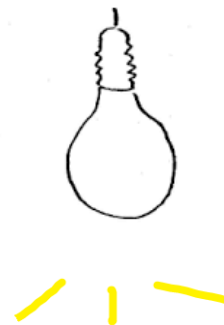
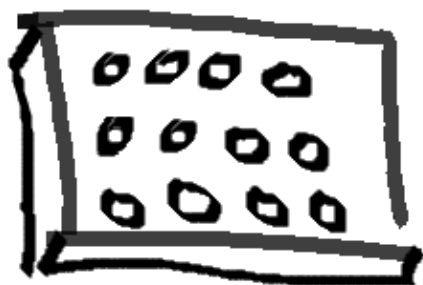
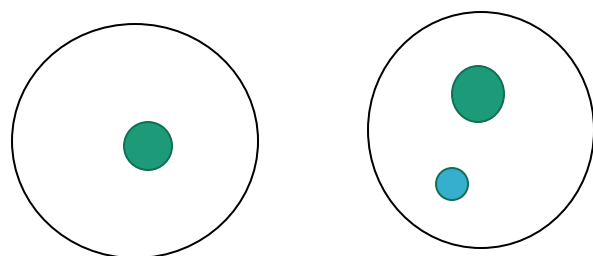
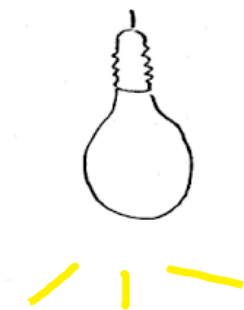


Using Species-Specific and Broad Range
Primers in PCR to Identify Edaphic Strains
of *Chlamydomonas reinhardtii*, *C. incerta*,
and Related Species

ISOLATION OF FOUR NEW STRAINS OF *CHLAMYDOMONAS REINHARDTII*
(CHLOROPHYTA) FROM SOIL SAMPLES¹

*Lawren Sack, Clifford Zeyl,² Graham Bell, Timothy Sharbel, Xavier Reboud,
Torsten Bernhardt, and Hans Koelewyn*

Department of Biology, McGill University, 1205 Docteur Penfield, Montréal, Québec, Canada H3A 1B1

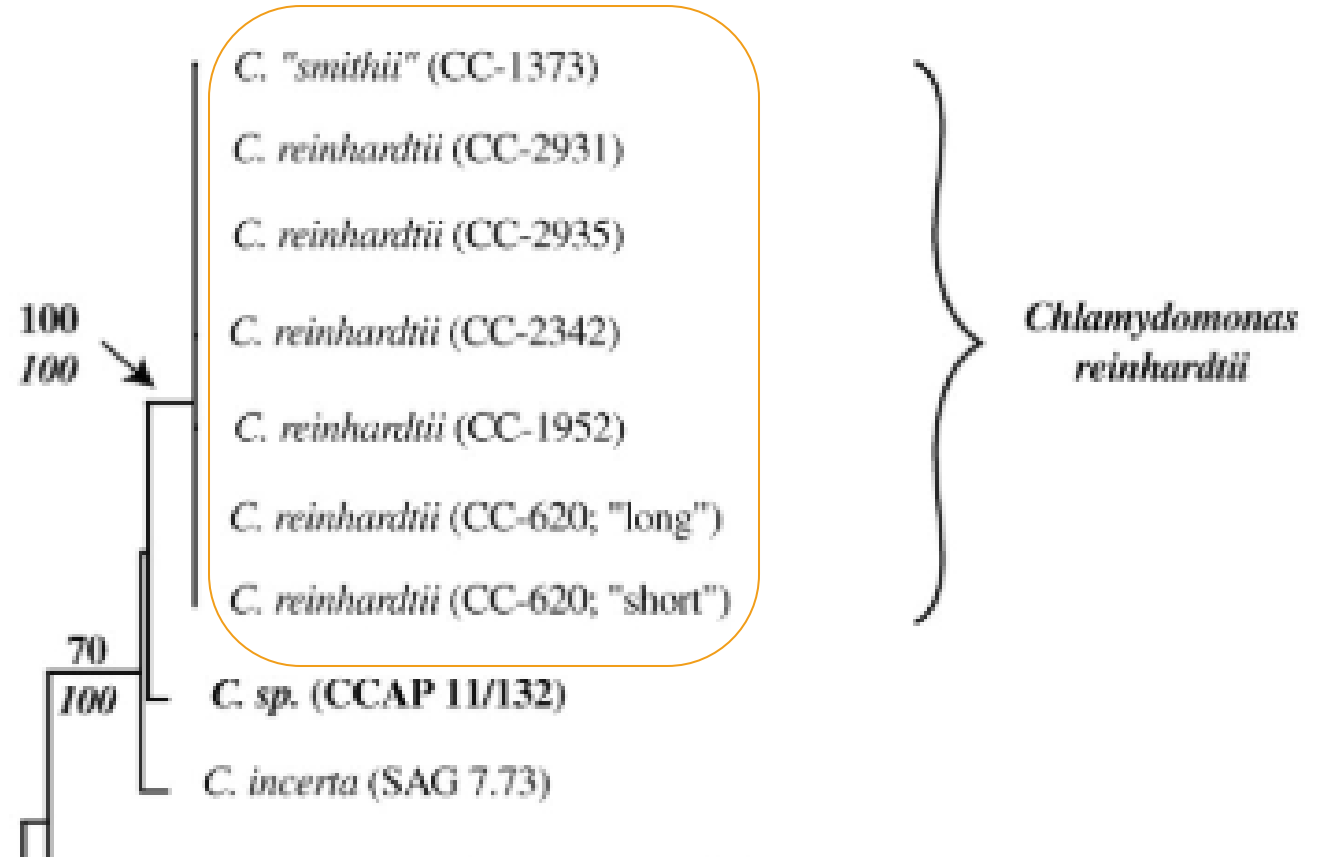


- M. Wilson, Kim & Schembri, Mark & D. Baker, Peter & Saint, Christopher. (2000). *Molecular Characterization of the Toxic Cyanobacterium Cylindrospermopsis raciborskii and Design of a Species-Specific PCR*. *Applied and Environmental Microbiology*. 66. 10.1128/AEM.66.1.332-338.2000.
- D Atkins, Simon & Clark, Ian & Pande, Sonal & Hirsch, Penny & R Kerry, Brian. (2005). The use of real-time PCR and species-specific primers for the identification and monitoring of *Paecilomyces lilacinus*. *FEMS microbiology ecology*. 51. 257-64. 10.1016/j.femsec.2004.09.002.
- Bérard, Annette & Dorigo, U & Humbert, Jean-François & Martin-Laurent, Fabrice. (2005). Microalgae community structure analysis based on 18S rDNA amplification from DNA extracted directly from soil as a potential soil bioindicator. *Agronomy for Sustainable Development - AGRON SUSTAIN DEV*. 25. 285-291. 10.1051/agro:2005004.

Primer Design

C.reinhardtii-Specific Primers

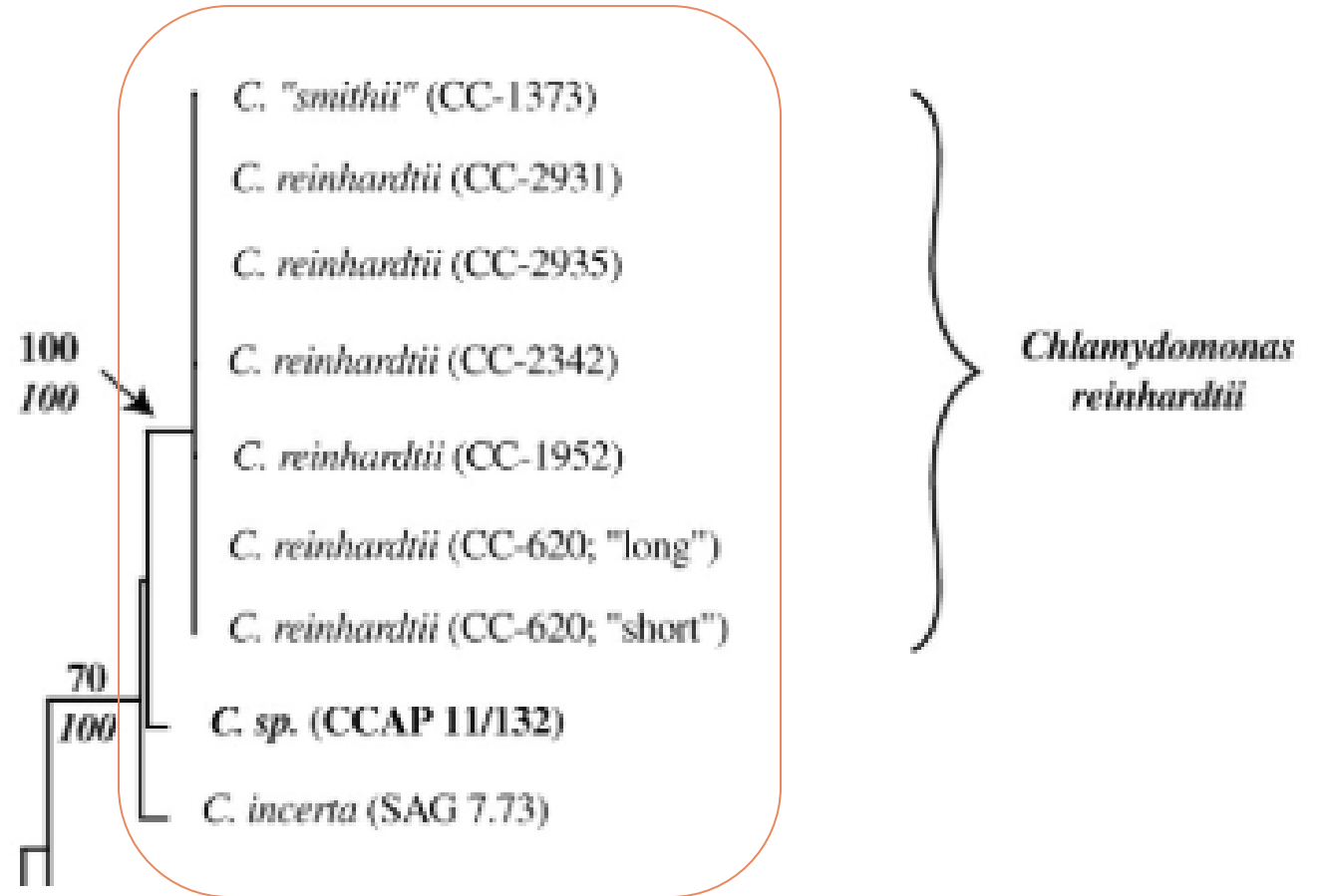
B



Broad-Range Primers

Ask rory for less prehis
tric figure?

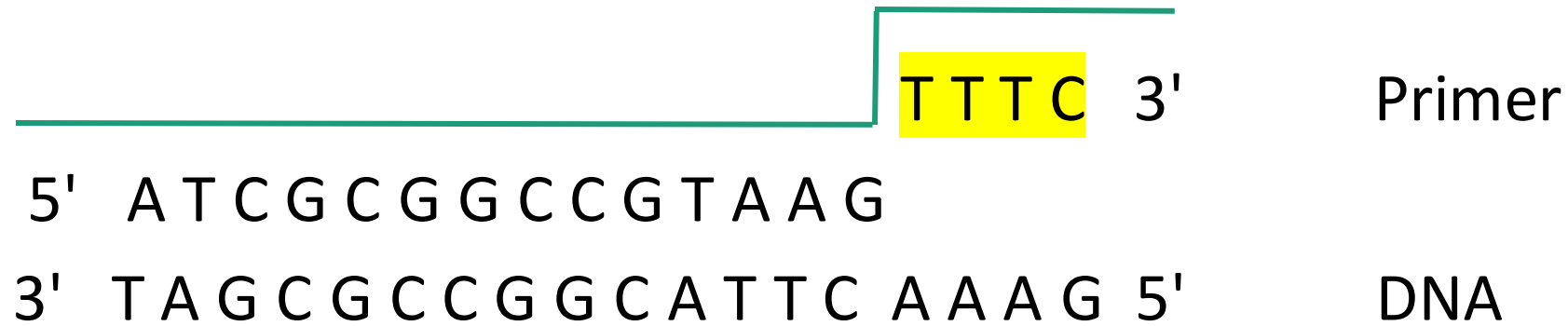
B



Portrait of a Species: *Chlamydomonas reinhardtii*,
Thomas Pröschold, Elizabeth H. Harris and Annette W. Coleman
GENETICS August 1, 2005 vol. 170 no. 4 1601-1610; <https://doi.org/10.1534/genetics.105.044503>

A Primer on Good Primers

- | | | | | | | | | | | | | | | | | | | | | |
|----|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|----------|----------|----------|----|-----------------|
| 5' | A | T | C | G | C | G | G | C | C | G | T | A | A | G | T | G | G | C | 3' | Primer Sequence |
| 3' | T | A | G | C | G | C | C | G | G | C | A | A | T | C | A | C | C | G | 5' | DNA |



A Primer on Good Primers

A) Primer self annealing



B) Primer-primer annealing



C) Primer-dimer formation



A Primer on Good Primers

- T_m of both primers close enough to each other
- GC content

Primer3

Primer3Plus

pick primers from a DNA sequence

[More...](#)[Source Code](#)[Help](#)[About](#)[Load server settings:](#)

Default

[Activate Settings](#)

Select primer pairs to detect the given template sequence. Optionally targets and included/excluded regions can be specified.

[Task:](#)

generic

[Pick Primers](#)[Reset Form](#)[Main](#)[General Settings](#)[Advanced Settings](#)[Internal Oligo](#)[Penalty Weights](#)[Advanced Sequence](#)[Sequence Id:](#)[Paste template sequence below](#)

Or upload sequence file:

[Choose File](#)

No file chosen

[Upload File](#)

Mark selected region:

< >

[]

{ }

[Clear](#)[Save Sequence](#)[Excluded Regions:](#)

<

>

[Targets:](#)

[

]

[Included Region:](#)

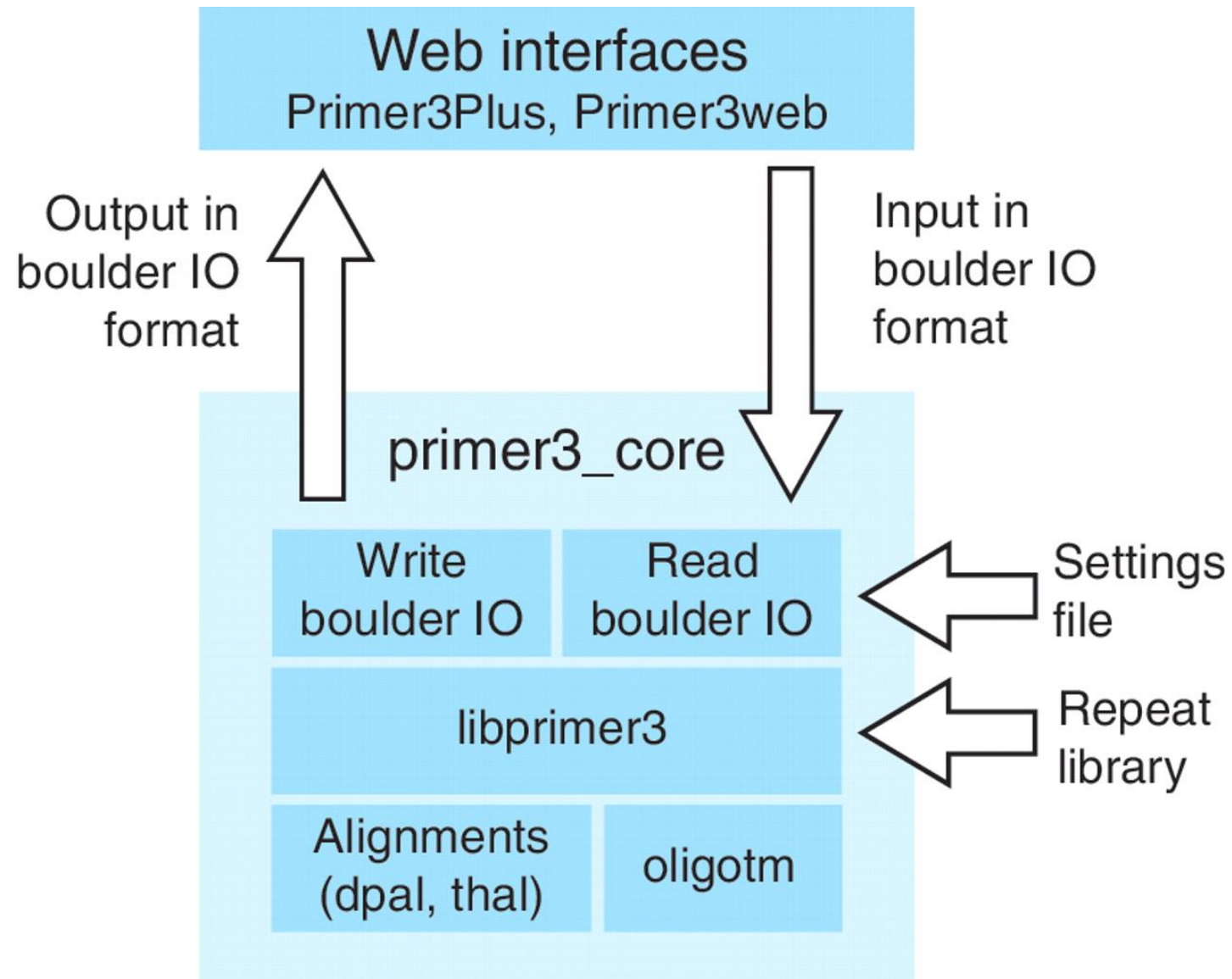
{

}

[Primer overlap positions:](#)

-

[Pair OK Region List:](#)☒ [Pick left primer](#)☐ [Pick hybridization probe](#)☒ [Pick right primer](#)or use [left primer](#) below.(internal oligo) or use [oligo](#) below.or use [right primer](#)



- Andreas Untergasser, Ioana Cutcutache, Triinu Koressaar, Jian Ye, Brant C. Faircloth, Mado Remm, Steven G. Rozen; Primer3—new capabilities and interfaces, *Nucleic Acids Research*, Volume 40, Issue 15, 1 August 2012, Pages e115, <https://doi.org/10.1093/nar/gks596>

```
1 PRIMER_MISPRIMING_LIBRARY=incerta_mispriming_library.fasta
2 PRIMER_OPT_SIZE=22
3 PRIMER_MAX_LIBRARY_MISPRIMING=16.00
4 PRIMER_MAX_TEMPLATE_MISPRIMING=12
5 PRIMER_LIB_AMBIGUITY_CODES_CONSENSUS=0
6 PRIMER_INTERNAL_MIN_GC=50
7 PRIMER_TASK=generic
8 PRIMER_PICK_LEFT_PRIMER=1
9 PRIMER_PICK_INTERNAL_OLIGO=0
10 PRIMER_PICK_RIGHT_PRIMER=1
11 PRIMER_PRODUCT_SIZE_RANGE=250-900
12 PRIMER_MAX_SIZE=22
13 PRIMER_GC_CLAMP=3
14 PRIMER_MAX_END_GC=3
15 PRIMER_THERMODYNAMIC_PARAMETERS_PATH=/scratch/research/tmp_apps/primer3-2.3.7/src/primer3_config/
16 SEQUENCE_ID=g2524.t1|PACid:26899432
17 SEQUENCE_TEMPLATE=ATGGCGAGTTTGCAGTCCGCGGGTGCCCCGGGCAGCGTCCCTCCCCTCCTGGATCCCAGCAAGCTAAGCATTTGGACGAGTGCTTGGCCGGGGCGCCTTTGCCACAGTGTAC
CTTGCAACTTACGAAGGCATCGGCCCAGCAGCGCTCAAGGTTGCCGTCCCCGCCTCTAAGCGCCTACCGCCTGTGTTTGAAAGACTGTTTCATGAACGAGGTTTCCCTCCTTTGTCGCCACAGGCACAGG
CGCCTGGTGAAATGCTATGGGCTTCTAAGGCTGCCGCCGGGGTGGCTGCCTGGAATCCAAGCTCAGGCAGCTCCGATACCCGCATGCCTGCTGGAGCTGTGTGAGGGTGAGACGCTGCGGGATGTGGTA
CTACACGCCAAGGCCGGCCGCCGTCCCTACAGCGACGCCACAGCGCTGTCGCTGCTGCGTGACGTGACAGAGGCGCTGGCGGCGTTGCACGCGGCGTCGCCGCCGCCATTCACCGCGACGTGAAAGCT
GAGAACATTCTGCTCAAGCGCGAGGGACTTACGCCGGCAGCAGCGCGCGGCGGTGGCTGCGCCGCGTCATCGCTGCCGGCGGTGGCATCTGGAGGTGGCGAGGGGCAGGAGCAGCCGTACCAGCAG
CTGACGGCGCGGGTGGCGGATCTGGGTCTGCACGTGCGGGTGGAGCAGCATCGCTCCGTGATGCTGCGACGCCGCGGCGCGGCCTCGCTGGACGGCTCCGCCTTTGTGGCCATCTCGCGCCACAACCTCC
```

18 =
19 SEQUENCE_ID=g6136.t1|PACid:26892997
20 SEQUENCE_TEMPLATE=ATGAAGCGCCCTGACACCTTGGGTGACAGGATATGGCGGTCAGTCGATAAGAGCGTGGACCAGAGGTACCGCGGCGTGCTGCTGGTGGTGTCCATCCCGGTAATTCTAATC
CTGGCGGTCATCACGCTGGTGCCACGTTCAACGCCCTTCCACGCGGAGCGCACCCACACGCTGAAGGAGCTGCGCCATACAGGCGCGGCGCCAAAGTTCGCGATTGTGTTTGATGCGGGCAGCACAGGC
AGCCGCATCCACGTGTTCAAGTTTGAGCAGGCCGGCGGAGAGCTCAAGCTCATTAGCGACACCTTTGAGCAGCTGAAGCCCGGGTTGAGCTCCTTCGCGGACTCCCCGAGAAGGCGGCCGCTCCCTG
CAGCCACTAATTGACACCGCGCTGAAGACGGTGCCCCAGAACTTGACAGCTCCACTCCCATCTCCCTCAAAGCCACGGCCGGCCTGCGCCTGCTGCCCCGGTGATAAGGCTGACAATATCCTGAAGGCC
GTGGAGGCGCTGCTGCGGAAGCAGCCCTTTAAGCTTGCCCCCTGGGGGCGTGGCCATCATGGACGGCAAGGACGAGGGTTCGCTTCGCTGGCTGACGCTCAACTACCTCCTTGCCCGCCTGTCTGGGCGG
GTGGGCAAGACCGTGGCTGCCATTGACATGGGTGGCGGCTCGATTGAGGAGCGTTTGCGCTGGAGGACACGAATGCAAACTGGCGCCACGGACTACGTTGTTTACGCTGCACGGAGGCGGCAAGACC
TTCAACGTGTACGTGCACTCGTACCTGGGCTACGGCCTCATGGCCGGCCGCGCCAAGCTGATTGATGCCGCGGAGAAGGGCAAGCCGCACAGCTGCATCCATGAGGGCGCTGCCGGCACGTACGCCTAC
GCGGGCAAGGAGTACACCTTCTCGGGCGGCGAGACGGACTTCGATGCGTGCGCCGAGATCGGCGGCAGCGCGCTGCAAGCCAACTTGACCTGCGGCACCAAGCCGCAACGGAGTGCTCTTTCAACGGT
GCGTGGCGTGGCAACGGCCTAAGCAAGGGCCGTGACTACTTTGTGTCGTGATTTCTGGGACCGCGCCTCCGAGACCGGCATTATTGAGGATGAGCAGGCGGCTATGTGGGAGGTCACCGCGCGGGAC
TTTGCTGACAAGGCCGATGAGGTCTGCGTCCAGTCCGTGGACGACATTGGCAAGGTGTATAAGAAGGTGCAGGGCGAGAACACCAAGTTCCTGTGCTTGGACCTGACCTACTGCCACGTCATGCTGACC
CAGGGCTTCAAGCTGGACGAGAAGATGAAGCTGACGCTGGTGAAGCAGGTGGAATACAACGGACAGCGCATTGAGGCGGCGTGGCCCCCTGGGCGCTGCCATTAACGACCTCTCCTCGTGA

21 =
22 SEQUENCE_ID=Cre06.g290250.t1.3|PACid:26893818
23 SEQUENCE_TEMPLATE=ATGAAGGACGCGGGCCCCCTGCCCCCTCCCGGCGCCCCGCGGGCGAGGGCGTGGCGCCAGCACGCGCTGGTGCCTGTGGCACGTGCTGAGCAAGCACCCGGAGGGGCTGCCG
TTGGGCAAGCTGGTGCAGCTGGTGACGGAAGAGAAAAAGGAGTTCAACTATAAGAACCCACGGGCGCGATCTCTGGGGAGCTATCGCGCGACAAGGAGCACTTTTGTAAGGCCGGCGGGCGGCCGCTGG
GCGCTCAGCGTGTTTGGGCCGTTTCGAGGGAGCGGGCGGCAGCGGCGGCGGCGCGCCGCCATCAGGCGCTGCAGTCGGCGTCAGCACGCGCGCGCACGCGCACGCCGCCACCGCGGTGCCAGGGGCTGCT
GGCGGGGAGGGCGGCGGGGAGGCCATGGACTTGGATGAGGCAGCAGCCGGGCATGAGGGCAACGGCGATGCCATGACGGGGGAGGACGCCGGCGCTGTAGAAGCCGGCGGTGCGGCGGCAGCGGCAGC
GATAGTGATGATGATGACCTGCCAATCGGGGCCGCGCCGCGGGCGGCGGTGGCCGCTGAGGGCGCGGCGGCGGCGGCTGCGGCGGCGGCGACGGTGAAGCGGGCGGCCAGCACATCAGCAGAGCCCGGG
GACGGAGCGGCCGAGGCGGCTGGGGCGGCGGCTGGGGCGGCGGCTGGGGCGGCAAGGCGCCGCCGCCAAGCGCCAGCGCACCGACGGCGCCGCCGGCGACGCCGGGGAAGGCGTAGCGGCAGCGCCG
CCTGCGGAAGGCGAAGGGGCGGCGCGGAGGCCGGGGCGGGGCCAGCGGCGCCACAGCCGCGGTGCCGGTGGCGGCTCCGGCTGCAGCTGGGGGCGGCGGGGCGGGGGCGGGGGCGAGGCCGGGGC

C.reinhardtii-Specific Primers

- highly specific to C. reinhardtii --> used C. incerta as mispriming library

```
1 PRIMER_MISPRIMING_LIBRARY=incerta_mispriming_library.fasta
2 PRIMER_OPT_SIZE=22
3 PRIMER_MAX_LIBRARY_MISPRIMING=16.00
4 PRIMER_MAX_TEMPLATE_MISPRIMING=12
5 PRIMER_LIB_AMBIGUITY_CODES_CONSENSUS=0
6 PRIMER_INTERNAL_MIN_GC=50
7 PRIMER_TASK=generic
8 PRIMER_PICK_LEFT_PRIMER=1
9 PRIMER_PICK_INTERNAL_OLIGO=0
```

```
##bash
```

```
#primer3 doing its work on reinhardtii input for C.reinhardtii-specific primers
```

```
/scratch/research/tmp_apps/primer3-2.3.7/src/primer3_core < reinhardtii_input2.txt > primer3_output_reinhardtii_selection
```

```
24 PRIMER_LEFT_NUM_RETURNED=3
25 PRIMER_RIGHT_NUM_RETURNED=5
26 PRIMER_INTERNAL_NUM_RETURNED=0
27 PRIMER_PAIR_NUM_RETURNED=5
28 PRIMER_PAIR_0_PENALTY=5.801451
29 PRIMER_LEFT_0_PENALTY=0.158696
30 PRIMER_RIGHT_0_PENALTY=5.642755
31 PRIMER_LEFT_0_SEQUENCE=AGGCTGACAATATCCTGAAGGC
32 PRIMER_RIGHT_0_SEQUENCE=AAGAGCACTCCGTTTGCG
33 PRIMER_LEFT_0=475,22
34 PRIMER_RIGHT_0=1005,18
35 PRIMER_LEFT_0_TM=60.159
36 PRIMER_RIGHT_0_TM=58.357
37 PRIMER_LEFT_0_GC_PERCENT=50.000
38 PRIMER_RIGHT_0_GC_PERCENT=55.556
39 PRIMER_LEFT_0_SELF_ANY_TH=0.00
40 PRIMER_RIGHT_0_SELF_ANY_TH=0.00
41 PRIMER_LEFT_0_SELF_END_TH=0.00
42 PRIMER_RIGHT_0_SELF_END_TH=0.00
43 PRIMER_LEFT_0_HAIRPIN_TH=34.44
44 PRIMER_RIGHT_0_HAIRPIN_TH=45.15
45 PRIMER_LEFT_0_LIBRARY_MISPRIMING=14.00, startgeneg5578
46 PRIMER_RIGHT_0_LIBRARY_MISPRIMING=11.00, comp startgeneg20636
47 PRIMER_PAIR_0_LIBRARY_MISPRIMING=24.00, startgeneg5578
48 PRIMER_LEFT_0_END_STABILITY=4.3500
49 PRIMER_RIGHT_0_END_STABILITY=4.8500
50 PRIMER_LEFT_0_TEMPLATE_MISPRIMING=7.0000
51 PRIMER_RIGHT_0_TEMPLATE_MISPRIMING=8.0000
52 PRIMER_PAIR_0_COMPL_ANY_TH=0.00
53 PRIMER_PAIR_0_COMPL_END_TH=0.00
54 PRIMER_PAIR_0_PRODUCT_SIZE=531
```

.txt

Broad Range Primers

```

109 Query= startgeneg5578
110
111 Length=1392
112
113                                     Score      E
114 Sequences producing significant alignments:   (Bits)    Value
115
116 g6136.t1|PACid:26892997                                1906       0.0
117
118 >g6136.t1|PACid:26892997
119 Length=1392
120
121     Score = 1906 bits (1032), Expect = 0.0
122     Identities = 1272/1392 (91%), Gaps = 0/1392 (0%)
123     Strand=Plus/Plus
124
125
126 Query    1          ATGAAGCGCCCTGACACCTTGGGTGACAGGATATGGCGGTCAGTCGATAAGAACGTGGAC  60
127           |||||
128 Sbjct    1          ATGAAGCGCCCTGACACCTTGGGTGACAGGATATGGCGGTCAGTCGATAAGAGCGTGGAC  60
129
130 Query    61          CAGAGGTACCGTGGCGTGCTGCTGGTGGTGTCCATCCCGGTCATTCTCATCCTGGCGGTC  120
131           |||||
132 Sbjct    61          CAGAGGTACCGCGGCGTGCTGCTGGTGGTGTCCATCCCGGTAATTCTAATCCTGGCGGTC  120
133
134 Query    121         ATCATGCTAGTACCCCGCTCTACGCCTTTCCATGCGGAGCGCCCGCACACCTTGAAGGAG  180
135           |||||
136 Sbjct    121         ATCACGCTGGTGCCACGTTCAACGCCTTTCCACGCGGAGCGCACCCACACGCTGAAGGAG  180
137
138 Query    181         CTGCGCCATACGGGCGCGGCCGCGAAGTATGCAATTGTGTTTCGATGCGGGCAGCACCGGC  240
139           |||||
140 Sbjct    181         CTGCGCCATACAGGCGCGGCCGCAAAGTTCGCGATTTGTGTTTTCGATGCGGGCAGCACAGGC  240

```

Primer3Plus

pick primers from a DNA sequence

[More...](#)[Source Code](#)[Help](#)[About](#)[Load server settings:](#)

Default



Activate Settings

Select primer pairs to detect the given template sequence. Optionally targets and included/excluded regions can be specified.

Task:

generic

[Pick Primers](#)[Reset Form](#)**Main****General Settings****Advanced Settings****Internal Oligo****Penalty Weights****Advanced Sequence**[Sequence Id:](#)[Paste template sequence below](#)

Or upload sequence file:

[Choose File](#)

No file chosen

[Upload File](#)

Mark selected region:

< >

[]

{ }

[Clear](#)[Save Sequence](#)[Excluded Regions:](#)

<

>

[Targets:](#)

[

]

[Included Region:](#)

{

}

[Primer overlap positions:](#)

-

[Pair OK Region List:](#)☒ [Pick left primer](#)☐ [Pick hybridization probe](#)☒ [Pick right primer](#)or use [left primer](#) below.(internal oligo) or use [oligo](#) below.or use [right primer](#)

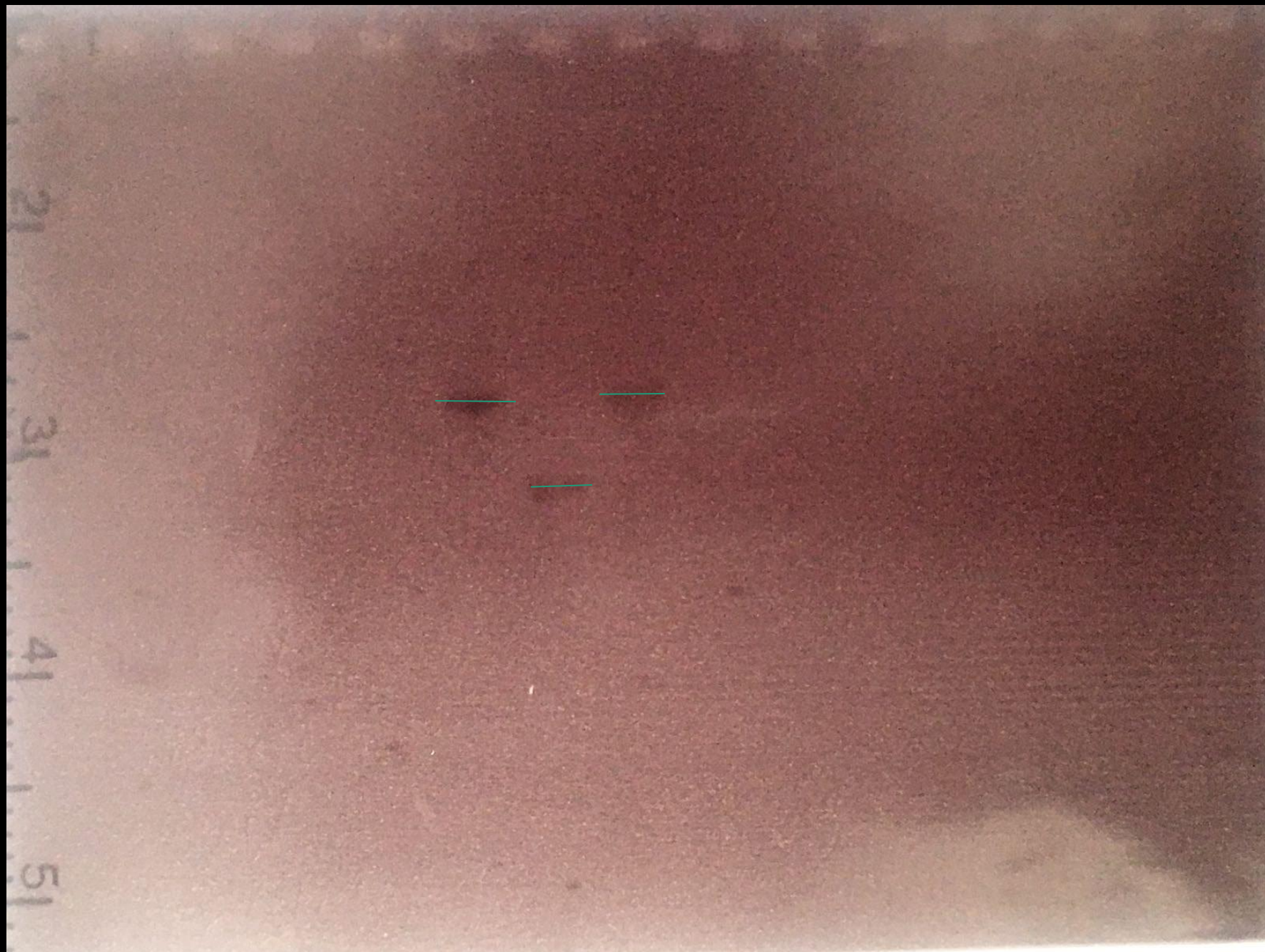
- Query= startgeneg7179
- Cre03.g161400.t1.2 | PACid:26901868

OLIGO	<u>start</u>	<u>len</u>	<u>tm</u>	<u>gc%</u>	<u>any</u>	<u>3'</u>	<u>seq</u>
LEFT PRIMER	573	18	59.27	66.67	5.00	2.00	GACGCCCCTGTACTACGC
RIGHT PRIMER	1026	18	59.55	61.11	7.00	0.00	CACCATCATGGGGTAGGG

6616					
6617	Query	555	GGACTACGTGGGCCGCGAGACGCCCTGTACTACGCGGAGCGCCTGTCGGCGAACTACAA	614	
6618					
6619	Sbjct	555	GGACTACGTGGGTCGCGAGACGCCCTTTACCACGCCGAGCGCCTGTCGGCGCACTACAA	614	
6620					
6644					
6645	Query	975	CCACTACATCCTGGGCTCGGCCGCCGGCCCCCACCCTACCCCATGATGGTGCGCGAGTT	1034	
6646					
6647	Sbjct	975	CCACTACATCCTGGGGTCGGCCGCTGGCCCCCACCCTACCCCATGATGGTGCGCGAGTT	1034	
6648					

PCR

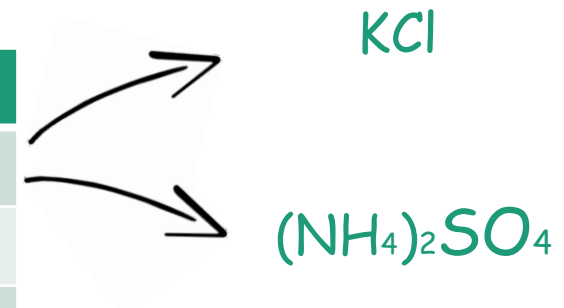
I. Primer Eval with Lab Strain DNA extract



PCR

I. Primer Eval with Lab Strain DNA extract

Final Concentration/ Ratio	Reagent
2.5 uL in 25 uL	Buffer
2.0 or 2.5 mM	Mg ²⁺
0.125 uL in 25 uL	Taq Polymerase
200 mM	dNTPs
1 ng/uL	gDNA
0.5 uL in 25 uL	Forward Primer
0.5 uL in 25 uL	Reverse Primer
1 M	Betaine - Additive



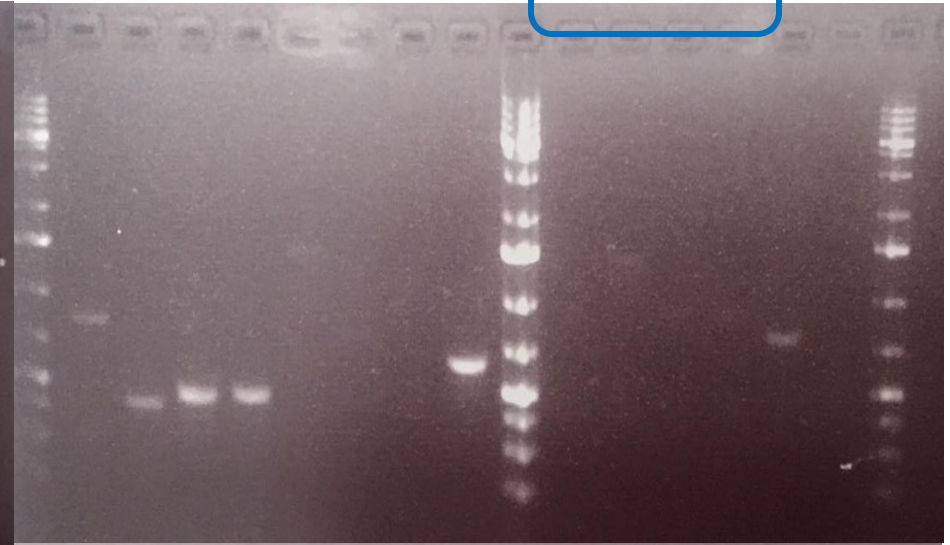
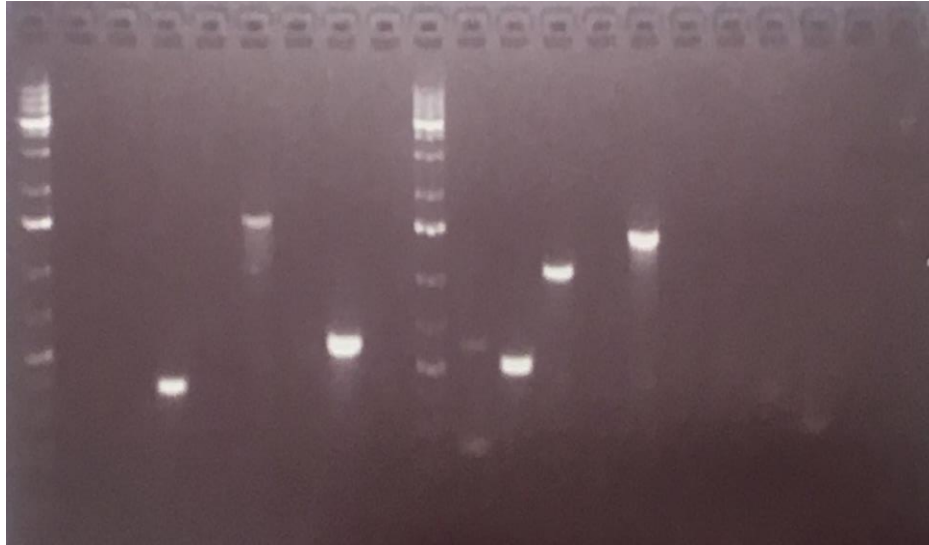
C.reinhardtii-specific

Broad Range

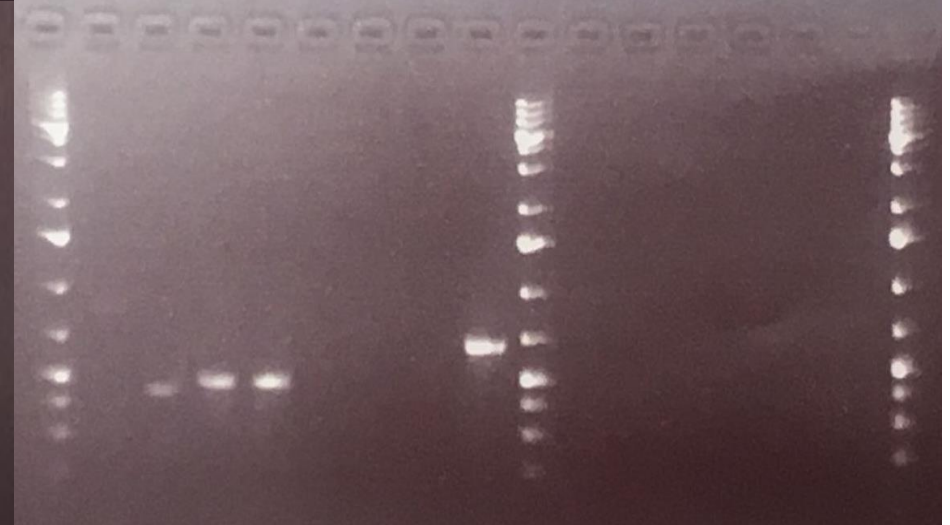
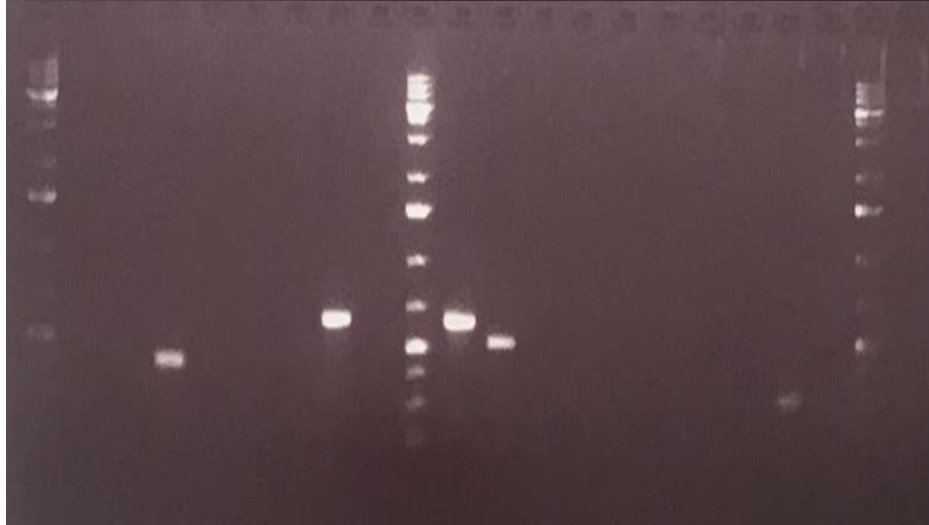
1 1 2 2 3 3 4 4 6 6 7 7 8 8 9 9 10 10

1 1 3 3 4 4 6 6 7 7 8 8 9 9

KCl,
2 mM MgCl

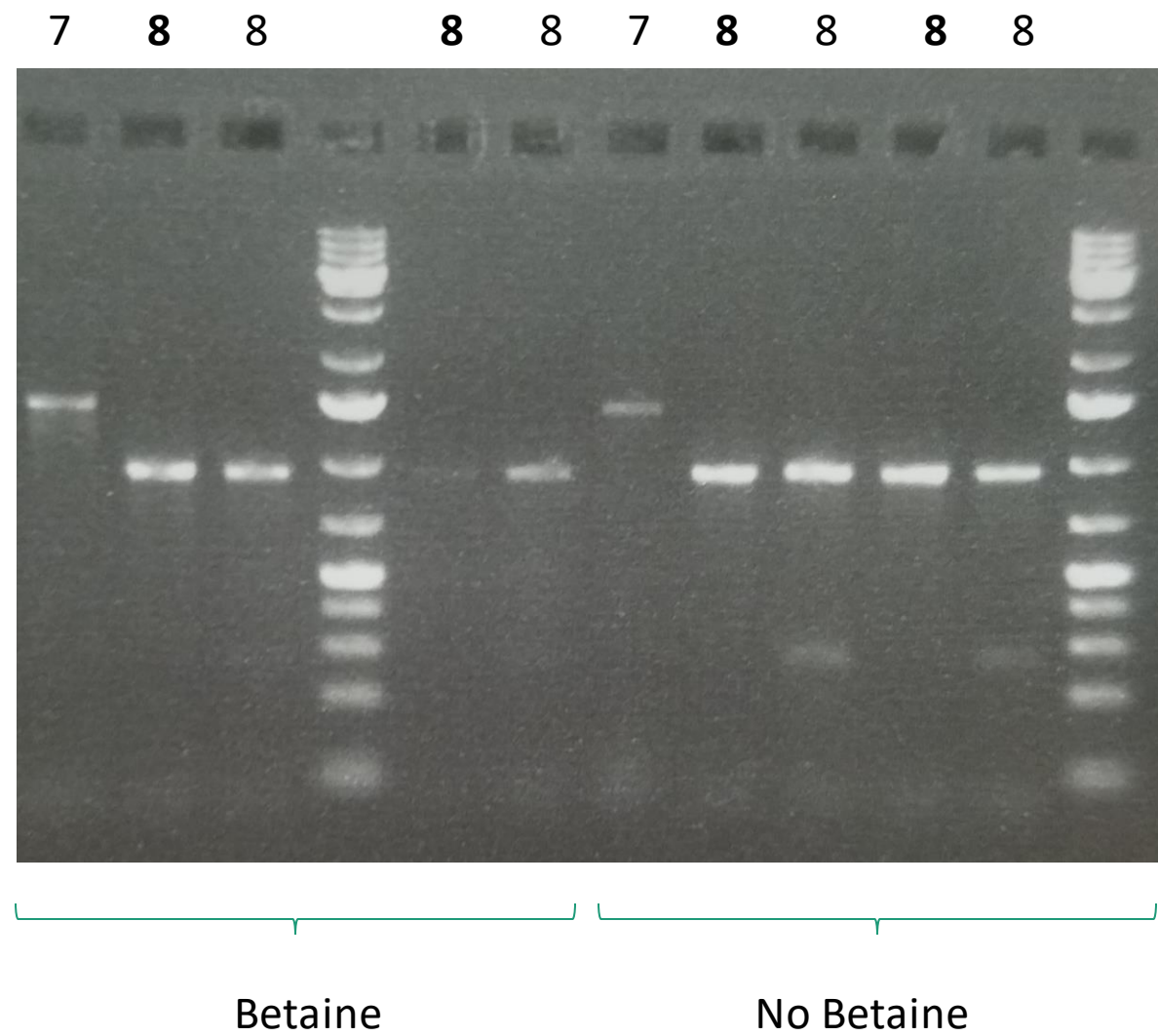


(NH₄)₂SO₄,
2.5 mM MgCl



Bold numerals: C. reinhardtii amplicon

Else: C. incerta amplicon



Soil Sampling

Riverwood Conservancy, Mississauga

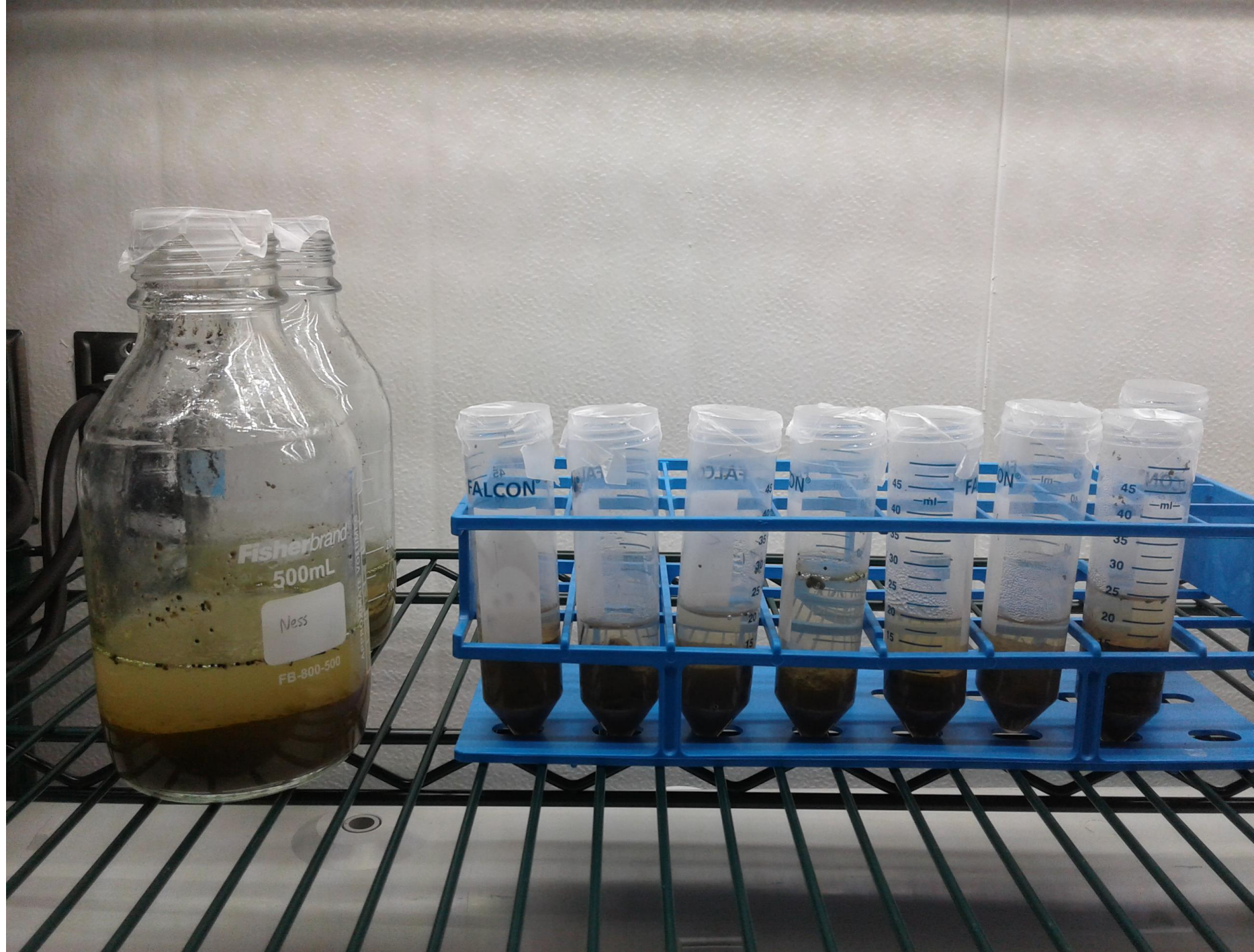
- Culham trail
- Two samples from two very different soil types
- Upper Culham (UC) : clay/loam like
- Lower Culham (LC) : drier, shallower.

A- Raspberry Fields
B- Sunflower Fields
C- Seedlings
D- Blueberry Fields
E- x



Andrew's (very) Scenic Acres, Milton

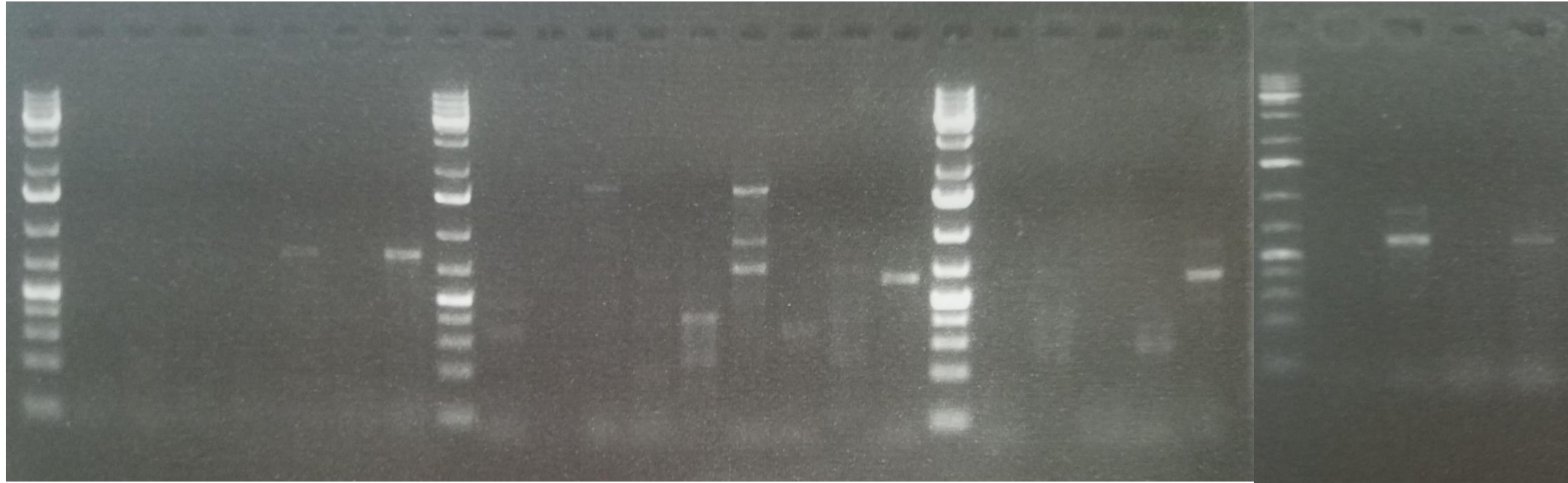
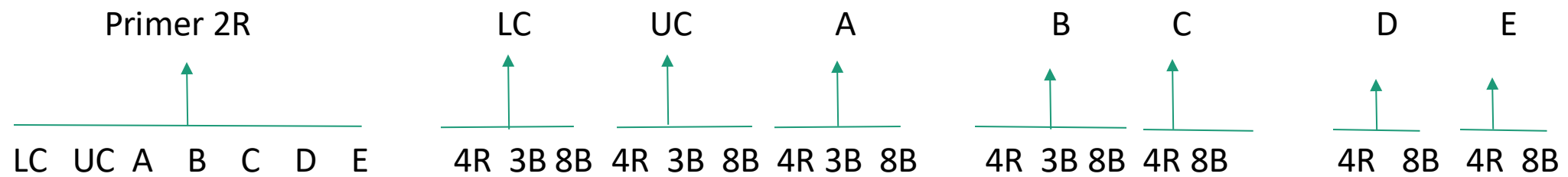




To Culture or Not to Culture



Direct DNA extraction from Soil	Culture
Point of using PCR?	Will I get enough DNA?
Faster- contamination not an issue	Primer specificity requirement less stringent than it would be with indigenous soil DNA composition
Chlorella may outcompete	



Prime r	Expec ted FS	FS- 1690 &/or 3871	Fragm ent Size- E	Fragm ent Size- C	FS UC	FS A	FS D	FS LC
2R	267	4 (400 bp)	6.5 (700-	6.5				