

Protocol 4: PCR with HOT FIREPol® (Solis BioDyne) for amplification of rRNA and protein-coding genes (Archidona-Yuste *et al.*, 2018)

- Add 1–4 µl of extracted DNA to a 0.2 ml Eppendorf tube containing 5 µl of 5× HOT FIREpol® Blend Master Mix (with 10 mM MgCl₂), 0.15 µl of each primer (1.0 µg µl⁻¹), and distilled water to a final volume of 25 µl.
- Put the tube in the PCR machine with the following thermal profile: an initial denaturation at 95°C for 15 min, 35 cycles of 45 s at 94°C, 45 s at 55°C, and 45 s at 72°C, followed by a final step of 5 min at 72°C. For some specific A + T rich gene region as the *coxII*-16S used for identification of *Meloidogyne* species: an initial denaturation at 95°C for 15 min, 40 cycles of 1 min at 95°C, 1 min at 54°C, and 2 min 30 s at 66°C, followed by a final step of 7 min at 68°C.

Primer combination and code (direction) ^a	Primer sequence (5'-3')	Amplified region	References
G18SU (f)	GCT TGC CTC AAA GAT TAA GCC	18S rRNA	Blaxter <i>et al.</i> (1998)
R18Tyl1 (r)	GGT CCA AGA ATT TCA CCT CTC		Chizhov <i>et al.</i> (2006)
F18Tyl2 (f)	CAG CCG CGG TAA TTC CAG C	18S rRNA	Chizhov <i>et al.</i> (2006)
R18Tyl2 (r)	CGG TGT GTA CAA AGG GCA GG		
988F (f)	CTC AAA GAT TAA GCC ATG C	18S rRNA	Holterman <i>et al.</i> (2006)
1912R (r)	TTT ACG GTC AGA ACT AGG G		
1096F (f)	GGT AAT TCT GGA GCT AAT AC	18S rRNA	Holterman <i>et al.</i> (2006)
1912R (r)	TTT ACG GTC AGA ACT AGG G		
1813F (f)	CTG CGT GAG AGG TGA AAT	18S rRNA	Holterman <i>et al.</i> (2006)
2646R (r)	GCT ACC TTG TTA CGA CTT TT		
SSUF_04	GCT TGT CTC AAA GAT TAA GCC	18S rRNA	Blaxter <i>et al.</i> (1998)
SSUR_09	AGC TGG AATTAC CGC GGC TG		
SSUF_22	TCC AAG GAA GGC AGC AGG C	18S rRNA	Blaxter <i>et al.</i> (1998)
SSUR_13	GGG CAT CAC AGA CCT GTT A		
SSUF_23	ATT CCG ATA ACG AGC GAG A	18S rRNA	Blaxter <i>et al.</i> (1998)
SSUR_81	TGA TCC WKC YGC AGG TTC AC		
designated	CGC GAA TRG CTC ATT ACA	18S rRNA	Floyd <i>et al.</i> (2005)
Nem_18S_F	ACAGC		
Nem_18S_R	GGG CGG TAT CTG ATC GCC		
18S-CL-F3	CTT GTC TCA AAG ATT AAG CCA TGC AT	18S rRNA +	Carta and Li (2018, 2019)
28S-CL-R	CAG CTA CTA GAT GGT TCG ATT AGT C	ITS1-5.8S- ITS2 rRNA + 28S rRNA	
18S (f)	TTG ATT ACG TCC CTG CCC TT	ITS1-rRNA	Vrain <i>et al.</i> (1992)
rDNA1.58S (r)	ACG AGC CGA GTG ATC CAC CG		Szalanski <i>et al.</i> (1997)
TW81 (f)	GTT TCC GTA GGT GAA CCT GC	ITS1-rRNA	Curran <i>et al.</i> (1994)
5.8SM5 (r)	GGC GCAATG TGC ATT CGA		Zheng <i>et al.</i> (2000)
18S (f)	TTG ATT ACG TCC CTG CCC TT	ITS1-5.8S	Vrain <i>et al.</i> (1992)
26S (r)	TTT CAC TCG CCG TTA CTA AGG	ITS2 rRNA	
F194 (f)	CGT AAC AAG GTA GCT GTA G	ITS1-5.8S	Ferris <i>et al.</i> (1993)
F195 (r)	TCC TCC GCT AAA TGA TAG G	ITS2 rRNA	
TW81 (f)	GTT TCC GTA GGT GAA CCT GC	ITS1-5.8S	Curran <i>et al.</i> (1994)
AB21 (r)	AFA TGC TTA AGT TCA GCG GGT	ITS2 rRNA	
D2A (f)	ACA AGT ACC GTG AGG GAA AGTTG	D2-D3 of 28S rRNA	Nunn (1992)
D3B (r)	TCG GAA GGA ACC AGC TAC TA		
D2Tyl (f)	GAG AGA GTT AAA NAG BAC GTG A	D2-D3 of 28S rRNA	Chizhov <i>et al.</i> (2012)
D3B (r)	TCG GAA GGA ACC AGC TAC TA		Nunn (1992)
D2A (f)	ACA AGT ACC GTG AGG GAA AGTTG	D2 of 28S rRNA	Nunn (1992)
D2B (r)	GAC CCG TCT TGA AAC ACG GA		

^af, forward; r, reverse.

Nematode species	Primer code and sequence (5'-3')	Gene fragment	Amplified size	References
<i>Aphelenchoides besseyi</i>	AbF5 – ATG TGT AAG TAG AGC GTT ATA AbR5 – ATT CGC CGT TTT TAA GGC G- A	18S rRNA	~ 340 bp	Devran et al. (2017)
<i>Aphelenchoides fragariae</i>	AfragF1 – GCA AGT GCT ATG CGA TCT TCT AfragR1 – GCC ACA TCG GGT CAT TAT TT	ITS rRNA	~ 169 bp	McCuiston et al. (2007)
<i>Aphelenchoides ritzemabosi</i>	B5F – TCG ATG AAG AAC GCA GTG AATT ArtR – CTC CAC ACG CCG ACC GA	ITS rRNA	~ 208 bp	Cui et al. (2010)
<i>Bursaphelenchus cocophitus</i>	B2C1F – AAC TAC CGT CTT CCG CTG TCG B2C1R – TTG AGC ACC AAC ACG CCG TCA	ITS rRNA	~ 528 bp	Silva et al. (2016)
<i>Bursaphelenchus fraudulentus</i>	FF – GTG ATG GGT TTG CGG GCG GCG FR – CAA CCA ATG CAC ACC AAC CAA	ITS rRNA	~ 617 bp	Filipiak et al. (2010)
<i>Bursaphelenchus mucronatus</i>	MF – TCCG GCCAATCTCTACGAC MR – GTTCAACCAATCCGAACC	ITS rRNA	~ 210 bp	Matsuriaga and Togashi (2004)
<i>Bursaphelenchus xylophilus</i>	XF – ACGATGATGCGATTGGTGAC XR – TATTGGTCGCGGAACAAACC	ITS rRNA	~ 557 bp	Matsuriaga and Togashi (2004)
<i>Bursaphelenchus mucronatus</i>	Y01F – AGT CCG TGC CTT TGC TCT AGC Y01R – CCG AAG TGT CTC CAG CGA AAT	SCAR	~ 809 bp	Chen et al. (2011)
<i>Bursaphelenchus xylophilus</i>	B22 – TCA CGA TGA TGC GAT TGG TG B23 – AGA AGA TCT TGG TCG CGG AA	ITS rRNA	~ 580 bp	Jiang et al. (2005)
<i>Ditylenchus destructor</i>	D2 – TGG ATC ACT CGG CGG CTC GTAGA D1 – ACT GCT CTG CGT TTG GCT TCA	D2-D3 of 28S rRNA	~ 346 bp	Liu et al. (2007)
<i>Ditylenchus dipsaci</i>	D1NF1 – TTA TGA CAA ATT CAT GGC GG rDNA2 – TTT CAC TCG CCG TTA CTA AGG	ITS rRNA	~ 263 bp	Subbotin et al. (2005)
<i>Ditylenchus dipsaci</i>	U831 – AAY AAR ACM AAG CCN TYT GGA C Dipsaci-hsp90R – GWG TTA WAT AAC TTG GTC RGC	Hsp90	~ 182 bp	Madani et al. (2015)
<i>Ditylenchus dipsaci</i>	H05 – TCA AGG TAA TCT TTT TCC CCA CT H06 – CAACTG CTA ATG CGT GCT CT	SCAR	~ 242 bp	Esquibet et al. (2003)
<i>Ditylenchus dipsaci</i>	DdpS1 – TGG CTG CGT TGA AGA GAA CT rDNA2 – TTT CAC TCG CCG TTA CTA AGG	ITS rRNA	~ 517 bp	Kerkoud et al. (2007)
<i>Ditylenchus dipsaci</i>	DITunF – CTG TAG GTG AAC CTG C DITdipR – GAC ATC ACC AGT GAG CAT CG	ITS rRNA	~ 148 bp	Jeszke et al. (2015)
<i>Ditylenchus gigas</i>	D09 – CAA AGT GTT TGA TCG ACT GGA D10 – CAT CCC AAA ACA AAG AAA GG	SCAR	~ 198 bp	Esquibet et al. (2003)

Nematode species	Primer code and sequence (5'-3')	Gene fragment	Amplified size	References
<i>Ditylenchus gigas</i>	DITuniF – CTG TAG GTG AAC CTG C DITgigR – GAC CAC CTG TCG ATT C	ITS rRNA	~ 270 bp	Jeske et al. (2015)
<i>Globodera rostochiensis</i>	PITSi3 – AGC GCA GAC ATG CCG CAA ITS5 – GGA AGT AAA AGT CGT AAC AAG G	ITS rRNA	~ 434 bp	Bulman and Marshall (1997)
<i>Globodera rostochiensis</i>	GGT GAC TCG ACG ATT GCT GT GCA GTT GGC TAG CGA TCT TC	ITS rRNA	~ 391 bp	Mulholland et al. (1996)
<i>Globodera pallida</i>	PITSp4 – ACA ACA GCA ATC GTC GAG ITS5 – GGA AGT AAA AGT CGT AAC AAG G	ITS rRNA	~ 265 bp	Bulman and Marshall (1997)
<i>Globodera pallida</i>	GGT GAC TCG ACG ATT GCT GT GCA GTT GGC TAG CGA TCT TC	ITS rRNA	~ 238 bp	Mulholland et al. (1996)
<i>Heterodera avenae</i>	AVEN-COIF - GGG TT T TCG GTT ATT TGG AVEN-COIR - CGC CTA TCT AAA TCT ATA CCA	COI	~ 109 bp	Toumi et al. (2013a)
<i>Heterodera filipjevi</i>	FILI-COIF - GTA GGA ATA GAT TTA GAT AGT C FILI-COIR - TGA GCA ACA ACA TAA TAAG	COI	~ 245 bp	Toumi et al. (2013a)
<i>Heterodera filipjevi</i>	HfF1 – CAG GAC GAA ACT CAT TCA ACCAA HfR1 – AGG GCG AAC AGG AGA AGA TTAGA	SCAR	~ 646 bp	Peng et al. (2013)
<i>Heterodera latipons</i>	Hlat-actF - ATG CCA TCA TTA TTC CTT Hlat-actR - ACA GAG AGT CAA ATT GTG	actin	~ 204 bp	Toumi et al. (2013b)
<i>Heterodera glycines</i>	JBG1 – TGG TT T AGT TAG ATT AAC TAT C JB3R – TCC AAA CTW GCG TTA CTY AG	COI	~ 339 bp	Ko et al. (2017)
<i>Heterodera glycines</i>	SCNFI – GGA CCC TGA CCA AAA AGT TTCCGC SCNRI – GGA CCC TGA CGA GTT ATG GGCCCG	SCAR	~ 477 bp	Qu et al. (2008)
<i>Heterodera glycines</i>	GlyF1 – TTA CGG ACC GTA ACT CAA 26S – TTT CAC TCG CCG TTA CTA AGG	ITS rRNA	~ 181 bp	Subbotin et al. (2001)
<i>Heterodera schachtii</i>	JBS1 – GGA TAA TT T ATG CTA TTA TC JB3R – TCC AAA CTW GCG TTA CTY AG	COI	~ 339 bp	Ko et al. (2017)
<i>Heterodera schachtii</i>	SHF8 – GTT CTT ACG TTA CTT CCA AB28 – ATA TGC TTA AGT TCA GCG GGT	ITS rRNA	~ 200 bp	Anisii et al. (2002)
<i>Hoplolaimus columbus</i>	Hoc-1f – AAC CTG CTG CTG GAT CAT TA HC-1r – TCA GCA CACAAT GGTACC TTT	ITS1 rRNA	~ 580 bp	Bae et al. (2009)
<i>Hoplolaimus galeatus</i>	Hoc-1f – AAC CTG CTG CTG GAT CAT TA HG-2r – TCC TCG TTC ACA CAT TGACA	ITS1 rRNA	~ 120 bp	Bae et al. (2009)

Nematode species	Primer code and sequence (5'-3')	Gene fragment	Amplified size	References
<i>Hoplolaimus magnistylus</i>	Hoc-1f – AAC CTG CTG CTG GAT CAT TA HM-3r – AGA CTG GAC GGC CAA AGTT	ITS1 rRNA	~ 340 bp	Bae et al. (2009)
<i>Longidorus attenuatus</i>	GenF – TTG ATT ACG TCC CTG CCC TTT GT Latten3 – TTC CCT TTT CCC TGA TTA TAA TTT TCT ATC	ITS1 rRNA	~ 419 bp	Hübschen et al. (2004)
<i>Longidorus elongatus</i>	GenF – TTG ATT ACG TCC CTG CCC TTT GT Lelong1 – TTA TCG TAC GTA TTC CCA GTT CT	ITS1 rRNA	~ 847 bp	Hübschen et al. (2004)
<i>Longidorus macrosoma</i>	GenF – TTG ATT ACG TCC CTG CCC TTT GT Lmacro2 – GTT CCC GAC GAT TAT TTT TGT	ITS1 rRNA	~ 705 bp	Hübschen et al. (2004)
<i>Longidorus helveticus</i>	GenF – TTG ATT ACG TCC CTG CCC TTT GT Lhel1 – CCG CAT CTC TTT ATT TCC GAC CAT CAA CC	ITS1 rRNA	~ 369 bp	Hübschen et al. (2004)
<i>Longidorus profundorum</i>	GenF – TTG ATT ACG TCC CTG CCC TTT GT Lprof2 – TTA TTA TTT TTC AGG CTC TACCTTTTCGC	ITS1 rRNA	~ 1071bp	Hübschen et al. (2004)
<i>Longidorus sturhani</i>	GenF – TTG ATT ACG TCC CTG CCC TTT GT Lstur – TTT TCC CCA CTA ATA CTC CCT CGTT	ITS1 rRNA	~ 667 bp	Hübschen et al. (2004)
<i>Meloidogyne arenaria</i>	Far – TCG GCG ATA GAG GTA AAT GAC Rar – TCG GCG ATA GAC ACT ACA AACT	SCAR	~ 420 bp	Zijlstra et al. (2000)
<i>Meloidogyne chitwoodi</i>	Fc – TGG AGA GCA GCA GGA GAA AGA Rc – GGT CTG AGT GAG GAC AAG AGTA	SCAR	~ 800 bp	Zijlstra (2000)
<i>Meloidogyne enterolobii</i>	Me-F – AAC TT TGTGAAAGTGCCGCTG Me-R – TCAGTTCAGGCAGGATCAACC	IGS rRNA	~ 200 bp	Long et al. (2006)
<i>Meloidogyne exigua</i>	Ex-D15-F – CAT CCG TGC TGT AGC TGCGAG Ex-D15-R – CTC CGT GGG AAG AAA GACTG	SCAR	562 bp	Randig et al. (2002)
<i>Meloidogyne fallax</i>	Ff – CCA AAC TAT CGT AAT GCA TTA TT Rf – GGA CAC AGT AAT TCA TGA GCTAG	SCAR	~ 515 bp	Zijlstra et al. (2000)
<i>Meloidogyne hapla</i>	Fh – TGA CGG CGG TGA GTG CGA Rh – TGA CGG CGG TAC CTC ATAG	SCAR	610 bp	Zijlstra (2000)
<i>Meloidogyne incognita</i>	Finc – CTC TGC CCA ATG AGC TGTCC Rinc – CTC TGC CCT CAC ATT AGG	SCAR	~ 1200 bp	Zijlstra et al. (2000)

Nematode species	Primer code and sequence (5'-3')	Gene fragment	Amplified size	References
<i>Meloidogyne incognita</i>	MI-F – GTG AGG ATT CAG CTC CCC AG MI-R – ACG AGG AAC ATA CTT CTC CGTCC	SCAR	~ 955 bp	Meng et al. (2004)
<i>Meloidogyne incognita</i>	F – TAG GCA GTA GGT TGT CGG G R – CAGA A TCT CTG CAT TGG TGC	SCAR	~ 1350 bp	Dong et al. (2001)
<i>Meloidogyne incognita</i>	Inc-K14-F – GGG ATG TGT AAA TGCTCCTG Inc-K14-R – CCC GCT ACA CCC TCA ACTTC	SCAR	~ 399 bp	Randig et al. (2002)
<i>Meloidogyne javanica</i>	Fjav – GGT GCG CGA TTG AAC TGAGC Rjav – CAG GCC CTT CAG TGG AAC TAT AC	SCAR	~ 620 bp	Zijlstra et al. (2000)
<i>Meloidogyne naasi</i>	N-ITS – CTC TTT ATG GAG AAT AAT CGTR195 – CCT CCG CTT ACTGAT ATG	ITS rRNA	433 bp	Zijlstra et al. (2004)
<i>Nacobbus</i> spp	NacF – GAT CAT TAC ACG TAC CGT GAT GGTC NacR – CTG CTC AAC CAC GCA TAG ACG	ITS rRNA	141-173 bp	Atkins et al. (2005)
<i>Paralongidorus maximus</i>	GenF – TTG ATT ACG TCC CTG CCC TT TGT Pmax1 – TGCATT TCA CCA CTT CTC ACTC	ITS1 rRNA	~ 649 bp	Hübschen et al. (2004)
<i>Paratrichodorus allius</i>	BL18 – CCC GTC GMT ACT ACC GATT PAR2 – CCG TYC AAA CGC GTA TAT GAT C	ITS rRNA	~ 432 bp	Riga et al. (2007)
<i>Paratrichodorus teres</i>	BL18 – CCC GTC GMT ACT ACC GAT T PTR4 – CCT GAC AAG CTT GCA CTAG C	ITS rRNA	~ 677 bp	Riga et al. (2007)
<i>Pratylenchus brachyurus</i>	18S – TTG ATT ACG TCC CTG CCC TTT ACM7R – GCW CCA TCC AAA CAA YGA G	ITS1 rRNA	~ 267 bp	Machado et al. (2007)
<i>Pratylenchus bolivianus</i>	TW81 – GTT TCC GTA GGT GAA CCT GC P-bolivR1 – ATA GCG CAC TGG CGC AGCATA	ITS rRNA	~ 295 bp	Troccoli et al. (2016)
<i>Pratylenchus crenatus</i>	PCR22 (f) – AAA GCC TGA ATG CCC TGA G PCR22 (r) – AAA TTG AAA GAG GTC GGTCGT	ITS rRNA	~ 610 bp	Mekete et al. (2011)
<i>Pratylenchus jaehni</i>	Pj1F – TGG TCA ATG AAT GTT ACG 5818 – ACG ARC CGA GTG ATC CAC	ITS1 rRNA	~ 476 bp	Consoli et al. (2012)
<i>Pratylenchus neglectus</i>	PNEG – ATG AAA GTG AAC ATG TCC TC D3B – TCG GAA GGA ACC AGC TAC TA	D3 of 28S rRNA	~ 290 bp	Al-Banna et al. (2004)
<i>Pratylenchus neglectus</i>	PNEG-F1 – CGC AAT GAA AGT GAA CAATGTC D3B5 – AGT TCA CCA TCT TTC GGG TC	D3 of 28S rRNA	~ 144 bp	Yan et al. (2008)

Nematode species	Primer code and sequence (5'-3')	Gene fragment	Amplified size	References
<i>Pratylenchus oleae</i>	Poleae-fw1 – GAC AGA TTA GAA TGG AAT CTG TTCG Poleae-iv1 – ATC GCT TTT GGA TTC AATAAT ATA	ITS rRNA	~ 547 bp	Palomares-Rius <i>et al.</i> (2014)
<i>Pratylenchus parazeae</i>	PpzF – CTG CTG CTG GAT CAT TAC ATT PpzR – TCA AAT AGA CAT GCC CCA AT	ITS rRNA	~ 570 bp	Wang <i>et al.</i> (2015)
<i>Pratylenchus penetrans</i>	PPEN – TAA AGA ATC CGC AAG GAT AC D3B -TCG GAA GGA ACC AGC TAC TA	D3 of 28S rRNA	~ 278 bp	Al-Banna <i>et al.</i> (2004)
<i>Pratylenchus penetrans</i>	PP5 (f) –ACA TGG TCG ACA CGG TGA TA PP5 (r) - TGT TGC GCA AAT CCT GTT TA	beta-1,4 endoglu- canase	~ 520 bp	Mekete <i>et al.</i> (2011)
<i>Pratylenchus penetrans</i>	PpenA – TGA CTA TAT GAC ACA TTT RAACTTG AB28 -ATA TGCTTAAGT TCA GCG GGT	ITS rRNA	~ 660 bp	Waeyenberge <i>et al.</i> (2009)
<i>Pratylenchus penetrans</i>	PP1 – ATG ATG GAA GTG TCC GCC T PP2 – CCC AAC GAC GGT CAAAG G	ITS rRNA	~ 462 bp	Uehara <i>et al.</i> (1998)
<i>Pratylenchus scribneri</i>	PSCR – AAA GTG AAC GTT TCC ATT TC D3B -TCG GAA GGA ACCAGC TAC TA	D3 of 28S rRNA	~ 286 bp	Al-Banna <i>et al.</i> (2004)
<i>Pratylenchus scribneri</i>	PsF7 – AGT G1T GCT ATA ATT CAT GTAAAG TTGC PsR7 –TGG CCA GAT GCG ATT CGA GAG GTGT	ITS rRNA	~ 136 bp	Huang and Yan (2017)
<i>Pratylenchus speijeri</i>	TW81 - GTT TCC GTA GGT GAA CCT GC speijeri-specific – GTG CAC TGA TGT TAT TAT GTA TGG	ITS rRNA	~ 102 bp	De Luca <i>et al.</i> (2012)
<i>Pratylenchus thomei</i>	PTH – GAA AGT GAA GGT ATC CCT CG D3B -TCG GAA GGA ACCAGC TAC TA	D3 of 28S rRNA	~ 288 bp	Al-Banna <i>et al.</i> (2004)
<i>Pratylenchus thomei</i>	Pthf - TTC GGA AGA CAA TAA ATC Pthr - TCC AAA ATG AAA TAA TAA A	SCAR	~ 1078 bp	Carrasco-Ballesteros <i>et al.</i> (2007)
<i>Pratylenchus vulnus</i>	PVUL – GAA AGT GAA CGC ATC CGC AA D3B -TCG GAA GGA ACC AGC TAC TA	D3 of 28S rRNA	~ 287 bp	Al-Banna <i>et al.</i> (2004)
<i>Pratylenchus zeae</i>	TW81 - GTT TCC GTA GGT GAA CCT GC P-zeaeR1 - TAC GCA TAC RGT TCT GCT CAT	ITS rRNA	~ 560 bp	Troccoli <i>et al.</i> (2016)
<i>Radopholus similis</i>	PF – CTA CAA ATG TGA CGC GAA PR – CAA TCT GCA CAA TGA ACA TAC	ITS rRNA	~ 500 bp	Liu <i>et al.</i> (2011)
<i>Radopholus similis</i>	RsimF – GAT TCC GTC CTT TGG TGG GCA RsimR – GAA CCA GGC GTG CCA GAG G	ITS rRNA	~ 398 bp	Ravindran <i>et al.</i> (2011)

Nematode species	Primer code and sequence (5'-3')	Gene fragment	Amplified size	References
<i>Rotylenchulus reniformis</i>	Ren240F – ACC GGC TTA ATT GCA ATGGT Ren240R – ACA ACT GCT CAA CAA CGCAG	ITS rRNA	~ 240 bp	Sayler <i>et al.</i> (2012)
<i>Rotylenchulus reniformis</i>	D2A - ACA AGT ACC GTG AGG GAA AGTTG Rrenit-R1A - GAA AAG GCC TAC CCA ATGTG	D2-D3 of 28S rRNA	~ 142 bp	Van den Berg <i>et al.</i> (2016)
<i>Rotylenchus robustus</i>	TW81 - GTT TCC GTA GGT GAA CCT GC CCT GC R-robustus - GACGTGGACATCAATACAGTC	ITS rRNA	~ 438 bp	Cantalapiedra-Navarrete <i>et al.</i> (2013)
<i>Scutellonema bradys</i>	TW81 - GTT TCC GTA GGT GAA CCT GC S-bradys – GTG ATG GCT AAA CCA CAT TC	ITS rRNA	~ 250 bp	Van den Berg <i>et al.</i> (2013)
<i>Scutellonema brachyurus</i>	TW81 - GTT TCC GTA GGT GAA CCT GC S-brachyurustype A – GCT GAA GTG ACA GCC CAA CTT	ITS rRNA	~ 185 bp	Van den Berg <i>et al.</i> (2013)
<i>Tylenchutius semipenetrans</i>	TW81 - GTT TCC GTA GGT GAA CCT GC Semipenetrans – GGA CTC TGC TCA ACCTGG TAGA	ITS rRNA	~ 113 bp	Tanha Maañ <i>et al.</i> (2012)
<i>Xiphinema diversicaudatum</i>	TW81 - GTT TCC GTA GGT GAA CCT GC Xip-diver-ITS - GAA TAA ACA CCT TTC AAC GCTC	ITS rRNA	~ 864 bp	Chizhov <i>et al.</i> (2014)
<i>Xiphinema index</i>	I27 – GAG TCG TAA CGT TTC TCG TCT ATCAGG A-ITS1 – GAA TAG CCA CCT AGT GAG CCGAGCA	ITS rRNA	~ 340 bp	Wang <i>et al.</i> (2003)
<i>Xiphinema vuittenezi</i>	V18 – GTG GAA CGA AAA GACCTC A-ITS1 – GAA TAG CCA CCT AGT GAG CCGAGCA	ITS rRNA	~ 591 bp	Wang <i>et al.</i> (2003)
<i>Xiphinema italiae</i>	ITA26 – GAA ATA AGA ACC CTG AAA AAG ATA GG A-ITS1 – GAA TAG CCA CCT AGT GAG CCGAGCA	ITS rRNA	~ 414 bp	Wang <i>et al.</i> (2003)