

# R implementation

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1 Loaded functions:

† Project started July 10 2015

```
#source("/media/Data/Dropbox/humanR/01funcs.R")
rm(list=ls())
#setwd("/media/Data/Dropbox/humanR/PD/")
#setwd("~/Dropbox/humanR/PD/")
###load("PD.Rdata", .GlobalEnv)
#lsos(pat="")
```

## 2 1 Data preprocessing

3 Load packages.

```
pkgs <- c('xlsx', 'caret', 'leaps', 'glmnet', 'lattice', 'latticeExtra',
          'ggplot2')
lapply(pkgs, require, character.only = TRUE)

[[1]]
[1] TRUE

[[2]]
[1] TRUE

[[3]]
[1] TRUE

[[4]]
[1] TRUE

[[5]]
[1] TRUE

[[6]]
[1] TRUE

[[7]]
[1] TRUE
```

4 Load file with read counts per sample.

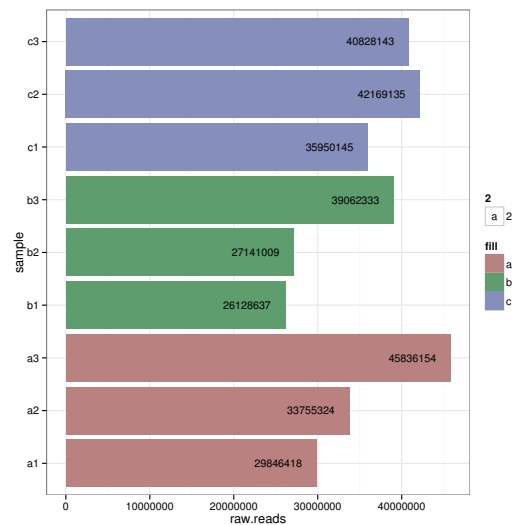
- 5 • A samples as nodule tissue
- 6 • B samples as non nodule diseased tissue
- 7 • C samples as non nodule non diseased tissue

8 Raw reads is the number of reads that have been trimmed, mapped to reference genome (Steve Roberts  
9 v15 with 21280 contigs), sorted by position on the genome, and cleaned from duplicated reads.

† Hypothetically these raw reads includes specific QPX reads

```
reads.counts <- read.xlsx("./data/mappedNodules.xlsx", sheetIndex = 1)
```

```
reads.counts$fill <- gl(3, 3, 9, labels = c("a", "b", "c"))
ggplot(reads.counts,
  aes(x = sample,
      y = raw.reads,
      fill = fill)) +
  coord_flip() +
  theme_bw() +
  geom_bar(stat = "identity") +
  geom_text(aes(x = sample,
                y = raw.reads,
                ymax = raw.reads,
                label = raw.reads,
                size = 2,
                hjust = 1.3)) +
  scale_fill_hue(c = 40, l = 60)
```

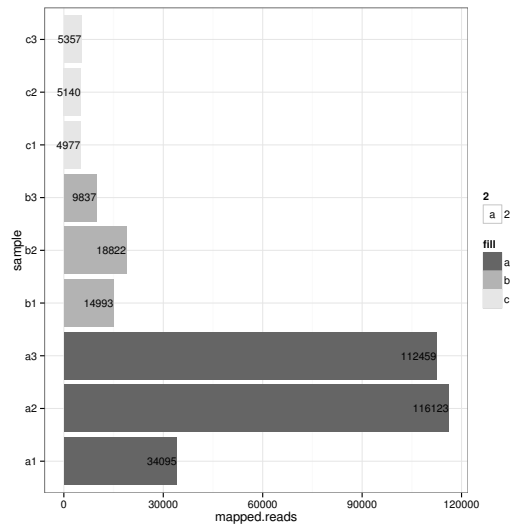


10

11 Number of reads that mapped to the reference genome of QPX.

↑ These reads are probably those of QPXs'

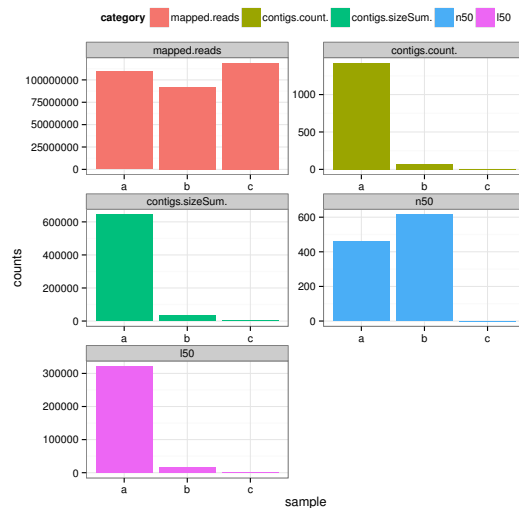
```
ggplot(reads.counts,
  aes(x = sample,
      y = mapped.reads,
      fill = fill)) +
  geom_bar(stat = "identity") +
  geom_text(aes(x = sample,
                y = mapped.reads,
                ymax = mapped.reads,
                label = mapped.reads,
                hjust = 1,
                size = 2)) +
  coord_flip() +
  theme_bw() +
  scale_fill_grey(start = .4, end = .9)
```



12 Mapped reads to the QPX reference where than assembled into contigs (ie, the reads showing in the  
 13 chart above).  
 14

† These contigs must be specific transcripts to QPX

```
reads.counts <- read.xlsx("./data/mappedNodules.xlsx", sheetIndex = 2)
reads.counts <- gather(reads.counts, "category", "counts", 3:7)
ggplot(reads.counts,
  aes(x = sample,
    y = counts,
    fill = category)) +
  geom_bar(stat = "identity") +
  theme_bw() +
  facet_wrap(~ category,
    ncol = 2,
    scales = "free") +
  theme(legend.position = "top")
```

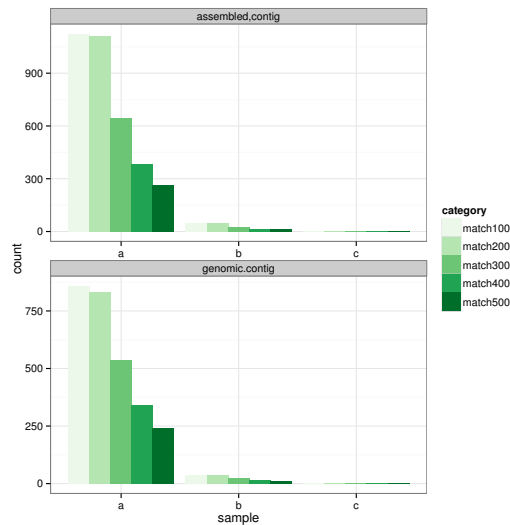


15 All contigs were than aligned to the reference genome QPX. The chart shows the number of contigs (both  
 16 genomic and mRNA sequenced) that align with an increasing length of 100>200>300>400>500.  
 17

† Helps discard misassembled contigs or non QPX ones

```
reads.counts <- read.xlsx("./data/mappedNodules.xlsx", sheetIndex = 4)
```

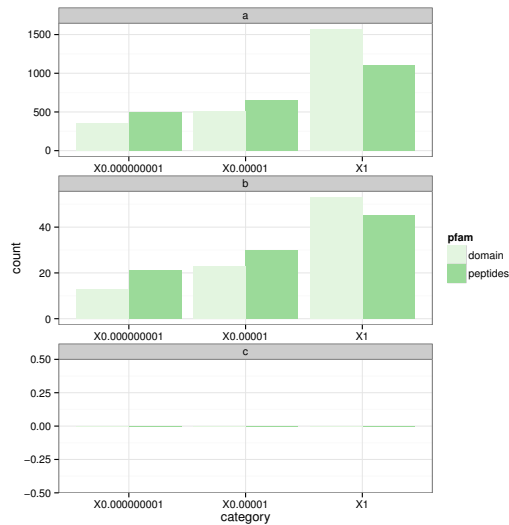
```
reads.counts <- gather(reads.counts, "category", "count", 3:7)
ggplot(reads.counts,
  aes(x = sample,
      y = count,
      fill = category)) +
  theme_bw() +
  geom_bar(stat = "identity",
    position = "dodge") +
  facet_wrap(~ blat, ncol = 1, scale = "free") +
  scale_fill_manual(values = brewer.pal(5, "Greens"))
```



18  
19 All contigs were then translated to peptides in 6 frames. Each frame peptide was then aligned to the  
20 whole PFAM library (v28, date: Jul 14 2015).

```
reads.counts <- read.xlsx("./data/mappedNodules.xlsx", sheetIndex = 3)
reads.counts <- gather(reads.counts, "category", "count", 3:5)
ggplot(reads.counts,
  aes(x = category,
      y = count,
      fill = pfam)) +
  geom_bar(stat = "identity",
    position = "dodge") +
  theme_bw() +
  facet_wrap(~ sample, ncol = 1,
    scales = "free") +
  scale_fill_manual(values = brewer.pal(2, "Greens"))
```

Warning in brewer.pal(2, "Greens"): minimal value for n is 3, returning requested palette with 3 different levels



21

## 22 **2 System Information**

23 The version number of R and packages loaded for generating the vignette were:

```
###save(list=ls(pattern=".*|.*)" , file="PD.Rdata")
sessionInfo()

R version 3.1.2 (2014-10-31)
Platform: x86_64-unknown-linux-gnu (64-bit)

locale:
 [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
 [3] LC_TIME=en_US.UTF-8      LC_COLLATE=en_US.UTF-8
 [5] LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
 [7] LC_PAPER=en_US.UTF-8     LC_NAME=en_US.UTF-8
 [9] LC_ADDRESS=en_US.UTF-8   LC_TELEPHONE=en_US.UTF-8
[11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=en_US.UTF-8

attached base packages:
[1] grid      stats      graphics  grDevices  utils      datasets
[7] methods   base

other attached packages:
 [1] glmnet_1.9-8      Matrix_1.1-4      leaps_2.9
 [4] caret_6.0-37      ggbiplot_0.55     scales_0.2.4
 [7] plyr_1.8.1        tidyr_0.1         vegan_2.2-0
[10] permute_0.8-3     dplyr_0.3.0.2     ggplot2_1.0.0
[13] latticeExtra_0.6-26 RColorBrewer_1.0-5 lattice_0.20-29
[16] xlsx_0.5.7        xlsxjars_0.6.1    rJava_0.9-6
[19] knitr_1.8

loaded via a namespace (and not attached):
 [1] assertthat_0.1      BradleyTerry2_1.0-5 brglm_0.5-9
 [4] car_2.0-22          cluster_1.15.3     codetools_0.2-9
 [7] colorspace_1.2-4    compiler_3.1.2     DBI_0.3.1
[10] digest_0.6.4        evaluate_0.5.5     foreach_1.4.2
[13] formatR_1.0         gtable_0.1.2       gtools_3.4.1
[16] highr_0.4           iterators_1.0.7     labeling_0.3
[19] lazyeval_0.1.9      lme4_1.1-7         magrittr_1.5
[22] MASS_7.3-35         mgcv_1.8-4         minqa_1.2.4
[25] munsell_0.4.2       nlme_3.1-118       nloptr_1.0.4
[28] nnet_7.3-8          parallel_3.1.2     proto_0.3-10
[31] Rcpp_0.11.3         reshape2_1.4       splines_3.1.2
[34] stringr_0.6.2       tcltk_3.1.2        tools_3.1.2
```