `rca1dm.m`:

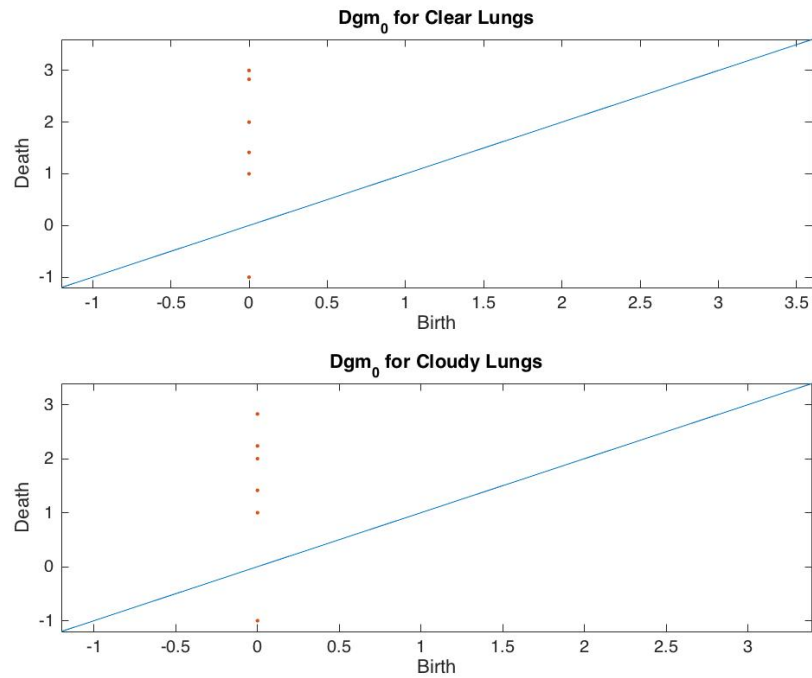


Figure 2.8: 0-Dimensional Persistence Diagram

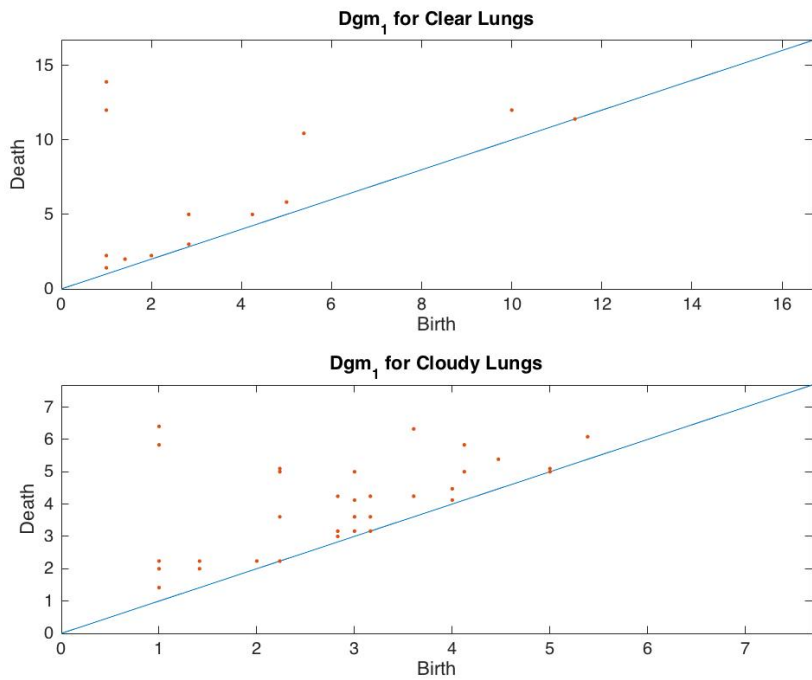


Figure 2.9: 1-Dimensional Persistence Diagram

As can be seen in the persistence plots, there seem to be some differences (in this very specific case) between lungs with nodules and lungs without nodules in terms of topological features of their CT scans. This is a very promising start and while we are looking into a few different filtrations to try.

### **2.1.4 Quantifying Topological Features of an Entire Scan from Individual Slides**

Some headway has been made in exploring ways to summarize the information of  $N$  slides for a particular CT scan. In the Appendix, `testScript3.m` and `testScript4.m` are early attempts at computing persistence using the method of the previous section and tiling up metrics on those values into a vector. For example, by looking at some persistence diagrams of different slides by using the method of 2.1.3, one can see that the "clear" lungs have one or two very persistent 1-cycles and a couple very insignificant 1-cycles. However, for slides of "cloudy" lungs, there is a more even distribution. Therefore, `testScript3.m` and `testScript4.m` are concatenating the standard deviation of the 1-dimensional persistence intervals and comparing them for healthy and cloudy lungs. More ways of using the data from each slide will be implemented to see what can maximize the difference between CT scans of patients with clear and cloudy lungs.

## **2.2 Future Directions**

### **2.2.1 Exploring Different Filtrations and Point Cloud Constructions**

### **2.2.2 Quantifying Topological Features of an Entire Scan from Individual Slides (cont.)**

More work needs to be done in exploring how the topological features of one image's point cloud can be combined with the other images in a CT scan to create a vector of information that represents one CT scan (see 2.1.4).

### **2.2.3 Data Storage and Handling**

One tedious but important obstacle is how we are going to:

1. Store the 125 GB of CT scan data
2. Divide the data into different blocks for processing
3. Keeping track of each CT scan / how they will be labeled

### **2.2.4 Training and Machine Learning**

The final goal of the project is to train a classifier to identify generally whether a patient has nodes in the lungs or does not as well as a regression model that predicts the degree of tumor proliferation in the lungs based on some metric calculated on the CT scan using our TDA techniques.

# Appendices



# Appendix A

## Appendix

### A.1 testScript.m

```
%% Test Script to Load and Examine CT Scan Jpegs  
% Nicholas von Turkovich
```

```
pathHealthy = '/Users/nbv3/Desktop/Math_Projects/topoCAT/ctScans/306/Ct_Lung  
filenameHealthy = 'IM-0001-0060.jpg';
```

```
pathUnhealthy = '/Users/nbv3/Desktop/Math_Projects/topoCAT/ctScans/321/Ct_Ch  
filenameUnhealthy = 'IM-0001-0060.jpg';
```

```
image = rgb2gray(imread([pathHealthy filenameHealthy]));  
image2 = rgb2gray(imread([pathUnhealthy filenameUnhealthy]));
```

```
figure(1)  
subplot(2,1,1)  
imagesc(image);  
title('healthy');  
colormap gray
```

```
subplot(2,1,2)  
imagesc(image2);  
title('unhealthy');
```

```

colormap gray

% Need to condense the image

smallerImage = imresize(image,0.125);
smallerImage2 = imresize(image2,0.125);

figure(2)
subplot(2,1,1)
mesh(smallerImage);

subplot(2,1,2)
mesh(smallerImage2);

% processing for image 1

croppedImage = double(smallerImage);

indicesToFloor = find(croppedImage < mean(croppedImage(:)));
croppedImage(indicesToFloor) = 0;

% processing for image 2

croppedImage2 = double(smallerImage2);

indicesToFloor2 = find(croppedImage2 < mean(croppedImage2(:)));
croppedImage2(indicesToFloor2) = 0;

figure(3)
subplot(2,1,1)
imagesc(croppedImage);

subplot(2,1,2)
imagesc(croppedImage2);

```

```

% Cropped image ready in uni-valued intensity grayscale
start = tic;
S1 = generateS(croppedImage);
S2 = generateS(croppedImage2);
%
stop = toc(start)

% compute persistence diagrams

[I, J] = rcalmfscm(S1, 270);
[I2, J2] = rcalmfscm(S2, 270);
figure(4)
subplot(2,1,1)
plotpersistencediagram(I);
title('Healthy');

subplot(2,1,2)
plotpersistencediagram(I2);
title('Unhealthy');

figure(5)
subplot(2,1,1)
plotpersistencediagram(J);
title('Healthy');

subplot(2,1,2)
plotpersistencediagram(J2);
title('Unhealthy');

% figure(4)

```

```
% I = sortbypersistence(I)
% plot(I(:,3));
```

## A.2 generateS.m

```
function [ sMat ] = generateS( mat )

% This function performs triangulation and generates S matrix

dim = size(mat);
dim = dim(1);
numPixels = dim^2;

% below is the vertex row array for the S matrix, initialized
vertices = zeros(numPixels, 3);

% filling the vertices array with each pixel of mat
for i = 1:numPixels
    vertexRow = [i-1, i-1, mat(i)];
    vertices(i,:) = vertexRow;
end

% now have all the vertex rows that contain their filtration values
% (intensity vals)

% find the edge rows

edges = [];
edgeRelations = [1 dim (dim + 1)];

% iterate through all points

for i = 1:numPixels
```

```

% generate all the possible edge pairs (not lower left and upper right
% diagonals)

for j = 1:length(edgeRelations)
    neighborIndex = i + edgeRelations(j);
    if neighborIndex < 1 || neighborIndex > numPixels || (mod(i, dim) ==
        continue;
    end
    newEntry = [i-1, neighborIndex-1, max(mat(i), mat(neighborIndex))];
    edges = [edges; newEntry];
end

end

sMat = vertcat(vertices, edges);

end

```

### A.3 testScript2.m

```

%% Test2
% Nicholas von Turkovich

pathHealthy = '/Users/nbv3/Desktop/Math_Projects/topoCAT/ctScans/306/Ct_Lung
filenameHealthy = '/IM-0001-0050.jpg';

pathUnhealthy = '/Users/nbv3/Desktop/Math_Projects/topoCAT/ctScans/321/Ct_Ch
filenameUnhealthy = '/IM-0001-0060.jpg';

image = rgb2gray(imread([pathHealthy filenameHealthy]));
image2 = rgb2gray(imread([pathUnhealthy filenameUnhealthy]));

```

```

figure(1)
subplot(2,1,1)
imagesc(image);
title('healthy');
colormap gray
subplot(2,1,2)
imagesc(image2);
title('unhealthy');
colormap gray

% Need to condense the image

smallerImage = imresize(image,0.0625);
smallerImage2 = imresize(image2,0.0625);

figure(2)
subplot(2,1,1)
mesh(smallerImage);

subplot(2,1,2)
mesh(smallerImage2);

% processing for image 1
% xcropAmount = 10;
% xstartCrop = xcropAmount;
% xendCrop = length(smallerImage) - xcropAmount;
%
% % processing for image 2
% xcropAmount = 5;
% xstartCrop = xcropAmount;
% xendCrop = length(smallerImage2) - xcropAmount;
%
% ycropAmount = 5;
% ystartCrop = ycropAmount;

```

```

% yendCrop = length(smallerImage2) - ycropAmount;
%
% croppedImage2 = double(smallerImage2(xstartCrop:xendCrop, ystartCrop:yendCrop));
% ycropAmount = 1;
% ystartCrop = ycropAmount;
% yendCrop = length(smallerImage) - ycropAmount;
%
% croppedImage = double(smallerImage(xstartCrop:xendCrop, ystartCrop:yendCrop));

croppedImage = double(smallerImage);
croppedImage2 = double(smallerImage2);

figure(3)
subplot(2,1,1)
imagesc(croppedImage);
colormap gray;

subplot(2,1,2)
imagesc(croppedImage2);
colormap gray;

m = mean2(croppedImage);
m2 = mean2(croppedImage2);
dev = std(croppedImage);
dev2 = std(croppedImage2);

% indicesToFloor = find(croppedImage < m);
% croppedImage(indicesToFloor) = 0;
% indicesToFloor2 = find(croppedImage2 < m2);
% croppedImage2(indicesToFloor2) = 0;

figure(4)
subplot(2,1,1)
imagesc(imadjust(croppedImage));

```

```

colormap gray;

subplot(2,1,2)
imagesc(imadjust(croppedImage2));
colormap gray;

% compute topology features on a cloud composed of the nonzero 2D vertices

[row col] = find(croppedImage);
[row2 col2] = find(croppedImage2);

figure(5)
subplot(2,1,1);
plot(row, col, 'k. ');

subplot(2,1,2);
plot(row2, col2, 'k. ');

cloud = [row col];
cloud2 = [row2 col2];

distances = pdist(cloud);
distances2 = pdist(cloud2);

dmat = squareform(distances);
dmat2 = squareform(distances2);

distanceBound = max(distances);
distanceBound2 = max(distances2);

init;
[I, J] = rcldm(dmat, distanceBound*0.25);

```



```
[I2, J2] = rcaldm(dmat2, distanceBound2*0.25);
```

```
figure(5)
subplot(2,1,1)
plotpersistencediagram(J);
title('Dgm-{0} for Clear Lungs');
xlabel('Birth');
ylabel('Death');

subplot(2,1,2)
plotpersistencediagram(J2);
title('Dgm-{0} for Cloudy Lungs');
xlabel('Birth');
ylabel('Death');
```

```
figure(6)
subplot(2,1,1)
plotpersistencediagram(I);
title('Dgm-{1} for Clear Lungs');
xlabel('Birth');
ylabel('Death');

subplot(2,1,2)
plotpersistencediagram(I2);
title('Dgm-{1} for Cloudy Lungs');
xlabel('Birth');
ylabel('Death');
```

## A.4 testScript3.m

```
%% Test3: Clear Lungs
% Nicholas von Turkovich
```

```
imageNum = 123;
```

```

pers0 = [];
pers1 = [];
pathHealthy = '/Users/nbv3/Desktop/Math_Projects/topoCAT/ctScans/306/Ct_Lung

for i = 1:5:imageNum

    i

    string = '';

    if i < 10
        string = sprintf('000%d', i);
    end

    if i >= 10 && i < 100
        string = sprintf('00%d', i);
    end

    if i >= 100
        string = sprintf('0%d', i);
    end

    filenameHealthy = strcat('/IM-0001-', string, '.jpg');

    image = rgb2gray(imread([pathHealthy filenameHealthy]));

    % Need to condense the image

    smallerImage = imresize(image, 0.0625);

```

```

xcropAmount = 5;
xstartCrop = xcropAmount;
xendCrop = length(smallerImage) - xcropAmount;

ycropAmount = 5;
ystartCrop = ycropAmount;
yendCrop = length(smallerImage) - ycropAmount;

croppedImage = double(smallerImage(xstartCrop:xendCrop, ystartCrop:yendCrop));

m = mean2(croppedImage);

indicesToFloor = find(croppedImage < m);
croppedImage(indicesToFloor) = 0;

%     figure(1)
%     imagesc(croppedImage);

% compute topology features on a cloud composed of the nonzero 2D vertices

[row col] = find(croppedImage);

cloud = [row col];

distances = pdist(cloud);

dmat = squareform(distances);

distanceBound = max(distances);

init;

```

```

[I, J] = rcaldm(dmat,distanceBound*0.25);

pers0 = horzcat(pers0, std(J(:,2)));
pers1 = horzcat(pers1, std(I(:,2)));

end

```

## A.5 testScript4.m

```

%% Test4: Cloudy Lungs
% Nicholas von Turkovich

```

```

imageNum = 131;

```

```

p0 = [];

```

```

p1 = [];

```

```

pathHealthy = '/Users/nbv3/Desktop/Math_Projects/topoCAT/ctScans/84/Unnamed_...

```

```

for i = 1:5:imageNum

```

```

    i

```

```

    string = '';

```

```

    if i < 10

```

```

        string = sprintf('000%d', i);

```

```

    end

```

```

    if i >= 10 && i < 100

```

```

        string = sprintf('00%d', i);

```

```

    end

```

```

if i >= 100
    string = sprintf('0%d',i);
end

filenameHealthy = strcat('/IM-0001-', string , '.jpg ');

image = rgb2gray(imread([pathHealthy filenameHealthy]));

% Need to condense the image

smallerImage = imresize(image,0.0625);

xcropAmount = 5;
xstartCrop = xcropAmount;
xendCrop = length(smallerImage) - xcropAmount;

ycropAmount = 5;
ystartCrop = ycropAmount;
yendCrop = length(smallerImage) - ycropAmount;

croppedImage = double(smallerImage(xstartCrop:xendCrop, ystartCrop:yendCr

m = mean2(croppedImage);

indicesToFloor = find(croppedImage < m);
croppedImage(indicesToFloor) = 0;

figure(1)
imagesc(croppedImage);

```

```

% compute topology features on a cloud composed of the nonzero 2D vertices

[row col] = find(croppedImage);

cloud = [row col];

distances = pdist(cloud);

dmat = squareform(distances);

distanceBound = max(distances);

init;
[I, J] = rcaldm(dmat, distanceBound*0.2);

p0 = horzcat(p0, std(J(:,2)));
p1 = horzcat(p1, std(I(:,2)));

end

```