CATEGORICAL DATA ANALYSIS NOTE EDGARD MABOUDOU

Chapter 4. Logistic Regression

1. Introduction to Logistic Regression

- \bullet Assume a single explanatory variable X, quantitative, and a binary response variable
- $\pi(x)$ is the probability of success at value x
- the logistic linear regression model is defined as $logit(\pi(x)) = log\left(\frac{\pi(x)}{1-\pi(x)}\right) = \alpha + \beta x$.
- This implies that

$$\pi(x) = \frac{e^{\alpha + \beta x}}{1 + e^{\alpha + \beta x}}.$$

a. Interpretation of β

- The logit increases by β for every 1 unit increase in x
- it determines the ratio of increase or decrease of the S-shaped curve for $\pi(x)$
- The median effective level (EL_{50}) is the level at which each outcome has a 50%
- It is obtained by solving $\log \left(\frac{\pi(x)}{1-\pi(x)}\right) = \alpha + \beta x$ where $\pi(x) = 0.5$.
- this gives $x = -\alpha/\beta$

b. Example: Horseshoe Crabs data (p. 101)

- Let $\pi(x)$ be the probability that a female horseshoe crab of width x has a satellite,
- the simplest logistic model is $logit(\pi(x)) = \alpha + \beta x$
- The fit can be done easily in R as

```
> data<-read.table("crab.txt",header=T)</pre>
> attach(data)
> weight<-weight/1000; color=color-1;</pre>
> names(data)
[1] "color" "spine" "width" "satell" "weight"
> satell<-ifelse(satell>0,1,0)
> fit<-glm(satell~width,family=binomial(logit))</pre>
> summary(fit)
Call:
glm(formula = satell ~ width, family = binomial(logit))
Deviance Residuals:
    Min
              10
                 Median
                                3Q
                                        Max
-2.0281 -1.0458 0.5480 0.9066
                                     1.6941
Coefficients:
            Estimate Std. Error z value Pr(>|z|)
(Intercept) -12.3508
                         2.6287 -4.698 2.62e-06 ***
                                  4.887 1.02e-06 ***
width
              0.4972
                         0.1017
Signif. codes: 0 *** 0.001 ** 0.01 * 0.05 . 0.1
(Dispersion parameter for binomial family taken to be 1)
    Null deviance: 225.76 on 172 degrees of freedom
Residual deviance: 194.45 on 171 degrees of freedom
AIC: 198.45
Number of Fisher Scoring iterations: 4
> coef(fit)
(Intercept)
                  width
-12.3508177
              0.4972306
```

- From R output, our logistic model is

- The estimate probability of a satellite is

– The median effective level is

- The interpretation of the logistic regression uses the odds and the odds
- The odds of response (1) (odds of success) are $\frac{\pi(x)}{1-\pi(x)} = \exp(\alpha + \beta x) =$ $e^{\alpha}(e^{\beta})^x$
- so, for every 1 unit increase in x, the odds of response 1 are multiplied by
- For our example,

2. Inference for Logistic Regression

a. Confidence Interval

- A large sample Wald CI for β is $\hat{\beta} \pm z_{\alpha/2} SE(\hat{\beta})$ Exponentiating the endpoints yields an interval for e^{β}
- In our example,

– The Wald CI can also be obtained from R as follows:

```
> ### Wald CI
> beta<-coef(fit)[2];beta
    width
0.4972306
> vb<-vcov(fit)
> seb<-sqrt(vb[2,2]);seb
[1] 0.1017355
> logwald.ci<-beta+c(-1,1)*qnorm(1-alpha/2)*seb;wald.ci
[1] 0.2978326 0.6966286
> wald.ci<-exp(logwald.ci);waldci
[1] 1.346936 2.006975</pre>
```

- In the same way, the likelihood based CI for β is obtained from R by
- > ### Likelihood ratio CI
 > library(MASS)
 > lr<-confint(fit)
 Waiting for profiling to be done...
 > lrci<-lr[2,];lrci
 2.5 % 97.5 %
 0.3083806 0.7090167
 > elrci<-exp(lrci);elrci
 2.5 % 97.5 %
 1.361219 2.031992</pre>

doing:

– The likelihood based CI for β is

Interpretation of CI (Wald or Likelihood Ratio)

– For example, for Wald

b. Significance Testing

- For this example, want to test $H_0: \beta = 0$ vs $H_a: \beta \neq 0$
- LRT is more powerful than Wald test.

- Wald test is obtained from R by
- > ### Wald test

```
> waldt<-beta/seb; waldt
   width
4.887482
> pval=2*pnorm(waldt,lower.tail=F);pval
      width
1.02134e-06
   and the likelihood ratio test (LRT) test is performed in R by doing
> ### Likelihood Test
> nul<-fit$null.deviance;nul
[1] 225.7585
> res<-fit$deviance;res
[1] 194.4527
> lrt<-nul-res;lrt
[1] 31.30586
   - To make a prediction for a new value of x, use the command predict()
from R. Here is an example:
> ### Prediction of a new data
> new <- data.frame(width = 26.5)</pre>
> prednew<-predict(object =fit, newdata = new, type = "response", se = TRUE)
> prednew
$fit
0.6954646
$se.fit
         1
0.03996454
$residual.scale
[1] 1
> alpha<-.05
> lower<-prednew$fit-qnorm(1-alpha/2)*prednew$se;lower
        1
0.6171356
> upper<-prednew$fit+qnorm(1-alpha/2)*prednew$se;upper
0.7737937
> ci<-prednew$fit+c(-1,1)*qnorm(1-alpha/2)*prednew$se;ci</pre>
[1] 0.6171356 0.7737937
```

c. Model Checking

- Need to be able to check whether our model fits.
- If not, a more complicated model may be needed.
- For instance, we may need to include other term like a quadratic term (X^2) .
- We may need other explanatory variable in addition to X.
- We use goodness-of-fit statistic to check model fit.
 - Pearson χ^2
 - Likelihood Ratio G^2

d. Residuals

- Let y_i be the number of successes for the n_i observation at the *i*th setting of the explanatory variable.
- Let $\widehat{\pi}_i$ be the predicted probability of success using the fitted model.
- If the model is good, y_i should be close to $n_i \hat{\pi}_i$.
- The *i*th (Pearson) residual is

$$e_i = \frac{y_i - n_i \widehat{\pi}_i}{\sqrt{n_i \widehat{\pi}_i (1 - \widehat{\pi}_i)}} \sim N(0, 1).$$

- When the model doesn't fit well, inspection of the residuals may help identify the problem or suggest alternative models to fit.
- Residuals behave similarly to Z scores.
- Lack of fit will be values very far from $0 \ (\pm 2, 3)$.

3. Logit Models for Qualitative Predictors

- Our explanatory variables have been continuous variables so far.
- When X is continuous, we look at models like

$$\log\left(\frac{\pi}{1-\pi}\right) = \alpha + \beta x.$$

– When X is categorical, having levels $1, 2, \ldots, I$:

$$\log\left(\frac{\pi_i}{1-\pi_i}\right) = \alpha + \beta_i,$$

where X is at level i.

- Similar to one-way ANOVA model.
- Suppose X has I levels (this is a $I \times 2$ table),

$$\log\left(\frac{\pi_i}{1-\pi_i}\right) = \alpha + \beta_i, \quad \text{for } i = 1, 2, \dots, I.$$

Put restriction on β_i : (1) $\beta_1 = 0$, (2) $\beta_I = 0$, (3) $\sum_{i=1}^{I} \beta_i = 0$. Assume restriction (2), then $\alpha = \log \left(\frac{\pi_I}{1-\pi_I}\right)$. So

$$\beta_{1} = \log\left(\frac{\pi_{1}}{1 - \pi_{1}}\right) - \log\left(\frac{\pi_{I}}{1 - \pi_{I}}\right)$$

$$= \log\left(\frac{\pi_{1}(1 - \pi_{I})}{(1 - \pi_{1})\pi_{I}}\right) = \log\left(\frac{\Omega_{1}}{\Omega_{I}}\right)$$

$$\vdots$$

$$\beta_{I-1} = \log\left(\frac{\Omega_{I-1}}{\Omega_{I}}\right)$$

- An alternative approach uses dummy variables

$$X_i = \begin{cases} 1 & \text{if observation are at ith level of } X, \\ 0 & \text{otherwise.} \end{cases}$$

$$\log\left(\frac{\pi_i}{1-\pi_i}\right) = \alpha + \beta_1 x_1 + \dots + \beta_{I-1} x_{I-1}.$$

– As many parameters as categories for $X \Rightarrow$ model will fit the data perfectly. This model is called the saturated model.

Remark

$$\beta_i = \log\left(\frac{\pi_i}{1 - \pi_i}\right) - \log\left(\frac{\pi_I}{1 - \pi_I}\right)$$
$$= \log\left(\frac{\pi_i(1 - \pi_I)}{(1 - \pi_i)\pi_I}\right) = \log\left(\frac{\Omega_i}{\Omega_I}\right)$$

To find

$$\log\left(\frac{\Omega_1}{\Omega_2}\right) = \beta_1 - \beta_2.$$

To find odds ratio of level 3 and level 5, we have

$$\frac{\Omega_3}{\Omega_5} = e^{\beta_3 - \beta_5}$$

- Now, assume that we have

$$\log\left(\frac{\pi}{1-\pi}\right) = \alpha.$$

This means the levels of X has no effect on Y. We have the independent model.

- We can also have more than one explanatory variables.
- For example, Suppose X has levels $1,\ldots,I$ and Z has levels $1,\ldots,K$. More variables implies more models to consider.

Some potential models

$$\log \left(\frac{\pi(X=i,Z=k)}{(1-\pi(X=i,Z=k))} \right) = \alpha$$

$$= \alpha + \beta_i^X + \beta_k^Z$$

$$= \alpha + \beta_i^X + \beta_k^Z + \beta_{ik}^{XZ}.$$

4. Multiple Logistic Regression

- As with ordinary regression, logistic regression generalizes to more than one explanatory variable.

Continuous

For example, with k explanatory variables X_1, \ldots, X_k , we may fit the model

$$\log\left(\frac{\pi}{1-\pi}\right) = \alpha + \beta_1 x_1 + \dots + \beta_k x_k.$$

We need to first see the interpretation of the β 's:

 e^{β_i} gives the multiplicative factor/effect on the odds of a unit increase in the *i*th explanatory variable (X_i) when the other variables are held fixed.

AZT and AIDS Example (p111)

Assume X and Z each takes values 0 and 1 to represent the 2 levels of each explanatory variable.

$$logit_1(P(Y=1)) = \alpha + \beta_1 x + \beta_2 z.$$

The fit of this example using dummy variables is obtained from R.

```
> ### Example 1: AZT on AIDS
> y < -cbind(c(14,32,11,12),c(93,81,52,43))
> x<-c(1,0,1,0)
> z < -c(1,1,0,0)
> fitd<-glm(y~x+z,family=binomial)</pre>
> summary(fitd)
Call:
glm(formula = y ~ x + z, family = binomial)
Deviance Residuals:
      1
         2
                        3
-0.5547 0.4253 0.7035 -0.6326
Coefficients:
            Estimate Std. Error z value Pr(>|z|)
-0.71946 0.27898 -2.579 0.00991
0.05548 0.28861 0.192 0.84755
                        0.27898 -2.579 0.00991 **
Х
Z
Signif. codes: 0 *** 0.001 ** 0.01 * 0.05 . 0.1
(Dispersion parameter for binomial family taken to be 1)
    Null deviance: 8.3499 on 3 degrees of freedom
Residual deviance: 1.3835 on 1 degrees of freedom
AIC: 24.86
Number of Fisher Scoring iterations: 4
– An alternative for this fit is to use ANOVA. In R, this is done easily by:
> ### Example 1 using ANOVA
> table.4.3<-expand.grid(azt=factor(c("yes","no"),levels=c("no","yes")),</pre>
+ race=factor(c("white", "black"), levels=c("black", "white")))
> table.4.3<-data.frame(table.4.3,yes=c(14,32,11,12),no=c(93,81,52,43))
> options(contrasts=c("contr.treatment","contr.poly"))
> fita<-glm(cbind(yes,no)~azt+race,family=binomial,data=table.4.3)
> summary(fita)
```

Call:

glm(formula = cbind(yes, no) ~ azt + race, family = binomial,
 data = table.4.3)

Deviance Residuals:

1 2 3 4 -0.5547 0.4253 0.7035 -0.6326

Coefficients:

Estimate Std. Error z value Pr(>|z|)

Signif. codes: 0 *** 0.001 ** 0.01 * 0.05 . 0.1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 8.3499 on 3 degrees of freedom Residual deviance: 1.3835 on 1 degrees of freedom

AIC: 24.86

Number of Fisher Scoring iterations: 4

- Note that the deviance of the fit can be obtained in R by

> fita\$deviance

[1] 1.38353

> sum(residuals(fita,type="pearson")^2)

[1] 1.391026

> nulc<-fita\$null.deviance;nulc

[1] 8.349946

> resc<-fita\$deviance;resc</pre>

[1] 1.38353

Conditional independence between X and Y controlling for Z if $\beta_1 = 0$:

$$logit_2(P(Y=1)) = \alpha + \beta_2 z.$$

To test for conditional independence of AZT treatment and the development of AIDS symptoms, controlling for race. i.e.

 $H_0: \beta_1 = 0$ v.s. $H_a: \beta_1 \neq 0$. We want to compare model 2 (reduced model) to model 1 (full model).

- The reduced model is obtained in R as

```
> ### Fitting the reduced model
> fit1<-glm(chind(yes no)~race</pre>
```

> fit1<-glm(cbind(yes,no)~race,family=binomial,data=table.4.3)

> summary(fit1)

Call:

glm(formula = cbind(yes, no) ~ race, family = binomial, data = table.4.3)

Deviance Residuals:

Coefficients:

Signif. codes: 0 *** 0.001 ** 0.01 * 0.05 . 0.1 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 8.3499 on 3 degrees of freedom Residual deviance: 8.2544 on 2 degrees of freedom

AIC: 29.731

Number of Fisher Scoring iterations: 4

The LR statistic

 $LRT_{2/1}$ = residual deviance (2)-residual deviance (1) = 8.2544-1.3825 = 6.87 and the $df_{2/1}$ = 2 - 1 = 1. There is evidence of association. In R, we can do

> ### Testing the impact of AZT

> nulc1<-fit1\$null.deviance;nulc1</pre>

[1] 8.349946

> resc1<-fit1\$deviance;resc1</pre>

[1] 8.254436

> lrtm<-resc1-resc;lrtm</pre>

[1] 6.870906

– Note that in this specific case, we can also look at the Wald statistic P-value = 0.0099 .

Horseshoe Crabs with Color and Width Example (p116)

- Use the horseshoe crabs data with color and width as predictors.
- Color has 5 categories (light, medium light, medium, medium dark, dark).
- Sample contains no light crabs, so we used the other 4 categories.
- Treat color in the regression model using 3 dummy variables. (0, 0, 0) gives the 4th one.

$$logit(P(Y = 1)) = \alpha + \beta_1 c_1 + \beta_2 c_2 + \beta_3 c_3 + \beta_4 x.$$

– Assuming a model with no interaction, the fit is done using R.

```
> ### Example 2: Horseshoes crabs with colors and width predictors
> colfac<-factor(data\$color,levels=c("5","4","3","2"),labels=c("dark","med-dark
+ ,"med","med-light"))
> satell<-ifelse(satell>0,1,0)
> crabfit<-glm(satell~colfac+width,family=binomial)</pre>
> summary(crabfit)
Call:
glm(formula = satell ~ colfac + width, family = binomial)
Deviance Residuals:
    Min
              10
                   Median
                                 3Q
                                         Max
-2.1124 -0.9848
                   0.5243
                           0.8513
                                      2.1413
```

```
Estimate Std. Error z value Pr(>|z|)
                                      -4.604 4.14e-06 ***
(Intercept)
                -12.7151
                              2.7617
colfacmed-dark
                  1.1061
                              0.5921
                                       1.868
                                                0.0617 .
colfacmed
                  1.4023
                              0.5484
                                       2.557
                                                0.0106 *
colfacmed-light
                  1.3299
                              0.8525
                                       1.560
                                                0.1188
                                       4.434 9.26e-06 ***
width
                  0.4680
                              0.1055
Signif. codes: 0 *** 0.001 ** 0.01 * 0.05 . 0.1
```

(Dispersion parameter for binomial family taken to be 1)

```
Null deviance: 225.76 on 172 degrees of freedom Residual deviance: 187.46 on 168 degrees of freedom
```

AIC: 197.46

Number of Fisher Scoring iterations: 4

– The model has a different intercept parameter for crabs of different colors.

For example, the logit model for dark crab is

For example, the logit model for medium crab is

[–] However, the slope on width is always the same. So Regardless of the color, a 1 cm increase in width has a multiplicative effect of $e^{.468} = 1.60$ on the odds of having a satellite.

⁻ The exponentiated difference between two-color parameter estimates is an

odds ratio comparing those colors.

- So, at any given width, the estimated odds that a medium crab has a satellite are $e^{1.4023-1.1061}=1.34$ times the estimated odds for a medium dark crab.
- The confidence intervals for the parameters are obtained from R

```
> library(MASS)
```

> confint(crabfit)

Waiting for profiling to be done...

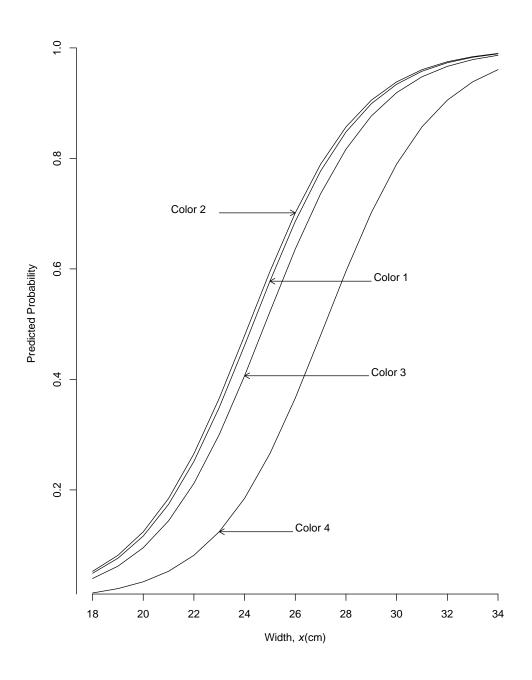
```
2.5 % 97.5 % (Intercept) -18.45674069 -7.5788795 colfacmed-dark -0.02792233 2.3138635 colfacmed 0.35269965 2.5260703 colfacmed-light -0.27377584 3.1356611 width 0.27128167 0.6870436
```

– For the graph, predict the probability at widths from 18 to 34cm each of the colors.

```
> ### Plot of the predicted values
> res1<-predict(crabfit, type="response", newdata=data.frame(width=seq(18,34,1),
+ colfac="med-light"))
> res2<-predict(crabfit, type="response", newdata=data.frame(width=seq(18,34,1),
+ colfac="med"))
> res3<-predict(crabfit, type="response", newdata=data.frame(width=seq(18,34,1),
+ colfac="med-dark"))
> res4<-predict(crabfit, type="response", newdata=data.frame(width=seq(18,34,1),
> plot(seq(18,34,1),res1,type="l",bty="L",ylab="Predicted Probability", axes=F,
+ xlab=expression(paste("Width, ", italic(x), "(cm)")))
> axis(2, at=seq(0,1,.2))
> axis(1, at=seq(18,34,2))
> lines(seq(18,34,1),res2) # add colors 2-4
> lines(seq(18,34,1),res3)
> lines(seq(18,34,1),res4)
> # add arrows and text
> arrows(x0=29, res1[25-17], x1=25, y1=res1[25-17], length=.09)
> text(x=29.1, y=res1[25-17], "Color 1", adj=c(0,0))
> arrows(x0=23, res2[26-17],x1=26, y1=res2[26-17], length=.09)
> text(x=21.1, y=res2[26-17], "Color 2", adj=c(0,0))
> arrows(x0=28.9, res3[24-17], x1=24, y1=res3[24-17], length=.09)
> text(x=29, y=res3[24-17], "Color 3", adj=c(0,0))
```

> arrows(x0=25.9, res4[23-17], x1=23, y1=res4[23-17], length=.09)

> text(x=26, y=res4[23-17], "Color 4", adj=c(0,0))



5. Model Checking

a. Definition

• There are typically two test statistics for goodness of fit.

$$X^2 = \sum \frac{(\text{observed} - \text{fitted})^2}{\text{fitted}}$$

$$G^2 = 2 \sum (\log \text{ observed/fitted}).$$

• There is a lack of fit when X^2 or G^2 is too big.

b. Likelihood Ratio Model Comparison Tests

• The log likelihood-ratio statistic equals

$$l = -2(L_0 - L_1)$$

- L_1 = the maximized log of the likelihood function from a complex model, say M_1 .
- L_0 = the maximized log of the likelihood function from a simpler (nested) model, say M_0 .
- The goodness of model fit statistic G^2 is a special case of the likelihood ratio test statistic where
 - $-M_0=M$, the model we're testing.
 - $-M_1 = M_S$, the most complex model possible or the "saturated" model.
- For Generalized Linear Models such as Poisson and logistic regression, G^2 is equal to "deviance" of the model.
 - $-L_S = \text{maximized log of the likelihood function from the saturated model } M_S.$
 - $-L_0$ = maximized log of the likelihood function from the simpler model M_0 .
 - $-L_1$ = maximized log of the likelihood function from the complex model M_1 .

• We want to compare the fit of the model M_0 and M_1 .

deviance for
$$M_0 = G^2(M_0) = 2(L_0 - L_S)$$

deviance for $M_1 = G^2(M_1) = 2(L_1 - L_S)$

• The likelihood ratio statistic is

$$G^2(M_0|M_1) = G^2(M_0) - G^2(M_1)$$

• and

$$df = df_0 - df_1.$$

- Assuming that M_1 holds, this statistic tests
 - Whether the lack of fit of M_0 is significantly larger than that of M_1 .
 - Whether the parameters in M_1 that are not in M_0 equal zero.
- In R, Null Deviance = $2(L_N L_S)$ where L_N is the "Null Model" and
- Residual Deviance = $2(L_P L_S)$ where L_P is the "Proposed Model"
- The Null Model assumes one parameter for all of the data points, which means you only estimate 1 parameter, the intercept.
- The null deviance shows how well the response is predicted by the model with nothing but an intercept.
- The Proposed Model assumes you can explain your data points with p parameters + an intercept term, so you have p+1 parameters.
- The residual deviance shows how well the response is predicted by the model when the predictors are included.

Example: Horseshoe crab on width

- Using the Horseshoe crab on width example, we can test $H_0: \beta = 0$ vs $H_a: \beta \neq 0$ by doing the following.
- M_0 = Model with only an intercept, i.e. $logit(\pi(x) = \alpha)$
- $G^2(M_0) = 225.76$ with $df_0 = 172$.
- $M_1 = \text{Model}$ with an intercept and width, i.e. $logit(\pi(x) = \alpha + \beta x)$
- $G^2(M_1) = 194.45$ with $df_1 = 171$.
- $G^2(M_0|M_1) = 225.76 194.45 = 31.31$ with df = 172 171 = 1