

Analysis of long Non-Coding RNAs from RNA-seq Data of Leishmania-Infected Human Macrophages

Flavia Regina Florencio de Athayde, Flavia Lombardi Lopes

FMVA-Unesp

Abstract

Long non-coding RNAs (lncRNAs) are RNAs greater than 200 nucleotides in length, that accomplish a remarkable variety of biological functions. They function as inhibitors or activators of transcription/translation, but with no protein-coding capacity. Macrophages are the primary host cells of *Leishmania* spp., and constitute a first line of defense against these trypanosomatids responsible for the prevalent zoonotic disease, leishmaniasis. Little is known about the regulatory function of lncRNA in human cells harboring intracellular pathogens. We conducted an analysis using RNA-seq data to identify annotated lncRNAs and alterations in their expression in *L. amazonensis* and *L. major* infected macrophages, compared to macrophages exposed to latex beads, as a control for phagocytosis. The main cloud-computing server of Galaxy (usegalaxy.org) was used to align eleven datasets with paired-end reads (GSE-PRJNA290995) to the human genome (version 38) using hierarchical indexing for spliced alignment of transcripts (Hisat2 - Galaxy version 2.1.0). Transcriptome assembly was performed with StringTie (Galaxy version 1.3.4) using annotation Gencode (version 29) to identify transcripts in the data. Next, using StringTie merge (Galaxy version 1.3.4), we created a single assembly GTF file from each group. To characterize their coding potential, we used the software FEXible Extraction of Long non-coding RNAs (FEELnc), and featureCounts (version 1.6.3) was employed to estimate the number of candidate lncRNAs fragments in all paired-end libraries. Abundance of reads were used in differential expression analysis with DESeq2 (R version 3.5), results were filtered to 3107 known lncRNAs, of which 311 were differentially expressed between treatments with FDR-adjusted p-value<0.05 and fold change>2.0. Of 218 differentially expressed lncRNAs in macrophages infected with *L. amazonensis* versus control, 153 were upregulated and 65 were downregulated. In macrophages infected with *L. major*, we found 217 differentially expressed lncRNAs, 123 upregulated and 94 downregulated in macrophages, as a result of *L. major* infection. This study characterizes lncRNA expression signatures in macrophages following infection by *Leishmania* spp, and suggests a role for non-coding RNAs in immune response to *Leishmania* infection.

Funding: FMVA - UNESP