

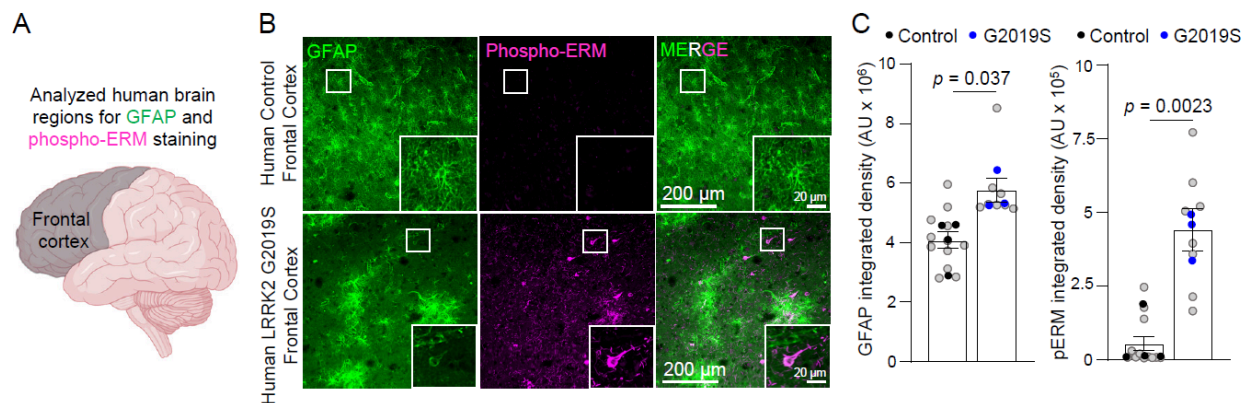
Interactive example 1 - Manuscript text

To study how the LRRK2 G2019S mutation impacts astrocytes in the prefrontal cortex, a cortical region linked to PD, we stained sections from the frontal cortices of age and sex-matched control and LRRK2 G2019S mutation-carrying PD patients with glial fibrillary acidic protein (GFAP, a marker for astrocyte branches) and phosphorylated ERM (Phospho-ERM, which are enriched in PAPs).

Interactive example 1 - Figure legend

Figure 1: ERM phosphorylation is impaired in PD patients carrying LRRK2 G2019S mutation. (A) Schematic of human frontal cortex regions analyzed for phospho-ERM staining. (B) Representative confocal images of GFAP (green) and phospho-ERM (purple) in the frontal cortex of human controls (n = 4; 3M, 1F) and LRRK2 G2019S mutation carriers (n = 3; 2M, 1F), aged > 80 years. Scale bar, 200 μ m. (C) Quantification of GFA integrated density in (B). GFAP: t (5) = 2.822, p = 0.037; phospho-ERM: t (5) = 5.695, p = 0.0023. Grey dots: individual images; black dots: control averages; blue dots: mutation carrier averages.

Interactive example 1 Figure



Interactive example 2- Manuscript text

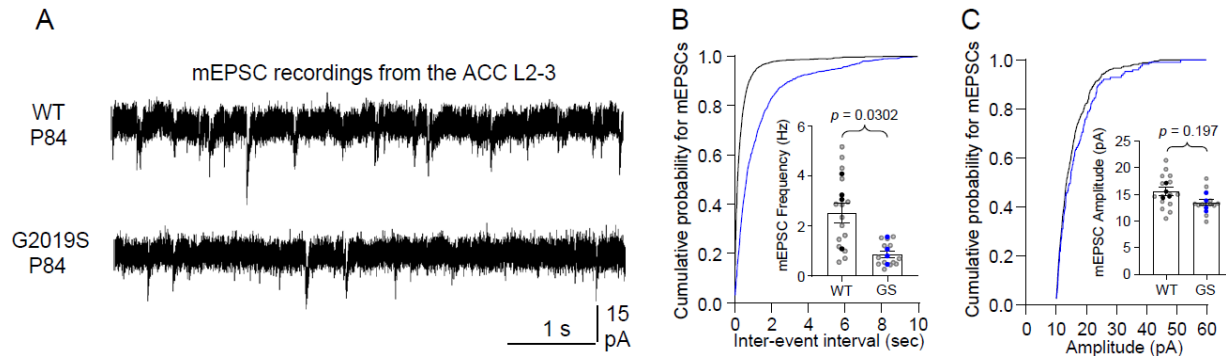
Next, we used electrophysiology to assess how altered synapse density in the ACC and MOP of LRRK2 G2019Ski/ki mice affects synaptic function. We recorded miniature excitatory postsynaptic currents (mEPSCs) in the ACC L2-3 pyramidal neurons and miniature inhibitory postsynaptic currents (mIPSCs) in the MOP L2-3 pyramidal neurons from acute brain slices of P84 WT and LRRK2 G2019Ski/ki mice. LRRK2 G2019Ski/ki neurons displayed mEPSC frequency reduced by ~65% and a corresponding right shift in the cumulative distributions of mEPSC inter-event intervals (Figure 3A-B) when compared to WT neurons, with no change in mEPSC amplitude (Figure 3C).

Interactive example 2 - Figure legend

Figure 3: LRRK2 G2019S affects excitatory and inhibitory synapse function in the

ACC and MOp. (A) Representative mEPSC traces from ventral ACC L2-3 pyramidal neurons in WT and LRRK2 G2019Ski/ki mice. (B) Cumulative probability and quantification of mEPSC frequency: $n = 15$ (WT), 13 (LRRK2 G2019S ki/ki) neurons, 4 mice/genotype. Kolmogorov-Smirnov test: $D = 0.504$, $p < 0.001$. Mean frequency: WT (2.878 ± 0.6355), LRRK2 G2019S ki/ki (0.9673 ± 0.2328). Unpaired t-test: $t(6) = 2.823$, $p = 0.0302$.

Interactive example 2 - Figure



Interactive example 3 - Manuscript text

In vivo, Atg7 knockdown in WT astrocytes reduced astrocyte territory volume and morphological complexity, resembling the LRRK2 G2019Ski/ki astrocytes transfected with shControl (Figure 7I-K). In LRRK2 G2019S ki/ki astrocytes, Atg7 knockdown improved morphological complexity but did not further reduce territory volume (Figure 7J-K). These findings highlight Atg7's role in regulating astrocyte morphology and suggest its dysfunction in LRRK2 G2019Ski/ki astrocytes contributes to altered morphology.

Interactive example 3 - Figure legend

(I) Images of ACC and MOp L2-3 astrocytes at P21 expressing shControl or shAtg7-PB-mCherry-CAAX. Scale bar, 10 μ m. (J) Quantification of astrocyte territories. Nested ANOVA [$F(3,28) = 9.484$, $p = 0.0002$] with Bonferroni tests: WT shControl vs. LRRK2 G2019Ski/ki shControl ($p = 0.0036$), WT shControl vs. WT shAtg7 ($p = 0.0003$), WT shAtg7 vs. LRRK2 G2019Ski/ki shAtg7 ($p = 0.0109$). $n = 20$ – 25 cells from 4–5 mice/group. (K) Astrocyte branching complexity. Two-way ANOVA revealed significant condition [$F(2,324) = 43.1$, $p < 0.05$], radius [$F(9, 250) = 301.8$, $p < 0.05$], and interaction effects [$F(27, 610) = 4.539$, $p < 0.05$]. Bonferroni tests showed significant differences between groups, including WT shControl vs. LRRK2 G2019Ski/ki shControl ($p < 0.0001$) and WT shControl vs. WT shAtg7 ($p < 0.0001$).

Interactive example 3 - Figure

