

Volatile Organic Compound Detection Using Insect Odorant-Receptor Functionalised Field-Effect Transistors

by

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1. Vapour Sensing System for Transistor Biosensing

1.1. General Remarks

Through the adaptation of an existing setup, a custom vapour delivery system was developed to measure the response of field-effect biosensors to vapour. To achieve this goal, the new system needed to meet three requirements:

- The ability to automatically deliver a vapour to an enclosed environment in a controlled manner.
- The ability to collect measurements from a sensor device within that environment.
- The ability to collect data from off-the-shelf reference sensors monitoring the same environment, for comparison with data collected by the novel biosensor.

The previously existing system had a limited ability to meet the first two requirements, but was not able to take reference measurements of vapour flow. To implement new elements that would enable the system to fulfill all three requirements, a two-step development approach was taken across the course of the thesis. The changes made with each step of the redesign are outlined in Section 1.3.

Three mass flow controllers (MFC) were used to precisely control and monitor the flow of nitrogen into the system in units of standard cubic centimeters per minute (sccm). The manner in which these controllers were configured in the system is discussed in Section 1.2.1. The reference sensors chosen were a photoionisation detector (Ametek Mocon) and relative humidity and temperature indicator (Telaire). The photoionisation detector is able to monitor a wide range of volatile organic compounds, but cannot monitor compounds with an ionisation energy exceeding 10.6 eV. This includes nitrogen, oxygen, carbon dioxide, argon and water [1], [2]. Therefore, the photoionisation detector (PID) should not respond to either ambient air or standard nitrogen flow through the detector. As we would also like to monitor the presence of water vapour in the system, we use the relative humidity indicator (RHI). The operation of these reference sensors is discussed further in Section 1.2.2.

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1.2. Technical Notes

1.2.1. Delivery System

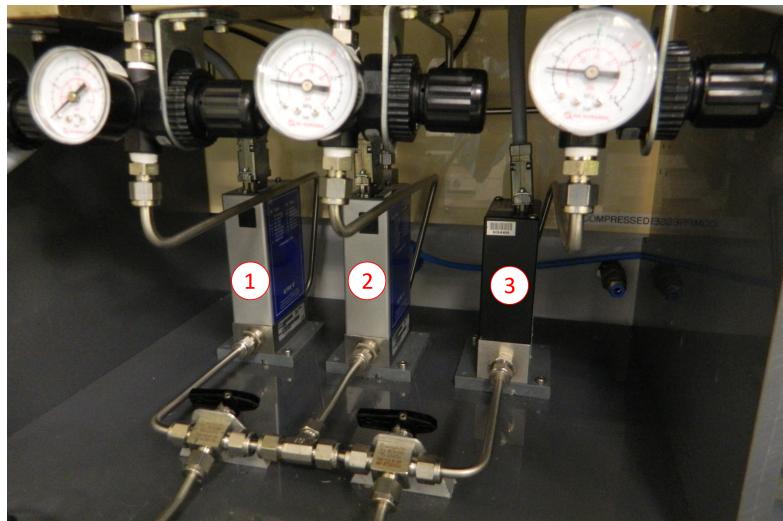


Figure 1.1.: Image of the three mass flow controllers (MFCs) of the Te Herenga Waka - Victoria University of Wellington cleanroom vapour delivery system, each with a regulator to set the pressure at the MFC inlet. (1) is the 20 sccm full-scale flow MFC, (2) is the 200 sccm full-scale flow MFC, and (3) is the 500 sccm full-scale flow MFC.

Three mass flow controllers and their associated regulators sit in a covered enclosure, seen from the front in Figure 1.1. These are used to control the nitrogen flow rate through two different lines towards the chamber, the carrier line and dilution line. These lines reconnect at a mixing point about a metre before the chamber. Flow through the carrier line passes through the analyte bottle and carries analyte vapour to the mixing point. The dilution line separately delivers flow to the mixing point, modifying the flow pushing the analyte vapour towards the chamber. The 10 mL analyte Schott bottle (Duran) is shown connected to the carrier line in Figure 1.2. Each line consist of a mix of stainless steel and flexible PVC tubing, with various Swagelok fittings and valves. These valves include check valves, to ensure there is no backflow of vapour mass within the system. The electronic integration and programming of the mass flow controllers is described in Section 1.2.3.

1.2.2. Reference Sensors

Two reference sensors were added to the vapour delivery setup to compare the response to vapour by the fabricated sensor device with some reference signal. These reference

1.2. Technical Notes

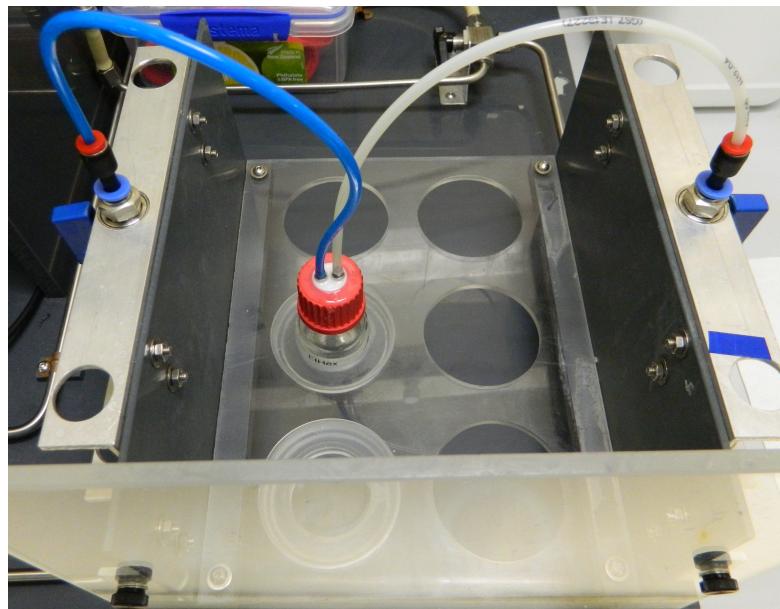


Figure 1.2.: Image of the analyte bottle, used to hold a volatile compound which provides vapour to the carrier line, either through bubbling or headspace sampling.

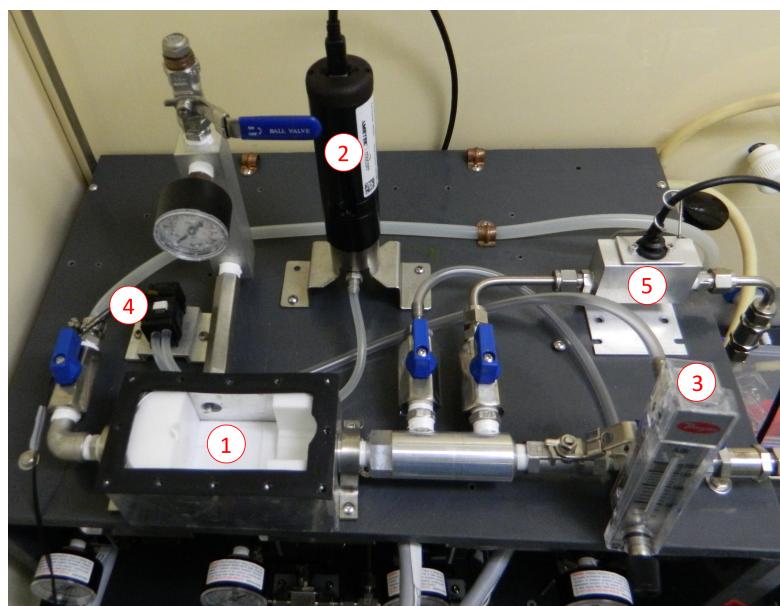


Figure 1.3.: Image of the device chamber, reference sensors and other related components. The components are labelled as follows: (1) Device chamber, (2) Photoionisation detector (PID), (3) Flowmeter from chamber to PID, (4) Micropump from chamber to PID, (5) Relative humidity and temperature monitor.

1. Vapour Sensing System for Transistor Biosensing

sensors are a photoionisation detector (Ametek Mocon) and a relative humidity and temperature indicator (Telaire). The layout of these reference sensors (and their associated peripherals) relative to the device chamber is shown in Figure 1.3. These components are on a raised platform directly above the mass flow controller enclosure. Vapour flowing through the device chamber passes into a cylindrical manifold with three outlets. One outlet is the system exhaust, one flows into relative humidity indicator chamber, and one flows into the photoionisation detector. A dial-controlled micro diaphragm pump is used to set the flow rate from the manifold into the photoionisation detector, with a flowmeter used to monitor this flow rate.

The electronic integration and programming of the relative humidity and temperature indicator is described in Section 1.2.3. The photoionisation detector was connected to a laptop directly via USB, then controlled and monitored using the supplier-provided VOC-TRAQ II software package.

Relative Humidity and Temperature Indicator

The relative humidity and temperature indicator used here is a capacitive humidity sensor [3]. It consists of a capacitor with a hygroscopic polymer as the capacitor dielectric. As room temperature water has a much larger dielectric constant than the polymer dielectric, absorption of water by the polymer leads to increased sensor capacitance [4]. The sensor capacitance, corresponding to the amount of moisture absorbed by the polymer and therefore the relative humidity, is then translated by the sensor into a calibrated electronic output. This output is then processed using the hardware and software described in Section 1.2.3 to give a value for the relative humidity. The sensor has a quoted relative humidity (RH) accuracy of $\pm 2.0\%$ when RH is below 80%, and has a quoted temperature accuracy of 0.5°C [3].

The absolute humidity (AH), the mass of water vapour within a set volume, can be calculated using Equation 1.1, where $C = 2.16679 \text{ gKJ}^{-1}$, P_W is the water vapour pressure (in Pa) and T is the temperature (in K) [5].

$$AH = C \frac{P_W}{T} \quad (1.1)$$

For temperatures between -20°C and 50°C , water vapour pressure P_W (in hPa) can be approximated using Equation 1.2, where RH is relative humidity, T is temperature in $^\circ\text{C}$, $A = 6.116441 \text{ hPa}$, $m = 7.591386$ and $T_n = 240.7263^\circ\text{C}$ [5].

$$P_W = RH \times A \times 10^{(mT/(T+T_n))} \quad (1.2)$$

Photoionisation Detector

A photoionisation detector (PID) can be used to continuously monitor volatile organic compounds by measuring the extent to which vapour molecules passing through the detector can be ionised. A small percentage of vapour molecules flowing into the detector diffuse into a sensor cavity. This cavity is bounded on each side by a pair of electrodes. A lamp in the body of the detector radiates UV light through a window into this cavity. The vapour molecules have their outer-most electrons excited and removed when struck with these high-energy photons. The ionised molecules then drift towards the sensor cathode, while free electrons drift towards the sensor anode. This results in a current proportional to the concentration of vapour molecules in the chamber. The current can then be amplified for a signal readout. To be detected, the ionisation energy of the molecules being monitored cannot exceed the energy of the incident UV light. Therefore, molecules of clean air will not be detected. Likewise, volatile organic compounds with high ionisation energy – such as methane – will not be recognised by the PID. Conversely, if the energy is required to ionise a volatile of interest is relatively low, the PID will generally show a relatively large response to that volatile [1], [2].

The photoionisation detector used in this work had a lamp energy of 10.6 eV, with a quoted response time of less than 2 seconds. Photoionisation detectors are designed to sensitively detect within a particular concentration range. These ranges can be stated in terms of the sensitivity of the detector to isobutylene, which is typically used to calibrate PID sensors. The range of the PID sensor used was 10 ppb – 200 ppm isobutylene. With constant temperature and pressure, the quoted accuracy of the sensor is $\pm 3\%$.

The vapour of interest can be delivered to the PID either through diffusion or by means of a low-power pump. A micro diaphragm pump (Xavitech) was selected to pump the vapour from the chamber into the PID detector. This pump was selected to have a low maximum flow rate as the PID requires an inlet flow of less than 300 sccm. As the pump is controlled using an unlabelled dial, a flowmeter was used to independently measure the flow rate through the micropump into the PID.

1.2.3. Control System

The vapour delivery system was controlled and monitored from a laptop connected to a National Instruments USB-6009 multifunction data acquisition system (DAQ). This USB-6009 DAQ connected to the mass flow controllers and relative humidity and temperature indicator via a custom-designed circuit board manufactured by PCBway. The outputs and inputs of the USB-6009 DAQ were set using custom LabView software. These electronic and software components of the vapour delivery control system are described in more detail below.

1. Vapour Sensing System for Transistor Biosensing

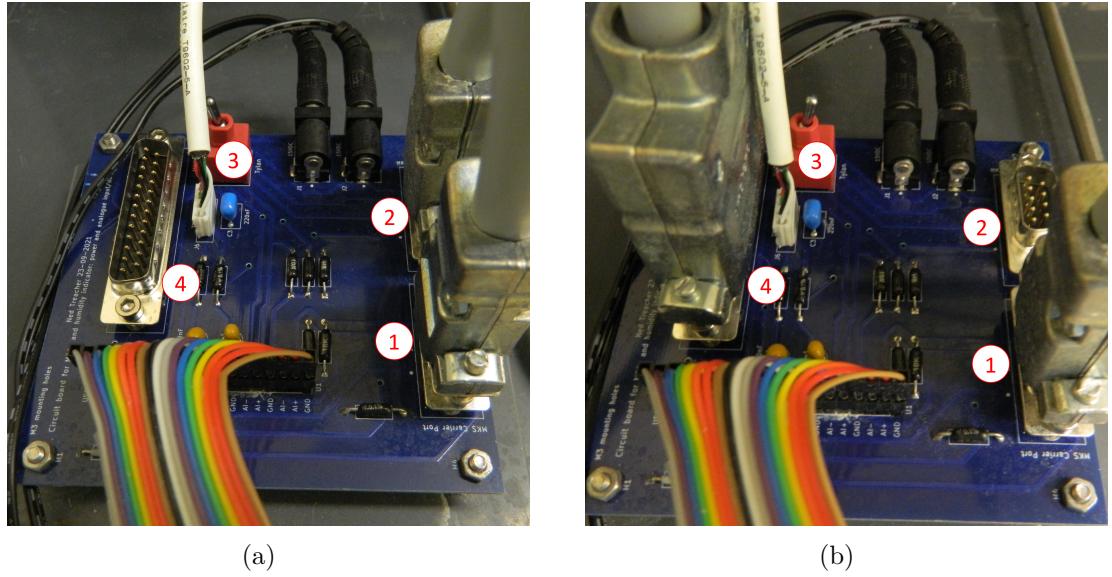


Figure 1.4.: Images of the vapour delivery control system circuit board, where (a) shows the low-flow configuration and (b) shows the high-flow configuration. Components are labelled as follows: (1) 9-pin carrier line port, (2) 9-pin dilution line port, (3) dilution port switch (determines which dilution line port is active), (4) 25-pin dilution line port. In (a), the 500 sccm full-scale MFC is connected at the 25-pin dilution line port, the 200 sccm full-scale MFC is connected at the 9-pin carrier line port and the red dilution port switch is towards “Tylan” (to the right). In (b), the 200 sccm full-scale MFC is connected at the 9-pin dilution line port, the 20 sccm full-scale MFC is connected at the 9-pin carrier line port and the red dilution port switch is towards “MKS” (to the left).

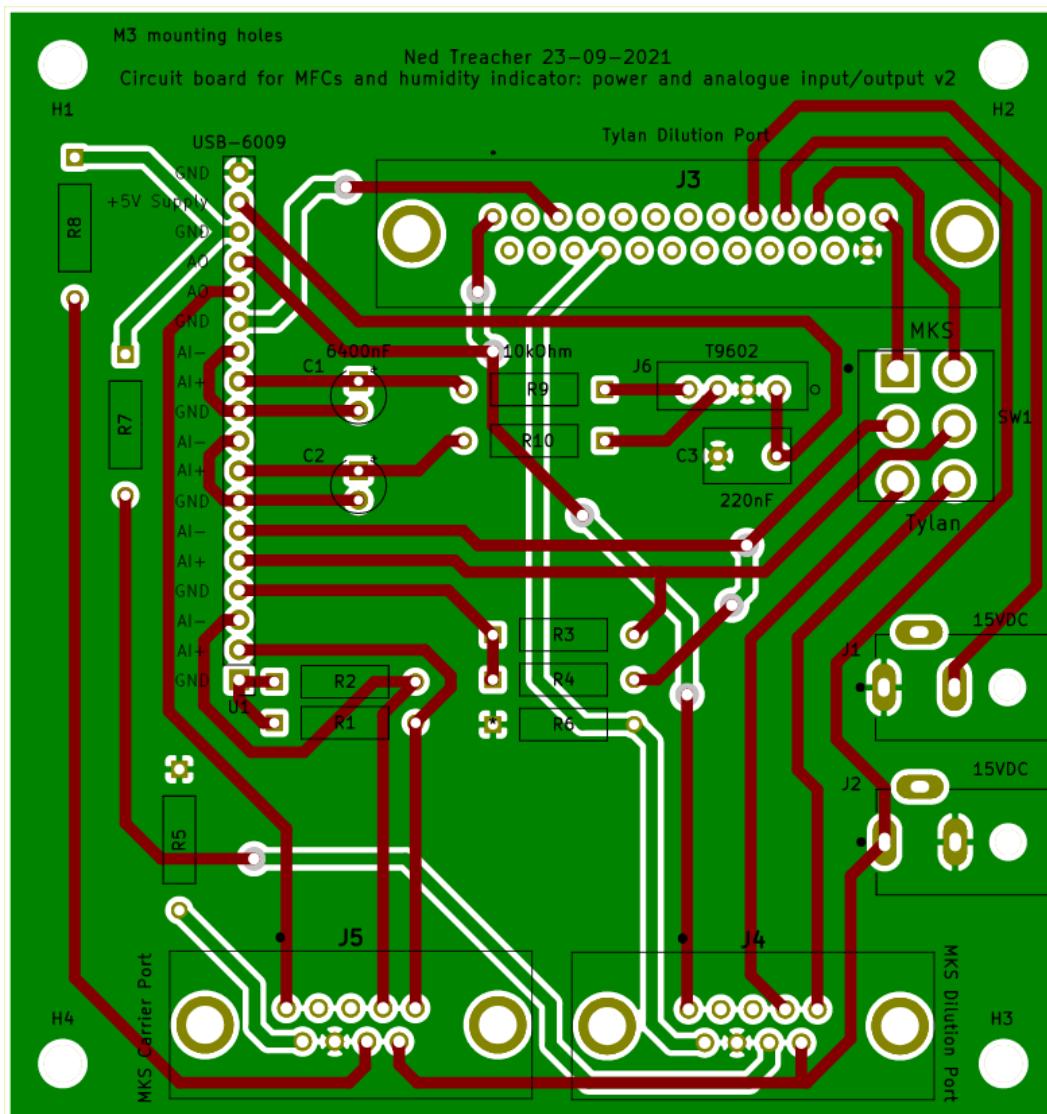


Figure 1.5.: Circuit board schematic for controlling and monitoring both the mass flow controllers and the relative humidity and temperature sensor. The relative humidity and temperature sensor is connected to the circuit board via the T9602 footprint. The mass flow controllers connect to the board in two configurations. The first (“high-flow”) configuration has the Tylan dilution and MKS carrier ports connected, with switch SW1 in the Tylan direction. The second (“low-flow”) configuration has both MKS ports connected, with switch SW1 connected in the MKS direction. Resistors R1-R6 are all 10 kOhm, while R7-R8 are both 0 Ohm. The circuit board was designed using the KiCad Layout Editor.

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Electronics

The control circuit board used to connect the mass flow controllers and relative humidity and temperature indicator to the NI USB-6009 is shown in Figure 1.4. Only one mass flow controller can be set to provide flow to a specific line, and so only two mass flow controllers can be operational simultaneously during testing with the vapour delivery system. The control circuit board allows the user to set which two mass flow controllers are to be used during a specific test run. Figure 1.4b shows the “high-flow” configuration, where the 500 sccm full-scale MFC is connected to the dilution line and the 200 sccm full-scale MFC is connected to the carrier line. Figure 1.4a shows the “low-flow” configuration, where the 200 sccm full-scale MFC is connected to the dilution line and the 20 sccm full-scale MFC is connected to the carrier line. The design for the circuit board is shown in Figure 1.5, showing the pinout to the USB-6009 and the various components used to connect the MFCs, relative humidity indicator, and the power supply for the MFCs.

Software

Two LabView Virtual Instruments (VIs) were adapted from pre-existing VIs for operating the mass flow controllers and monitoring vapour flow into the device chamber, as well as monitoring temperature and humidity in the vapour delivery system’s manifold. These VIs were named “vapour-sensor-basic.vi” and “temp-and-humidity-basic.vi”. A third VI was developed in parallel which combined the first two Virtual Instruments and allowed the user to set a sequence of values for the output flow from the mass flow controllers before an experimental run. This VI was named “vapour-sensor-sequence-timestamped.vi”. Flow rate, relative humidity and temperature data were then saved as .lvm files. The LabView VIs described here are available on request.

1.3. Design

1.3.1. Initial Design

The initial design of the vapour delivery system, as shown in Figure 1.6, was relatively simple. No reference sensors were included in the setup, and only one channel could be characterised without opening the chamber and changing the position of the device. However, as constructed it worked well as a self-contained system, which was able to deliver vapour to a device channel while measuring current across the channel. The original system is shown in Figure 1.7, and the circuit board used to control it is shown in @fig-original-pcb A 500 sccm full-range MFC (Tylan) was placed on the dilution line, and a 200 sccm full-range MFC (Tylan) was placed on the carrier line. A glass container for analyte was present on the carrier line, with a vapour trap upstream to collect any

1.3. Design

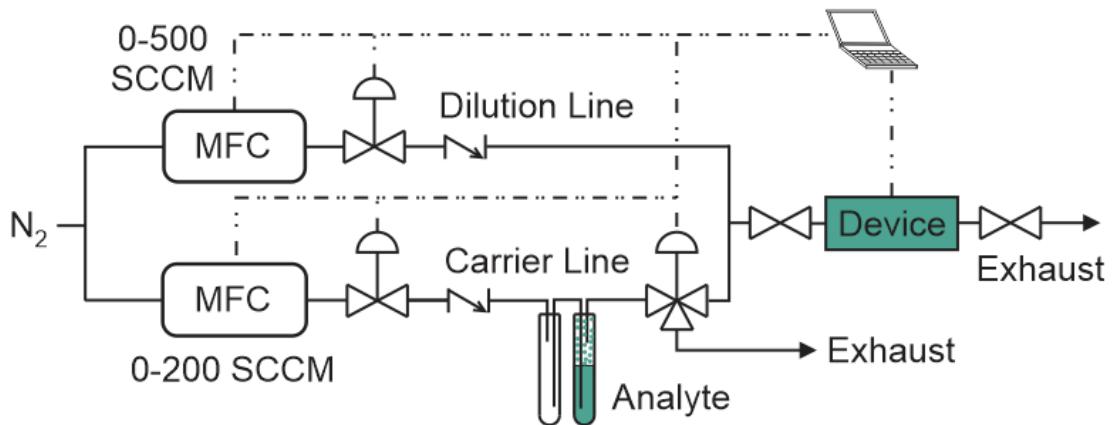


Figure 1.6.: P&ID of the original vapour delivery system

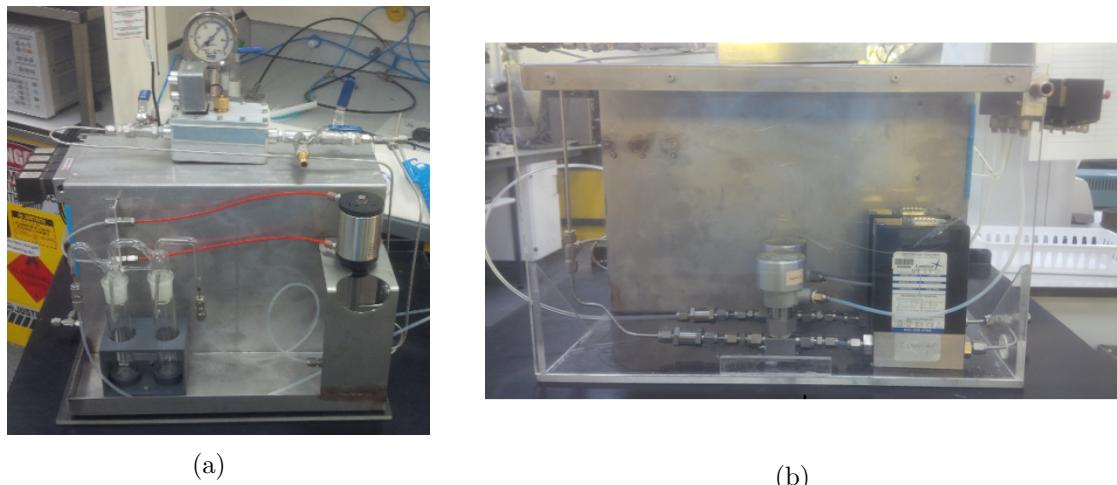


Figure 1.7.: The original vapour delivery system setup, where (a) shows the front of the system, including the device chamber, analyte bottles and four-way valve, and (b) shows the back of the system, including the mass flow controllers and solenoid valves.

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Figure 1.8.: The control circuit board for the original vapour delivery system.

backflow. The vapour trap was removed in later iterations due to the presence of a check valve to prevent backflow. The device chamber and mass flow controllers were connected to a laptop and an Agilent 4156C semiconductor parameter analyser and controlled using LabView.

1.3.2. Stage I Design

The first stage of the vapour delivery system redesign, as shown in Figure 1.9 and Figure 1.10 was implemented in Nov 2021. This system introduced the ability to use a 20 sccm full-range MFC (MKS Instruments) for carrier line flow and a 200 sccm full-range MFC (MKS Instruments) for either carrier or dilution line flow, to give better control when using low flow rates. The reference sensors were also implemented, with each sensor connected in parallel to the chamber exhaust. Through testing the system with ethanol and acetone as analytes, the following issues with this implementation of the setup were identified:

- With the system connected to the lab supply of nitrogen, pressure changes in the line due to nitrogen use elsewhere in the lab impacted the pressure at the MFCs and the flow through the lines.
- The pressure indicator used for the device chamber had a much wider range than the pressure reached before nitrogen began to leak out of the PVC tubing; this meant pressure changes in the chamber, resulting from closing the exit valves while nitrogen flow entered the chamber, did not register on the indicator.
- The PID responded unexpectedly slowly to changes in vapour concentration in the chamber. For example, after acetone or ethanol vapour had been run through

1.3. Design

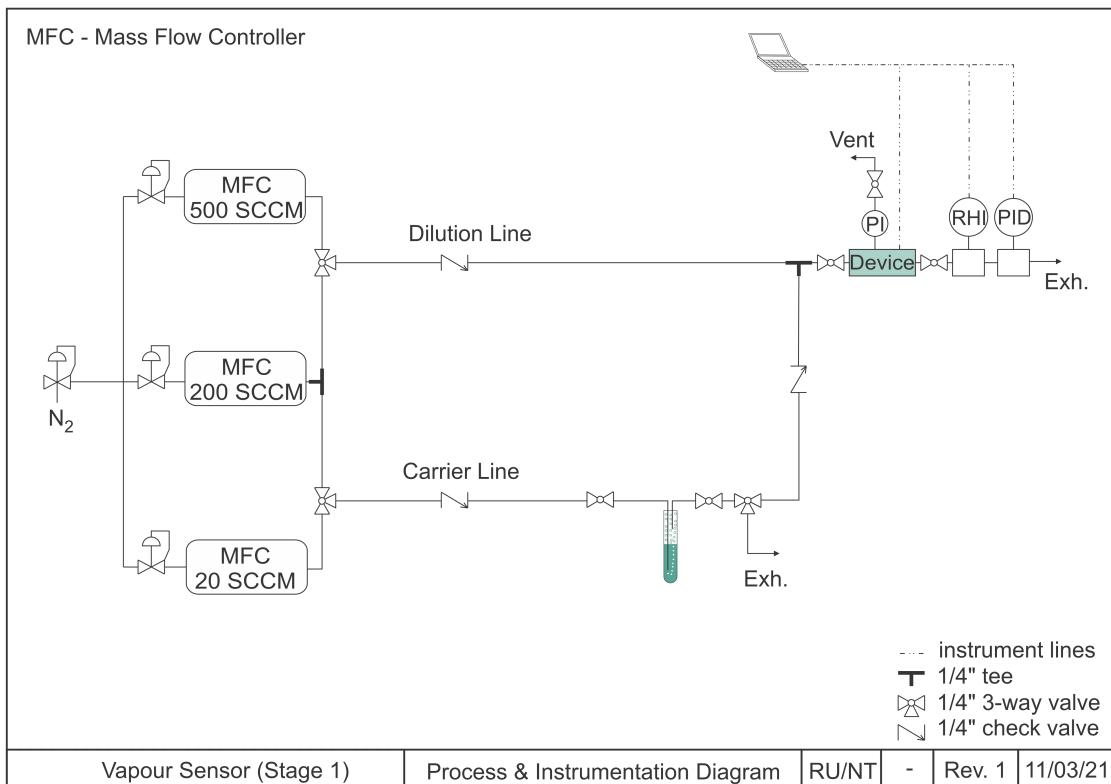


Figure 1.9.: P&ID of the Stage I vapour delivery system.

1. Vapour Sensing System for Transistor Biosensing

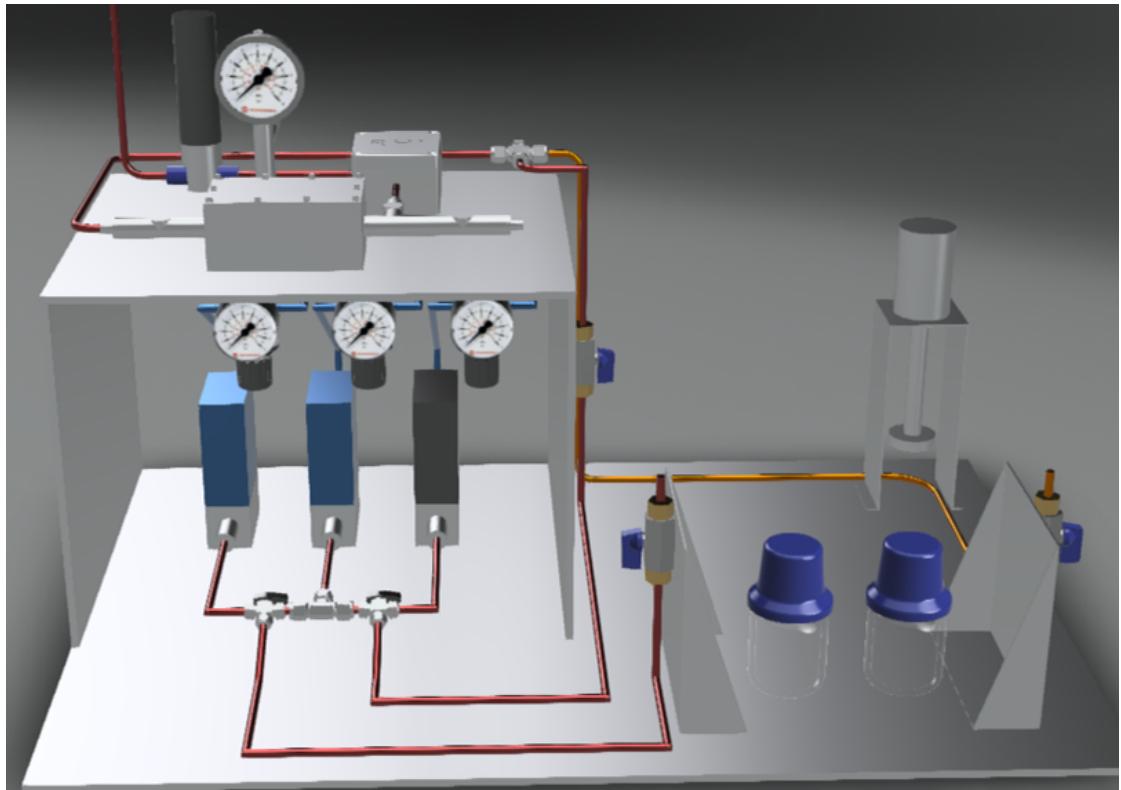


Figure 1.10.: Conceptual 3D Model of Stage I vapour delivery system. The model was made in Autodesk 360 by Alex Puglisi, School of Chemical and Physical Sciences, Te Herenga Waka - Victoria University of Wellington.

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the chamber, running clean nitrogen through the system for 3 hours was required before the PID returned to a constant baseline reading.

- There was no way to ensure the device chamber was free of analyte vapour before an experimental run aside from running nitrogen through the dilution line. After prolonged use, condensed analyte was sometimes visible in the PVC lines of the delivery system.

These issues, along with various minor structural and design issues, were addressed in the second-stage implementation of the system.

1.3.3. Stage II Design

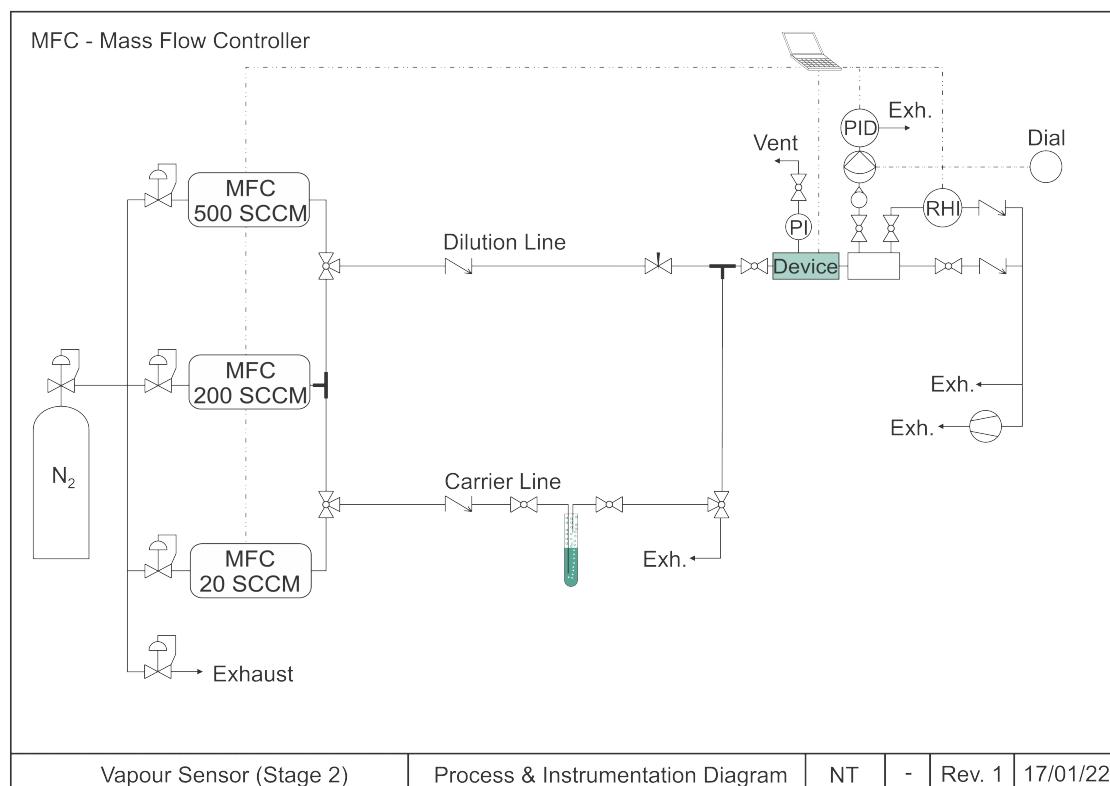


Figure 1.11.: Process & instrumentation diagram (P&ID) of the second-stage design for the vapour delivery system.

Figure 1.11 gives an overview of the second-stage design for the vapour delivery system setup. This stage of the redesign was implemented between Jan and May 2022. Changes from the first stage included:

- The addition of a N₂ cylinder (152D size) as the source of nitrogen for the system to replace the lab supply.

1. Vapour Sensing System for Transistor Biosensing

- A pressure indicator with a lower pressure range was used, which could register pressure changes within the device chamber.
- A chamber manifold was placed before the exhaust with outlets into the PID and RHI.
- A micro diaphragm pump was introduced between the manifold and PID to supply the PID with vapour from the chamber, and a flowmeter was placed before the pump to measure the flow rate out of the chamber to the PID. The PID was then seen to respond quickly to system changes (discussed further in Section 1.4).
- A piece of PVC tubing was placed at the PID outlet to limit air from the fumehood entering the PID when the micropump was off.
- Valves were placed before all system components so that the device chamber and post-analyte bottle carrier line could be evacuated with a roughing pump without potentially affecting components.
- Check valves were placed at the exhaust to prevent backflow from the roughing pump into the delivery system.

These changes broadly addressed the issues identified in Section 1.3.2.

1.4. Calibration and Measurements of Vapour Flow

1.4.1. Chamber Flow Calibration

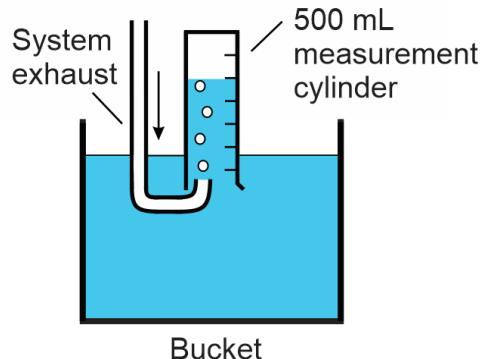


Figure 1.12.: Setup for calibration of mass flow controllers using the water displacement method.

A water displacement test was carried out to determine the relationship between the flow rate measured by the mass flow controllers and the actual flow rate passing through the chamber. All valves were set so that both the dilution and carrier lines followed a

1.4. Calibration and Measurements of Vapour Flow

single path. Both these paths went through the device chamber and out through the system exhaust. An empty analyte bottle was placed on the carrier line. The system exhaust was placed into a bucket filled with tap water, with the outlet sitting beneath an upturned 500 mL measurement cylinder, as pictured in Figure 1.12. The cylinder was used to measure the volume of displaced water over time, which is equivalent to the rate of change of nitrogen volume entering the cylinder from the exhaust. As leaks in the manifold and exhaust line were not detected when leak testing with bubble solution, it can be safely assumed that the rate at which nitrogen exits the exhaust is equivalent to the nitrogen flow rate through the device chamber.

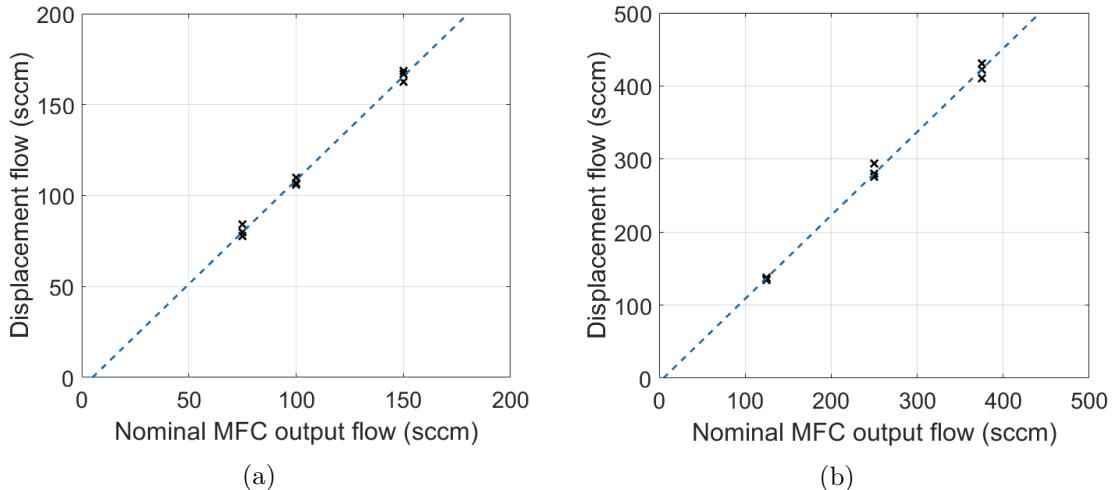


Figure 1.13.: Calibration curves for the 200 sccm full-scale MFC through the carrier line (a) and the 500 sccm full-scale MFC through the dilution line (b).

The time taken to displace a fixed volume of water was measured three times for a series of constant flow rates, both for the 200 sccm MFC (MKS) on the carrier line and the 500 sccm MFC (Tylan) on the dilution line. The displacement flow rate corresponding to each measurement could then be found by dividing volume by time. These measurements, of displacement flow relative to nominal flow through the MFC, are shown in Figure 1.13. The flow through the chamber was offset from the nominal flow reading from the mass flow controllers, with a strong linear relationship between the two measurements. A linear least-squares fit was performed, where coefficients a_1 and a_2 were found for the linear relationship $D = a_1d + a_2$, where d is nominal flow from the MFC and D is measured displacement flow. A 95% confidence interval for each fit was also obtained. For the 200 sccm MFC flow through the carrier line, values of $a_1 = 1.14 \pm 0.07$ and $a_2 = -6 \pm 8$ were obtained, while for the 500 sccm MFC flow through the dilution line, values of $a_1 = 1.14 \pm 0.06$ and $a_2 = -6 \pm 16$ were obtained.

It appears that the offset between the measured displacement flow and nominal output flow is not due to leaks in the system, since the offset indicates measured flow exceeds the nominal flow. Instead, the offset appears to be a systematic error introduced by the

1. Vapour Sensing System for Transistor Biosensing

electronics or software used to record the output flow from the MFCs. The identical offset between measured and nominal flow observed for each MFC, even when placed on different lines to the chamber, further strengthens the likelihood of the offset being due to the control side of the system. Furthermore, due to the identical offset for each of the carrier and dilution MFCs, the same offset should apply to a mixture of flows on each line. For example, a 200 sccm nominal flow through the dilution line from the 500 sccm full-scale MFC should have a roughly identical actual flow rate to a 50 sccm nominal flow through the dilution line and a 150 sccm flow through the carrier line. In this work, tests performed with the vapour delivery system have flow rate stated in terms of their nominal value, but the reader should keep in mind the $1.14 \times$ offset from the actual chamber flow.

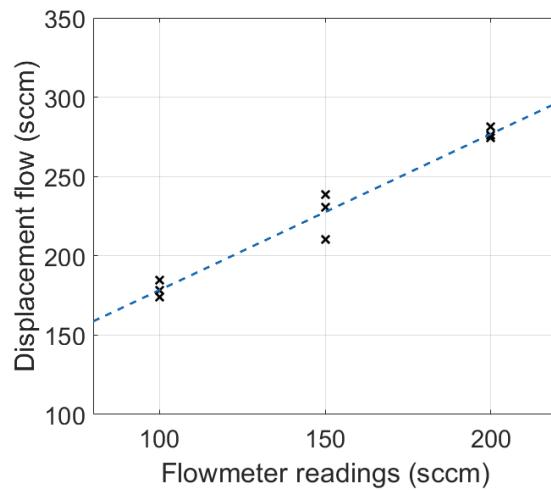


Figure 1.14.: Calibration curve for the PID flowmeter, comparing flowmeter readings with flow rate of water displacement.

The time taken to displace a fixed water volume was also measured three times for a series of constant flow rates through the flowmeter from the chamber to exhaust. A linear relationship was obtained between flowmeter readings and actual displacement, as shown in Figure 1.14. Expressing the relationship as $D = b_1 f + b_2$, where f is the flowmeter reading and D is measured displacement flow, values of $b_1 = 1.0 \pm 0.2$ and $b_2 = 80 \pm 25$ were obtained. A 200 sccm flow rate through the dilution line from the Tylan MFC, corresponding to a ~ 230 sccm actual flow rate through the chamber, would therefore be measured as a ~ 150 sccm flow rate by the flowmeter. Subsequent measurements showed that the linear relationship in Figure 1.14 breaks down for flowmeter readings $\lesssim 75$. For example, a 50 sccm flow on the flowmeter was found to correspond to 85 sccm of measured displacement flow. By disconnecting the flowmeter from the PID micropump and closing all valves out of the manifold except the valve exiting to the flowmeter (see Figure 1.11), the nominal flow on the MFC could be directly compared to the flowmeter reading. In this manner, the actual flow rate of readings $\lesssim 75$ sccm on the flowmeter could be quickly obtained.

1.4.2. Sensor Responses to Vapour Flow

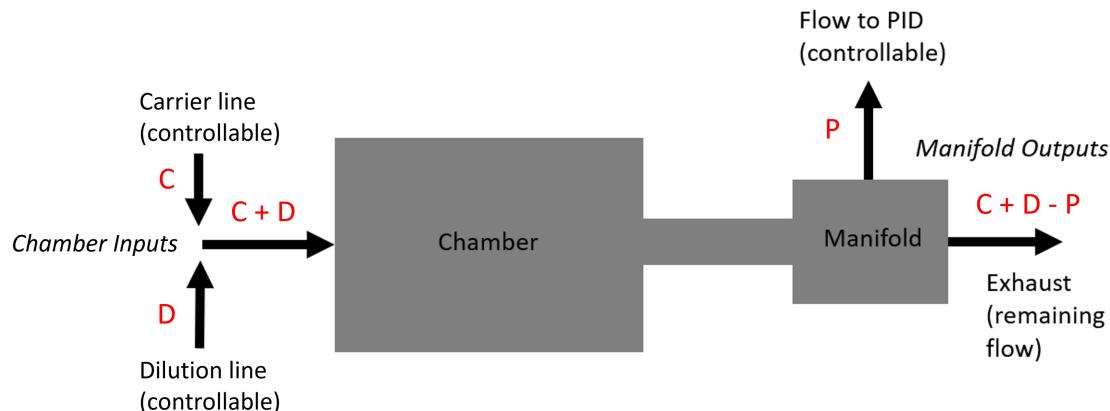


Figure 1.15.: Simplified schematic showing the flow into and out of the device chamber and manifold of the delivery system. The input flows from the carrier and dilution line are represented by C and D, and the output flow through the PID is represented by P. The exhaust can either flow past the relative humidity indicator or straight to the fumehood. This diagram assumes that flow through leaks in the chamber and manifold is low enough to be considered negligible, which was confirmed by leak testing with bubble solution.

Once the rate of flow through the device chamber had been calibrated, the next step was to verify the correct operation of the reference sensors used in the system. Various flow rates in and out of the chamber were used to calibrate and verify the reference sensors. These flows in and out of the chamber are labelled on the simplified schematic in Figure 1.15.

Relative Humidity Indicator

Table 1.1.: Flow sequence for testing relative humidity indicator.

t (s)	C (sccm)	D (sccm)
200	0	400
200	100	300
200	200	200
200	200	100
200	200	0

To test the relative humidity indicator (RHI), all valves out of the chamber were sealed

1. Vapour Sensing System for Transistor Biosensing

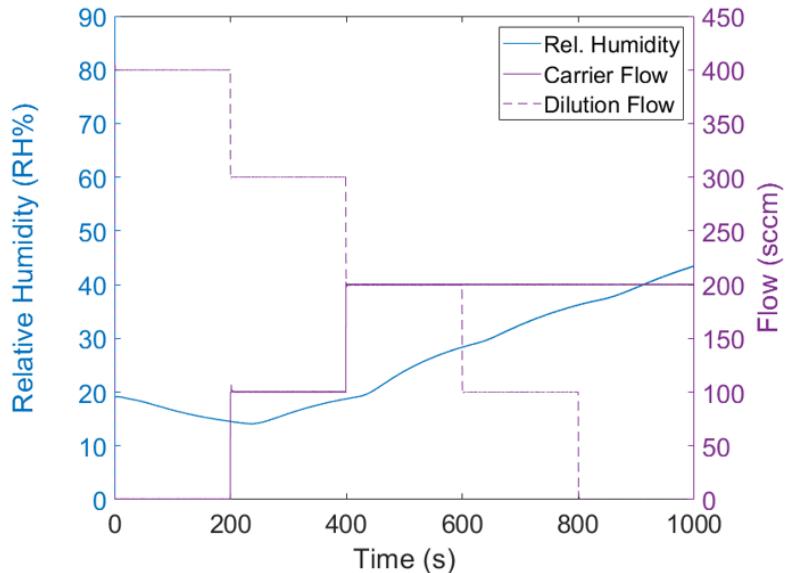


Figure 1.16.: Relative humidity readouts from the relative humidity indicator juxtaposed with flow rates from the dilution line and carrier lines of the vapour system, with 10 mL deionised water in the carrier line analyte bottle.

except for the valve for the relative humidity indicator chamber. This meant all flow coming out of the system would pass by the relative humidity indicator ($P = 0$ sccm and exhaust goes to RHI in Figure 1.15). Continuous nitrogen flow was then placed through the chamber until relative humidity dropped to about 20%. 10 mL of deionised water was placed into the analyte bottle. The following sequence of flows shown in Table 1.1 was placed through the chamber ($t = \text{time}$, $C = \text{carrier line flow rate}$, $D = \text{dilution line flow rate}$). Note that between 230 s and 630 s, the total flow rate remains the same, but the ratio of dilution to carrier flow differs.

From Figure 1.16, it is clear that with increased flow from the analyte bottle containing deionised water, the relative humidity registered by the Telaire sensor also increases. The increases in relative humidity occur about 50 s after corresponding increases in flow through the carrier line, which appears to be the time taken for water vapour to first reach the relative humidity indicator chamber. Over the full 800 s of carrier line flow, relative humidity increases from a minimum of 14.0% to a maximum of 44.3%.

Photoionisation Detector

First year report: ““First, a 200 sccm flow of N₂ gas was sent through the dilution line to the device chamber until 1000 s. Then, the flow controller three-way valves were manually adjusted so that the same 200 sccm flow was directed through 50 mL of EtOH analyte in the carrier line. This continued until 2200 s, where the valves were again

1.5. Summary

manually adjusted so that 200 sccm clean N₂ again flowed through the device chamber. The resulting current across the device channel was monitored over this time, and is shown in Figure 19. A response to EtOH exposure and removal is visible.” ”

1.5. Summary

A. Vapour System Hardware

Table A.1.: Major components used in construction of the vapour delivery system described in this thesis.

Description	Part No.	Manufacturer
Mass flow controller, 20 sccm full scale	GE50A013201SBV020	MKS Instruments
Mass flow controller, 200 sccm full scale	GE50A013202SBV020	MKS Instruments
Mass flow controller, 500 sccm full scale	FC-2901V	Tylan
Analogue flowmeter, 240 sccm max. flow	116261-30	Dwyer
Micro diaphragm pump	P200-B3C5V-35000	Xavitech
Analogue flow controller, for micro diaphragm pump	X3000450	Xavitech
10 mL Schott bottle	218010802	Duran
PTFE connection cap system	Z742273	Duran
Baseline VOC-TRAQ flow cell, red	043-951	Mocon
Humidity and temperature sensor	T9602	Telaire
Enclosure, for humidity and temperature sensor	MC001189	Multicomp Pro

B. Python Code for Data Analysis

B.1. Code Repository

The code used for general analysis of field-effect transistor devices in this thesis was written with Python 3.8.8. Contributors to the code used include Erica Cassie, Erica Happe, Marissa Dierkes and Leo Browning. The code is located on GitHub and the research group OneDrive, and is available on request.

B.2. Atomic Force Microscope Histogram Analysis

The purpose of this code is to analyse atomic force microscope (AFM) images of carbon nanotube networks in .xyz format taken using an atomic force microscope and processed in Gwyddion (see [?@sec-afm-characterisation](#)). It was originally designed by Erica Happe in Matlab, and adapted by Marissa Dierkes and myself for use in Python. The code imports the .xyz data and sorts it into bins 0.15 nm in size for processing. To perform skew-normal distribution fits, both *scipy.optimize.curve_fit* and *scipy.stats.skewnorm* modules are used in this code.

B.3. Raman Spectroscopy Analysis

The purpose of this code is to analyse a series of Raman spectra taken at different points on a single film (see [?@sec-raman-characterisation](#)). Data is imported in a series of tab-delimited text files, with the low wavenumber spectrum ($100\text{ cm}^{-1} - 650\text{ cm}^{-1}$) and high wavenumber spectrum ($1300\text{ cm}^{-1} - 1650\text{ cm}^{-1}$) imported in separate datafiles for each scan location.

B.4. Field-Effect Transistor Analysis

The purpose of this code is to analyse electrical measurements taken of field-effect transistor (FET) devices. Electrical measurements were either taken from the Keysight 4156C Semiconductor Parameter Analyser, National Instruments NI-PXIe or Keysight B1500A Semiconductor Device Analyser as discussed in [?@sec-electrical-characterisation](#);

B. Python Code for Data Analysis

the code is able to analyse data in .csv format taken from all three measurement setups. The main Python file in the code base consists of three related but independent modules: the first analyses and plots sensing data from the FET devices, the second analyses and plots transfer characteristics from channels across a device, and the third compares individual channel characteristics before and after a modification or after each of several modifications. The code base also features a separate config file and style sheet which govern the behaviour of the main code. The code base was designed collaboratively by myself and Erica Cassie over GitHub using the Sourcetree Git GUI.

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