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# Introduction to Bash

## **pwd: Find out where we are**

After installing and starting the terminal, let's orient ourselves by typing our first command, `pwd`, into the terminal and pressing enter.

`pwd`

`pwd` prints the location of the current working directory and tells you where exactly you are in the file system. When we login we typically start from what is called our home directory.

💡 Tip: finding the desktop on different user systems

Your home directory will be something like `/Users/YourUserName` but the path might be slightly different depending on your operating system. Below you find some help to orient yourself better for different terminal interfaces:

For MAC users:

- The home directory should be `/Users/YourUserName`
- To access the current folder in Finder you can try using `open .`
- Your desktop should be here `/Users/YourUserName/Desktop`

For Mobaxterm users:

- Your home directory is `/home/mobaxterm`
- By default this home directory is in a temporary folder, which gets deleted every time you exit Mobaxterm, To give this folder a persistent home, do the following:
  - Settings -> Configuration -> General
  - In there set Persistent home directory to a folder of your choice
- To access the file explorer and get used to where you are you can type `explorer.exe .`

- The path to the desktop would be something like this `/mnt/c/Users/YourUserName/OneDrive/Desktop` or `/mnt/c/Users/YourUserName/Desktop`

For WSL2 users:

- The home directory is `/home/YourUserName`
- To access the file explorer and get used to where you are you can type `explorer.exe` .
- The path to the desktop would be something like this `/mnt/c/Users/YourUserName/OneDrive/Desktop` or `/mnt/c/Users/YourUserName/Desktop`

## ls: List the contents of a directory

Now that we know where we are, let's find out what files and folders exist in our home directory. For this we can use the `ls` command, which allows us to list directory contents:

```
ls
```

In my case this returns something like this:

The colors will look different depending on the cli use but in my case I see a list of files (in bold text) and folders (green-highlighted text) in my home directory.

Since this output can easily become over-whelming if we deal with a lot of files and folders, lets look a bit closer into how we can optimize our commands.

## The structure of a command

Let's start with looking at the general structure of a command, which generally consists of three elements, the command itself and some optional options and arguments:

Understanding better what a command looks like, let's use the `ls` command together with the option `-l`. This option results in `ls` printing the content of a folder in a long listing format.

```
ls -l
```

After running this, we should see our files and folders again but in a long format, which gives more detailed information and structures our output a bit better:

## Getting help

If you want to know what options are available for a command it is always a good idea to check out the manual. You can do this with:

```
man ls
```

You can exit the manual by pressing **q**.

In case you want to check what a program does or what options there are, depending on the program there might be different ways to access the manual. These most common ways are:

- `man ls`
- `ls --help`
- `ls -h`

## cd: Move around folders

Most of the time you do not want to perform your analyses in the home directory but in a dedicated folder for your project. To get started, we will learn about the **cd** command that allows us to move around the file system.

The file system is a hierarchical system used to organize files and directories. It is a tree-like structure that starts with a single directory called the root directory, which is denoted by a forward slash (/). All other files are “descendants” of the root. To move from the root into other folders, we can go via the descendants to reach the john folder as follows: `/users/john`.

If we specify the location of a folder or file starting from the root directory, we use what is called an absolute path. If we specify the location relative to our current directory, such as our home directory, we use a relative path.

To start moving around, let's begin by moving relative to our working directory by moving into any of the folders that you saw listed after you have used `ls -l`. In my case I want to move into the source directory:

```
cd source/
```

If you use `pwd` after moving around directories, you should see that a different location is printed to the screen.

We can also move back to our original directory using `cd ..`, which will move us back one directory (and move us out of the source and back into the home directory).

```
cd ..
```

We can also move around multiple levels. In the example below, I am going into the source folder, then back to the home directory and then into the docs folder.

```
cd source/../docs
```

Another useful way to move around quickly is using the tilde symbol, `~`, which can be used as a shortcut to move directly into our home directory from wherever you are on the file system:

```
cd ~
```

#### Exercise

Explore your current location with `pwd` and `ls` and move around with `cd` and try to get used to these three commands. If you are more comfortable, try finding your Desktop based on the tip in the section about `pwd`.

## **mkdir: Make new folders**

Now that we know how to explore our surroundings, let's make a new folder in which we start our data analysis. For this we use the `mkdir` command.

To do this, we will first move into our home directory and then create and move into a new folder called `data_analysis` as follows:

```
#go into the home directory
cd ~

#in the home directory make a new folder and name it data_analysis
mkdir data_analysis

#check if new folder was generated correctly
ls

#move into the newly generated folder
cd data_analysis

#check if we correctly changed our location
pwd
```

#### 💡 Tip: commenting your code

Notice, how in the example below I added the commands to run as well as some explanation about what I did?

Here, I used a specific character `#` in front of a line of text to denote the beginning of a single-line comment. Anything coming after the character is considered a comment and won't be executed by the shell.

In the above example I definitely commented the code too much as my comments basically duplicate the code and that should be avoided, however, it is useful to add comments in code to ensure that the purpose of the code is clear to others.

You could, for example, add a comment above functions to describe what the function does and you can also add comments to “tricky” code where it is not immediately obvious what you are trying to do. Basically, you want to add comments to any section where you think that the future you might get confused a month later.

[Here](#) you find some examples for python and R but the same logic applies when writing code for bash, some examples for that can be found [here](#).

#### 💡 Tip: Command-line completion

Most command-line interpreters allow to automatically fill in partially typed commands, file paths or file names.

So instead of having to type out `data_analysis` completely when changing the directory, we can let the interpreter do the work for us.

To do this, start from the home directory (or wherever you ran the `mkdir` command)

and type `cd data_` and press the Tab-key on your keyboard. The cli should have auto-completed the folder name automatically.

If there are multiple options, such as `data` and `data_analysis`, the cli can not auto-complete the folder name, however, by pressing Tab twice you will see all options to extend the name.

## wget: Download data

Next, let's download our sequencing data into a new `data` folder. One way to do this is using `wget`. In the command below we add the option `-P` to specify into which folder we want to download the data. You can also see how I documented my code here in order to add some more specifics about where the link came from to help future me in case I, for example, need to look for some e-mails about the data later on.

```
mkdir data

#download data using link provided by the sequencing center on
↪ 23.01.2023
wget -P data
↪ https://github.com/ndombrowski/cli_workshop/raw/main/data/seq_project.tar.gz
```

### Exercise

1. Check with `ls` if the file was downloaded correctly
2. Check the manual if there are extra options that we can use for `ls` to check the file size (advanced)

Click me to see an answer

```
#question 1:
ls -l data/

#question 2:
ls -lh data/seq_project.tar.gz
```

## tar: Work with tar files

The data we downloaded is provided as a tar file:

- tar is short for Tape Archive, and sometimes referred to as tarball
- The TAR file format is commonly used when storing data
- TAR files are often compressed after being created and then become TGZ files, using the `tgz`, `tar.gz`, or `gz` extension.

We can decompress the data into our data folder as follows:

```
tar -xvf data/seq_project.tar.gz -C data
```

The options we use with the `tar` command are:

- `x` tells it to extract files from the archive
- `v` display verbose information and provide detailed information while creating the tarball
- `f` specify the file name
- `C` tells tar to change directory (so the package content will be unpacked there)

 Tip: how to generate a tarball

If you ever want to generate a tarball you can do the following:

```
tar -cvzf my_tarball.tar.gz folder_to_tar
```

The options we use are:

- `c` create an archive by bundling files and directories together
- `z` use gzip compression when generating the tar file

### Exercise

1. After extracting the tarball use `ls` to explore the content of the folder we just extracted. Ensure that you explore the content of potential sub-directories.
2. Find the path for at least one sequence file that ends on `fastq.gz`. Hint, to make your life easier, check the **Tip: Command-line completion** above to not have to type every single word.

[Click me to see an answer](#)



```
#question 1:
ls -l data/seq_project

#question 2:
ls -l data/seq_project/barcode001_User1_ITS_1_L001/
```

## rm: Remove files and folders

To remove files and folders, we use the `rm` command. We want to remove the `seq_project.tar.gz` file since we do not need it anymore once we have extracted the data:

```
#rm a file
rm data/seq_project.tar.gz

#check if that worked
ls -l data
```

If we want to remove a folder, we need to tell `rm` that we want to remove folders using an option. To do this, we use `-r`, which allows us to remove directories and their contents recursively.

### ! Important

#### **Unix does not have an undelete command.**

This means that if you delete something with `rm`, it's gone. Therefore, use `rm` with care and check what you write twice before pressing enter!

## Wildcards

After we have explored the folder with our sequencing data with `ls`, we have seen that `data/seq_project` contains 4 folders, 2 folders for two different users who each generated two replicates.

You also might have also noticed that it gets tedious to figure out how many files are in each folder because we would need to run `ls` on each single folder and view its content individually.

Luckily, the shell provides special characters to rapidly specify groups of filenames or patterns. **Wild-cards** are characters that can be used as a substitute for any class of characters in a search.

The `*` wildcard is the wildcard with the broadest meaning of any of the wildcards, it can represent 0 characters, all single characters or any string of characters. It allows us to list all files in the `seq_project` folder as follows:

```
ls data/seq_project/*
```

You should have seen before that the sequencing files all end with `.fastq.gz` we can make this command a bit more specific and at the same time make the output a bit more readable:

```
ls data/seq_project/*/*.gz
```

If we wanted to print the absolute and not the relative path, we could do the following (beware that the absolute path will be slightly different depending in you system):

```
ls /home/User_name/data/seq_project/*/*.gz
```

Now we can easily see for each folder how many files we have.

#### Exercise

1. Use a wildcard to list files for barcode001 only
2. Use a wildcard to list files only for user1

Click me to see an answer

```
#question 1:
ls data/seq_project/barcode001/*/*.gz

#question 2:
ls data/seq_project/*_User1_*/*.gz
```

### Tip: More wildcards

\* is not the only wildcard we can use and a full list can be found [here](#). Different wildcards can be useful in different contexts, not only when listing files but also finding patterns in files. To make such searches as specific as possible there are different wildcards available. A few examples are:  
The [0-9] wildcard = matches any number exactly once and allows us to extract a range of files.

```
ls data/seq_project/barcode00[0-9]*/*.gz
```

The [012] wildcard = matches 1 or 2 exactly once and allows us to extract a range of files.

```
ls data/seq_project/barcode00[12]*/*.gz
```

[A-Z] matches any letter in capitals occurring once  
[a-z]\* matches any letter in non-capital letters occurring many times

```
ls data/seq_project/barcode001_User1_ITS_1_[A-Z]001*/*.gz
```

## I/O redirection to new files

We have seen by now that by default our commands direct print the standard output to the terminal. For example, when we use `ls` the list of files and folders is printed. However, we can also redirect the standard output to a file by using the `>` character.

For example, we might want to generate a list with all fastq files and can do this as follows:

```
ls data/seq_project/*/* > fastq_paths.txt
```

## Exploring the content of text files

Next, let's view the content of the list we just generated. Viewing the actual files we work with is often important to ensure the integrity of our data.

### **head: View the first rows of a file**

`head` can be used to check the first 10 rows of our files:

```
head fastq_paths.txt
```

### **tail: View the last rows of a file**

If you want to check the last 10 rows use `tail`:

```
tail fastq_paths.txt
```

### **less: View the full file**

`less` is a program that let's you view a file's contents one screen at a time. This is useful when dealing with a large text file (such as a sequence data file) because it doesn't load the entire file but accesses it page by page, resulting in fast loading speeds.

```
less -S fastq_paths.txt
```

Once opened, `less` will display the text file one page at a time.

- You can use the arrow Up and Page arrow keys to move through the text file
- To exit `less`, type `q`

#### Tip: Editing text files

You can also edit the content of a text file and there are different programs available to do this on the command line, the most commonly used tool is `nano`, which should come with most command line interpreters. You can open any file as follows:

```
nano fastq_paths.txt
```

Once the document is open you can edit it however you want and then

- Close the document with `control + X`
- Type `y` to save changes and press enter

## wc: Count things

Another useful tool is the `wc` (= wordcount) command that allows us to count the number of lines in a file. It is an useful tool for sanity checking and here allows us to count how many files we work with:

```
wc -l fastq_paths.txt
```

After running this we see that we work with 8 files.

We could of course easily count this ourselves, however, if we work with hundreds of files its a quick and easy way to get an overview about how much data we work with.

One down-side of this approach is that to be able to count the number of files, we first need to generate a file in which we count the number of files. This (i) can create files we do not actually need and (ii) we use two commands while ideally we want to get the information with a single command.

## Pipes

Pipes are a powerful utility to connect multiple commands together. Pipes allow us to feed the standard output of one command, such as `ls` as input into another command such as `wc -l` and as such combine multiple commands together.

Therefore, lets use `ls` to first list all fastq files and then pipe the output from `ls` into the `wc` command in order to count with how many files we work with:

```
ls data/seq_project/*/* | wc -l
```

We should see again that we work with 8 files, but now we did not have to generate an intermediate text file and have a more condensed command.

### Exercise

1. Use `ls` and `wc` to count how many R1 files we have
2. Count how many R2 files we have
3. Count how many files were generated for User1?

[Click me to see an answer](#)

```
#question 1: We see 4 files
ls data/seq_project/*/*R1*/*.gz | wc -l

#question 2: We see 4 files
ls data/seq_project/*/*R2*/*.gz | wc -l

#question 3: We see 4 files
ls data/seq_project/*_User1_*/*.gz | wc -l
```

Checking whether we have the same number of R1 and R2 files is a good sanity check, to ensure that we have received all the files from the sequencing center whenever we generate paired sequencing data.

## cut: Extract elements from strings

Remember, how we stored a list of sequencing files and the path leading to these files (relative to our home directory in a text file called `fastq_paths.txt`)?

Imagine that we only wanted to have a list of files but not the path, how would we do that?

There are different ways to do this, the simplest one is to use `cd` and go into the folder with our sequence files and generate a list in there.

However, another way in which we do not have to move around (and learn some other concepts) is to use the `cut` command. `cut` allows us to separate lines of text into different elements using any kind of delimiter, for example the `/` that we use in the file path. To ensure that `/` is seen as a separator we use the `-d` option and with `-f4` we tell `cut` to print the fourth element of each separated field.

```
head fastq_paths.txt

#extract the file name (i.e. the fourth field when using a /
↪ separator)
cut -f4 -d "/" fastq_paths.txt

#do the same as above but this time save the output in a new file
cut -f4 -d "/" fastq_paths.txt > fastq_files.txt
```

We can also combine this with pipes in order to extract different pieces of information. Let's assume we start with extracting a folder name, such as `barcode002_User1_ITS_9_L001`, and from that we want to extract some other information, such as a list with all the barcode IDs. We can easily do this as follows:

```
cut -f3 -d "/" fastq_paths.txt | cut -f1 -d "_"
```

Nice, we now only have a list with barcodes. However, it's not yet ideal since we have duplicated barcodes. If you want to extract this information for some metadata file this is not ideal and we need to know two more commands to make this work:

## sort and uniq: Create unique lists

- **sort**: sort lines in a file from A-Z and is useful for file organization.
- **uniq**: remove or find duplicates. For this command to work you need to provide it with a sorted file

Let's first ensure that our barcode list is sorted to then extract only the unique information:

```
cut -f3 -d "/" fastq_paths.txt | cut -f1 -d "_" | sort | uniq
```

### Exercise

1. Use `cut` to print the folder names that lead to the `fastq.gz` files
2. Print the folder names as in 1 but then also extract the User names
3. Ensure that when running the code from 2 that we print a unique list of User names

[Click me to see an answer](#)

```
#question 1:
cut -f3 -d "/" fastq_paths.txt

#question 2:
cut -f3 -d "/" fastq_paths.txt | cut -f2 -d "_"

#question 3:
cut -f3 -d "/" fastq_paths.txt | cut -f2 -d "_" | sort | uniq
```

## cat: Read and combine files

cat can do different things:

1. Create new files
2. Display the content of a file
3. Concatenate, i.e. combine, several files

To view files we do:

```
cat fastq_paths.txt
```

To combine files we do:

```
#combine files
cat fastq_paths.txt fastq_files.txt > combined_files.txt

#view new file
cat combined_files.txt
```

### Exercise

Fastq.gz files can easily be combined using `cat` and while this is not strictly necessary in our case, you might need to do this with your sequence files in the future. For example if you sequenced a lot of data and for space reasons the sequencing center send you multiple files for a single sample.

1. Using wildcards and `cat`, combine all R1 fastq.gz files into a single file
2. Use `ls` to judge the file size of the individual R1 files and the combined file to assess whether everything worked correctly

[Click me to see an answer](#)



```
#question 1:
cat data/seq_project/*/*R1.fastq.gz > combined.fastq.gz

#question 2:
ls -lh data/seq_project/*/*R1.fastq.gz

#question 3:
ls -lh combined.fastq.gz
```

If you compare the numbers from part 2 and 3, you should see that the combined.fastq.gz file is roughly the sum of the individual files.

## Exploring the content of compressed fastq files

### Exercise

1. View the first few rows of any one of the fastq.gz sequence files
2. Does this look like a normal sequence file to you?

Click me to see an answer

```
head
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq.gz
```

After running this command you will see a lot of random letters and numbers but nothing that looks like a sequence, so what is going on?

## gzip: (De)compress files

After downloading and exploring the content of the downloads folder, you have seen that the file we downloaded ends with **gz**. This indicates that we work with a gzip-compressed file. Gzip is a tool used to (de)-compress the size of files.

In order to view the content of such files, we sometimes need to de-compress them first. We can do this using the gzip command together with the decompress **-d** option:

```
gzip -d
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq.gz

head
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq
```

After running this, we finally see a sequence and some other information. Notice that for fastq files we always should see 4 rows with information for each sequence:

### Exercise

1. After running the gzip command above, make a list of each sequence file with `ls`
2. Use `ls` with an option to also view the file size and compare the size of our compressed and uncompressed files.

[Click me to see an answer](#)

```
#question 1:
ls -l data/seq_project/*/*

#question 2:
ls -lh data/seq_project/*/*
```

We see that our file has a file size of about ~25M after compression while the compressed files are around ~5M.

When working with sequencing files the data is usually much large and its best to keep the files compressed to not clutter your computer.

To keep our files small its best to work with the compressed files. If we want to compress a file, we can do this as follows:

```
gzip
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq

ls data/seq_project/*/*
```

## zcat: Decompress and print to screen

We have seen that to view the content of a compressed file and make sense of the content we had to first decompress the file. However, sequence files tend to get rather large and we might not want to decompress our files to not clutter our system.

Luckily, there is one useful tool in bash to decompress the file and print the content to the screen called **zcat**, the original compressed file is kept while doing this. We combine this command with the head command, since do not want to print millions of sequences to the screen but only want to explore the first few rows:

```
zcat
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq.gz
↪ | head
```

## grep: Find patterns in data

The grep command is used to search text. It searches the given file for lines containing a match to the given strings or words. Also this command is simple but very useful for sanity checks after file transformations.

Let's first search for some things in the text files we generated, for example, we might want to only print information for the R1 files:

```
#grep a pattern, here R1, in fastq_paths.txt
grep "R1" fastq_paths.txt
```

We see the list of files that match our pattern. If we simply are interested in the number of files that match our pattern, we could add the option **-c** for counting.

```
grep -c "R1" fastq_paths.txt
```

We can also combine this with wildcards, for example, if we look for all samples from User1 we could do the following:

```
grep "User1_ITS_*_" fastq_paths.txt
```

### Exercise

1. Grep for paths containing the pattern “ITS\_9” in fastq\_paths.txt
2. Grep for the pattern “AAGACG” in any of the fastq.gz files
3. Count how often “AAGACG” occurs in any in any of the fastq.gz files

Click me to see an answer

```
#1
grep "ITS_9" fastq_paths.txt

#2
zcat
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq.gz
↪ | grep "AAGACG"

#3
zcat
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R1.fastq.gz
↪ | grep -c "AAGACG"
```

The last two commands can be very useful to check if the adapters or primers are still part of your sequence.

## wc: Count things

Another useful tool is the `wc` (= wordcount) command that allows us to count the number of lines in a file. It is an easy way to perform sanity checks and in our case allows us to count how many sequences we work with:

```
zcat
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R2.fastq.gz
↪ | wc -l
```

After running this we see that we work with 179,384 / 4 sequences. We need to divide the number we see on the screen by 4 since each sequence is represented by 4 lines of information in our fastq file.

### 💡 Advanced Tip: better counting

We have seen that we need to divide the output of `wc` by four to get the total number of sequences. We can do this with a calculator but actually, some intermediate bash can also be used to do this for us:

```
zcat
↪ data/seq_project/barcode001_User1_ITS_1_L001/Sample-DUMMY1_R2.fastq.gz
↪ | echo $((`wc -l`/4))
```

In the command above we have some new syntax:

- The `echo` command is used to display messages or print information to the terminal. In our case it will print whatever is going on in this section `$(( `wc -l` / 4 ))`
- `$((...))`: This is an arithmetic expansion in Bash. It allows you to perform arithmetic operations inside the brackets and substitute the result into the command line.
- The backticks (“”) around `wc -l` indicate command substitution. This means that the `wc -l` command is executed, and its output (the number of lines counted) is used in the overall command. Without command substitution (`wc -l` alone), you would not capture the output; instead, you would only see the literal text “`wc -l`”. Command substitution allows you to use the actual result of the command.
- `/4`: This is dividing the result obtained from `wc -l` by 4. Since each sequence in a FASTQ file is represented by four lines (identifier, sequence, separator, and quality scores), dividing the total number of lines by 4 gives the number of sequences.

## For-loops

Imagine we want to count the lines not only from one but all files. Could we do something like the code below?

```
zcat data/seq_project/*/*.gz | wc -l
```

When running this, we see that the command prints a single number, 869 944, but not the counts for each file, so something did not work right.

The problem with this command is that it prints the text from all 8 fastq files and only

afterwards performs the counting. We basically concatenated all files and then calculated the sum of all 8 files. However, what we want to do is to repeat the same operation over and over again:

- Decompress a first file
- Count the lines in the first file
- Decompress a second file
- Count the lines in the second file
- ...

A **for loop** is a bash programming language statement which allows code to be repeatedly executed. I.e. it allows us to run a command 2, 3, 5 or 100 times.

Let's start with a simple example but before that let's introduce a simple command `echo`. `echo` is used to print information, such as `Hello` to the terminal:

```
echo "Welcome 1 time!"
```

We can use for-loops to print something to the screen not only one, but two, three, four ... times as follows:

```
for i in 1 2 3; do
    echo "Welcome ${i} times"
done
```

An alternative and more condensed way of writing this that you might encounter in the wild is the following:

```
for i in 1 2 3; do echo "Welcome ${i} times"; done
```

Here, you see what this command does step by step:

Let's try to do the same but for our files by storing the individual files found in `data/seq_project/*/*.gz` in the variable `i` and print `i` to the screen in a for-loop.

```
for i in data/seq_project/*/*.gz; do
    echo "I work with: File ${i}"
done
```

- `for i in data/seq_project/*/*.gz; do`: This part initializes a loop that iterates over all files matching the pattern `data/seq_project/*/*.gz`. The variable `i` is assigned each file in succession.

We can then use these variables stored in `i` to perform more useful operations, for example for each file, step-by-step, count the number of lines in each file by using some of the tools we have seen before:

```
for i in data/seq_project/*/*.gz; do
    zcat ${i} | wc -l
done
```

- `zcat ${i} | wc -l`: This is the action performed inside the loop. `zcat` is used to concatenate and display the content of compressed files (\*.gz). The `|` (pipe) symbol redirects this output to `wc -l`, which counts the number of lines in the uncompressed content.

### Exercise

1. Use a for loop and count how often `grep` finds the pattern “TAAGA”

Click me to see an answer

```
for i in data/seq_project/*/*.gz; do
    zcat ${i} | grep -c "TAAGA"
done
```

Sometimes it is useful to not store the full path in `i`, especially when we want to store the output of our loop in a new file. Luckily, we can use the list of files we stored in `fastq_files.txt` to rewrite this command a bit and make use of the fact that we can use `cat` to print something in a text file to the screen:

```
for i in `cat fastq_files.txt`; do
    zcat data/seq_project/*/${i} | wc -l
done
```

- `for i in `cat fastq_files.txt`; do`: This initiates a loop that iterates over each item in the file `fastq_files.txt`. The backticks ``` are used to execute the `cat` command within the backticks to read the content of the text file and assign its output to the variable `i`.

- `zcat data/seq_project/*/${i} | wc -l`: Like before, this line uncompresses (`zcat`) and counts (`wc`) the number of lines in the specified file. However, in this case, the file is determined by the content of `fastq_files.txt`, which contains a list of file names.

After this modification of the code, we can very easily store the output in a file instead of printing the results to the screen.

```
#Count the number of lines in each sequencing file
#Store the output in a new folder
mkdir counts

for i in `cat fastq_files.txt`; do
    zcat data/seq_project/*/${i} | wc -l > counts/${i}.txt
done

ls counts/*.txt

head counts/Sample-DUMMY1_R1.fastq.gz.txt
```

#### 💡 Tip: Advanced: better counting in for loops

Let's get a bit more advanced to show you some powerful features of bash. For this imagine that you would do this for 100 files. In this case it would be useful to see the file names next to the counts. We can achieve this by using what we have learned in the **Better counting tip** where we have learned about `echo` and command substitution.

```
for i in data/seq_project/*/*.gz; do
    echo "$i: $(zcat $i | wc -l)"
done
```

- The `echo` command is used to display messages or print information to the terminal. In our case it will print whatever is going on here `"$i: $(zcat $i | wc -l)"`
- `$(...)`: These parentheses are used for command substitution. It means that the command `zcat $i | wc -l` is executed, and its output (the line count of the uncompressed content) is substituted in that position.
- The double quotes (“”) perform what is called a string concatenation. It concatenates the filename (`$i`), a colon (:), a space, and the line count obtained from the command substitution. The entire string is then passed as a single



argument to the echo command.

Almost perfect, now we only want to divide this by 4:

```
for i in data/seq_project/**/*.gz; do
    echo "$i: $(( $(zcat $i | wc -l) /4 ))"
done
```

Notice, how we use both single brackets and double brackets?

- `$((...))` in contrast to `$(...)` is an arithmetic expansion in Bash. It allows you to perform arithmetic operations and substitute the result into the command line.