# Identifying HCC subtypes and the B2 subtype

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```
##load libraries
pacman::p_load(tidyverse,data.table,ComplexHeatmap,knitr,GSVA,
               fgsea,png,grid,dendextend,gridExtra,EnvStats,patchwork,
               kableExtra,unikn,patchwork,sjPlot,sjmisc,fastDummies,lsa)
setDTthreads(6)
##source the masterfile which contains several clinical and molecular features
Data <- fread("data/Masterfile_TCGA.tsv") %>% as.data.frame() %>%
  mutate_if(grepl("mut",.)==TRUE,function(x) factor(x,c("wt","mut"))) ##color palettes
asian_col <- usecol("pal_unikn_light")[c(5)]</pre>
caucasian col <- usecol("pal unikn light")[c(7)]</pre>
subtypes col pal <- c("B"="#d73027", "G"="#2166ac", "B1"="#ce1256",
                       "B2"="#ff7a00", "G1"="#1d91c0", "G2"="#41ab5d")
#as.character(Reduce(paste, deparse(formula(fit1))))
layout_mat = rbind(c(1,1,2,2,3,3),
                   c(1,1,2,2,3,3))
# source Functions file
source("Functions.R")
```

#### NMF output: Num of cluster selection

NMF was run in R package "NMF" using the Brunet algorithm. Top 3000 most variable genes were used to run NMF. Algorithm number of runs is 200. Below we will analyze transcriptomic subtypes in Asian and European patients in the TCGA cohort.

```
# load NMF results
Asian_NMF <- "data/asianTCGA_158_Asian_deseq2_top_3000.txt_200_nmfncbrunet_nc.rds"
Caucasian_NMF <- "data/caucasianTCGA_184_Caucasian_deseq2_top_3000.txt_200_nmfncbrunet_nc.rds"

# load top3000 most variables genes
Asian_top3000 <- fread("data/TCGA_158_Asian_deseq2_top_3000.txt") %>%
    tibble::column_to_rownames("V1")
Caucasian_top3000 <- fread("data/TCGA_184_Caucasian_deseq2_top_3000.txt") %>%
    tibble::column_to_rownames("V1")

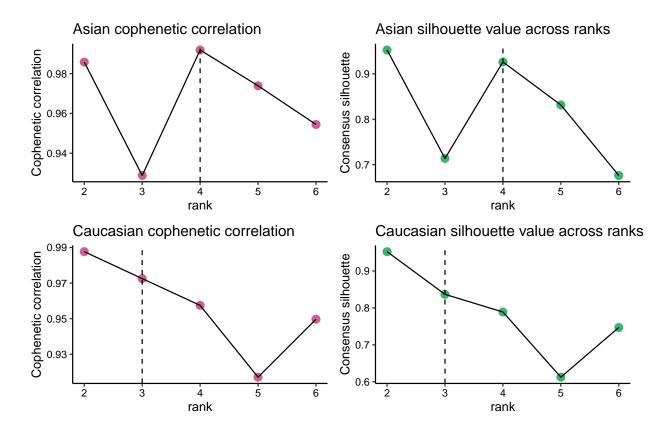
# load all normalized genes
Asian_all_genes <-fread("data/TCGA_158_Asian_deseq2.tsv") %>%
    tibble::column_to_rownames("V1")
Caucasian_all_genes <-fread("data/TCGA_184_Caucasian_deseq2.tsv") %>%
```

## [1] "The best fit for Asian is: 4"

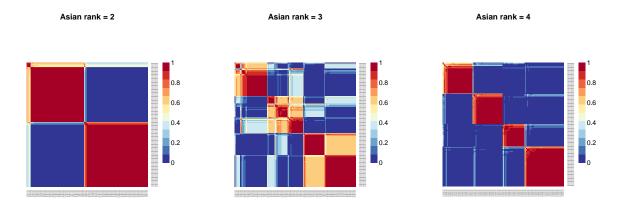
```
## [1] "The best fit for European is: 3"
```

We run NMF Brunet algorithm for different ranks from 2 to 6 and 200 runs for each. Let's look at cophenetic correlation and silhouette values across ranks to choose optimal number of clusters.

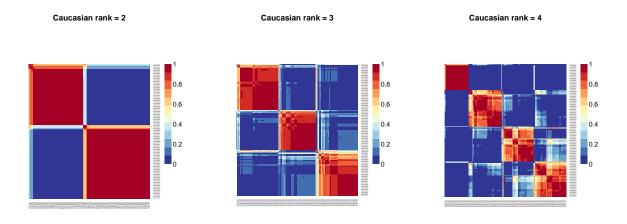
```
cop_sil_asian <- plotRanksNMF(Asian_NMF,"Asian")
cop_sil_caucasian <- plotRanksNMF(Caucasian_NMF,"Caucasian")
cop_sil_asian/cop_sil_caucasian</pre>
```



For Asians, 4 is the optimal number of cluster. For Caucasians 3 is the optimal number. Now let's plot consensus matrices.



For Asians, 4 looks quite delineated.

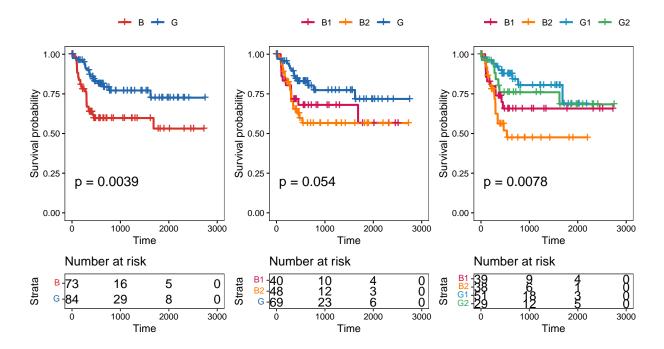


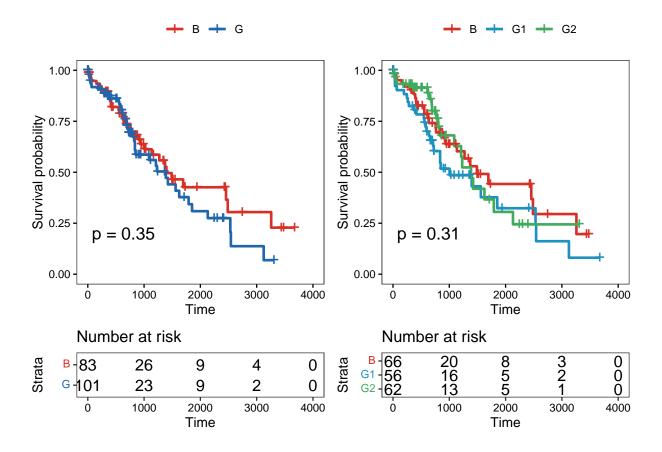
For Caucasian, 3 is better than 4.

```
## swap function for swapping subtype names
swap <- function(vec, from, to) {</pre>
  tmp <- to[ match(vec, from) ]</pre>
  tmp[is.na(tmp)] <- vec[is.na(tmp)]</pre>
 return(tmp)
}
#renaming was done after mapping to literature pathways.
asian2 <- getSubtypesNMF(Asian NMF,2) %>%
  mutate(subtype=swap(swap(subtype,1,"B"),2,"G"))
asian3 <- getSubtypesNMF(Asian_NMF,3) %>%
  mutate(subtype=swap(swap(swap(subtype,3,"B1"),2,"B2"),1,"G"))
asian4 <- getSubtypesNMF(Asian_NMF,4) %>%
  mutate(subtype=swap(swap(swap(swap(subtype,2,"B1"),1,"B2"),3,"G1"),4,"G2"))
caucasian2 <- getSubtypesNMF(Caucasian_NMF,2) %>%
  mutate(subtype=swap(swap(subtype,2,"B"),1,"G"))
caucasian3 <- getSubtypesNMF(Caucasian_NMF,3) %>%
  mutate(subtype=swap(swap(swap(subtype,2,"B"),3,"G2"),1,"G1"))
```

I have selected 2, 3 or 4 clusters for each cohort. Let's analyze survival differences of these subtypes. Note: We have changed B2 subtype naming to P2 later on. In this documents these two (P2/B2) could have been used to define Asian enriched subtype.

```
Data.survival <- fread("data/Masterfile_TCGA_survival.tsv")</pre>
asian subtypes <- asian2 %>% rename(S2="subtype") %>%
  inner_join(.,asian3 %>% rename(S3="subtype")) %>%
  inner_join(.,asian4 %>% rename(S4="subtype")) %>%
  mutate(sample=substr(sample,1,12)) %>%
 left join(.,Data.survival)
surv2 <- survPlot(asian subtypes, Time = "OS.time", Event = "OS", var = "S2", risktable = T,</pre>
          palette = c(subtypes_col_pal[["B"]],subtypes_col_pal[["G"]]))
surv3 <- survPlot(asian_subtypes, Time = "OS.time", Event = "OS", var = "S3", risktable = T,</pre>
          palette = c(subtypes_col_pal[["B1"]],subtypes_col_pal[["B2"]],
                      subtypes_col_pal[["G"]]))
surv4 <- survPlot(asian_subtypes, Time = "OS.time", Event = "OS", var = "S4", risktable = T,</pre>
          palette = c(subtypes_col_pal[["B1"]],subtypes_col_pal[["B2"]],
                       subtypes_col_pal[["G1"]],subtypes_col_pal[["G2"]]))
survs_asian <- (surv2$plot+surv2$table+plot_layout(ncol = 1,heights = c(5,1)))|</pre>
  (surv3$plot+surv3$table+plot_layout(ncol = 1,heights = c(5,1)))|
  (surv4$plot+surv4$table+plot_layout(ncol = 1,heights = c(5,1)))
survs_asian
```



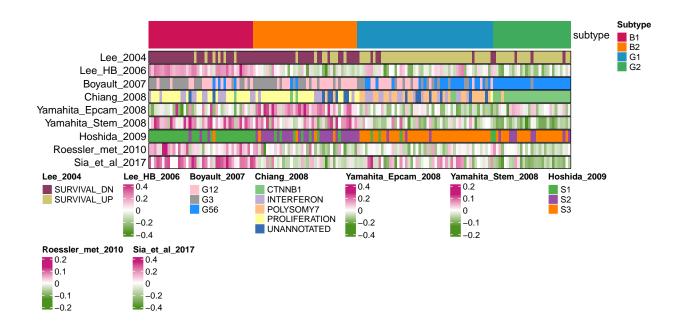


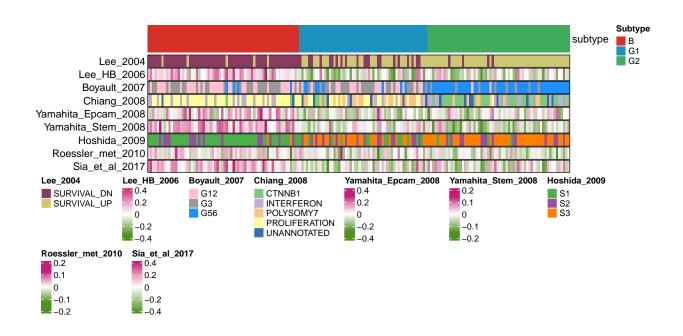
### Activity score calculation

Using gene signatures from previous HCC subtyping papers, I have calculated a pathway activity score. Patients were assigned to the subtype with highest pathway activity score for some subtypes (Hoshida, Boyault, Chiang, Lee survival) and raw activity scores were used for gene set (lee hepatoblast, Roessler Metastasis).

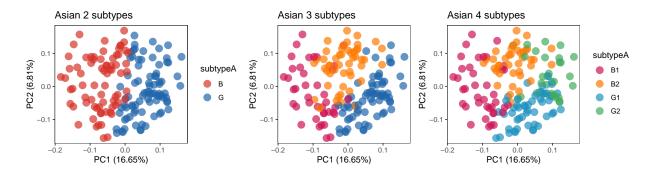
Steps (method from Robinson et al.)

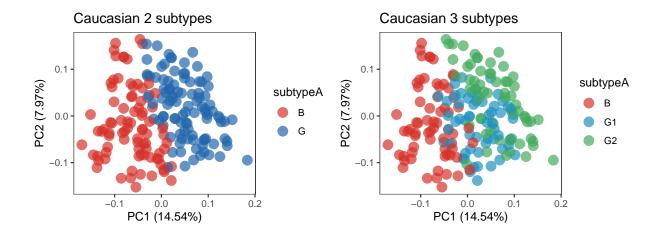
- Collect gene sets from MSigDB.
- Convert gene expression of all genes in a gene set to percentiles across patients.
- Take the mean percentile transformed value across genes for each patient.
- Assign patients to max pathway activity for subtypes (Hoshida, Lee, Chiang, Boyault), or keep pathway activity score (Roessler metastasis and Lee Hepatoblast subtype (HB))



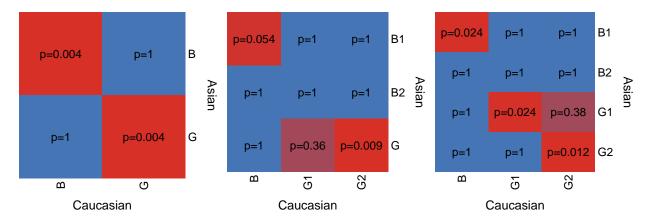


# **PCA** Analysis





### Mapping subtypes: SubMap



Now I will compare clinical and molecular features between Good (G) and Bad (B) subypes in Asian and Caucasian separately. I will also compare G1 and G2 in both cohorts. I can only compare B1 vs B2 in Asian.

#### Comparison of features between subtypes

```
#Feature list###
Clinical_features <- c("age", "gender", "race", "stage", "AFP", "vas_invasion", "HBV", "HCV")
Molecular features <- c("GII", "ploidy", "SCNA.focal", "SCNA.arm", "GD", "TMB", "immuneGr", "purity")
Selected drivers <- fread("data/TCGA driver binary.tsv") %>% select(-sample) %>%
  select if(colSums(.)>10) %>% colnames()
Frequent_CNV_events_arm <- c("AMP_1q","AMP_5p","AMP_6p",</pre>
                              "AMP_8q","DEL_13q","DEL_16q",
                              "DEL_17p", "DEL_21p", "DEL_4q", "DEL_8p")
Frequent_CNV_events_focal <- c("AMP_16q11.2", "AMP_1q21.1",
                                "AMP_2p11.1","DEL_1p36.13",
                                "DEL_1p36.31","DEL_16q11.2")
Clonality_features <- c("MATH", "Clone_no", "Simpson", "Shannon",
                         "pLM", "neutrality", "RNA_ITH")
Other_molecular_feature <- c("GII_amp", "GII_del", "SCNA.focal.amp",
                              "SCNA.focal.del", "SCNA.arm.amp", "SCNA.arm.del",
                              "SCNA.focal.now", "SCNA.arm.now", "SigGr")
Feat_list_for_RNA <- c(Clinical_features, Molecular_features, Selected_drivers)</pre>
##function to format Data for comparison of molecular and
##clinical feature between subtypes
manipulate_for_feat_comparison <- function(subtype_df,racem){</pre>
return(Data %>% left join(subtype df %>% mutate(sample=substr(sample,1,12))) %>%
  select(c(Feat_list_for_RNA,subtype)) %>% filter(race==racem) %>% select(-race) %>%
    rename(Purity="purity", Stage="stage", Age="age", Ploidy="ploidy") %%
  mutate(Female=ifelse(gender=="FEMALE","1","0")) %>%
fastDummies::dummy_cols(c("Stage"),remove_selected_columns = T,
                         remove_first_dummy = F,ignore_na = T) %>%
  mutate_at(.,vars(matches("stage_")),as.character) %>%
  mutate_at(.,vars(matches("AMP_|DEL_")), function(x) ifelse(x%in%c("AMP","DEL"),"1","0")) %%
  mutate_at(.,vars(matches(Selected_drivers)),function(x) ifelse(x=="mut","1","0")) %>%
  mutate(GD=ifelse(GD=="GD","1","0"), #remove immune qr qender
         Hot=ifelse(immuneGr=="hot","1","0"),
         HBV=ifelse(HBV=="HBV+","1","0"),
```

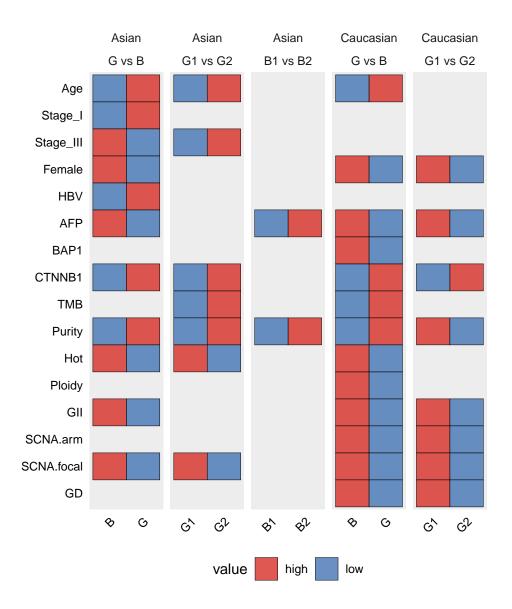
```
HCV=ifelse(HCV=="HCV+","1","0"),
         MVI=ifelse(vas_invasion=="Yes","1","0")) %>%
  select(-c(gender,immuneGr,vas_invasion)))
asian_difference2 <- manipulate_for_feat_comparison(asian2, "ASIAN")</pre>
caucasian difference2 <- manipulate for feat comparison(caucasian2, "CAUCASIAN")
cl <- c(Clinical_features, "Stage_I", "Stage_II",</pre>
        "Stage_III", "Stage_IV", "Age", "Female", "MVI")
ml <- c(Molecular_features, "Ploidy", "Purity", "Hot")</pre>
## differences between Good and Bad subtypes.
asian_p_values <- sapply(colnames(asian_difference2)[!colnames(asian_difference2)%in%
                                                        c("subtype")],
                         function(x) calculate_p_value(asian_difference2, "subtype", x)) %>%
  as.data.frame() %>% tibble::rownames_to_column("feature") %% rename(asian_p_value=2) %>%
  mutate(categ=ifelse(feature%in%cl, "Clinical",
                      ifelse(feature%in%ml, "Molecular",
                              ifelse(feature%in%Selected_drivers,"Driver",feature)))) %>%
  group_by(categ) %>%
                        mutate(asian_p.adj=p.adjust(asian_p_value, "BH")) %>%
  as.data.frame() %>% select(-categ)
caucasian p values <- sapply(colnames(caucasian difference2)[!colnames(caucasian difference2)%in%
                                                                c("subtype")],
                             function(x) calculate_p_value(caucasian_difference2, "subtype", x)) %%
  as.data.frame() %>% tibble::rownames_to_column("feature") %>%
  rename(caucasian_p_value=2) %>%
  mutate(categ=ifelse(feature%in%cl, "Clinical",
                      ifelse(feature%in%ml, "Molecular",
                             ifelse(feature%in%Selected_drivers, "Driver", feature)))) %>%
  group_by(categ) %>%
  mutate(caucasian_p.adj=p.adjust(caucasian_p_value, "BH"))%% as.data.frame() %>%
  select(-categ)
## annotate the subtype with higher value for each feature
asian_directions_num <- (asian_difference2 %>% group_by(subtype) %>%
                           summarise_if(is_numeric,function(x) mean(x,na.rm=T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Asian=2) %>%
  mutate(Asian=ifelse(Asian==2, "G", "B"))
asian_directions_char <- (asian_difference2 %>% group_by(subtype) %>%
                            summarise_if(is.character,
                                          function(x)
                                            sum(x=="1",na.rm = T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Asian=2)%>%
  mutate(Asian=ifelse(Asian==2, "G", "B"))
```

```
caucasian_directions_num <- (caucasian_difference2 %>% group_by(subtype) %>%
                               summarise_if(is_numeric,function(x)
                                 mean(x, na.rm=T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Caucasian=2) %>%
  mutate(Caucasian=ifelse(Caucasian==2, "G", "B"))
caucasian directions char <- (caucasian difference2 %>% group by(subtype) %>%
                                summarise_if(is.character,function(x)
                                  sum(x=="1",na.rm = T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Caucasian=2)%>%
  mutate(Caucasian=ifelse(Caucasian==2, "G", "B"))
asian_directions <- rbind(asian_directions_num,asian_directions_char)
caucasian_directions <- rbind(caucasian_directions_num,caucasian_directions_char)</pre>
assoc_resultGvsB <- inner_join(asian_p_values, caucasian_p_values) %>%
  mutate(asian_p.adj=ifelse(asian_p.adj < 0.1, "sig", NA)) %>%
  mutate(caucasian_p.adj=ifelse(caucasian_p.adj < 0.1, "sig", NA)) %>%
  select(1,3,5) %>%
  inner_join(.,asian_directions) %>%
  inner join(.,caucasian directions) %>%
  mutate(Asian=ifelse(asian_p.adj=="sig",Asian,NA)) %>%
  mutate(Caucasian=ifelse(caucasian_p.adj=="sig",Caucasian,NA)) %>%
  filter(asian_p.adj=="sig"|caucasian_p.adj=="sig") %>%
  select(-c(asian_p.adj,caucasian_p.adj)) %>%
  melt(.,id.vars=c("feature"))
asian_difference4 <- manipulate_for_feat_comparison(asian4, "ASIAN") %%
  filter(subtype%in%c("G1","G2"))
caucasian_difference3 <- manipulate_for_feat_comparison(caucasian3,"CAUCASIAN") %>%
  filter(subtype%in%c("G1","G2"))
## differences between Good and Bad subtypes.
asian_p_values_2 <- sapply(colnames(asian_difference4)[!colnames(asian_difference4)%in%
                                                          c("subtype")].
                           function(x) calculate_p_value(asian_difference4, "subtype", x)) %>%
  as.data.frame() %>% tibble::rownames_to_column("feature") %>% rename(asian_p_value=2) %>%
  mutate(categ=ifelse(feature%in%cl, "Clinical",
                      ifelse(feature%in%ml, "Molecular",
                             ifelse(feature%in%Selected_drivers, "Driver", feature)))) %>%
  group_by(categ) %>%
  mutate(asian_p.adj=p.adjust(asian_p_value,"BH")) %>% as.data.frame() %>% select(-categ)
caucasian_p_values_2 <- sapply(colnames(caucasian_difference3)[!colnames(caucasian_difference3)%in%
                                                                  c("subtype")],
                               function(x)
                                 calculate_p_value(caucasian_difference3, "subtype",x)) %>%
  as.data.frame() %>% tibble::rownames_to_column("feature") %>%
  rename(caucasian_p_value=2) %>%
```

```
mutate(categ=ifelse(feature%in%cl,"Clinical",
                      ifelse(feature%in%ml, "Molecular",
                             ifelse(feature%in%Selected_drivers,"Driver",feature)))) %>%
  group_by(categ) %>%
  mutate(caucasian_p.adj=p.adjust(caucasian_p_value, "BH"))%>%
  as.data.frame() %>% select(-categ)
## annotate the subtype with higher value for each feature
asian_directions_num2 <- (asian_difference4 %>% group_by(subtype) %>%
                            summarise if(is numeric, function(x)
                              mean(x,na.rm=T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Asian=2) %>%
  mutate(Asian=ifelse(Asian==2, "G2", "G1"))
asian_directions_char2 <- (asian_difference4 %>% group_by(subtype) %>%
                             summarise_if(is.character,function(x)
                               sum(x=="1",na.rm = T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Asian=2)%>%
  mutate(Asian=ifelse(Asian==2, "G2", "G1"))
caucasian_directions_num2 <- (caucasian_difference3 %% group_by(subtype) %>%
                                summarise if(is numeric, function(x)
                                  mean(x, na.rm=T)))[1:2,] \%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Caucasian=2) %>%
  mutate(Caucasian=ifelse(Caucasian==2, "G2", "G1"))
caucasian_directions_char2 <- (caucasian_difference3 %>% group_by(subtype) %>%
                                 summarise_if(is.character,function(x)
                                   sum(x=="1",na.rm = T)))[1:2,] %>%
  apply(.,2,function(x) which.max(x)) %>% unlist() %>% as.data.frame() %>%
  tibble::rownames_to_column("feature") %>% rename(Caucasian=2)%>%
  mutate(Caucasian=ifelse(Caucasian==2, "G2", "G1"))
asian_directions2 <- rbind(asian_directions_num2,asian_directions_char2)</pre>
caucasian_directions2 <- rbind(caucasian_directions_num2, caucasian_directions_char2)</pre>
assoc_resultG1vsG2 <- inner_join(asian_p_values_2,caucasian_p_values_2) %>%
  mutate(asian_p.adj=ifelse(asian_p.adj < 0.1, "sig", NA)) %>%
  mutate(caucasian_p.adj=ifelse(caucasian_p.adj < 0.1, "sig", NA)) %>% select(1,3,5) %%
  inner_join(.,asian_directions2) %>%
  inner_join(.,caucasian_directions2) %>%
  mutate(Asian=ifelse(asian_p.adj=="sig",Asian,NA)) %>%
  mutate(Caucasian=ifelse(caucasian_p.adj=="sig",Caucasian,NA)) %>%
  filter(asian_p.adj=="sig"|caucasian_p.adj=="sig") %>%
  select(-c(asian_p.adj,caucasian_p.adj)) %>%
  melt(.,id.vars=c("feature"))
```

When we look at B1 vs B2 in Asian, B2 has higher AFP and purity as well as higher frequency AXIN1 mutations. Also B2 is colder. WNT activation because of higher fraction of AXIN1 mutations might be cause of coldness.

```
g1vsg2 <- merged_assoc %>% filter(Comparison=="G1 vs G2") %>%
  rename(G1="value") %>% mutate(G1=ifelse(G1=="G1", "high", "low")) %>%
   mutate(G2=ifelse(G1=="low","high","low")) %>% rename(cohort="variable") %>%
  melt(.,id.vars=c("feature","Comparison","cohort"))
gvsb <- merged_assoc %>% filter(Comparison=="G vs B") %>%
  rename(G="value") %>% mutate(G=ifelse(G=="G", "high", "low")) %>%
    mutate(B=ifelse(G=="low", "high", "low"))%>% rename(cohort="variable") %>%
  melt(.,id.vars=c("feature","Comparison","cohort"))
b1vsb2 <- merged_assoc %>% filter(Comparison=="B1 vs B2") %>%
  rename(B1="value") %>% mutate(B1=ifelse(B1=="B1", "high", "low")) %>%
    mutate(B2=ifelse(B1=="low", "high", "low"))%>% rename(cohort="variable") %>%
  melt(.,id.vars=c("feature","Comparison","cohort"))
merged assoc 2 <- rbind(g1vsg2,gvsb,b1vsb2) %>%
  mutate(Comparison=factor(Comparison,c("G vs B", "G1 vs G2", "B1 vs B2")),
         variable=factor(variable,c("B","G","B1","B2","G1","G2")))
compare_feat_2 <- ggplot(merged_assoc_2,aes(variable,feature,fill=value))+</pre>
  geom_tile(alpha=0.8,colour="black")+
  scale_fill_manual(na.value="gray",values = c("#d73027","#4575b4"))+
  scale_shape_manual(values=c(24,25))+
 theme(panel.background=element_rect(fill = "gray93", color = NA),
   axis.title=element_blank(),
   panel.grid=element_blank(),
   axis.text.y=element_text(colour="black"),
   axis.ticks=element_blank(),
   legend.position = "bottom",
   legend.key = element_blank(),
    strip.background = element_rect(fill=NA,color=NA),
    axis.text.x=element_text(colour="black",angle=45,hjust=1))+
  facet_wrap(~cohort+Comparison,nrow = 1,scales="free_x")+
  guides(color = guide_legend(override.aes = list(size=4)))
compare_feat_2
```



```
Fgsea_asian_BvsG <- fread("data/FGSEA/FGSEA_B_vs_G_Asian_results.tsv")
Fgsea_caucasian_BvsG <- fread("data/FGSEA//FGSEA_B_vs_G_Caucasian_results.tsv")

BvsG_asian <- Fgsea_asian_BvsG %>% mutate(Cohort="Asian")
BvsG_caucasian <- Fgsea_caucasian_BvsG %>% mutate(Cohort="Caucasian")

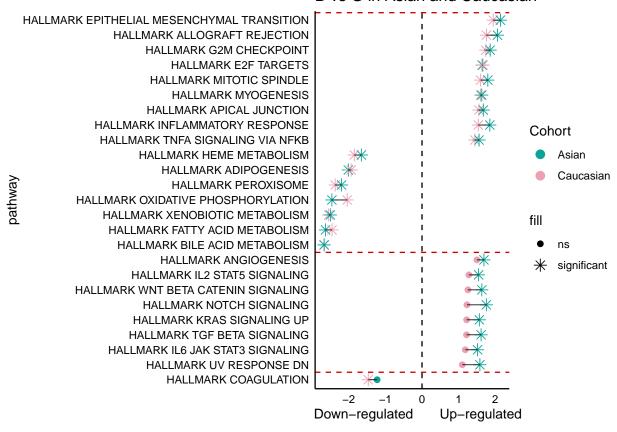
compare_DEG <- rbind(BvsG_asian,BvsG_caucasian)

group <- compare_DEG %>% group_by(pathway,Cohort) %>%
    summarise(sig=ifelse(padj<0.05,"sig","ns")) %>%
    group_by(pathway) %>% summarise(gr=toString(sig))

compare_DEG <- compare_DEG %>% left_join(.,group) %>% arrange(gr,NES,pathway) %>%
    mutate(pathway=factor(pathway,unique(.$pathway))) %>%
    mutate(fill=ifelse(padj<0.05,"significant","ns")) %>% filter(gr!="ns, ns")

DEG_1_plot <- ggplot(compare_DEG,aes(pathway,NES,col=Cohort,shape=fill))+geom_point(size=3)+</pre>
```

#### B vs G in Asian and Caucasian



```
Fgsea_asian_G1vsG2 <- fread("data/FGSEA//FGSEA_G1_vs_G2_Asian_results.tsv")
Fgsea_caucasian_G1vsG2 <- fread("data/FGSEA/FGSEA_G1_vs_G2_Caucasian_results.tsv")
G1vsG2_asian <- Fgsea_asian_G1vsG2 %>% mutate(Cohort="Asian")
G1vsG2_caucasian <- Fgsea_caucasian_G1vsG2 %>% mutate(Cohort="Caucasian")

compare_DEG_G1vsG2 <- rbind(G1vsG2_asian,G1vsG2_caucasian)
groupG1vsG2 <- compare_DEG_G1vsG2 %>% group_by(pathway,Cohort) %>%
    summarise(sig=ifelse(padj<0.05,"sig","ns")) %>%
    group_by(pathway) %>% summarise(gr=toString(sig))

compare_DEG_G1vsG2 <- compare_DEG_G1vsG2 %>% left_join(.,groupG1vsG2) %>%
```

```
arrange(gr,NES,pathway) %>%
  mutate(pathway=factor(pathway,unique(.$pathway))) %>%
  mutate(fill=ifelse(padj<0.05, "significant", "ns")) %>%
  filter(gr!="ns, ns")
DEG_2_plot <- ggplot(compare_DEG_G1vsG2,aes(pathway,NES,col=Cohort,shape=fill))+
  geom_point(size=3)+
  coord flip()+theme classic()+scale color manual(values = c(asian col,caucasian col))+
  geom_hline(yintercept = 0,lty=2)+theme(axis.ticks = element_blank(),
                                         axis.text = element text(colour="black"))+
  geom_vline(xintercept = cumsum(table(compare_DEG_G1vsG2$gr)/2 )+0.5 %>%
               as.vector(),lty=2,
             col="red3")+
  scale_shape_manual(values = c(20,8))+ggtitle("G1 vs G2 in Asian and Caucasian")+
  geom_line(aes(group = pathway),col="black",size=0.3)+
  ylab("
           Down-regulated
                               Up-regulated")
DEG_2_plot
```

### G1 vs G2 in Asian and Caucasian

```
HALLMARK TNFA SIGNALING VIA NFKB
              HALLMARK XENOBIOTIC METABOLISM
                          HALLMARK APOPTOSIS
             HALLMARK INFLAMMATORY RESPONSE
                HALLMARK ALLOGRAFT REJECTION
        HALLMARK INTERFERON GAMMA RESPONSE
  HALLMARK EPITHELIAL MESENCHYMAL TRANSITION
                                                                               Cohort
         HALLMARK INTERFERON ALPHA RESPONSE
                                                                                   Asian
                     HALLMARK MYC TARGETS V1
          HALLMARK WNT BETA CATENIN SIGNALING
                                                                                   Caucasian
oathway
                     HALLMARK MITOTIC SPINDLE
                         HALLMARK E2F TARGETS
                                                                               fill
                   HALLMARK MTORC1 SIGNALING
                     HALLMARK G2M CHECKPOINT
                                                                                   ns
                   HALLMARK SPERMATOGENESIS
                                                                                  significant
                    HALLMARK HEME METABOLISM
                HALLMARK ANDROGEN RESPONSE
                        HALLMARK COAGULATION
                       HALLMARK ADIPOGENESIS
                HALLMARK BILE ACID METABOLISM
               HALLMARK FATTY ACID METABOLISM
          HALLMARK OXIDATIVE PHOSPHORYLATION
                                                -2
                                                     -1
                                                                      2
                                              Down-regulated
                                                                 Up-regulated
```

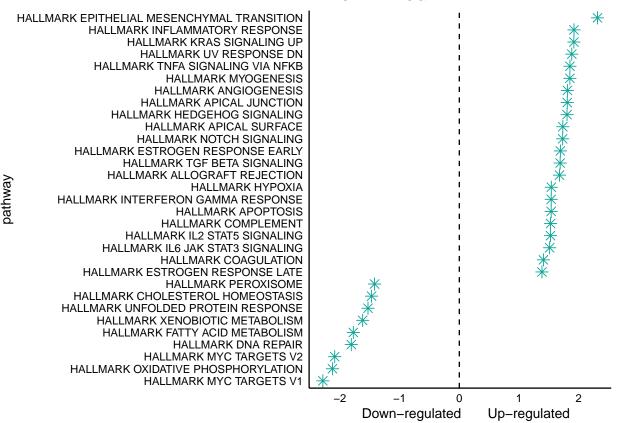
```
Fgsea_asian_B1vsB2 <- fread("data/FGSEA/FGSEA_B1_vs_B2_Asian_results.tsv")

B1vsB2_asian <- Fgsea_asian_B1vsB2 %>% mutate(Cohort="Asian")

compare_DEG_B1vsB2 <- B1vsB2_asian
```

```
groupB1vsB2 <- compare_DEG_B1vsB2 %>% group_by(pathway,Cohort) %>%
  summarise(sig=ifelse(padj<0.05, "sig", "ns")) %>%
  group by(pathway) %>% summarise(gr=toString(sig))
compare_DEG_B1vsB2 <- compare_DEG_B1vsB2 %>% left_join(.,groupB1vsB2) %>%
  arrange(gr, NES, pathway) %>%
  mutate(pathway=factor(pathway,unique(.$pathway))) %>%
  mutate(fill=ifelse(padj<0.05, "significant", "ns")) %>%
  filter(gr!="ns")
DEG_3_plot <- ggplot(compare_DEG_B1vsB2,aes(pathway,NES))+</pre>
  geom_point(size=3,col=asian_col,shape=8)+
  coord_flip()+theme_classic()+
  geom_hline(yintercept = 0,lty=2)+theme(axis.ticks = element_blank(),
                                          axis.text = element_text(colour="black"))+
  ggtitle("B1 vs B2 in Asian")+
  geom_line(aes(group = pathway),col="black",size=0.3)+
  ylab("
          Down-regulated
                                Up-regulated")
DEG_3_plot
```

#### B1 vs B2 in Asian



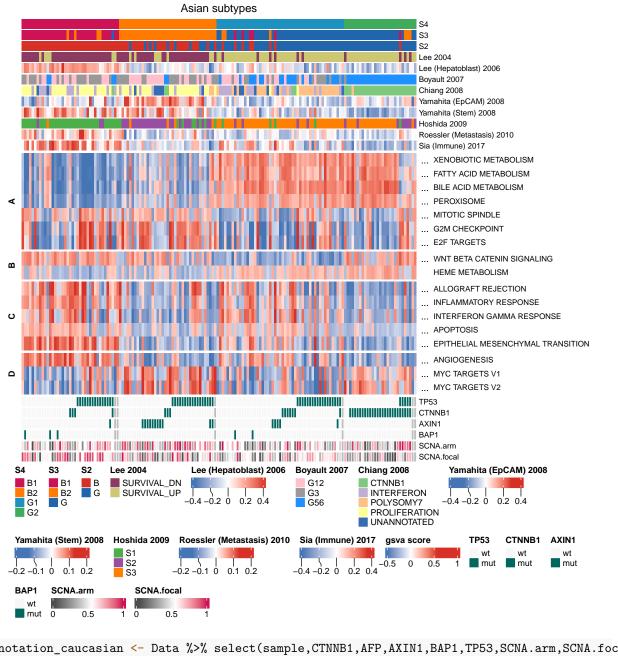
```
genesets <- gmtPathways("data/c2.cgp.v7.0.symbols.gmt")
genesets_h <- gmtPathways("data/h.all.v6.2.symbols.gmt")
names(genesets_h) <- lapply(names(genesets_h),function(x) gsub("HALLMARK_","",x)) %>% unlist
```

```
Lee genesets <- genesets[grep1("LEE",names(genesets))][c(32,33)]
Lee_hepatoblast <- genesets[grepl("LEE",names(genesets))][c(42)]
Roessler_met <- genesets[grepl("ROESSLER",names(genesets))][1]</pre>
YAMASHITA <- genesets[grep1("YAMASHITA", names(genesets))][c(3,5)]
Sia_immune_class <- fread("data/Sia_et_al_immune_class_signature.txt") %>%
    as.list()
Hoshida <- genesets[grepl("HOSHIDA", names(genesets))][5:7]</pre>
names(Hoshida) <- lapply(names(Hoshida),function(x) gsub("HOSHIDA ","",x)) %% unlist
Stemness <- c(YAMASHITA,Lee_hepatoblast)</pre>
survival <- Lee_genesets</pre>
Literature <- c(survival, Hoshida, Stemness )</pre>
normal_like_liver <- genesets_h[c(32:33,44:45)]</pre>
proliferation <- genesets_h[c(4,9,27)]</pre>
signalling <- genesets_h[c(5:7)]</pre>
immune <-c(genesets_h[c(46,31,19)])</pre>
G_vs_B <- c(normal_like_liver,proliferation)</pre>
G1 vs G2 <- c(signalling[1],genesets h[c(41)])
G1_vs_G2_and_B1_vs_B2 \leftarrow c(immune, genesets_h[c(10,30)])
B1_vs_B2 \leftarrow c(genesets_h[c(40,28,29)])
GS <- c(G_vs_B,G1_vs_G2,G1_vs_G2_and_B1_vs_B2,B1_vs_B2)
names(GS) <- lapply(names(GS), function(x) gsub("_"," ",x)) %>% unlist
mutation_color <- c("mut"="#01665e","wt"="#f7f7f7")</pre>
annotation_asian <- Data %>% select(sample,CTNNB1,AFP,AXIN1,BAP1,TP53,SCNA.arm,SCNA.focal,TMB)
es.max.asian <- GSVA::gsva(as.matrix(Asian_all_genes), GS, mx.diff=FALSE,
                            verbose=FALSE, parallel.sz=1)
subtype_df_asian <- inner_join(asian2 %>% rename(S2="subtype"),asian3 %>%
                                  rename(S3="subtype")) %>%
  inner join(.,asian4 %>% rename(S4="subtype")) %>%
  mutate(sample1=sample,sample=substr(sample,1,12)) %>%
  left_join(.,annotation_asian) %>%select(-sample) %>% rename(sample="sample1") %>%
  left_join(.,Asian_literature_Pathway$out)
subtype_df_asi <- subtype_df_asian %>% arrange(S4,TP53,CTNNB1,AXIN1)
es.max.asian <- es.max.asian[,subtype_df_asi$sample]</pre>
##color for annotations
col_funTMBasa <- circlize::colorRamp2(c(min(log2(subtype_df_asi$TMB),na.rm = T),</pre>
                                           median(log2(subtype_df_asi$TMB), na.rm = T),
                                           max(log2(subtype_df_asi$TMB), na.rm = T)),
                                         c("#01665e", "white", "#8c510a"))
col_funSCNAarmasa <- circlize::colorRamp2(c(min(subtype_df_asi$SCNA.arm,na.rm = T),</pre>
                                               median(subtype_df_asi$SCNA.arm,na.rm = T),
                                               max(subtype_df_asi$SCNA.arm,na.rm = T)),
                                             c("#4d4d4d", "white", "#ce1256"))
col_funSCNAfocalasa <- circlize::colorRamp2(c(min(subtype_df_asi$SCNA.focal,na.rm = T),</pre>
```

```
median(subtype_df_asi$SCNA.focal,na.rm = T),
                                                max(subtype_df_asi$SCNA.focal,na.rm = T)),
                                              c("#4d4d4d", "white", "#ce1256"))
col_fun_HB <- circlize::colorRamp2(c(min(subtype_df_asi$Lee_HB), 0, max(subtype_df_asi$Lee_HB)),</pre>
                                     c("#4575b4", "white", "#d73027"))
col_fun_MET <- circlize::colorRamp2(c(min(subtype_df_asi$Roessler_met), 0,</pre>
                                        max(subtype_df_asi$Roessler_met)),
                                      c("#4575b4", "white", "#d73027"))
col_fun_EPCAM <- circlize::colorRamp2(c(min(subtype_df_asi$Yamashita_EpCAM), 0,</pre>
                                          max(subtype_df_asi$Yamashita_EpCAM)),
                                        c("#4575b4", "white", "#d73027"))
col_fun_STEM <- circlize::colorRamp2(c(min(subtype_df_asi$Yamashita_Stem), 0,</pre>
                                         max(subtype_df_asi$Yamashita_Stem)),
                                       c("#4575b4", "white", "#d73027"))
col_fun_Sia <- circlize::colorRamp2(c(min(subtype_df_asi$ImmuneClassGenes), 0,</pre>
                                        max(subtype_df_asi$ImmuneClassGenes)),
                                      c("#4575b4", "white", "#d73027"))
#tick trial
Gs_vs_B_significance <- Fgsea_asian_BvsG %>% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(G_vs_B),function(x) gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
G1 vs G2 significance <- Fgsea asian G1vsG2 %>% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(G1_vs_G2),function(x) gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
G1_vs_G2__B1_vs_B2_significance <- Fgsea_asian_B1vsB2 %% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(G1_vs_G2_and_B1_vs_B2),function(x)
    gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
B1_vs_B2_significance <- Fgsea_asian_B1vsB2 %>% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(B1_vs_B2),function(x) gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
asian_row_anno_df <- rbind(Gs_vs_B_significance,G1_vs_G2_significance,
                           G1_vs_G2__B1_vs_B2_significance,B1_vs_B2_significance) %>%
  tibble::column to rownames("pathway")
asian_row_anno_df <- asian_row_anno_df[rownames(es.max.asian),]</pre>
#\u2713
asian_row_anno <- HeatmapAnnotation(significance = anno_simple(asian_row_anno_df,
                                                                col=c("significant"="white",
                                                                       "non-significant"="white"),
                                                                pt_gp = gpar(col = "black",fontsize=9),
                                                                pch =ifelse(asian_row_anno_df=="signific")
                                                                             "\u2713","")),
                                       which = "row", show_annotation_name = F,
                                       annotation_name_gp = gpar(fontsize = 9))
```

```
#top annot
annot top asi <- HeatmapAnnotation(S4=subtype df asi$S4,
                        S3=subtype df asi$S3,
                        S2=subtype_df_asi$S2,
                        `Lee 2004`=subtype_df_asi$Lee,
                       `Lee (Hepatoblast) 2006`=subtype_df_asi$Lee_HB,
                       `Boyault 2007`=subtype df asi$Boyault,
                       `Chiang 2008`=subtype_df_asi$Chiang,
                       Yamahita (EpCAM) 2008 = subtype_df_asi$Yamashita_EpCAM,
                       Yamahita (Stem) 2008 = subtype_df_asi$Yamashita_Stem,
                       `Hoshida 2009`=subtype_df_asi$Hoshida,
                       `Roessler (Metastasis) 2010`=subtype_df_asi$Roessler_met,
                       `Sia (Immune) 2017`=subtype_df_asi$ImmuneClassGenes,
                       col = list(`Lee 2004`=c("SURVIVAL_DN"="hotpink4",
                                                "SURVIVAL_UP"="khaki3"),
                              `Hoshida 2009`=c("S1"=brewer.pal(8, "Set1")[3],
                                               "S2"=brewer.pal(8, "Set1")[4],
                                               "S3"=brewer.pal(8, "Set1")[5]),
                              Chiang 2008 = c("CTNNB1"=brewer.pal(5, "Accent")[1],
                                              "INTERFERON"=brewer.pal(5, "Accent")[2],
                                            "POLYSOMY7"=brewer.pal(5, "Accent")[3],
                                            "PROLIFERATION"=brewer.pal(5, "Accent")[4],
                                            "UNANNOTATED"=brewer.pal(5, "Accent")[5]),
                                   Boyault 2007 = c("G12"="pink", "G3"="#999999",
                                                    "G56"="dodgerblue"),
                                   `Lee (Hepatoblast) 2006`=col_fun_HB,
                              `Sia (Immune) 2017`=col_fun_Sia,
                              `Roessler (Metastasis) 2010`=col_fun_MET,
                              Yamahita (EpCAM) 2008 = col_fun_EPCAM,
                              Yamahita (Stem) 2008 = col_fun_STEM,
                             S4=subtypes_col_pal,
                                  S3=subtypes_col_pal,
                                  S2=subtypes_col_pal),
                       annotation_name_gp = gpar(fontsize=9),
                       gap = unit(0.5, "mm"),
                       show annotation name = T,
                       border=F,
                       show_legend = T,
                       annotation name side = "right",
                       simple_anno_size = unit(4, "mm"),
                       annotation_legend_param = list(direction = "horizontal",
                                                       legend_height = unit(2, "mm"),
                                                       legend_width=unit(30,"mm")))
annot_asa_bottom <- HeatmapAnnotation( TP53=subtype_df_asi$TP53,</pre>
                        CTNNB1=subtype_df_asi$CTNNB1,
                        AXIN1=subtype_df_asi$AXIN1,
                        BAP1=subtype_df_asi$BAP1,
                       # TMB=subtype_df_asi$TMB,
                        SCNA.arm=subtype_df_asi$SCNA.arm,
```

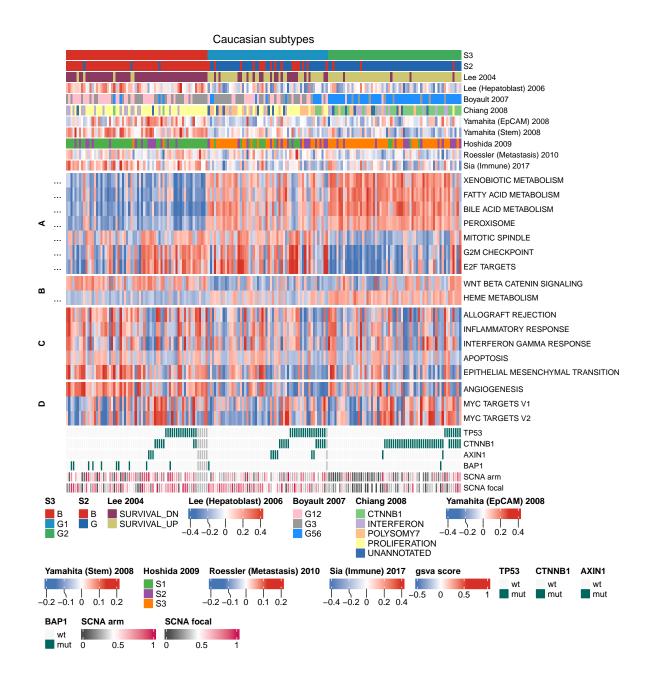
```
SCNA.focal=subtype_df_asi$SCNA.focal,
                         col=list(CTNNB1=mutation_color,
                                  BAP1=mutation_color,
                                  AXIN1=mutation_color,
                                  TP53=mutation_color,
                                  SCNA.arm=col funSCNAarmasa,
                                  SCNA.focal=col_funSCNAfocalasa),
                        gap = unit(0.5, "mm"),
                        border=F,
                        annotation_name_gp = gpar(fontsize=9),
                        gp = gpar(col = "white"),
                        simple_anno_size = unit(4, "mm"),
                       annotation legend param = list(direction = "horizontal",
                                                        legend_height = unit(2, "mm"),
                                                        legend_width=unit(30,"mm")))
col_fun <- circlize::colorRamp2(c(min(es.max.asian)/1.2, 0,</pre>
                                    \max(\text{es.max.asian}))/1.2, c("#4575b4", "white", "#d73027"))
hm <- Heatmap(es.max.asian, show_column_dend = F,
              show_row_dend = F,
              cluster_rows = F,
              row_names_gp = gpar(fontsize=9),
              cluster columns = F,
              show column names = F,
              right_annotation = asian_row_anno,
              top_annotation = annot_top_asi,
              bottom_annotation = annot_asa_bottom,
              row_split = c(rep("A",length(G_vs_B)),
                           rep("B",length(G1_vs_G2)),
                            rep("C",length(G1_vs_G2_and_B1_vs_B2)),
                           rep("D",length(B1_vs_B2))),
             row_names_side = "right",
             col=col_fun,
             heatmap_legend_param = list(direction = "horizontal",legend_width=unit(30,'mm')),
             #heatmap_height = unit(20, "cm"),
             row_title_gp = gpar(fontsize=10, font = 2, side="right "),
             name = "gsva score",
             column_title = "Asian subtypes")
plt <- draw(hm, padding = unit(c(2,10, 2, 10), "mm"), merge_legend=T, heatmap_legend_side="bottom")</pre>
```



```
##color for annotations
col funSCNAarmcau <- circlize::colorRamp2(c(min(subtype df cau$SCNA.arm, na.rm = T),
                                              median(subtype df cau$SCNA.arm,na.rm = T),
                                              max(subtype df cau$SCNA.arm, na.rm = T)),
                                            c("#4d4d4d", "white", "#ce1256"))
col_funSCNAfocalcau <- circlize::colorRamp2(c(min(subtype_df_cau$SCNA.focal,na.rm = T),</pre>
                                                median(subtype_df_cau$SCNA.focal,na.rm = T),
                                                max(subtype df cau$SCNA.focal,na.rm = T)),
                                              c("#4d4d4d", "white", "#ce1256"))
col_fun_HBcau <- circlize::colorRamp2(c(min(subtype_df_cau$Lee_HB), 0,</pre>
                                          max(subtype_df_cau$Lee_HB)),
                                        c("#4575b4", "white", "#d73027"))
col_fun_METcau <- circlize::colorRamp2(c(min(subtype_df_cau$Roessler_met), 0,</pre>
                                           max(subtype_df_cau$Roessler_met)),
                                         c("#4575b4", "white", "#d73027"))
col_fun_EPCAMcau <- circlize::colorRamp2(c(min(subtype_df_cau$Yamashita_EpCAM), 0,</pre>
                                             max(subtype df cau$Yamashita EpCAM)),
                                           c("#4575b4", "white", "#d73027"))
col_fun_STEMcau <- circlize::colorRamp2(c(min(subtype_df_cau$Yamashita_Stem), 0,</pre>
                                            max(subtype df cau$Yamashita Stem)),
                                          c("#4575b4", "white", "#d73027"))
col_fun_Siacau <- circlize::colorRamp2(c(min(subtype_df_cau$ImmuneClassGenes), 0,</pre>
                                           max(subtype df cau$ImmuneClassGenes)),
                                         c("#4575b4", "white", "#d73027"))
#tick trial
Gs_vs_B_significancecau <- Fgsea_caucasian_BvsG %>% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(G_vs_B),function(x) gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
G1_vs_G2_significancecau <- Fgsea_caucasian_G1vsG2 %>% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(G1_vs_G2),function(x) gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
G1_vs_G2__B1_vs_B2_significancecau <- Fgsea_caucasian_G1vsG2 %>% as.data.frame() %>%
  mutate(pathway=gsub("HALLMARK ","",pathway)) %>%
  filter(pathway%in% lapply(names(G1_vs_G2_and_B1_vs_B2),function(x)
    gsub("_"," ",x)) %>% unlist) %>%
  select(1,8)
caucasian_row_anno_df <- rbind(Gs_vs_B_significancecau,G1_vs_G2_significancecau,
                               G1_vs_G2__B1_vs_B2_significancecau) %>%
  rbind(.,data.frame(pathway=setdiff(rownames(es.max.caucasian),.$pathway),
                     fill="non-significant")) %>%
  tibble::column_to_rownames("pathway")
caucasian row anno df <- caucasian row anno df [rownames(es.max.caucasian),]
#\u2713
```

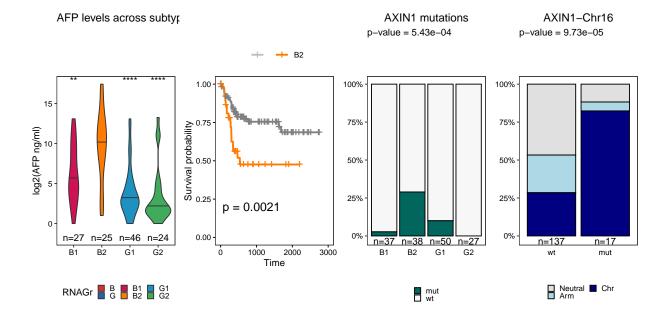
```
caucasian_row_anno <- HeatmapAnnotation(significance = anno_simple(caucasian_row_anno_df,</pre>
                                                                     col=c("significant"="white",
                                                                           "non-significant"="white"),
                                                                     pt_gp = gpar(col = "black",fontsize=
                                                                     pch =ifelse(caucasian_row_anno_df=="
                                                                                 "\u2713","")),
                                       which = "row", show_annotation_name = F,
                                       annotation name gp = gpar(fontsize = 9))
annot_top_cau <- HeatmapAnnotation( S3=subtype_df_cau$S3,</pre>
                        S2=subtype_df_cau$S2,
                         Lee 2004 = subtype_df_cau$Lee,
                       `Lee (Hepatoblast) 2006`=subtype_df_cau$Lee_HB,
                       `Boyault 2007`=subtype_df_cau$Boyault,
                       `Chiang 2008`=subtype_df_cau$Chiang,
                       `Yamahita (EpCAM) 2008`=subtype_df_cau$Yamashita_EpCAM,
                       `Yamahita (Stem) 2008`=subtype_df_cau$Yamashita_Stem,
                       `Hoshida 2009`=subtype_df_cau$Hoshida,
                       `Roessler (Metastasis) 2010`=subtype_df_cau$Roessler_met,
                       `Sia (Immune) 2017`=subtype_df_cau$ImmuneClassGenes,
                       col = list(`Lee 2004`=c("SURVIVAL_DN"="hotpink4",
                                                "SURVIVAL_UP"="khaki3"),
                              `Hoshida 2009`=c("S1"=brewer.pal(8,"Set1")[3],
                                               "S2"=brewer.pal(8, "Set1")[4],
                                               "S3"=brewer.pal(8, "Set1")[5]),
                              Chiang 2008 = c("CTNNB1"=brewer.pal(5, "Accent")[1],
                                              "INTERFERON"=brewer.pal(5, "Accent")[2],
                                            "POLYSOMY7"=brewer.pal(5, "Accent")[3],
                                            "PROLIFERATION"=brewer.pal(5, "Accent")[4],
                                            "UNANNOTATED"=brewer.pal(5, "Accent")[5]),
                                   Boyault 2007 = c("G12"="pink", "G3"="#999999",
                                                    "G56"="dodgerblue"),
                                   `Lee (Hepatoblast) 2006`=col_fun_HBcau,
                              `Sia (Immune) 2017`=col_fun_Siacau,
                              `Roessler (Metastasis) 2010`=col_fun_METcau,
                              Yamahita (EpCAM) 2008 = col_fun_EPCAMcau,
                              Yamahita (Stem) 2008 = col_fun_STEMcau,
                                  S3=subtypes_col_pal,
                                  S2=subtypes_col_pal),
                       annotation_name_gp = gpar(fontsize=9),
                       gap = unit(0.5, "mm"),
                       show annotation name = T,
                       border=F,
                       show_legend = T,
                       annotation_name_side = "right",
                       simple_anno_size = unit(4, "mm"),
                       annotation_legend_param = list(direction = "horizontal",
                                                        legend_height = unit(2, "mm"),
                                                        legend_width=unit(30,"mm")))
annot_cau_bottom <- HeatmapAnnotation( TP53=subtype_df_cau$TP53,</pre>
```

```
CTNNB1=subtype_df_cau$CTNNB1,
                        AXIN1=subtype_df_cau$AXIN1,
                        BAP1=subtype_df_cau$BAP1,
                        # TMB=subtype_df_asi$TMB,
                         `SCNA arm`=subtype_df_cau$SCNA.arm,
                        `SCNA focal`=subtype_df_cau$SCNA.focal,
                        col=list(CTNNB1=mutation color,
                                 BAP1=mutation color,
                                 AXIN1=mutation color,
                                 TP53=mutation_color,
                                  `SCNA arm`=col_funSCNAarmcau,
                                  `SCNA focal`=col_funSCNAfocalcau),
                       gap = unit(0.5, "mm"),
                       border=F.
                       annotation_name_gp = gpar(fontsize=9),
                       gp = gpar(col = "white"),
                       simple_anno_size = unit(4, "mm"),
                       annotation_legend_param = list(direction = "horizontal",
                                                       legend_height = unit(2, "mm"),
                                                       legend_width=unit(30,"mm")))
col_fun_cau <- circlize::colorRamp2(c(min(es.max.caucasian)/1.2, 0,</pre>
                                        max(es.max.caucasian))/1.2,
                                      c("#4575b4", "white", "#d73027"))
hm_cau <- Heatmap(es.max.caucasian, show_column_dend = F,</pre>
              show_row_dend = F,
              cluster_rows = F,
              row_names_gp = gpar(fontsize=9),
              cluster_columns = F,
              show_column_names = F,
              left_annotation = caucasian_row_anno,
              top_annotation = annot_top_cau,
              bottom_annotation = annot_cau_bottom,
              row_split = c(rep("A",length(G_vs_B)),
                           rep("B",length(G1_vs_G2)),
                           rep("C",length(G1_vs_G2_and_B1_vs_B2)),
                           rep("D",length(B1_vs_B2))),
             row_names_side = "right",
             col=col_fun,
             heatmap_legend_param = list(direction = "horizontal",
                                          legend width=unit(30,'mm')),
             #heatmap_height = unit(20, "cm"),
             row_title_gp = gpar(fontsize=10, font = 2,side="right "),
             name = "gsva score",
             column_title = "Caucasian subtypes")
plt_cau <- draw(hm_cau, padding = unit(c(2,10, 2, 10), "mm"),merge_legend=T,</pre>
                heatmap_legend_side="bottom")
```



### FOCUSING OB THE B2 Subtype

```
mutate_if(grepl("mut",.)==TRUE,function(x) factor(x,c("wt","mut"))) %>%
  mutate(DEL_Chr16=ifelse(DEL_16p=="DEL" & DEL_16q=="DEL", "Chr",
                      ifelse(DEL_16p=="DEL" & DEL_16q=="No-DEL","Arm",
                             ifelse(DEL_16p=="No-DEL" & DEL_16q=="DEL","Arm","Neutral"))))
#%>% mutate(DEL_Chr16=factor(DEL_Chr16,c("No-DEL","DEL")))
AFP <- ggplot(DataforB2 %>% filter(race=="ASIAN",!is.na(RNAGr)) %>%
                mutate(AFP=log2(AFP)),aes(RNAGr,AFP,fill=RNAGr))+
  geom violin(draw quantiles = c(0.5))+scale fill manual(values=subtypes col pal)+
  stat_compare_means(ref.group = "B2",method = "wilcox.test",label="p.signif")+
  xlab(NULL)+ylab("log2(AFP ng/ml)")+theme_bw()+
  stat_n_text()+theme(legend.key.size = unit(3,"mm"),
                      axis.text = element_text(colour="black"),
                      panel.grid = element_blank(),
                      legend.position = "bottom",
                      legend.direction = "horizontal")+
  ggtitle("AFP levels across subtypes")
Data_surv <- DataforB2 %>% filter(race=="ASIAN") %>% filter(!is.na(RNAGr)) %>%
  left_join(Data.survival) %>%
  mutate(B2_vs_rest=ifelse(RNAGr=="B2","B2","B1_G1_G2"))
survp <- survPlot(Data_surv, Time = "OS.time",</pre>
                  Event = "OS", var = "B2 vs rest",
                  risktable = T,palette = c("gray",subtypes_col_pal["B2"]))
mutation_color <- c("mut"="#01665e","wt"="#f7f7f7")</pre>
AXIN1 <- stacked_bar(DataforB2 %>% filter(race=="ASIAN"), "RNAGr", "AXIN1",
                     col = mutation_color,title="AXIN1 mutations")+
 theme(legend.direction = 1)
#WNT <- stacked_bar(Data %>% filter(race=="ASIAN"), "RNAGr", "WNT_",col = mutation_color,title="AXIN1 mu
Chr16_AXIN1 <- stacked_bar(DataforB2 %>% filter(race=="ASIAN") %>%
                             mutate(DEL_Chr16=factor(DEL_Chr16,
                                                      c("Neutral","Arm","Chr"))),
                           "AXIN1", "DEL_Chr16", col = c("gray88", "lightblue", "navy"),
                           title="AXIN1-Chr16")+
  theme(legend.direction =2)
AFP+survp$plot+AXIN1+Chr16_AXIN1+plot_layout(ncol=4)
```



#### Finding B2 specific genes (DEG B2 vs rest)

```
#
# Asian_raw <-fread("data/Asian_158_raw.tsv") %>%
    tibble::column to rownames("V1")
#
# colnames(Asian raw) <- substr(colnames(Asian raw),1,12)</pre>
# #
# AsianB2 <- Asian_raw[,Data %>% filter(race=="ASIAN",RNAGr=="B2") %>% pull(sample)]
# AsianRest <- Asian_raw[,Data %>% filter(race=="ASIAN",RNAGr!="B2") %>% pull(sample)]
# #
# Asian_B2vsRest <- DESeq2_DEG(AsianB2, AsianRest, "B2", "B1_G1_G2") %>% as.data.frame()
#write.table(Asian_B2vsRest, "data/asian_B2vsRestDEG.tsv", sep="\t", quote = F)
B2_vs_rest_DEG <- read.table("data/asian_B2vsRestDEG.tsv")</pre>
getSignaturesTop <- function(DEGoutput, n=10){</pre>
  return(DEGoutput %>% tibble::rownames_to_column("gene") %>%
           filter(log2FoldChange > 1 & padj <0.05) %>%
           arrange(padj) %>% head(n) %>% select(1) %>% pull(1))
}
## these are top 100 upregulated genes in the B2 subtype compared to the rest in Asian cohort
Top 100 up inB2 <- getSignaturesTop(B2 vs rest DEG, 100)
Top_100_up_inB2
     [1] "DUSP9"
                     "IGSF1"
                                "ARID3A"
                                            "MTMR7"
                                                       "EPO"
                                                                   "HIC2"
##
                                                                   "CD7"
##
     [7] "CYP19A1"
                     "AFP"
                                "GDPD3"
                                            "ACVR2B"
                                                       "PNCK"
##
    [13] "TNNI2"
                     "SLC1A7"
                                "CXCL17"
                                            "CSF3R"
                                                       "RPS7"
                                                                   "NREP"
##
    [19] "RPL9"
                     "NAALADL1" "HAVCR1"
                                            "PRDM15"
                                                       "LGI4"
                                                                   "PPARG"
##
    [25] "BEND3"
                     "JAML"
                                "RPSA"
                                            "C19orf48" "FLVCR1"
                                                                   "NR6A1"
```

```
[31] "RPS5"
                    "HDAC11"
                                "PIGZ"
                                           "RPL28"
                                                      "GLUD2"
##
                                                                  "NDRG1"
##
   [37] "PAQR9"
                    "RPS19"
                                "RPL32"
                                           "SFI1"
                                                      "BMF"
                                                                  "RPL18"
   [43] "RPS24"
                                                                  "SKP2"
##
                    "PDE9A"
                                "VSIG1"
                                           "OVGP1"
                                                      "MYCN"
   [49] "RPL13A"
                    "PACSIN1"
                                "RPL14"
                                           "TTLL4"
                                                      "ARHGEF2"
                                                                  "H2AFX"
##
##
   [55] "IGF2BP2"
                    "CDKN1C"
                                "TRNP1"
                                           "RPL36"
                                                      "TNNC1"
                                                                  "SPRN"
##
  [61] "PLA2G7"
                    "RPL8"
                                "SCML2"
                                           "CHST13"
                                                      "PNMA3"
                                                                  "B3GALT2"
  [67] "NTS"
                    "FABP3"
                                "RPS12"
                                           "OSBP2"
                                                      "MEP1A"
                                                                  "SSTR2"
##
   [73] "KISS1"
                    "H2AFY2"
                                "PEG3"
                                           "SSUH2"
                                                      "TSPAN7"
                                                                  "PSRC1"
##
##
    [79] "SULT1C4"
                    "GPC5"
                                "TMEM86B"
                                           "MSI1"
                                                      "KCTD17"
                                                                  "RPL36A"
##
   [85] "MARCKSL1" "PTHLH"
                                "SLC29A4"
                                           "CDC25A"
                                                      "UCHL1"
                                                                  "BLM"
   [91] "CYP26B1"
                    "TRIM71"
                                "PLCXD1"
                                           "LDOC1"
                                                      "S100A8"
                                                                  "PEG10"
   [97] "MYBL2"
                    "NPW"
                                "NEURL1"
                                           "LRRC1"
##
```

Now let's plot the expression of these top genes across patients.

