

Questions regarding developmental control of  
fibre diameter and fibre length growth rate in  
sheep

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# Contents

<b>1</b>	<b>Introduction</b>	<b>2</b>
<b>2</b>	<b>The papilla cell model</b>	<b>2</b>
<b>3</b>	<b>Competition between follicles</b>	<b>3</b>
3.1	The maths of competition and sharing . . . . .	3
3.2	Follicle function . . . . .	4
3.3	Closer look at Fraser and Short(1952) [4] . . . . .	5
3.4	Follicle groups . . . . .	8
3.5	Where to ? . . . . .	8
<b>4</b>	<b>Fibre length growth rate and fibre diameter</b>	<b>8</b>
4.1	Understanding how one follicle functions . . . . .	8
4.2	Fibre growth equations . . . . .	10
4.3	Data on length growth rate and diameter for individual fibres . .	10
4.4	Cell model for fibre growth . . . . .	10
<b>5</b>	<b>Discussion</b>	<b>10</b>

# 1 Introduction

This is not a research paper or a review. What we are trying to do is set down some ideas relevant to control of fibre diameter and fibre length growth rate in sheep.

The focus is on fibre diameter. We know that the number of papilla cells in a follicle bulb partly determines the diameter of the fibre grown. We want to understand what else is involved so that we can understand how variation in fibre diameter between and along fibres arises.

Fibre length growth rate is included, because it is felt that understanding the relationship between diameter and length will help to unravel the diameter control issue.

# 2 The papilla cell model

The fleece of a Merino sheep consists of something of the order of 100 million wool follicles each growing a fibre of approximate circular cross section whose diameter may be anything from 10 to 50 microns, and may vary along the length of the fibre. The length growth rate of individual fibres may be between 100 and 1000 microns per day.

The determination of follicle number is understood. The work of Moore and Jackson(1984) [10], Moore, Jackson and Lax(1989) [11], and Moore, Jackson, Isaacs, and Brown (1998) [12] suggests that a population of cells known as pre-papilla cells is responsible for follicle initiation at sites at which these cells aggregate. These pre-papilla cells end up in the papilla cavity of the follicle bulb, and the number of papilla cells in a bulb tend to relate to the size of the follicle and the diameter of the fibre it grows.

We can calculate the consequences for follicle number from assuming various combinations of parameters of the papilla cell population and its dynamics and differentiation. There is a writeup and software for this calculation (Jackson and Moore (2018) [9]). There is also a writeup (Swan(1999) [19]) which tackles the algebra of pre-papilla cell population numbers. If we assume a relation between papilla cell number and fibre diameter, we can even calculate mean fibre diameters.

We do have an empirical relationship between average no of papilla cells per follicle and mean fibre diameter. It is in Figure 4 of Jackson and Moore (2018) [9]. The correlation in Figure 4 is 0.81 - so only 67 percent of the variance in papilla cell number is explained by fibre diameter. There is room for some other effects to operate. Figure 4 is a mix of sheep from four selection lines. There is no evidence that any selection line deviates from the linear regression line.

However there is more to fibre diameter variation than the number of papilla cells can explain. Mean fibre diameter is a character with an enormous amount of phenotypic plasticity - it varies with seasonal conditions resulting in variation in diameter along the fibre. Papilla cells may be involved in controlling this

variation. In species which undergo a regular hair growth cycle, papilla cells direct the growth and decline of follicles as they go thru the various stages of the hair growth cycle, and fibre diameter varies with these stages. The physiological and hormonal signalling which affects follicles and fibre growth seems to operate via the papilla cells. In species like sheep in which follicles are almost permanently in Anagen phase, this effect is suppressed, but may not be entirely absent. One of the major modifications made to sheep in domestication is the shift from seasonal shedding to continuous fibre growth. There is also variation in fibre diameter ( and in papilla cell number) between follicles on the one sheep. This variance of fibre diameter between fibres is unexplained by the papilla cell model, and it is not known to what extent it depends on the number of papilla cells which aggregate in the bulb of each follicle, or to what extent it depends on other factors such as variation in the density or sizes of follicles. It is known that the type of follicle has an effect on fibre diameter in some sheep - primary follicles sometimes grow fibres of a larger diameter than those grown by secondary follicles.

### 3 Competition between follicles

The classic paper on this is Fraser and Short(1951) [3]. In Fraser and Short(1952) [4] evidence is presented in the form of

”..a negative correlation occurred between the size of a fibre and the number, size, and distance of fibres adjacent to it. ”

It is suggested that there is a maximum distance over which this negative correlation occurs. There is also an extensive discussion in Fraser and Short(1960) [6].

We focus here on the possibility of competition between adult follicles for some resource which is required to grow fibre. The issue of competition during follicle initiation and development is left aside.

#### 3.1 The maths of competition and sharing

There are two possibilities. Organs can actively compete for a resource, or they can passively share in it. Most organs seem to operate by passive sharing. In mammals, the circulation system is massively good at sharing nutritional resources around. The notable exceptions are the foetus, and cancer cells. These actively corner more than their share of everything. So, the big question, are follicles like most organs, or are they like cancer cells? We think the answer is obvious, the skin is like most other organs, it shares, both between the skin and other tissues, and between organelles within the skin.

To understand the subtle difference look at the maths. Sharing is the easiest. We all learnt about sharing in primary school - it is called division. If there are 12 lollies and 4 kids we all learnt to do  $12/4 = 3$ , and work out that they each get 3 lollies. Division is equal sharing.

Now, what if the sharing is not equal? There has to be some reason for a recipient receiving an unequal share. It may be that the recipient is more actively competitive, or it may be that some external agent is supervising the sharing and imposes a rule. In either case the maths behind it is something we all learnt a little later in primary school - fractions or ratios. A fraction or proportion specifies a share. A ratio specifies a rule for sharing - one for me, two for you, etc. Fractions and ratios don't identify the reasons behind the unequal sharing, they just say it exists.

Conclusion. We can not distinguish between competition and unequal sharing for other reasons. Not by looking at the maths anyway - we might get somewhere by knowing about the biology. We can probably compute each follicle's share of resources, simply by calculating each follicle's fibre output as a fraction of the whole fibre output of the sheep. It would be a rather small fraction, of the order of  $10^{-8}$ .

### 3.2 Follicle function

This is a huge topic. There are good reviews in .... What we are looking for here are things that might affect sharing. There are at least three levels

**between follicles within a sheep** some follicles might differ in a way which leads them to receive a greater or lesser share of nutritional resources than their neighbours. We know this is true of primary follicles. It may also operate within follicle groups or across the whole skin.

**between different parts of the fleece bearing surface** we know lots about this. Diameter and length growth rate variation over the body of the sheep has been extensively studied. Some key references are Chapman and Young(19..).... The issue of how this site variation relates to follicle structure and function is not well researched.

**between sheep** all the follicles on a sheep might be different in some way which leads the whole skin to receive a greater or a lesser share of nutritional resources

So let's make our tentative list of things that might affect sharing

**follicle attributes** this is actually quite difficult. Size apart, we can not imagine anything within a follicle that might cause it to consume more resources. So let's concentrate on size - if there are more bulb cells dividing to make fibre then we have a larger organ which might use more resources if they were available. It is another matter to say that if resources were limited a larger follicle might 'rob' its neighbours. If a follicle is simply 'more hungry' for resources, then, unless something actually supplies extra resources, it remains hungry. It can not go out and forage for extra on its own.

**skin attributes outside the follicle** the obvious issue here is blood supply to follicles. Studies of Ryder(1955) [17] show that follicle bulbs with a larger papilla cavity have more blood vessels inside the papilla. The work of Nay(1966) [13] shows that the arrangement of blood vessels in the papillary layer of sheep skin is different in sheep with tangled and straight follicles. This is a much more likely source of modifications to sharing. If follicles develop with different blood vessel supplies, then they may have different resource consuming abilities. This may be related to the number of papilla cells or it may be an independent issue.

**whole sheep attributes** some sheep simply make more resources available to all their tissues, wool follicles included. This would contribute to between sheep variation, but not to other levels within a sheep.

Tentative conclusion. There is likely to be an unequal sharing of resources between follicles. The follicle attributes that seem to be correlated with growth of larger fibres ( and therefore consumption of more resources) are likely to be developed in synchrony with skin attributes such as blood supply so that enhance supply of resources. Biology works like that. So while it might seem that certain follicle types grab a larger share of resources, the truth is they are actually given a larger share. Follicle biology seems to be an 'old school tie' system, not a 'rat race'.

### 3.3 Closer look at Fraser and Short(1952) [4]

If we are going to deviate from Fraser and Short(1952) [4] we need to go back over that paper, and see exactly where it is we differ. Fraser and Short(1952) [4] is an amazing piece of work. It is only three sheep, a Ryland, a Lincoln, and a medium strong Merino. The samples came from Dr Carter's collection. The measurements were as follows - choose a fibre at random, draw a circle around it of radius 147 micron and measure the distance to every follicle within that circle, and its fibre diameter. Repeat for 173 and 293 micron. Repeat for around 70 randomly chosen fibres.

The analyses consisted of regressions of the diameter of the central randomly chosen fibre on a measure of the total cross sectional area of growing fibre in the surrounding circle. The measure was intended to describe the total fibre output of the surrounding follicles. The regressions were done with and without weighting according to the distance of each follicle from the centre of the circle. We reproduce their Figure 1 in Figure 1, so we can see how good a fit the regressions were.

There is substantial variation around the regression line. This is not unexpected. Biological data are always noisy. This is the 'best' result - the Lincoln sample had the most significant regressions. We comment on this below. The weighting by distance did make some improvement. The regressions were better for the smaller diameter circle. That all makes sense.

What we now have to do is delineate our areas of concern. These are as follows

An infinite range of inverse functions of distance of separation could be analysed, and it is intended in a later paper to detail the determination of the function relating intensity of competition to distance of separation.

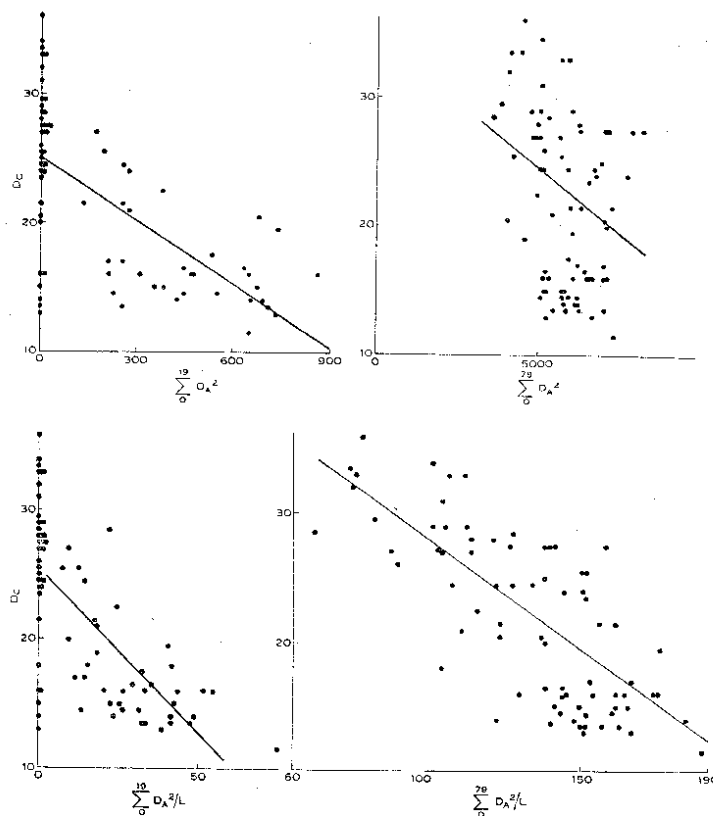


Fig. 1.—Scatter diagrams for the Lincoln sample of the diameters of the central fibres plotted against the sum of the diameters squared of the adjacent fibres. This summation is given from 0 to 79 and from 0 to 19. The two top diagrams are for summation without correction for distance. The two bottom diagrams are for summation after correction for distance.

Figure 1: Figure 1 from Fraser and Short(1952) [4]. These data are for the Lincoln sheep only. The left side graphs are for the 147 micron circle, the right graphs for the 173 micron circle. The lower graphs are weighted by distance between follicles, the upper graphs are not weighted.

**follicle positions** these data came from horizontal sections of skin at sebaceous gland level. What is sectioned at that level is the follicle shaft, not the

bulb. Bulbs deviate substantially from the position of the shaft, especially in sheep with high follicle curvature. This is why the Lincoln data have a better fit - the positional information reflects the bulb position more accurately in the Lincoln because they have straight follicles. The Ryland and Merino specimens would have some degree of follicle curvature. It is the position of the bulb which may matter in relation to competition, because the bulb is the active tissue involved in growing fibre.

**length growth rate** as a measure of fibre growth activity of the surrounding circle of skin, Fraser and Short used the sum of the squares of the diameters of all the follicles in the circle. This accounts for number of follicles, and fibre cross sectional area, but not fibre length growth rate. We do not know if diameter and length are correlated in this scenario. So one part of their measure of follicle output of fibre is missing.

**confounding fibre growth and development** we are going to take a punt and assert that most of the regression significance comes from follicle number, rather than from diameter. Follicle number is purely a development factor. It does not change as the follicles grow fibre in the adult. So the contribution of follicle number to the regression is not at all an indicator of competition between fibres while they are growing fibre. It may indicate competition during development, but more recent work (Moore et al (1998) [12]) indicates that it reflects variation in numbers of sites occupied by papilla cells, or, in the case of Merino sheep, follicles formed by branching from other follicles.

Diameter, on the other hand, could reflect either factors operating during development ( such as the number of papilla cells that aggregate at sites) or factors that operate during fibre growth ( such as the expansion and shrinkage of follicles that occurs over the hair growth cycle, or with nutritional and photoperiodic variations). So diameter mixes up developmental factors with adult growth factors. We suggest that the diameter effect on the regressions is likely to have been minimal - ie it is all about follicle number, which means it is all about development, not current growth.

Is there any way we can address these issues. Yes. The paper says

”The data from which the analysis detailed below was made are very bulky and have been lodged, with the accompanying statistical condensations and copies of the original drawings, with CSIRO Head Office Library, Melbourne, where they can be consulted. ”

Maybe we should request access to these data, and see if we can redo the analyses separately for follicle number and fibre diameter. That would at least address the third point above.



### 3.4 Follicle groups

One of the messages from the classic papers of H.B. Carter (Carter(1943) [1] and Carter and Hardy (1947) [2]) is that the *trio group* of three primary follicles and numbers of associated secondary follicles is the *biological unit* of sheep skin. What happens inside one trio group is repeated all over the woolgrowing surface, with some variations according to body sites. When you switch to a different sheep, you may get a slightly different trio group, but it is again repeated all over the body.

So it has to be said that a large part of variation in fibre diameter between fibres within a fleece is due to variations in follicle and skin structure within the trio group. So studying trio groups is an important approach to trying to understand what controls fibre diameter variation.

Unfortunately data on trio groups, particularly with fibre diameters measured, is rather limited.

### 3.5 Where to ?

The mere idea of unequal sharing of some resource among follicles does not help much sort out why the shares are unequal. That is what we need to know - who or what is dealing out the resource unequally, and on what basis.

One thing is certain. We need to get quantitative. That is what the next section is about. We try to get some real data on length and diameter, and to write some equations.

## 4 Fibre length growth rate and fibre diameter

This is about quantifying individual fibre length growth rates and fibre diameters and trying to write some equations defining how length growth rate and diameter are related to each other and to other things. So we are putting explanations on hold and trying to establish some facts.

### 4.1 Understanding how one follicle functions

We need to tackle this before looking at variations between follicles. For the present purpose, one fibre grown over a nominal time interval can be described as a cylinder of keratin with a length ( $L$ ) and diameter( $D$ ). We are neglecting internal structure and shape (curvature and cross section) and are not allowing diameter to vary along the length of the cylinder.

The amount of keratin material in the cylinder can be described by its volume ( $V$ ), which is related to length and diameter by an identity  $V = \frac{\pi}{4} D^2 L$ . So we have 3 variables,  $V$ ,  $D$ , and  $L$ . What the identity relationship says is that only 2 of the three could be under independent biological control. The third is automatically fixed by the other two.

So we already have a problem. Which 2 of the 3 are under biological control? There are three possibilities summarized in Figure 2.

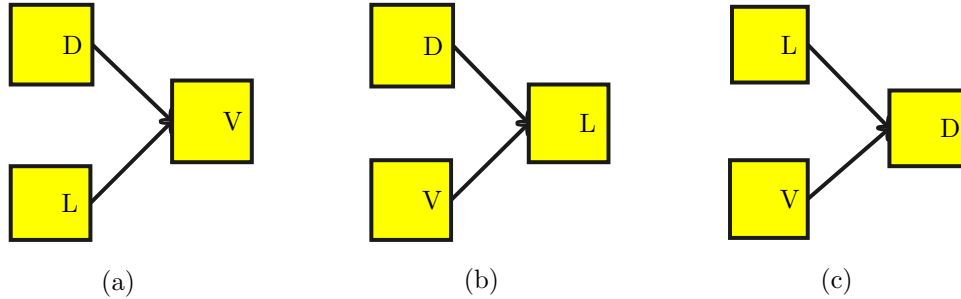


Figure 2: Path diagrams showing three possible causal relationships between fibre volume ( $V$ ), fibre diameter ( $D$ ) and fibre length ( $L$ ).

We think the correct causal model is Figure 2(b). This corresponds to transforming the volume equation to

$$L = \frac{V}{\frac{\pi}{4}d^2}$$

So we are saying that  $D$  and  $V$  are what is biologically determined, and  $L$  is simply a consequence of setting  $D$  and  $V$ .

How do we justify that? Lets start with an analogy. Take a beaker of glass beads and pour them into a measuring cylinder. The beads form a stack of diameter equal to the diameter of the measuring cylinder, and the height of the stack corresponds to  $L$ . Put a larger beaker of beads into the cylinder and the diameter stays the same, but  $L$  increases. Use a smaller cylinder with the same volume of beads and diameter of the stack will be less, but  $L$  will be greater.

So, to what extent is the growth of a wool fibre like making a cylindrical stack of glass beads? We say it is exactly like it mathematically, but not, of course, physically. To justify that we need to identify the biological factors which correspond to the volume of beads and the setting of the diameter of the cylinder. Here is our best guess

$V$  volume is an easy guess. Each glass bead corresponds to a cortical cell. So  $V$  is the total volume of cortical cells (per unit time). That in turn depends on the number of cells (per unit time) and the average cell volume. That in turn depends on the number of basal cells in the bulb (the cells which divide to form the cortical cells) and their mitotic rate, and certain cortical cell differentiation processes which determine their average volume.

$D$  diameter is more difficult. Some of the basal bulb cells divide to form cortical cells which migrate up to form fibre. Some of the basal cells divide to form cells which migrate up to form the inner root sheath, which is not part of the fibre, and is eventually resorbed. It would seem that follicles which produce larger fibres have larger bulbs, larger stems, larger papilla cavities, and more papilla cells in the papilla cavity. We might guess

that the papilla cells influence the size of the array of basal cells which become fibre. The only thing we know is that size can vary with time (eg nutritional effects, photoperiod effects, hair growth cycle effects) in an adult follicle and that the diameter of the fibre varies with it. There are also genetic differences in average follicle size between animals. There are also developmental differences in average size between P, So, and Sd follicles. There is also variation in size within each of these follicle types.

## 4.2 Fibre growth equations

## 4.3 Data on length growth rate and diameter for individual fibres

## 4.4 Cell model for fibre growth

# 5 Discussion

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