

Mass Spectrometry with M₃C

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1 Introduction

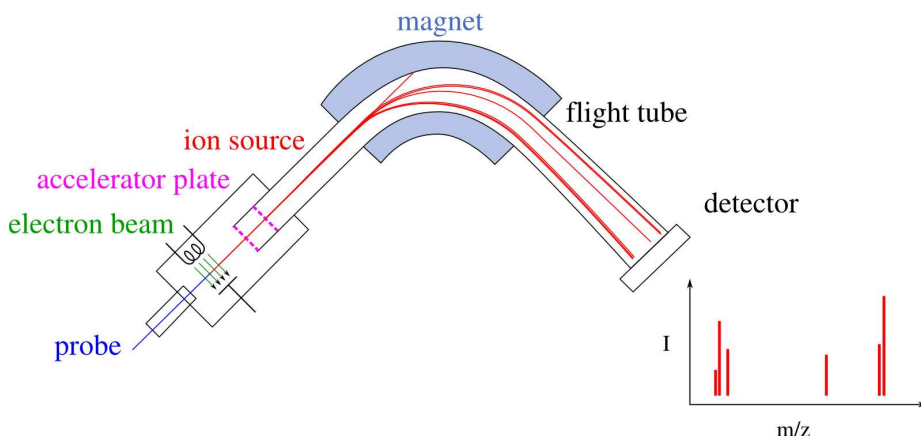


Figure 1: Schematic diagram of a mass spectrometer

A mass spectrometer consists of three components: an ion source, a mass analyzer, and a detector, as shown in Figure (1). The ionizer converts the sample (which may be solid, liquid, or gas) into ions by bombarding it with electrons (electron ionization). Only some of the collisions are energetic enough to knock one or more electrons out of the sample producing positive ions on the gas phase. This may cause some of the sample’s molecules to break into charged fragments. An extraction system removes ions from the sample, which are then trajected through the mass analyzer. The differences in masses of the fragments allows the mass analyzer to sort the ions by their mass/charge ratio, by accelerating them with an electric or magnetic field, until the fragments reach the detector. Results are displayed a spectrum of the relative abundance of detected ions as a function of the mass/charge ratio into a “stick diagram”. The atoms or molecules in the sample can be identified by correlating known masses to the identified masses or through a characteristic fragmentation pattern. In summary, the mass spectrum shows the mass of the ionized molecule and the masses of its corresponding ionic fragments.

When a highly energetic electron hits a neutral molecule, some of its energy is transferred to this molecule. If the transferred energy excess the *ionization energy* (IE) of the neutral molecule, then the ionization by ejection of one electron occurs, generating a molecular ion in an excited state.



This is the most desirable process. However there are several processes in competition that complicate this situation in practice. Some of them are shown in Figure (2). In principle, only unimolecular reactions are possible for the gaseous ions formed under the usual mass spectrometry operating conditions. As the energy of the electrons increases, the number of channels, abundance and variety of the ionized species will also increase, which gives rise to a fingerprint of the parent molecule’s spectrum.

Ionization of the sample molecules with 70 eV electrons produces molecular ions whose internal energy values (E) typically cover a broad range from 0 eV up to 20 eV. The nature and extent of these reactions depend only on the ion’s structure and internal energy irrespective of the ionization method.

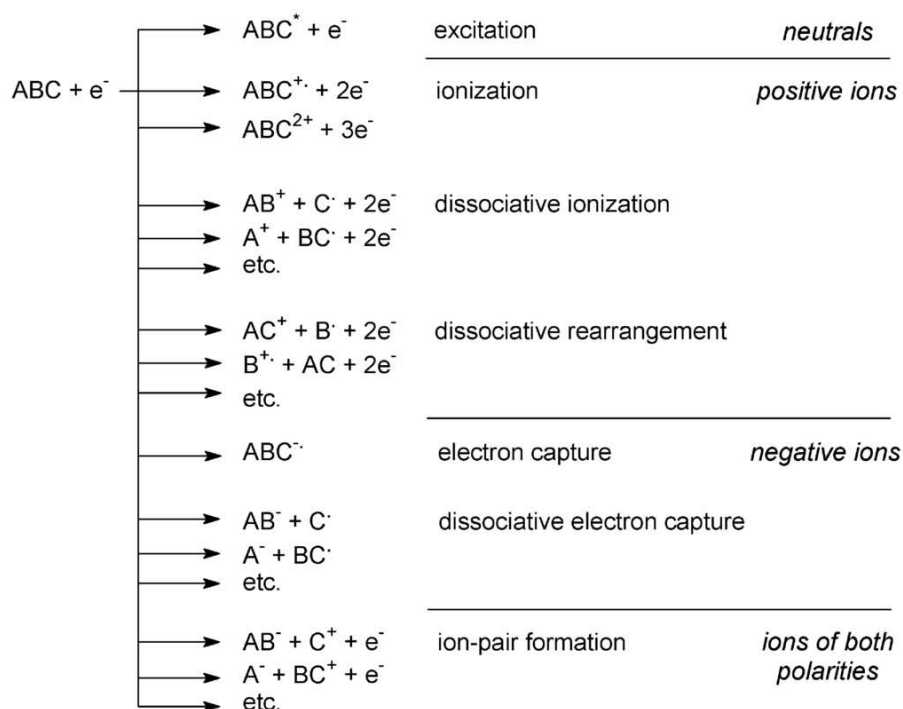


Figure 2: Processes under electron ionization conditions. Taken from: Jürgen H. Gross. Mass Spectrometry. A Textbook. Springer; 2nd ed. 2011 edition. Chapter 2, page 24.

The electron impact ionization of a molecule is a process which takes place in approximately 10^{-16} s and initially yields the excited molecular ion. The process is much more rapid than the time of one vibration, which is about 10^{-14} s. The distances between atoms thus do not change during the ionization. Thus, this ionization/excitation process can be seen as a vertical transition. After the ionization, the energy is distributed over the various molecule's degrees of freedom in a statistical fashion.

The fast exchange of internal energy occurs not only between the various degrees of freedom of the same electronic state but also between all the degrees of freedom of all the electronic states. These exchanges lead to the conversion of electronic energy acquired during ionization into vibrational and rotational energy of the ground electronic state of the molecular ion. It can be shown experimentally that the statistical energy distribution is carried out within a time span corresponding to a few vibrations, that is less than 10^{-10} s. Note that this time span is very short with respect to the time spent in the spectrometer source, at least 10^{-7} s. Then, fragmentation can be studied independently of the excitation process. Thus the probabilities of the various possible decompositions of an ion depend only on its structure and internal energy, and not on the method used for the initial ionization, or on the structure of the precursor for, or formation mechanism of, the ion undergoing decomposition. S. Weerasinghe *et al.* (*J. Chem. Phys.* **98** (1993) 4967) have shown that the dynamical evolution of a complicated many-body system is mainly guided by the accessible phase-space. Then, statistical mechanics provides the appropriate theoretical framework for conducting this kind of simulations.

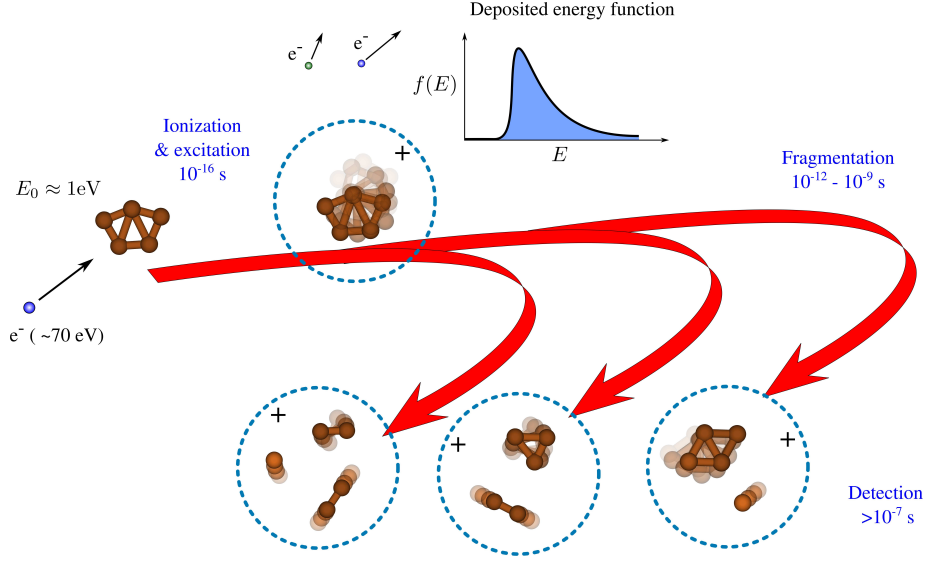


Figure 3: Schematic diagram of the fragmentation process induced by the electron impact ionization

2 Basics of Microcanonical Metropolis Monte-Carlo

The statistical Microcanonical Metropolis Monte-Carlo (M_3C) method is a theoretical approach that allows to describe the unimolecular decompositions of ions and hence their mass spectra. A better comprehension of the fragmentation mechanisms is the main goal. This theoretical description is based on the following premises:

1. There is no change of position or kinetic energy of the nuclei while the ionization and excitation processes take place (“vertical transition”).
2. The molecular ion will access to as many low-lying excited electronic states as necessary. Radiationless transitions then will result in transfer of electronic energy into vibrational, rotational or translational energy.
3. These low-lying excited electronic states will not be repulsive; hence, the molecular ions will not dissociate immediately, but rather remain together for a time long enough for the excess electronic energy to become randomly distributed over all internal degrees of freedom.
4. The deposited energy on the ion depends only on its structure and the experimental setup details. Thus, the probabilities of the different decomposition channels will not depend on the method used for the initial ionization.
5. The fragmentation channels are determined by the configuration of maximum entropy which is energetically accessible. It depends only on its structure and internal energy. Rearrangements of the ions would occur in the same fashion.

This description is focused on the fragmentation processes itself, irrespective of the excitation mechanism that leads to fragmentation. Furthermore the initial state of the system corresponds to an excited molecule where its excess of energy (E) is given by an unknown

energy deposited function $f(E)$, which contains all the information about the associated experimental details. The main information provided for the methodology developed in this work are the breaking-curves. Then, the mass spectrum can be obtained by summing the breaking-down curves over the distribution of internal energy imparted to the molecular ions by the electronic ionization process.

Let's do a short introduction of the statistical theory underlying this implementation. This tutorial is focused on how to get the mass spectra of two different molecules.

In the theory of thermodynamics several ensembles can be considered. A particular ensemble is defined by a set of magnitudes. In the microcanonical ensemble the physical system under study (atoms, molecules, clusters, spins...) has a fixed energy E .

In this ensemble, an isolated system at equilibrium is characterized by its microcanonical entropy, given by the Boltzmann's formula $\mathcal{S} = k_b \ln \Omega(E)$, where the number of accessible micro-states into a semiclassical description is proportional to the micro-canonical density of states (DOS),

$$\Omega(E) = \int d\mathbf{\Gamma} \delta[\mathcal{H}(\mathbf{\Gamma}) - E] \quad (2)$$

here $\mathcal{H}(\mathbf{\Gamma})$ represents the Hamiltonian of the system, and $\mathbf{\Gamma}$ its phase space, which consists of all the possible values of position and momentum variables. It is clear that the most important quantity in the microcanonical description is the DOS.

In our specific case, after some assumptions, Equation (2) can be factorized as follows:"

$$\Omega(E) \approx \frac{1}{\mathcal{N}} \sum_{i=1}^{N_c} \sum_{j=1}^{N_v} \sum_{k=1}^{\mathcal{N}} \Omega'(E, \mathbf{c}_i, \mathbf{E}_{v,ij}, \mathbf{R}_{ik}, \boldsymbol{\theta}_{ik}, \mathbf{J}_{ik}) \quad (3)$$

This means that the total DOS can be seen as an average of the **instantaneous DOS** (iDOS) $\Omega'(E, \boldsymbol{\mathcal{X}})$ which is a function of the sytem's **state vector**

$$\boldsymbol{\mathcal{X}} = (\mathbf{c}, \mathbf{E}_v, \mathbf{R}, \boldsymbol{\theta}, \mathbf{J}) \quad (4)$$

Here \mathbf{c} represents the composition of the system (number of molecules and their identity) and $\mathbf{E}_v, \mathbf{R}, \boldsymbol{\theta}, \mathbf{J}$ the vibrational energy, position (Cartesian coordinates of their centers of mass), orientation and angular momentum for the complete set of molecules or fragments.

The exact mathematical representation of $\Omega'(E, \boldsymbol{\mathcal{X}})$ is not important here, since the most important point that we have to keep in mind is how to generate the minimum number of state-vectors, in order to obtain a good approximation for the DOS according to the Equation (3). Here it is where we take advantage of the stochastic sampling methods. In particular, we use the Markov Chain Monte Carlo sampling algorithm (see Figure (4)).

The microcanonical average of a quantity $f(\mathbf{\Gamma})$ is expressed as,

$$\langle f(\mathbf{\Gamma}) \rangle = \frac{\int d\mathbf{\Gamma} f(\mathbf{\Gamma})}{\int d\mathbf{\Gamma}} \quad (5)$$

However, several components of the phase-space $\mathbf{\Gamma}$ can be integrated out, which allows to express this average in the space of the system's state-vectors as follows

$$\langle f(\boldsymbol{\mathcal{X}}) \rangle = \sum_{k=1}^N P(\boldsymbol{\mathcal{X}}_k) f(\boldsymbol{\mathcal{X}}_k), \quad (6)$$

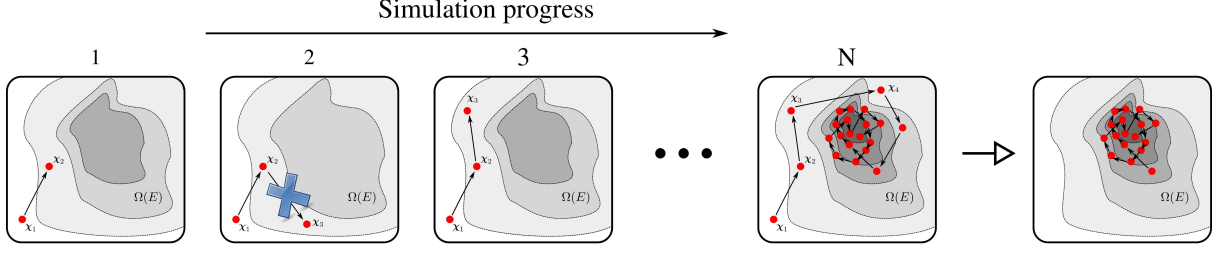


Figure 4: Schematic representation of the N -state Markov Chain sampling used to explore the state-vector space \mathcal{X} .

where the probability density of finding the system in the configuration \mathbf{x}_k is:

$$P(\mathbf{x}_k) = \Omega(E, \mathbf{x}_k) / \Omega(E) \quad (7)$$

This probability function can be used as the weighting factor in a microcanonical Markov chain Monte Carlo simulation to calculate averages according to equation (6). In this method we move in small steps $\{\mathbf{x}_1, \mathbf{x}_2, \dots, \mathbf{x}_k, \dots, \mathbf{x}_N\}$ (Markov chain) towards the most important region of the phase space, *i.e.* highest values for the $\Omega(E, \mathbf{x}_k)$. In the k th-step we generate a candidate \mathbf{x}_{k+1} which will be accepted or rejected depending of the acceptance ratio $p_E(\mathbf{x}_k \rightarrow \mathbf{x}_{k+1})$, given by

$$p_E(\mathbf{x}_k \rightarrow \mathbf{x}_{k+1}) = \min \left(1, \frac{P(E, \mathbf{x}_{k+1})}{P(E, \mathbf{x}_k)} \right) \quad (8)$$

It is important to highlight, that this expression for the acceptance ratio is specially convenient, because it does not depend on the normalization constant $\Omega(E)$. Then, the acceptance ratio can be simplified to

$$p_E(\mathbf{x}_k \rightarrow \mathbf{x}_{k+1}) = \min \left(1, \frac{\Omega(E, \mathbf{x}_{k+1})}{\Omega(E, \mathbf{x}_k)} \right) \quad (9)$$

At the end of the simulation, after the equilibration of the system (burn-in period), if we generate N randomly state-vectors (accepted or rejected) according to equation (8), expected values can be approximated by a simple arithmetic average, as follows

$$\langle f(\mathbf{x}) \rangle = \frac{1}{N} \sum_{k=1}^N f(\mathbf{x}_k), \quad (10)$$

where errors in $\langle f(\mathbf{x}) \rangle$ scale as $1/\sqrt{N}$. Figure (4) represents a schematic representation of this algorithm, note the removing of the burn-in period.

In summary, the theoretical description behind this method/implementation is a specific random way to move in the state-vectors space until a region of maximum entropy is reached, where the physical observables are measured by performing a statistical average in this region. In the current version of M₃C the available observables includes: channels/species distributions, energy components distributions, temperature and heat capacity among others. Additionally, by providing a deposited energy function $f(E)$ it is possible also to calculate channels' or species' branching ratios and the associated mass spectra.

3 Goals

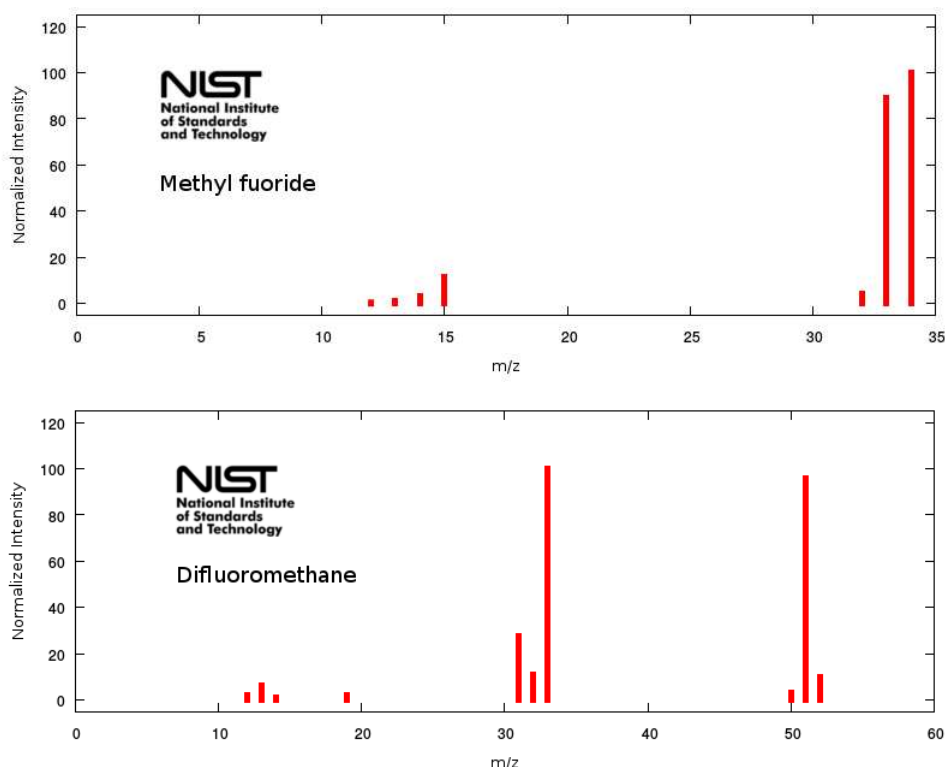


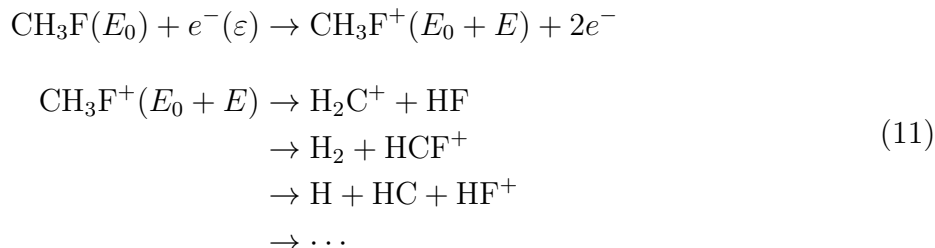
Figure 5: Data from NIST Standard Reference Database 69: NIST Chemistry WebBook. NIST Mass Spec Data Center, S.E. Stein, director, "Mass Spectra" in NIST Chemistry WebBook, NIST Standard Reference Database Number 69, Eds. P.J. Linstrom and W.G. Mallard, National Institute of Standards and Technology, Gaithersburg MD, 20899, <http://webbook.nist.gov>, (retrieved January 3, 2015).

In this tutorial we will show you how to build a mass spectrum from scratch. The minimum information that M3C needs is the electronic energy, molecular geometry and vibrational frequencies for all possible molecules (local minima) which are involved in the fragmentation process. As more molecules you consider, better results you will obtain. So, taking into account that the most expensive computational part corresponds to obtain these parameters, we will dedicate an important part of this tutorial to show, how to use the scripts provided by the M3C program, to carry out this task.

We have chosen two related systems to illustrate how M3C works: fluoromethane (also called methyl fluoride) CH₃F and difluoromethane CH₂F₂. The experimental mass spectra for these two molecules are shown on the Figure (5). The most remarkable difference between them is that in the second one, the molecular ion peak does not corresponds to the base peak (the most abundant ion), in contrast with the first one. After reading this tutorial you will be able to clarify the origin of this effect.

4 Example 1: Fluoromethane (guided tour)

Our main hypothesis is that fragmentation process occurs in two steps:



The first one corresponds to the electronic ionization, which leads to the associated cation with an energy excess E . E is distributed according to a specific energy deposited function $f(E)$ which we assume contains all information about the experimental conditions. The second one is the cation's fragmentation process itself. We suppose that the first step is much faster than the second one, therefore the measured fragmentation patterns when the fragments reach the detectors depend mainly on the fragmentation process. This means, simulating the mass spectrum for the CH_3F is equivalent to simulating the fragmentation process of its cation CH_3F^+ , convoluted by an energy deposited function $f(E)$.

The first step in our simulation is to get all geometries for the possible fragments and their isomers. First we will make a stochastic search by using a molecular electronic structure code, these calculations will be done by using a semi empirical method, due its high computational time consuming. Then, the first guess of molecular structures will be refined at DFT-B3LYP/6-311+G* level of theory. The vibrational frequencies will be obtained at the end by using the same level of theory. Once all structures and vibrational frequencies are available for all possible fragments, they will be used to build the M3C input file.

At the end of this document you will find a step-by-step summary without descriptions. We recommend you follow these steps first and then return to this document to understand their meaning.

4.1 Stochastic search for isomers

How many fragments can we get by the fragmentation of the CH_3F^+ molecule? This is a combinatorial problem which is equivalent to get the different combinations of the three elements {H,C,F} with repetitions (maximum one for C and F, and three for H). M3C offers an automatic way to calculate it by using the command `M3C.fragments`, as follows:

```
1 user@hostname$ M3C.fragments H3,C,F
2
3 H, C, F, H2, HC, HF, CF, H3, H2C, H2F, HCF, H3C, H3F, H2CF, H3CF
```

Fifteen possible fragments are obtained. Here, we could discard some of them by chemical arguments of stability or based on the peaks which appear in the experimental mass spectrum. This could be very important for molecular systems which consist of a vast number of particles, because search for isomers it is the most expensive computational part of this methodology. However, we are going to continue considering all possible combinations for this system.

Now, it is necessary to build trial geometries, one for each fragment. To do it you can use any of the free available molecular editors in the web, as **avogadro**¹, **molden**², **pymol**³, among others. Figure (6) shows the trial geometries we used. Note that these geometries do not necessarily correspond to stable molecules, these only are an initial guess. These geometries are stored in the directory `init`.

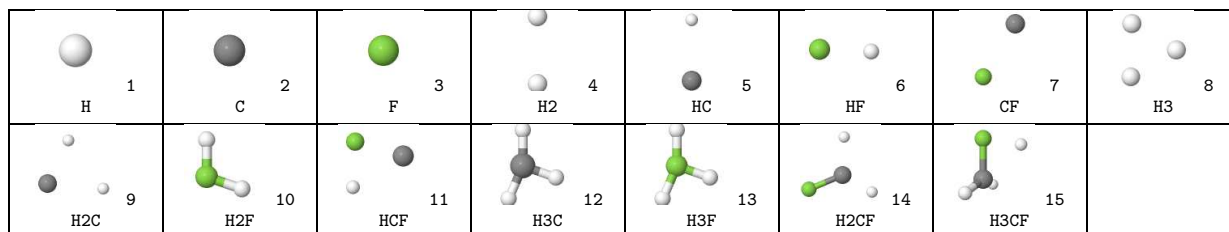


Figure 6: Trial geometries used to represent the possible fragments in the fragmentation of the CH_3F^+ molecule.

The procedure can begin with any structure for the fragments. It is submitted to GAMESS⁴ program optimization. The minimum energy structure obtained is then stored. The initial structure (without optimization) is then subject to an operation called a *kick*, each atom is moved a random distance in a random direction. The constraints are the maximum distance the atoms are going to be moved and the maximum radius allowed of the system R_{sys} (`systemRadius`), to generate a configuration where their atoms are non-overlapping. Each atom is kicked to a position within a sphere of radius R (`randomWalkStepRadius`) around its initial position, where R is the maximum kick distance. After all the atoms are randomly moved in this way, quantum mechanical optimization is carried out again. This algorithm is typically referred to as the **random walker algorithm**. Figure (7) shows an example of the trajectories obtained by a system of three particles after thousands steps.

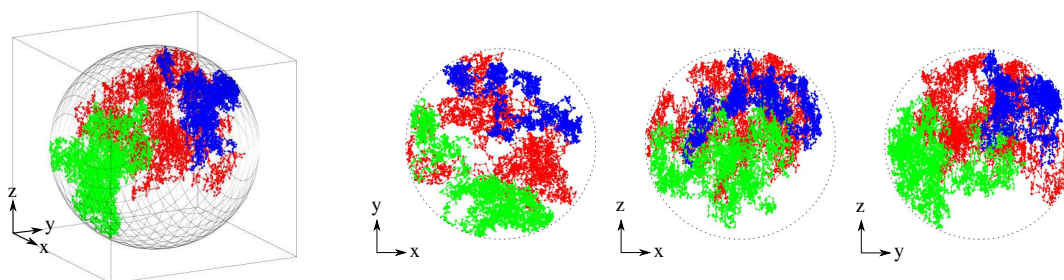


Figure 7: Example of the trajectories followed by 3 particles with different masses by using the random walker algorithm described in the text.

For each step, there are two possible results: the structure can go back to some previous state or it can go to a different structure. Then at the end of the algorithm, a filter removes duplicate isomers. If this procedure is repeated enough times, eventually all isomeric structures for the molecule will be found.

¹http://avogadro.cc/wiki/Main_Page

²<http://www.cmbi.ru.nl/molden/>

³<http://www.pymol.org/>

⁴<http://www.msg.ameslab.gov/gamess/>

The controlling parameter in the operation of stochastic searching is the size of the kick. Small kicks will result in return to the kicked isomer. It is easy to start with a small kick and gradually increase it to see when isomerization starts to occur with some reasonable probability. This probability will become larger with further increase of the kick size. With very large kicks, molecules often break into separate pieces. These pieces do not usually come back together into the optimization process to form bonds and the optimization usually stops at some point. With experience, one can fairly readily find the range of kick size, which gives a reasonable probability of isomerization and yet does not cause fragmentation to occur too frequently.

M3C offers a way to execute the above explained algorithm in an automatic way by interfacing with GAMESS through the command `M3C-gamess.geniso`. `M3C-gamess.geniso` requires three files as parameters: 1) A GAMESS template to control the geometry optimizations, 2) A M3C input file to control each step of the geometries' random search, and 3) and a file containing the charges, multiplicities and initial geometries to use. We will describe briefly each one:

- **GAMESS template.** First we need a GAMESS template for the optimization processes like the following. For each step, variables `@CHARGE`, `@MULT` and `@GEOMETRY` will be substituted by the corresponding charge, multiplicity and by the geometry block respectively. In this example geometry optimization is carried out at the PM3 semiempirical level.

Input File 1: GAMESS template for geometry optimization at PM3 level (pm3.optg-GAMESS.inp)

```

1  $contrl runtyp=optimize icharg=@CHARGE mult=@MULT $end
2  $basis gbasis=pm3 $end
3  $statpt projct=.f. nstep=50 $end
4  $system timlim=600000 memory=2500000 $end
5  $data
6  pm3
7  c1
8  @GEOMETRY
9  $end

```

- **M3C input file.** This input file will control the generation of the next non-overlapping geometry. The input file is divided in blocks, in the `GOPTIONS` block you can change the system radius and the maximum kick distance. The `REACTOR` block defines a geometric-translational operation (`type=T`). The reactor will read the geometry from `products.xyz`, it will modify it in a random way and it will save it by using the same file name. `FRAGMENTS_DATABASE` block defines the parts of the molecule that will be moved, in this case it will correspond to the atoms, however, as it will be shown later, it also can be molecules. M3C is case sensitive for input files, comments start with `#` and length units in angstroms.

Input File 2: M3C input file for random walker algorithm (reactorT.m3c)

```

1  BEGIN GOPTIONS
2      systemRadius = 2.0
3      overlappingRadius = 0.3
4
5      randomWalkStepRadius = 1.5
6      useRandomWalkers = TRUE
7  END GOPTIONS
8
9  BEGIN REACTOR
10     type = T
11
12     reactives = file:products.xyz

```

```

13      excitationEnergy = 10.0
14
15      geomProductsFile = products.xyz
16  END REACTOR
17
18  BEGIN FRAGMENTS_DATABASE
19      #-----
20      # Label      Z  M  L  SYM      geomFile      Elec
21      #              Angs      eV
22      #-----
23      C      0  1  0  0      C.xyz      0.000000
24      H      0  1  0  0      H.xyz      0.000000
25      F      0  1  0  0      F.xyz      0.000000
26      #-----
27  END FRAGMENTS_DATABASE

```

- **Configuration file.** This file presents a simple table format. The first column is the file with the initial trial geometry, the second one is the charge (@CHARGE) and the last one the multiplicity (@MULT). Each row represents an electronic configuration for a chosen stoichiometry as given in the XYZ file. In this file, we have included only fragments with charge up to one and the lowest multiplicity state, in order to obtain better results. It could include states with higher multiplicity.

Input File 3: Configuration file (fragments.inp)

```

1  #-----
2  # XYZfile  charge  mult
3  #-----
4  F.xyz      0      2
5  C.xyz      0      1
6  CF.xyz     0      2
7  H.xyz      0      2
8  HF.xyz     0      1
9  HC.xyz     0      2
10 HCF.xyz    0      1
11 H2.xyz     0      1
12 H2F.xyz    0      2
13 H2C.xyz    0      1
14 H2CF.xyz   0      2
15 H3.xyz     0      2
16 H3F.xyz    0      1
17 H3C.xyz    0      2
18
19 F.xyz      1      1
20 C.xyz      1      2
21 CF.xyz     1      1
22 H.xyz      1      0
23 HF.xyz     1      2
24 HC.xyz     1      1
25 HCF.xyz    1      2
26 H2.xyz     1      2
27 H2F.xyz    1      1
28 H2C.xyz    1      2
29 H2CF.xyz   1      1
30 H3.xyz     1      1
31 H3F.xyz    1      2
32 H3C.xyz    1      1
33 H3CF.xyz   1      2

```

Once the above files have been prepared, the command `M3C-gamess.geniso` can be executed as follows

```

1  user@hostname$ ls
2  CH3F+.m3c  fragments.inp  init
3
4  user@hostname$ M3C-gamess.geniso fragments.inp ../pm3.optg-GAMESS.inp ../reactorT.m3c 10 init results
5
6  Running:  F,    C,    CF,    H,    HF,    HC,    HCF,    H2    ... OK    Time elapsed: 0h 3m 59s
7  Running:  H2F, H2C, H2CF, H3,    H3F, H3C,    F,    C    ... OK    Time elapsed: 0h 13m 55s
8  Running:  CF,    H,    HF,    HC,    HCF,    H2,    H2F,    H2C    ... OK    Time elapsed: 0h 7m 49s
9  Running:  H2CF, H3,    H3F, H3C,    H3CF    ... OK    Time elapsed: 0h 13m 8s
10                                     Total: 0h 38m 51s
11
12 user@hostname$ ls
13 CH3F+.m3c  fragments.inp  init  results

```

In this example, ten random configurations have been generated for each stoichiometry, and all successful optimizations have been stored into directory **results**. Geometry files are coded with the format <label>.q<charge>.m<mult>.xyz. Total elapsed time was around forty minutes.

```

1 user@hostname$ cd results
2 user@hostname$ ls
3
4 CF.q0.m2-1.xyz   H3CF.q1.m2-1.xyz   H3F.q1.m2-6.xyz   history-C.q1.m2*   history-H3.q0.m2*
5 CF.q1.m1-1.xyz   H3CF.q1.m2-2.xyz   H3.q0.m2-1.xyz   history-F.q0.m2*   history-H3.q1.m1*
6 ...
7 H2.q0.m1-10.xyz  H3F.q1.m2-2.xyz   history-CF.q1.m1*  history-H3F.q0.m1*
8 H2.q1.m2-10.xyz  H3F.q1.m2-3.xyz   history-C.q0.m1*   history-H3F.q1.m2*

```

All optimized geometries can be easily visualized by using the command **M3C.viewXYZ** as follows

```

1 user@hostname$ M3C.viewXYZ
2 CF.q0.m2-1.xyz ... OK
3 CF.q1.m1-1.xyz ... OK
4 ...
5 H.q0.m2-1.xyz ... OK
6 H.q1.m0-1.xyz ... OK
7
8 user@hostname$ gwenview .

```

where one should get a diagram like the one shown in Figure (8).

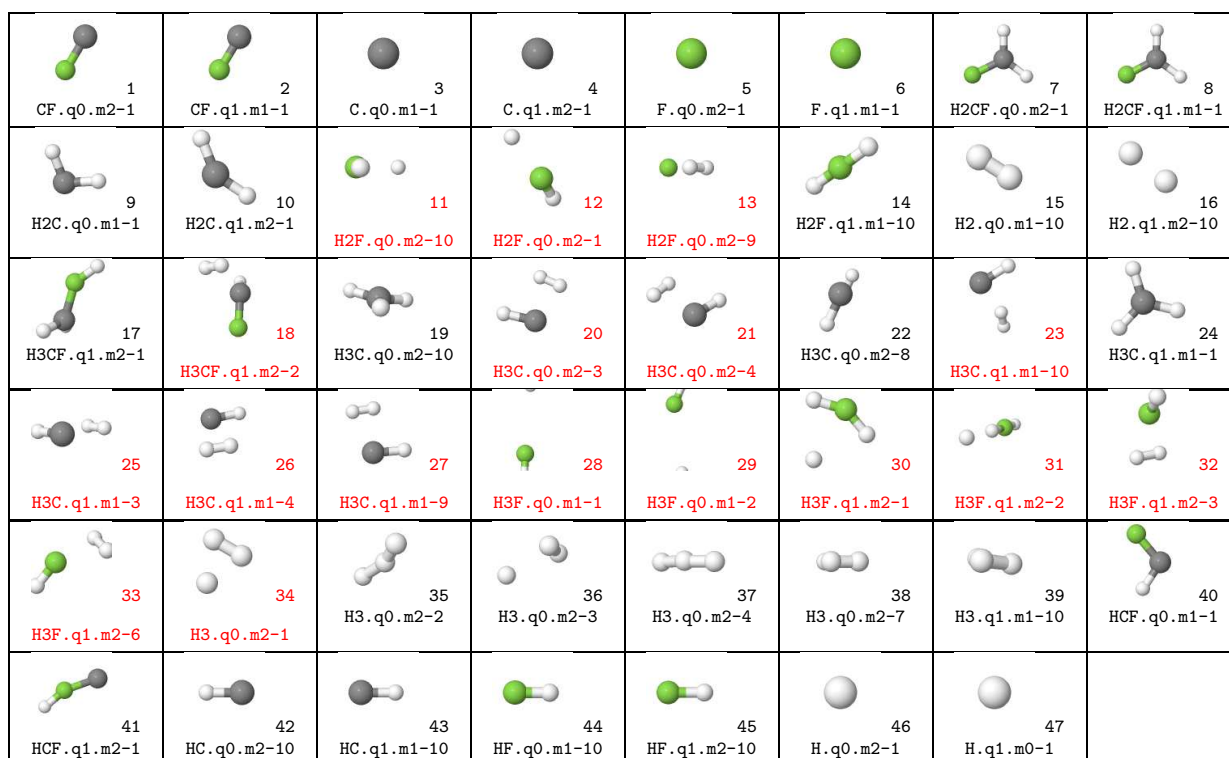


Figure 8: First set of molecules obtained by the random-walkers algorithm, as implemented in **M3C-gamess.geniso** command.

As it is possible to appreciate in Figure (8), there are molecules separated into two or more pieces. Such structures were not rejected by the program automatically. So, we have to

remove these molecules by hand. The molecules that have been removed appear highlighted in red. After that, we obtain the set of molecules shown in Figure (9).

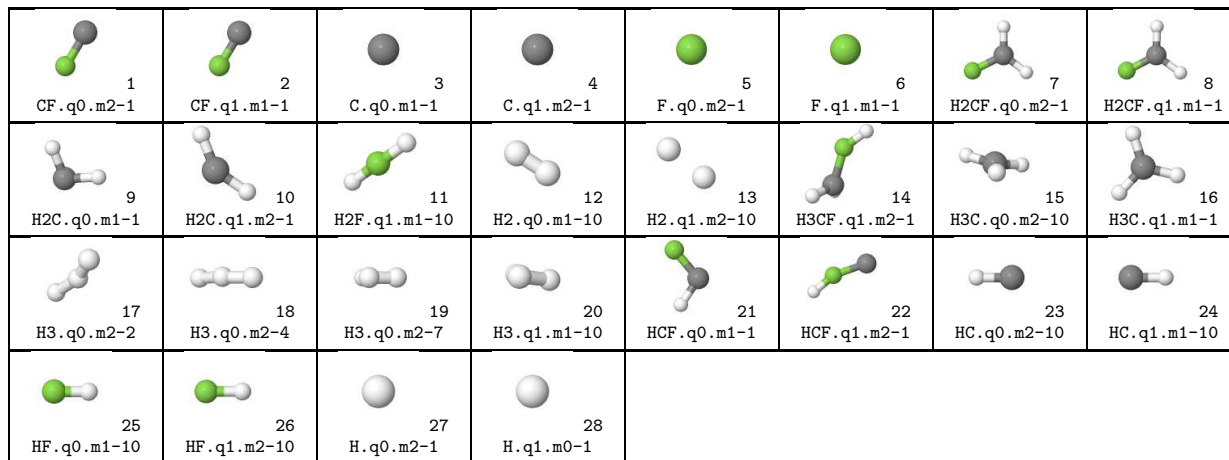


Figure 9: Filtered set of molecules obtained by the random-walkers algorithm with $N = 10$ trials, after removal of molecules separated into two or more pieces.

For each stoichiometry, we do not know how many isomers exist in advance. Then, one could increase the number of the steps in the random-walkers algorithm, in order to verify that the number of isomers not change. Specifically we use fifty steps with the following command,

```

1 user@hostname$ M3C-gamess.geniso fragments.inp ../pm3.optg-GAMESS.inp ../reactorT.m3c 50 init results
2
3 Running:  F,   C,   CF,   H,   HF,   HC,   HCF,   H2 ... OK   Time elapsed: 0h 23m 4s
4 Running:  H2F, H2C, H2CF, H3,   H3F, H3C,   F,   C ... OK   Time elapsed: 1h 21m 24s
5 Running:  CF,   H,   HF,   HC,   HCF,   H2, H2F, H2C ... OK   Time elapsed: 0h 41m 38s
6 Running: H2CF, H3,   H3F, H3C, H3CF ... OK   Time elapsed: 0h 57m 33s
7                                     Total: 3h 23m 39s

```

Total elapsed time was around three hours and a half (*i.e.* it scales approximately linearly). The final set of molecules we obtained after filtering are show in Figure 10. By using 50 steps, it has emerged one more isomer for the H2CF.q0.m2 and H2CF.q1.m1, H3CF.q1.m2 and HCF.q1.m2 configurations. We will continue our exercise taking this last set of molecules.

4.2 Geometry refinement

The semiempirical results provide just a preliminary overview of the interactions in the molecules. For this reason, the set of filtered molecules or local minima must be refined using a higher level of theory, in our case we employ DFT-B3LYP. M3C offers a way to do it automatically by interfacing with GAMESS through the command `M3C-gamess.optg`. `M3C-gamess.optg` requires one file as parameter: A GAMESS template to control the geometry optimizations. We use the following file for a geometry optimization at the B3LYP/6-311+G* level of theory.

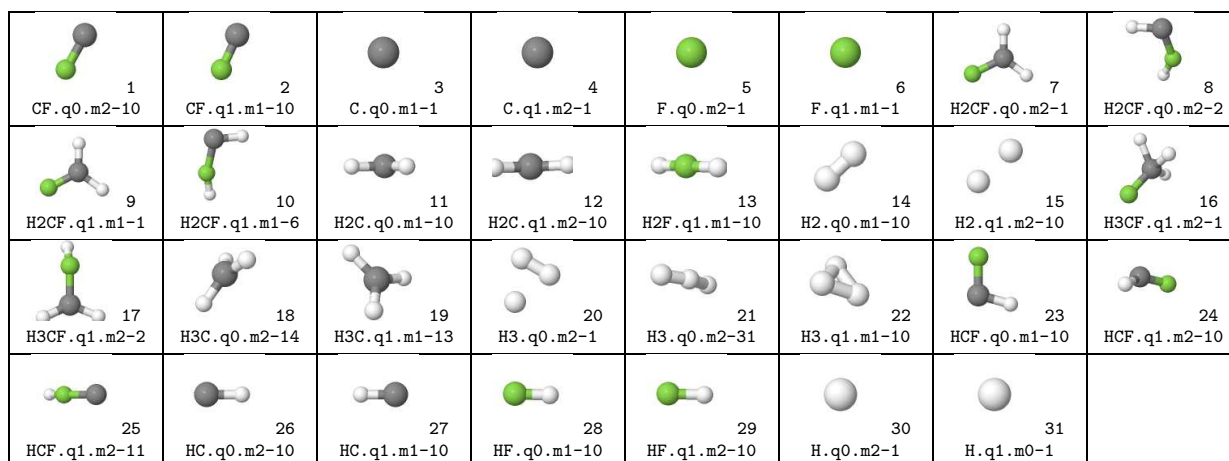


Figure 10: Filtered set of molecules obtained by the random-walkers algorithm with $N = 50$ trials, after removal of molecules separated into two or more pieces.

Input File 4: GAMESS template for geometry optimization at B3LYP level (b3lyp.optg-GAMESS.inp)

```

1 $contrl dfttyp=b3lyp runtyp=optimize
2   maxit=200 icharg=@CHARGE mult=@MULT $end
3 $basis gbas=N311 ngauss=6 diffsp=.T. ndfunc=1 $end
4 $statpt projct=.f. nstep=200 $end
5 $system timlim=600000 memory=2500000 $end
6 $data
7 b3lyp
8 c1
9 @GEOMETRY
10 $end

```

Then, the M3C-gamess.optg command is executed as follows:

```

1 user@hostname$ M3C-gamess.optg ../b3lyp.optg-GAMESS.inp
2
3 Running: CF.q0.m2-10, CF.q1.m1-10, ... , H2CF.q0.m2-1, H2CF.q0.m2-2 ... OK Time elapsed: 0h 1m 56s
4 Running: H2CF.q1.m1-1, H2CF.q1.m1-6, ... , H2.q1.m2-10, H3CF.q1.m2-1 ... OK Time elapsed: 0h 1m 37s
5 Running: H3CF.q1.m2-2, H3C.q0.m2-14, ... , HCF.q0.m1-10, HCF.q1.m2-10 ... OK Time elapsed: 0h 5m 50s
6 Running: HCF.q1.m2-11, HC.q0.m2-10, ... , H.q1.m0-1 ... OK Time elapsed: 0h 1m 38s
7                                     Total: 0h 11m 1s
8
9 user@hostname$ ls
10 CF.q0.m2-10.xyz C.q1.m2-1.xyz0 ... HCF.q1.m2-11.xyz0 HF.q1.m2-10.xyz
11 CF.q0.m2-10.xyz0 F.q0.m2-1.xyz ... HC.q0.m2-10.xyz HF.q1.m2-10.xyz0
12 ...
13 C.q0.m1-1.xyz0 H2CF.q0.m2-1.xyz ... HF.q0.m1-10.xyz H.q1.m0-1.xyz0
14 C.q1.m2-1.xyz H2CF.q0.m2-1.xyz0 ... HCF.q1.m2-11.xyz HF.q0.m1-10.xyz0

```

Total elapsed time is around ten minutes. The original geometry files are renamed with the extension .xyz0, and the refined geometries are saved with the extension .xyz, by substitution of the original ones. Again, there are some molecules that are separated in several fragments during B3LYP optimization. Consequently, these fragments have to be filtered again. Figure (11) shows the final obtained geometries.

In this case the molecule H2CF.q0.m2-2 and the two H₃ isomers H3.q0.m2-1 and H3.q0.m2-31 have disappeared because they have been fragmented in several pieces during the optimization process. The rest of the molecules have been kept qualitatively invariant.

4.3 Calculation of Vibrational Frequencies

M3C offers a way to do it automatically by interfacing with GAMESS through the command M3C-gamess.freqs. M3C-gamess.optg not requires one file as parameter: A GAMESS

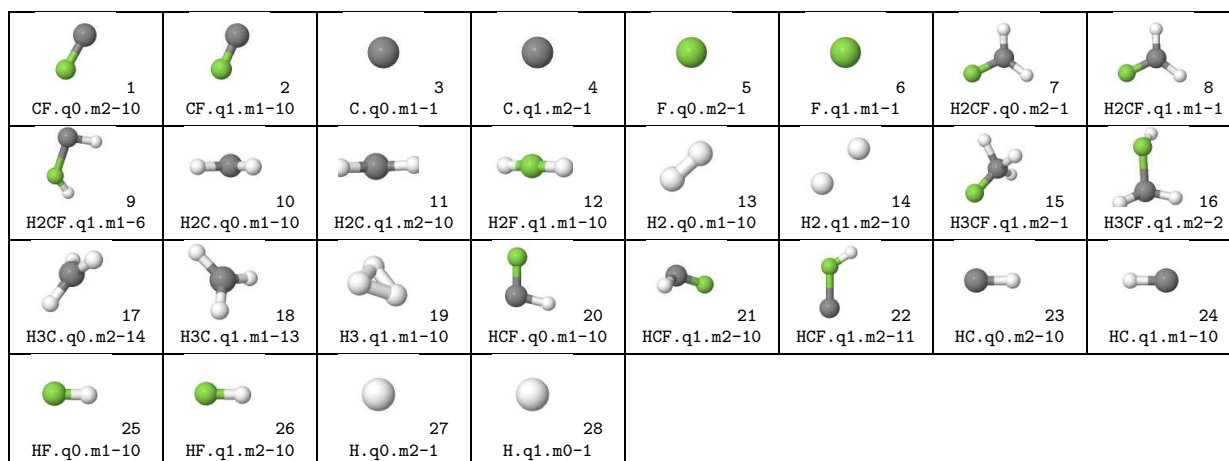


Figure 11: Refined geometries at B3LYP/6-311+G* level of theory. See Figure (10)

template to control the vibrational frequency calculation. We use the following file. Note that the frequencies are computed at the same level of theory as the one for the geometry optimization.

Input File 5: GAMESS template for frequency calculations at B3LYP level (b3lyp.freqs-GAMESS.inp)

```

1 $contrl dfttyp=B3LYP runtyp=hessian
2   maxit=100 icharg=@CHARGE mult=@MULT $end
3 $basis gbasis=N311 ngauss=6 diffsp=.T. ndfunc=1 $end
4 $system timlim=600000 memory=2500000 $end
5 $data
6 B3LYP
7 c1
8 @GEOMETRY
9 $end

```

Then, the `M3C-gamess.freqs` command is executed as follows:

```

1 user@hostname$ M3C-gamess.freqs ../../b3lyp.freqs-GAMESS.inp
2
3 Running:   CF.q0.m2-10,   CF.q1.m1-10,   ... , H2CF.q0.m2-1, H2CF.q1.m1-1 ... OK   Time elapsed: 0h 3m 24s
4 Running:   H2CF.q1.m1-6, H2C.q0.m1-10,   ... , H3CF.q1.m2-1, H3CF.q1.m2-2 ... OK   Time elapsed: 0h 8m 3s
5 Running:   H3C.q0.m2-14, H3C.q1.m1-13,   ... , HC.q0.m2-10, HC.q1.m1-10 ... OK   Time elapsed: 0h 3m 16s
6 Running:   HC.q1.m1-10, HF.q0.m1-10,   ...                                     ... OK   Time elapsed: 0h 0m 54s
7                                                    Total: 0h 15m 37s
8
9 user@hostname$ ls
10 CF.q0.m2-10.rxyz C.q1.m2-1.rxyz      ...   HC.q1.m1-10.rxyz H.q0.m2-1.rxyz
11 CF.q0.m2-10.xyz  C.q1.m2-1.xyz      ...   HC.q1.m1-10.xyz H.q0.m2-1.xyz
12 ...
13 C.q0.m1-1.xyz    F.q1.m1-1.xyz      ...   HC.q0.m2-10.xyz HF.q1.m2-10.xyz
14 C.q0.m1-1.xyz0   F.q1.m1-1.xyz0     ...   HC.q0.m2-10.xyz0 HF.q1.m2-10.xyz0

```

Total elapsed time is around fifteen minutes. Execution of the command generate the geometry files with the extension `.rxyz`. These files basically follow the same format than `.xyz` files, except that in the second line the value of the energy is given in atomic units (it is not only a simple comment!) and the calculated vibrational frequencies are added at the end of the file. The following is an example of the obtained `.rxyz` file for the molecule `H3CF.q1.m2-1`

```

1 user@hostname$ cat H3CF.q1.m2-1.xyz
2 5
3 Energy = -139.2622305495
4 C -0.9974192169 0.0295075834 -0.9233579316
5 F -1.8353908681 -0.3882844951 -1.8132708471
6 H -0.4374983257 0.9277189337 -1.1760712533
7 H -0.4136858960 -0.8796467265 -0.4806615335
8 H -1.4868576935 -0.0379892954 0.1345235654
9
10 FREQUENCIES 9
11 3202.42
12 2556.83
13 2210.40
14 1461.60
15 1285.89
16 1093.38
17 1060.52
18 967.56
19 725.60

```

The number of vibrational frequencies are automatically fixed with the right number of internal degrees of freedom ($3N - 6$ or $3N - 5$ for lineal molecules)

4.4 M3C execution

M3C can be executed by two different ways: 1) single-point energy calculation and 2) energy-scan calculation. We will describe both cases. However we will dedicate a first part to describe the input file and how to build it.

4.4.1 Input file description

The M3C input file consists of several text blocks:

- **GOPTIONS**

This block provides global control information for the calculation.

- **systemRadius**. Maximum system radius in angstroms R_{sys} .
- **overlappingRadius**. Around each fragment, we consider a semi-hard sphere of a given radius r (sum of covalent radii of their atoms). Hence, around each fragment there is a volume which is forbidden for all other fragments. This parameter controls the maximum overlapping of the fragments. Typical values range from 0.1 to 0.4 angstroms.
- **useRandomWalkers**. TRUE activates the use of random-walkers algorithm for sampling the configurational space. FALSE (default value) activates the completely random sampling search.
- **randomWalkStepRadius**. If **useRandomWalkers**=TRUE, this parameter modifies the maximum kick distance used in the random-walkers algorithm (given in angstroms).

- **ENERGY_RANGE**

- **grid**. Grid representing the excitation energies to be used. The format consists of three values: **<min energy>:<max energy>:<number of points>**. Energy values should be given in eV.

- **MARKOV_CHAIN**

- **task**. Allows to customize the Markov chain itself, by defining its irreducible part. It means that this irreducible part will be used cyclically up reach the chosen number of events (see **numberOfEvents** parameter). The format consists of several operations (reactors) separated by commas. Available operations are:
 - * **T**: Translational reactor. Changes the fragments' coordinates.
 - * **V**: Vibrational reactor. Changes the vibrational energies.
 - * **R**: Rotational reactor. Changes the rotational energies, by sampling new angular momentum values.
 - * **S:n:m**: Changes the chemical composition of the fragments. Parameters n and m represent the minimum and maximum values in the change of the number of fragments. For example, S:-1:1 will change the number of fragments in -1, 0 or 1.
- **burnInFraction**. Represents the burn-in period given in percentage of the chosen number of events (see **numberOfEvents** parameter)
- **reactives**. Label of the initial state. See **FRAGMENTS_DATABASE** block.
- **excitationEnergy**. Excitation energy given in eV. It will be taken into account when a single point calculation is carried out. Otherwise, it will be substituted internally by the appropriate value in the energy range which have been defined in the block **ENERGY_RANGE**.
- **tracking**. Track the calculation step-by-step through energy (**energy**), DOS values (**weight**) or neither of them (**none**). It is relevant only when a single-point calculation is carried out. See *output file* section for details.
- **numberOfExperiments**. Controls the number of replicas or experiments to perform. Each replica will consist of a different set of vibrational energies, angular momenta and electronic states which are chosen in a randomly way. At the end of the calculation, all observables will be reported with their errors which are estimated from the replicas' standard deviation.
- **historyFileFrequency**. Stores the calculated observables each **historyFileFrequency** steps.
- **energyHistoryFile**. File name where the track record of energy components will be saved.
- **weightHistoryFile**. File name where the track record of statistical weights (logarithm of the DOS) will be saved.
- **histogramFile**. File name where the histograms of several calculated observables will be saved.
- **FRAGMENTS_DATABASE** This block consists in a table that contains as many rows as number of molecules or fragments are going to be considered in the process. Each row in the table contains the following information:
 - *Label*. Represents a unique identifier for the molecule. The format is <group label>(<specifier label>). The program will sort the molecules in several

groups where each of those groups is identified by a **group label**. Additionally inside each group, each molecule is identified by a **specifier label**. This is specially advantageous to study observables which are to be discriminated by groups of molecules. For example: In a mass spectrum, a particular line represents the molecule A. However, this line is not a single signal but a superposition of signals produced by isomers or excited states of the same molecule A. In this sense, it is advantageous to label these isomers or excited states as A(s1), A(s2), A(tc), and so on, where the specifier label is arbitrary but useful for the user.

- *Charge (Z)* Assigns the charge of the molecule.
- *Multiplicity (M)* Assigns the multiplicity of the electronic state of the molecule.
- *Rotational symmetry number* Assigns the rotational symmetry number of the molecule. This is not relevant for this tutorial.
- *Geometry file* in RXYZ format, where coordinates are given in angstroms and frequencies in cm^{-1}
- *Electronic energy* given in eV
- *Maximum vibrational energy allowed.* This value is determined by the energy of the lowest transition state available, whereby the molecule can be breaking up, specifically by its energy barrier. This value may be written directly in the table (given in eV).

One simple way to estimate this value is to suppose that the reverse activation barrier is very small, then the maximum vibrational energy is equivalent to the difference between the electronic energy of the molecule and the electronic energy of the fragmentation products. In this case, you can write directly in the table, the chosen fragmentation channel. For example: A(s1)+B(st).

Generating this table may be too boring. So, you can use the command `M3C.makeDB`, to get a first version. The command reads the XYZ files available into the current directory, it will extract the relevant information and finally, it will print all this information in the right format.

The following is the M3C input file that we used to describe the fragmentation of CH_3F^+ molecule. It is important to point out that one row into the FRAGMENTS_DATABASE has been commented (H2Fp(s)). This is because, in particular this molecule shows two imaginary frequencies, indicating that the structure is a second order transition state and therefore it can not be included in our description.

Input File 6: M3C input file to describe the fragmentation of CH_3F^+ molecule (CH3F+.m3c)

```

1 BEGIN GOPTIONS
2   systemRadius = 8.0
3   overlappingRadius = 0.4
4
5   useRandomWalkers = FALSE
6   randomWalkStepRadius = 1.0
7 END GOPTIONS
8
9 BEGIN ENERGY_RANGE
10  grid = 0.0:30.0:91
11 END ENERGY_RANGE
12
13 BEGIN MARKOV_CHAIN

```

```

14 task = V,T,S:0,V,T,S:1:-1
15 burnInFraction = 0.1
16
17 reactivities = H3CFp(dt)
18 excitationEnergy = 10.0
19
20 tracking = energy
21 numberOfExperiments = 3
22 numberOfEvents = 20000
23 historyFileFrequency = 100
24
25 energyHistoryFile = energy.dat
26 weightHistoryFile = weight.dat
27 histogramFile = histogram.dat
28 END MARKOV_CHAIN
29
30 BEGIN FRAGMENTS_DATABASE
31 #-----
32 # Label Z M L SYM geomFile Eelec maxVib
33 #-----
34 H(d) 0 2 0 1 H.q0.m2-1.rxyz -13.572100
35 C(s) 0 1 0 1 C.q0.m1-1.rxyz -1027.790000
36 F(d) 0 2 0 1 F.q0.m2-1.rxyz -2713.690000
37 H2(s) 0 1 0 1 H2.q0.m1-10.rxyz -31.833900 H(d)+H(d)
38 HC(d) 0 2 0 1 HC.q0.m2-10.rxyz -1046.730000 H(d)+C(s)
39 HF(s) 0 1 0 1 HF.q0.m1-10.rxyz -2732.950000 H(d)+F(d)
40 CF(d) 0 2 0 1 CF.q0.m2-10.rxyz -3748.880000 C(s)+F(d)
41 H2C(s) 0 1 0 1 H2C.q0.m1-10.rxyz -1064.370000 H(d)+HC(d)
42 HCF(s) 0 1 0 1 HCF.q0.m1-10.rxyz -3765.840000 H(d)+CF(d)
43 H3C(d) 0 2 0 1 H3C.q0.m2-14.rxyz -1083.470000 H2(s)+HC(d)
44 H2CF(d) 0 2 0 1 H2CF.q0.m2-1.rxyz -3783.830000 H2(s)+CF(d)
45
46 Hp 1 0 0 1 H.q1.m0-1.rxyz 0.000000
47 Cp(d) 1 2 0 1 C.q1.m2-1.rxyz -1018.080000
48 Fp(s) 1 1 0 1 F.q1.m1-1.rxyz -2692.420000
49 H2p(d) 1 2 0 1 H2.q1.m2-10.rxyz -16.298600 Hp+H(d)
50 HCp(s) 1 1 0 1 HC.q1.m1-10.rxyz -1035.860000 H(d)+Cp(d)
51 HFp(d) 1 2 0 1 HF.q1.m2-10.rxyz -2716.870000 Hp+F(d)
52 CFp(s) 1 1 0 1 CF.q1.m1-10.rxyz -3739.460000 Cp(d)+F(d)
53 H3p(s) 1 1 0 1 H3.q1.m1-10.rxyz -35.977900 Hp+H2(s)
54 H2Cp(d) 1 2 0 1 H2C.q1.m2-10.rxyz -1054.540000 H2(s)+Cp(d)
55 # H2Fp(s) 1 1 0 1 H2F.q1.m1-10.rxyz -2737.320000 0.000
56 HCFp(dC) 1 2 0 1 HCF.q1.m2-10.rxyz -3755.850000 H(d)+CFp(s)
57 HCFp(dF) 1 2 0 1 HCF.q1.m2-11.rxyz -3753.040000 H(d)+CFp(s)
58 H3Cp(s) 1 1 0 1 H3C.q1.m1-13.rxyz -1073.720000 H(d)+H2Cp(d)
59 H2CFp(st) 1 1 0 1 H2CF.q1.m1-1.rxyz -3774.770000 H2(s)+CFp(s)
60 H2CFp(s) 1 1 0 1 H2CF.q1.m1-6.rxyz -3770.520000 H(d)+HCFp(dC)
61 H3CFp(dt) 1 2 0 1 H3CF.q1.m2-1.rxyz -3789.520000 H(d)+H2CFp(st)
62 H3CFp(d) 1 2 0 1 H3CF.q1.m2-2.rxyz -3789.280000 H(d)+H2CFp(st)
63 #-----
64 END FRAGMENTS_DATABASE

```

4.4.2 Single-point-energy calculation

First we are going to do a single-point-energy calculation. The excitation energy is that provided in the input file (see variable `excitationEnergy`, 10 eV). M3C is executed with the following command:

```

1 user@hostname$ M3C -i CH3F+.m3c > CH3F+.out
2 user@hostname$ cat CH3F+.out
3 ...
4 +-----+
5 | BEGIN MOLECULE DATABASE INITIALIZATION |
6 +-----+
7 ...
8         file name = H3C.q0.m2-14.rxyz
9         name      = H3C(d)
10 Moments of inertia = [ 1.76658 1.76695 3.53353 ] amu*angs**2
11         Radius   = 1.30619 A
12         Eelec    = -1083.470000 eV
13         Mass     = 15.0349997 amu
14         (fr, fv) = ( 3 6 )
15         maxEvib  = 4.9061000 eV
16 ...
17 +-----+
18 | END MOLECULE DATABASE INITIALIZATION |
19 +-----+
20
21 +-----+
22 | MARKOV CHAIN |
23 +-----+
24

```

```

25
26         reactives = H3CFp(dt)
27         excitationEnergy = 5.00000 eV
28         numberOfEvents = 20000
29         numberOfExperiments = 3
30         task = V,T,S:0,V,T,S:1:-1
31         geometryHistoryFilePrefix = geom
32         freqBlockingCheck = 4
33         track = energy
34
35 # -----
36 # ENERGY HISTORY
37 # -----
38 #
39 #          trans          elec          vib          rot          tot          formula
40 #          eV           eV           eV           eV           eV
41 #          -----
42 eV          0.48928      -3787.41000      1.82981      0.57091      -3784.52000      H3Cp(s)+F(d)
43 pT          1.19822      -3787.41000      1.68792      0.00387      -3784.52000      H3Cp(s)+F(d)
44 pT          1.13463      -3787.49000      1.70110      0.13427      -3784.52000      H2Cp(d)+HF(s)
45 aV          0.20951      -3788.34210      2.26303      1.34957      -3784.52000      H(d)+H2CFp(st)
46 ...
47
48 # -----
49 # Channels histogram
50 # -----
51 #
52 #          item          1          2          3          aver          desv
53 #          -----
54 H2Cp+HF          0.043      0.022      0.030      0.032      0.009
55 H3Cp+F          0.957      0.978      0.965      0.967      0.009
56 H+H2CFp          0.000      0.000      0.005      0.002      0.002
57
58 #          item          1          2          3          aver          desv
59 #          -----
60 H(d)+H2CFp(st)      0.000      0.000      0.005      0.002      0.002
61 H3Cp(s)+F(d)        0.957      0.978      0.965      0.967      0.009
62 H2Cp(d)+HF(s)       0.043      0.022      0.030      0.032      0.009
63
64 # -----
65 # Species histogram
66 # -----
67 #
68 #          item          1          2          3          aver          desv
69 #          -----
70 F          0.479      0.489      0.482      0.483      0.004
71 HF         0.021      0.011      0.015      0.016      0.004
72 H2CFp      0.000      0.000      0.003      0.001      0.001
73 H          0.000      0.000      0.003      0.001      0.001
74 H2Cp       0.021      0.011      0.015      0.016      0.004
75 H3Cp       0.479      0.489      0.482      0.483      0.004
76
77 #          item          1          2          3          aver          desv
78 #          -----
79 H(d)        0.000      0.000      0.003      0.001      0.001
80 H2CFp(st)   0.000      0.000      0.003      0.001      0.001
81 H3Cp(s)     0.479      0.489      0.482      0.483      0.004
82 F(d)        0.479      0.489      0.482      0.483      0.004
83 HF(s)       0.021      0.011      0.015      0.016      0.004
84 H2Cp(d)     0.021      0.011      0.015      0.016      0.004
85
86 # -----
87 # Temperature (eV)
88 # -----
89 #
90 #          1          2          3          aver          desv
91 #          -----
92 0.179      0.175      0.177      0.007      0.000
93
94 # -----
95 # Markov chain statistics
96 # -----
97 # Reactor type (ACCEPTED)
98 S:1:-1      0.00078
99 T          0.81750
100 V          0.17265
101 S:0        0.00907
102
103 # Reactor type (REJECTED)
104 S:1:-1      0.28086
105 V          0.44400
106 S:0        0.27514
107
108 # Reactor status
109 a.ACCEPTED      0.03695
110 e.REJECTED(E<0)   0.22360
111 p.ACCEPTED(p<PI) 0.37082
112 r.REJECTED        0.36863
113 ...

```

In the above frame, the main parts of the output file are also shown. First the program shows details about each molecule that have been loaded from the FRAGMENTS_DATABASE

block (Lines 5-22). The same is done for block `MARKOV_CHAIN` (Lines 24-35). Then, details of the simulation steps are shown (Lines 37-47). Here, because the variable `tracking` was chosen as energy, this block shows the energy components of the system for each step of the simulation, actually, each 100 steps (see `historyFile` frequency variable). Otherwise, if `tracking = weight`, the statistical-weights will be reported. After line 48, several calculated observables are shown. Among them, the probability for each fragmentation channel, by grouping them according with their group label (lines 52-57) and without it (lines 59-64). The probability for each specie is also shown in the same fashion (lines 69-77 and lines 79-87).

The results described above can be easily analyzed by using the `M3C.analysis` command. For example: We may generate a plot by typing the following command, to analyze the energy components through the simulation. See Figure (12).

```
1 user@hostname$ M3C.analysis CH3F+.m3c energy
```

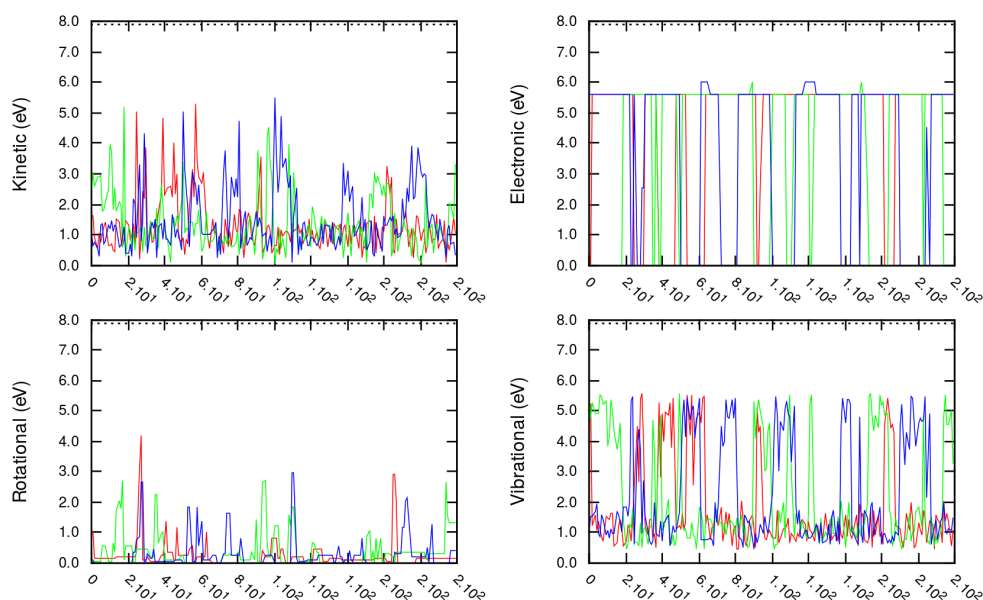


Figure 12: Energy components through the Markov chain, for the three numerical experiments. Each experiment is represented by different color.

It may also be possible to display histograms which correlate the energy components, by using the following commands See Figure (13).

```
1 user@hostname$ M3C.analysis CH3F+.m3c ecorr Et.vs.Ev
2 user@hostname$ M3C.analysis CH3F+.m3c ecorr Er.vs.Ev
3 user@hostname$ M3C.analysis CH3F+.m3c ecorr Er.vs.Et
```

It may also be possible to display histograms with the probabilities of the channels or fragments, by using the following commands. See Figure (14))

```
1 user@hostname$ M3C.analysis CH3F+.m3c species
2 user@hostname$ M3C.analysis CH3F+.m3c channels
```

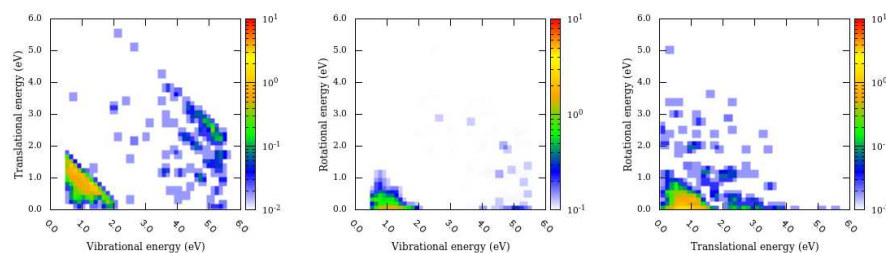


Figure 13: Histogram that represents the correlation between some energy component pairs.

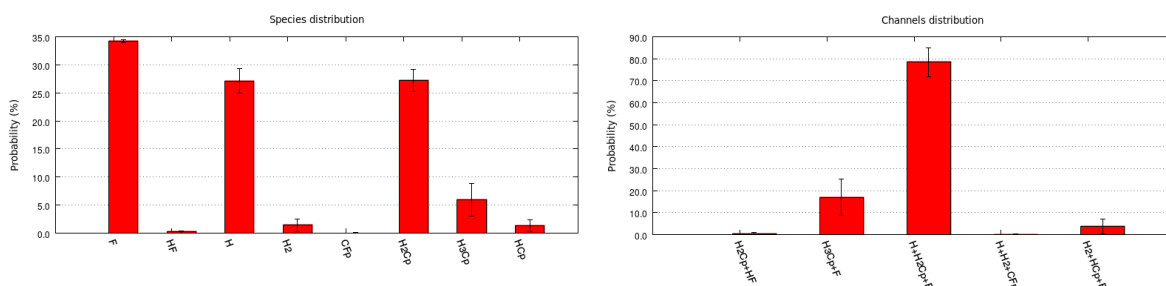


Figure 14: Species/channels probabilities

4.4.3 Scan-energy calculation

To carry out a scan-energy calculation, M3C provides the command `M3Cp`. `M3C.p` will perform as many calculations as energy values have been defined in `ENERGY_RANGE` block. `M3C.p` is executed as follows

```

1 user@hostname$ M3C.p -i CH3F+.m3c -n 8
2
3 Running: 0.00000, 0.33333, ... , 2.00000, 2.33333 ... OK Time elapsed: 0h 0m 32s
4 Running: 2.66667, 3.00000, ... , 4.66667, 5.00000 ... OK Time elapsed: 0h 0m 56s
5 ...
6 Running: 26.66667, 27.00000, ... , 28.66667, 29.00000 ... OK Time elapsed: 0h 2m 1s
7 Running: 29.33333, 29.66667, ... ... OK Time elapsed: 0h 1m 22s
8 Total: 0h 15m 49s

```

Total elapsed time is around of fifteen minutes. Execution of the command generates the directory `CH3F+.data`, which contains a lot of irrelevant information, because it will be handled by the `M3C.analysis` program. For example, the following commands produce Figure (15), which represents the probabilities for all channels and species/fragments in the fragmentation process as a function of the internal energy. The identity for each channel or fragment have been omitted for clarity.

```

1 user@hostname$ M3C.analysis CH3F+.m3c C.vs.E
2 user@hostname$ M3C.analysis CH3F+.m3c S.vs.E

```

4.4.4 Mass spectrum calculation

Having reached this point, it is important to point out, that in order to build the mass spectrum of this molecule, the only result we need from the above theoretical description

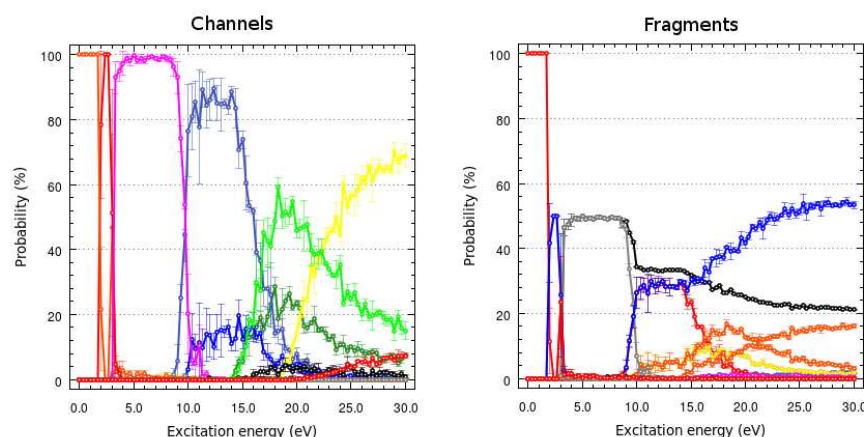


Figure 15: Species/channels probabilities as a function of the excitation energy.

is the breaking curve including all generated positive ions. This curve may be obtained by typing the next command, which selects only singly charged fragments. See upper panel in Figure (16).

```
1 user@hostname$ M3C.analysis CH3F+.m3c S.vs.E "p"
```

The next step is getting a deposited energy function from somewhere.

From a theoretical point of view, there are some ways to obtain this function, for example by carrying out stopping power calculations (See for example J. Postma, *et. al.*, *ApJ* **708** (2010) 435), however this kind of methodologies are too computationally expensive and normally infeasible for most molecules. There are some approximations where valence electrons in molecule are seen as an electron gas, thus the transferred energy by the electronic projectile can be expressed as a function of an effective friction coefficient which it depends of the electronic density of the molecule [See for example Schlathölder *et. al.* *Phys. Rev. Lett.* **82** (1999) 73]. This is a matter we will not treat here. However, if you have a deposited energy function, you may use it to get the mass spectrum based on the M3C breaking curves. This is already implemented.

From the experimental point of view, a variety of methodologies have been employed to estimate this function, for example, processing results of coincident two-electron energy analysis, from photo electron spectra or from breakdown graphs [See for example G. G. Meisels *et. al.* *J. Chem. Phys.* **56** (1972) 793]. When breakdown graphs are used, the incident electron is varied to effect changes in excitation energy, and the relative abundance of fragment ions at each energy which is measured. Breakdown graphs so obtained are multiplied by a series of assumed energy deposition functions. The function which gives the best fit to the mass spectrum observed with 70 eV electrons is then assumed correct one. This approach does not give unique results since different shapes will give almost indistinguishable results.

In this tutorial we will use a similar strategy like that used in experiments which is based on the breaking curves. Our advantage is that we do not need several trials by varying the electron projectile, because we already have the breaking curves.

The strategy we are going to use is take the percentage for each ion from the experimental mass spectrum and use them to estimate the best deposited energy function that fits with. Its shape is the only criteria we may use to decide if the obtained function is right or not. We hope obtaining a broad distribution which it has only one maximum and covering a range of energies between 0 up to 20 eV. Mathematical details about fitting process will not be given here.

The input file for the fitting process is as follows, which should be added at the end of the main input file.

Input File 7: Input file blocks to configure the fitting method

```

1 BEGIN EXPERIMENTAL_BRANCHING_RATIOS
2   error = absolute
3   diagram = S.vs.E
4
5   # -----
6   # Fragment      Intensity  error    m/z
7   # -----
8       Cp          0.5       0.0     12.0
9       HCp         1.2       0.0     13.0
10      H2Cp         3.1       0.0     14.0
11      H3Cp        11.6       0.0     15.0
12      CFp          7.7       0.0     31.0
13      HCFp         4.4       0.0     32.0
14      H2CFp        88.9       0.0     33.0
15      H3CFp       100.0       0.0     34.0
16      H2p           0.0       0.0      2.0
17      HFp           0.0       0.0     20.0
18      H3p           0.0       0.0      3.0
19   # -----
20 END EXPERIMENTAL_BRANCHING_RATIOS
21
22 BEGIN FIT_BRANCHING_RATIOS
23   method = NNLS
24   basis = 60,60
25   eDistfile = edist.out
26   BRfile = fitBR.out
27 END FIT_BRANCHING_RATIOS

```

Basically, it contains the peaks' size from experimental mass spectrum. Figure (5). Finally, mass spectrum for the CH_3F molecule and its deposited energy function can be visualized by typing the following commands. See Figure (16).

```

1 user@hostname$ M3C.analysis CH3F+.m3c fit_sfE
2 user@hostname$ M3C.analysis CH3F+.m3c fit_sBR

```

The deposited energy function for the CH_3F molecule shows a sharp peak around 2 eV and it drops off markedly in intensity at higher internal energies (see Figure 16). The breakdown graph for this molecule shows the molecular ion (CH_3F^+) as the dominant one up to 2 eV of internal energy and with the H_2CF^+ fragment ion dominating from 2 eV to 3 eV. This is the reason why these two ions are the highest peaks of the spectrum. The next peak in intensity which corresponds to the ion H_3C^+ , due to its breakdown curve which presents a broad band extending from 3 eV up to 10 eV that compensates the low contribution from the deposited energy distribution in this region. The rest of the ions do not contribute in a significant way.

As it is possible to appreciate, we found a relatively good agreement with the experiment. However the associated peak to H_2CF^+ is underestimated. It is a consequence that its probability distribution (Figure 16, upper panel, magenta line) is not enough wide, because the H_3C^+ molecule gets more stable above 4 eV. It may be due to the low level of theory we use in the electronic structure calculations or to the reduced number of excited states we use

to represent the different fragments. These are the two possible ways that one may follow to improve these results.

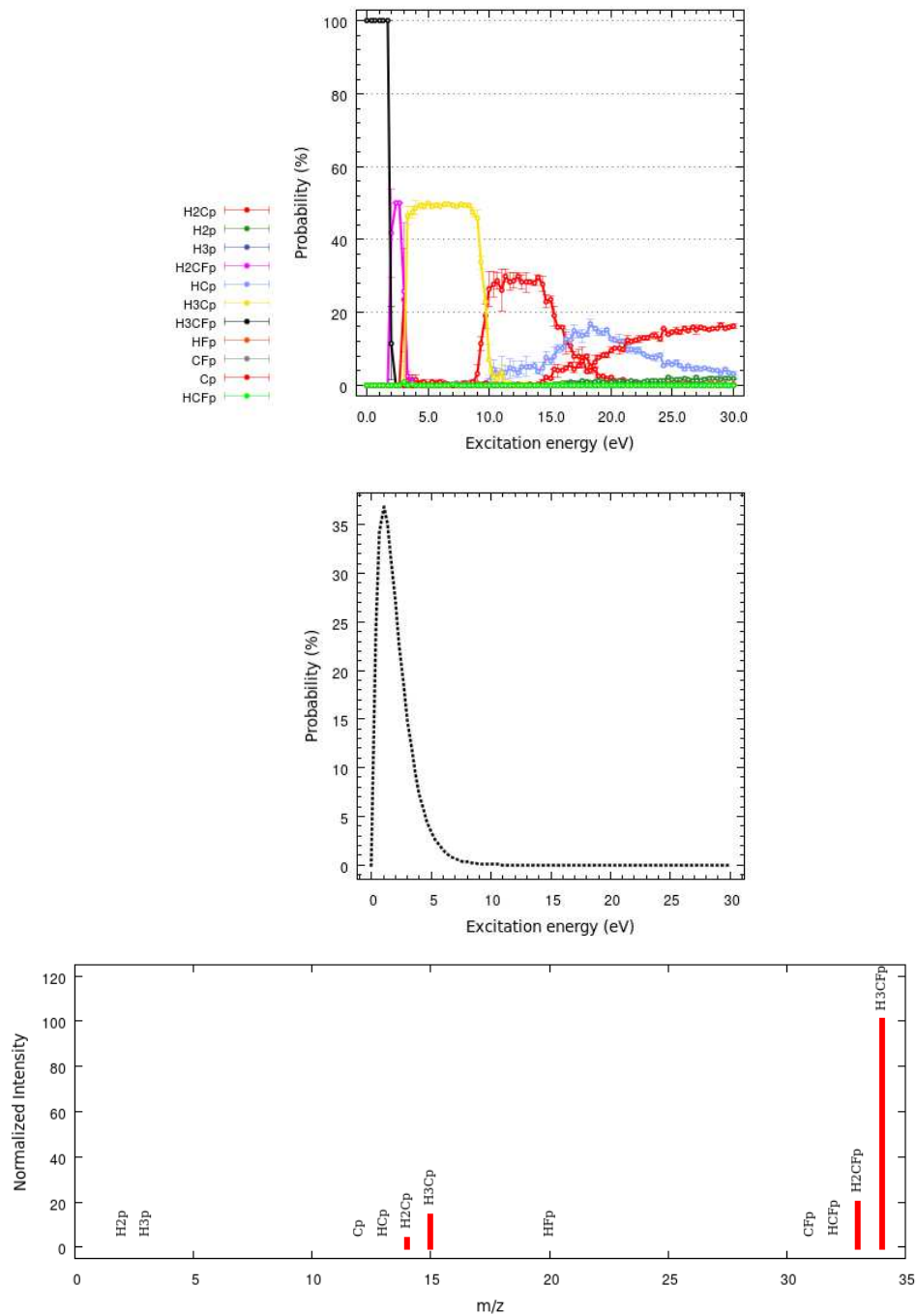
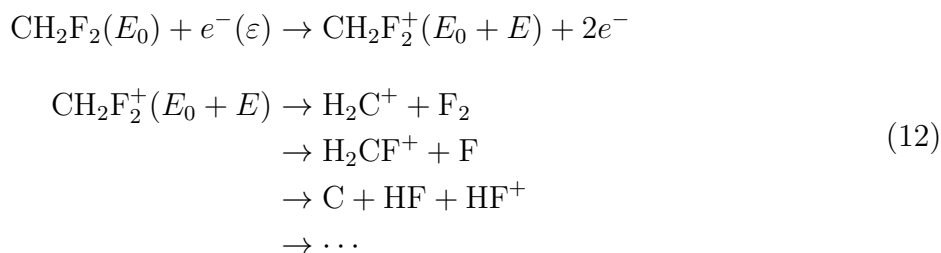


Figure 16: Upper panel, probabilities of singly-charged fragments (breakdown curves). Middle panel, fitted deposited energy function. Lower panel, theoretical mass spectrum for fluoromethane CH_3F .

5 Example 2. Difluoromethane

As in the previous example, our main hypothesis is that fragmentation process occurs in two steps:



This means, simulating the mass spectrum for the CH_2F_2 is equivalent to simulating the fragmentation process of its cation CH_2F_2^+ , convoluted by an energy deposited function $f(E)$.

First, how many fragments can we get by the fragmentation of the CH_2F_2^+ molecule?

```
1 user@hostname$ M3C.fragments H2,C,F2
2
3 H, C, F, H2, HC, HF, CF, F2, H2C, H2F, HCF, HF2, CF2, H2CF, H2F2, HCF2, H2CF2
```

As it is possible to appreciate, there are 17 possible fragments. However, the good news is that we already calculated eleven of these in the previous example. Then we have to search for isomers only for F_2 , HF_2 , CF_2 , H_2F_2 , HCF_2 and H_2CF_2 . Then, here we go

```
1 user@hostname$ ls
2 CH3F+.m3c fragments.inp init
3
4 user@hostname$ M3C-gamess.geniso fragments.inp ../pm3.optg-GAMESS.inp ../reactorT.m3c 10 init results
5
6 Running: CF2, F2, H2F2, HCF2, HF2, CF2, F2, H2CF2 ... OK Time elapsed: 0h 10m 48s
7 Running: H2F2, HCF2, HF2 ... OK Time elapsed: 0h 5m 3s
8 Total: 0h 15m 51s
9
10 user@hostname$ M3C-gamess.geniso fragments.inp ../pm3.optg-GAMESS.inp ../reactorT.m3c 50 init results
11
12 Running: CF2, F2, H2F2, HCF2, HF2, CF2, F2, H2CF2 ... OK Time elapsed: 0h 49m 46s
13 Running: H2F2, HCF2, HF2 ... OK Time elapsed: 0h 21m 22s
14 Total: 1h 11m 8s
15
16 user@hostname$ ls
17 CH2F2+.m3c fragments.inp init results
18
19 user@hostname$ cd results
20
21 <REMOVE MOLECULES WHICH ARE SEPARATED IN TWO OR MORE PIECES>
22
23 user@hostname$ M3C-gamess.optg ../b3lyp.optg-GAMESS.inp
24
25 Running: CF2.q0.m1-10, CF2.q1.m2-1, ... , H2F2.q1.m2-1, HCF2.q0.m2-1 ... OK Time elapsed: 0h 8m 38s
26 Running: HCF2.q1.m1-12, HCF2.q1.m1-1, ... OK Time elapsed: 0h 2m 19s
27 Total: 0h 10m 57s
28 <REMOVE MOLECULES WHICH ARE SEPARATED IN TWO OR MORE PIECES>
29
30 user@hostname$ M3C-gamess.freqs ../b3lyp.freqs-GAMESS.inp
31 Running: CF2.q0.m1-10, CF2.q1.m2-1, ..., HCF2.q1.m1-1, HF2.q1.m1-1 ... OK Time elapsed: 0h 11m 31s
32 Total: 0h 11m 31s
33
34 CF2.q0.m1-10.xyz ... OK
35 CF2.q1.m2-1.xyz ... OK
36 ...
37 HCF2.q1.m1-1.xyz ... OK
38 HF2.q1.m1-1.xyz ... OK
39
40 user@hostname$ gwenview .
```

The obtained molecules for each step in above commands execution are shown in Figure (17).

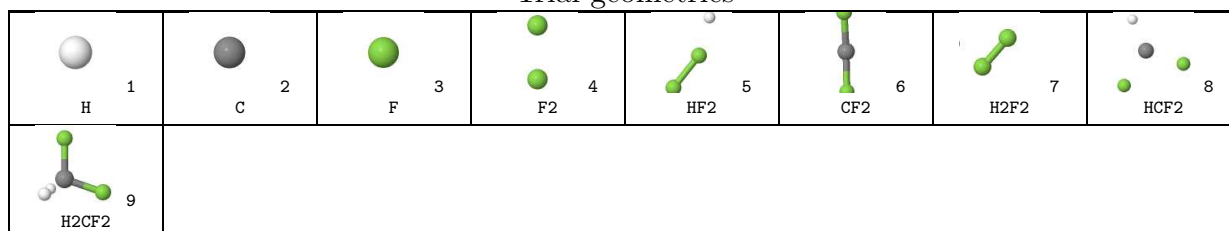
Then finally by using the M3C input file which is shown later, we can obtain the breaking curves, the deposited energy function and the experimental mass spectrum which are shown in Figure (18), after typing the following commands:

```

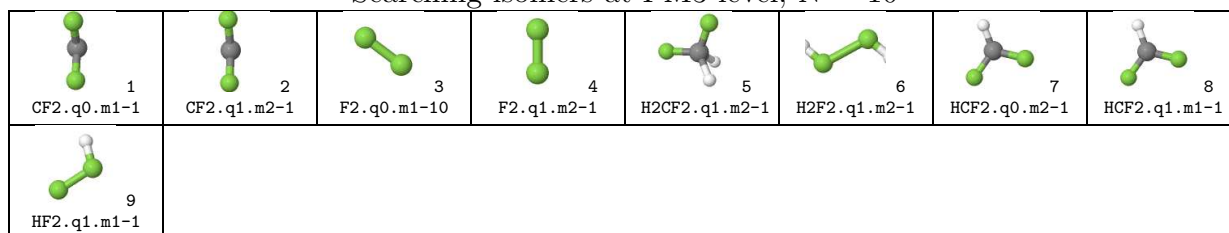
1 user@hostname$ M3C.p -i CH2F2+.m3c -n 8
2
3 Running: 0.00000, 0.33333, ... , 2.00000, 2.33333 ... OK Time elapsed: 0h 0m 32s
4 Running: 2.66667, 3.00000, ... , 4.66667, 5.00000 ... OK Time elapsed: 0h 0m 56s
5 ...
6 Running: 26.66667, 27.00000, ... , 28.66667, 29.00000 ... OK Time elapsed: 0h 2m 1s
7 Running: 29.33333, 29.66667, ... ... OK Time elapsed: 0h 1m 22s
8 Total: 0h 15m 49s
9
10 user@hostname$ M3C.analysis CH2F2+.m3c S.vs.E "p"
11 user@hostname$ M3C.analysis CH2F2+.m3c fit_sfE
12 user@hostname$ M3C.analysis CH2F2+.m3c fit_sBR

```

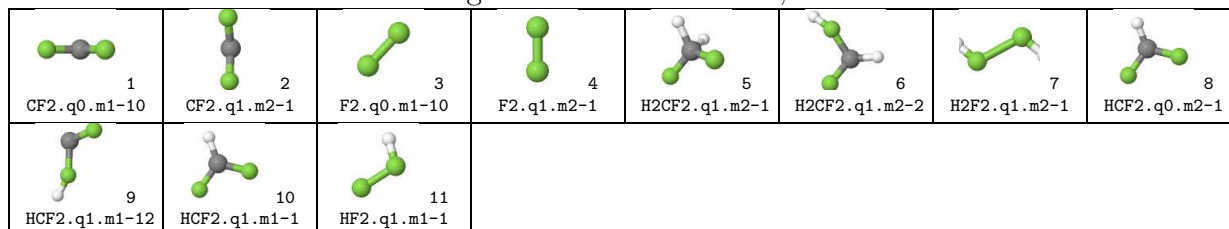
Trial geometries



Searching isomers at PM3 level, N = 10



Searching isomers at PM3 level, N = 50



Final set of molecules at B3LYP/6-311+G* level

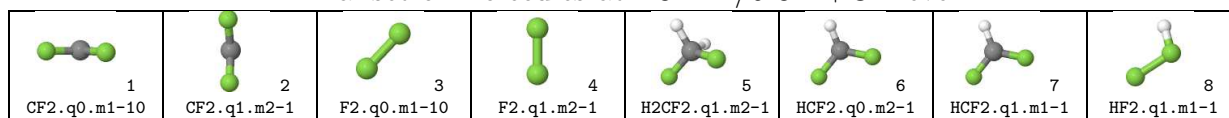


Figure 17: Stochastic search for isomers step by step.

The broad of the deposited energy function for CH_2F_2 is approximately twice than the one obtained for CH_3F , even if its maximum is at 3 eV too. The breakdown graph for this molecule shows the molecular ion (CH_2F_2^+) as the dominant one up to 1.5 eV of internal energy and with several fragment ions that compete between 2 eV and 10 eV, in contrast with the breakdown graph of molecule CH_3F (see Figures 16 and 18). These fragments

dominate the mass spectrum, because they are the most abundant over this region, where deposited energy function has its most significant contribution. The most intense peaks are those with highest probability in the breakdown graph, HCF_2^+ and H_2CF^+ . They are followed by the H_2CF_2^+ (molecular ion) and HCF^+ , and the last one corresponding to CF^+ , in which although it has a breakdown curve as broad as the previous one, its contribution is significantly reduced when it is weighted with the deposited energy function. The rest of ionic fragments does not have a significant contribution because they only exist for high excitation energies.

In general, we found a relatively good agreement with the experiment though we have used electronic energy calculations at a low level of theory. Again, it is important to highlight, that one way to improve these results is by increasing the number of isomers and electronic states for each fragment with more accurate electronic calculations.

Acknowledgments

I would like to thank Dr. Sergio Díaz-Tendero for his critical reading and effort to helping me to improve this tutorial. I am especially grateful to Dr. M. Merced Montero-Campillo for the english revision of this document.

If you find any errata or have any suggestions to improve this tutorial, please contact me at nestor.aguirre@uam.es

Input File 8: M3C input file CH2F2+.m3c

```

1 BEGIN GOPTIONS
2   systemRadius = 8.0
3   overlappingRadius = 0.4
4
5   useRandomWalkers = FALSE
6   randomWalkStepRadius = 1.0
7 END GOPTIONS
8
9 BEGIN ENERGY_RANGE
10  grid = 0:30:91 # dE = 1.0 eV
11 END ENERGY_RANGE
12
13 BEGIN RMJJ
14  task = V,T,S:0,V,T,S:1:-1
15  burnInFraction = 0.1
16
17  reactives = H2CF2p(d)
18  excitationEnergy = 5.0 # eV
19
20  tracing = none
21  numberOfExperiments = 3
22  numberOfEvents = 20000
23  historyFileFrequency = 100
24
25  energyHistoryFile = energy.dat
26  weightHistoryFile = weight.dat
27  histogramFile = histogram.dat
28 END RMJJ
29
30 BEGIN FRAGMENTS_DATABASE
31 #-----
32 #      Label      Z  M  L  SYM      geomFile      Elelec      maxVib
33 #-----
34      H(d)      0  2  0  1      H.q0.m2-1.rxyz      -13.572100
35      C(s)      0  1  0  1      C.q0.m1-1.rxyz      -1027.790000
36      F(d)      0  2  0  1      F.q0.m2-1.rxyz      -2713.690000
37      H2(s)      0  1  0  1      H2.q0.m1-10.rxyz      -31.833900      H(d)+H(d)
38      HC(d)      0  2  0  1      HC.q0.m2-10.rxyz      -1046.730000      H(d)+C(s)
39      HF(s)      0  1  0  1      HF.q0.m1-10.rxyz      -2732.950000      H(d)+F(d)
40      CF(d)      0  2  0  1      CF.q0.m2-10.rxyz      -3748.880000      C(s)+F(d)
41      F2(s)      0  1  0  1      F2.q0.m1-10.rxyz      -5428.820000      F(d)+F(d)
42      H2C(s)      0  1  0  1      H2C.q0.m1-10.rxyz      -1064.370000      H(d)+HC(d)
43      HCF(s)      0  1  0  1      HCF.q0.m1-10.rxyz      -3765.840000      H(d)+CF(d)
44      CF2(s)      0  1  0  1      CF2.q0.m1-10.rxyz      -6467.770000      F(d)+CF(d)
45      H3C(d)      0  2  0  1      H3C.q0.m2-14.rxyz      -1083.470000      H2(s)+HC(d)
46      H2CF(d)      0  2  0  1      H2CF.q0.m2-1.rxyz      -3783.830000      H2(s)+CF(d)
47      HCF2(d)      0  2  0  1      HCF2.q0.m2-1.rxyz      -6484.500000      HF(s)+CF(d)
48
49      Hp      1  0  0  1      H.q1.m0-1.rxyz      0.000000
50      Cp(d)      1  2  0  1      C.q1.m2-1.rxyz      -1018.080000
51      Fp(s)      1  1  0  1      F.q1.m1-1.rxyz      -2692.420000
52      H2p(d)      1  2  0  1      H2.q1.m2-10.rxyz      -16.298600      Hp+H(d)
53      HCP(s)      1  1  0  1      HC.q1.m1-10.rxyz      -1035.860000      H(d)+Cp(d)
54      HFP(d)      1  2  0  1      HF.q1.m2-10.rxyz      -2716.870000      Hp+F(d)
55      CFp(s)      1  1  0  1      CF.q1.m1-10.rxyz      -3739.460000      Cp(d)+F(d)
56      F2p(d)      1  2  0  1      F2.q1.m2-1.rxyz      -5413.010000      Fp(s)+F(d)
57      H3p(s)      1  1  0  1      H3.q1.m1-10.rxyz      -35.977900      Hp+H2(s)
58      H2Cp(d)      1  2  0  1      H2C.q1.m2-10.rxyz      -1054.540000      H2(s)+Cp(d)
59 #      H2Fp(s)      1  2  1  0  1      H2F.q1.m1-10.rxyz      -2737.320000      0.000
60      HCFp(dC)      1  2  0  1      HCF.q1.m2-10.rxyz      -3755.850000      H(d)+CFp(s)
61      HCFp(dF)      1  2  0  1      HCF.q1.m2-11.rxyz      -3753.040000      H(d)+CFp(s)
62      HF2p(s)      1  1  0  1      HF2.q1.m1-1.rxyz      -5432.360000      F(d)+HFp(d)
63      CF2p(d)      1  2  0  1      CF2.q1.m2-1.rxyz      -6456.410000      F(d)+CFp(s)
64      H3Cp(s)      1  1  0  1      H3C.q1.m1-13.rxyz      -1073.720000      H(d)+H2Cp(d)
65      H2CFp(st)      1  1  0  1      H2CF.q1.m1-1.rxyz      -3774.770000      H2(s)+CFp(s)
66      H2CFp(s)      1  1  0  1      H2CF.q1.m1-6.rxyz      -3770.520000      H(d)+HCFp(dC)
67      HCF2p(s)      1  1  0  1      HCF2.q1.m1-1.rxyz      -6475.650000      HF(s)+CFp(s)
68      H3CFp(dt)      1  2  0  1      H3CF.q1.m2-1.rxyz      -3789.520000      H(d)+H2CFp(st)
69      H3CFp(d)      1  2  0  1      H3CF.q1.m2-2.rxyz      -3789.280000      H(d)+H2CFp(st)
70      H2CF2p(d)      1  2  0  1      H2CF2.q1.m2-1.rxyz      -6490.040000      H(d)+HCF2p(s)
71 #-----
72 END FRAGMENTS_DATABASE
73
74 BEGIN EXPERIMENTAL_BRANCHING_RATIOS
75   error = absolute
76   diagram = S.vs.E
77
78   #-----
79   #      Channel      BR      error      q/n
80   #-----
81      Cp      0.9      0.0      12.0
82      HCP      2.4      0.0      13.0
83      H2Cp      0.5      0.0      14.0
84      HFP      0.2      0.0      20.0
85      CFp      10.6      0.0      31.0
86      HCFp      4.2      0.0      32.0
87      H2CFp      38.2      0.0      33.0
88      CF2p      1.3      0.0      50.0
89      HCF2p      36.6      0.0      51.0
90      H2CF2p      3.9      0.0      52.0
91      H2p      0.0      0.0      2.0
92   #-----
93 END EXPERIMENTAL_BRANCHING_RATIOS
94
95 BEGIN FIT_BRANCHING_RATIOS
96   method = NNLS
97   basis = 60,60
98   eDistfile = edist.out
99   BRfile = fitBR.out
100 END FIT_BRANCHING_RATIOS

```

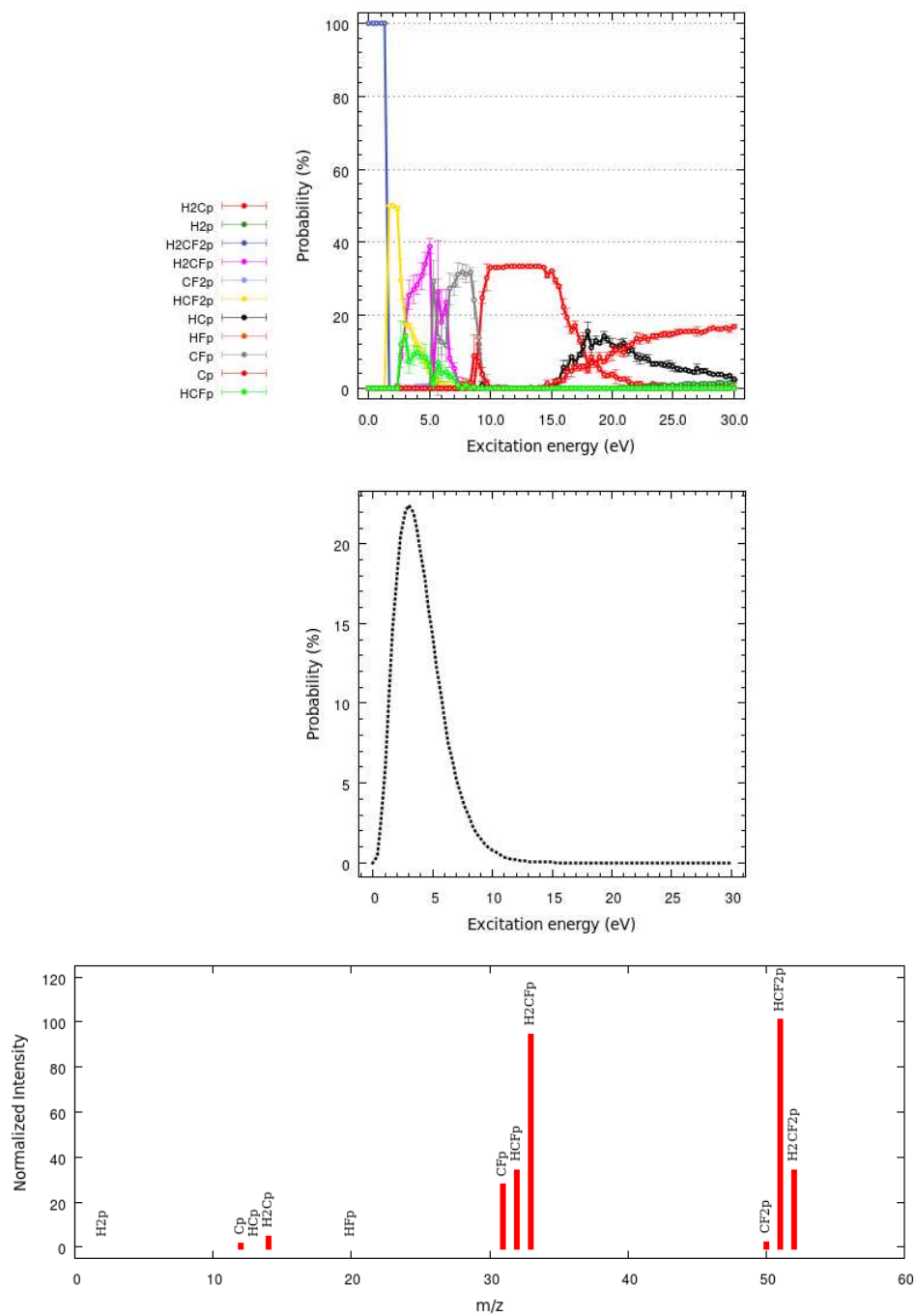


Figure 18: Upper panel, probabilities of singly-charged fragments (breakdown curves). Middle panel, fitted deposited energy function. Lower panel, theoretical mass spectrum for diMethylFluoride CH_2F_2 .

STEP BY STEP TUTORIAL. MASS SPECTRUM OF CH₃F MOLECULE

Operating system: QUBuntu

DEPENDENCIES

- 1) **jmol** (version ≥ 12.2)
This is already installed on QUBuntu
- 2) **gnuplot** (version ≥ 4.2)
This is already installed on QUBuntu
- 3) **gawk** (version ≥ 3.1)
This is **not** installed on QUBuntu
\$ sudo apt-get install gawk
- 4) **gwenview** or other image viewer for displaying a collection of images
This is **not** installed on QUBuntu
\$ sudo apt-get install gwenview
- 5) Download the following files from moodle:
M3C-1.0-intelc-14.0.3-i686.tar.gz
gamess-i686.tar.gz

```
user@hostname:~$ cd $HOME
user@hostname:~$ ls
gamess.tar.gz  M3C-1.0-intelc-14.0.3-i686.tar.gz
```

Installing GAMESS

```
user@hostname:~$ tar xzf gamess-i686.tar.gz
user@hostname:~$ ls
gamess  gamess-i686.tar.gz  M3C-1.0-intelc-14.0.3-i686.tar.gz
user@hostname:~$ mv gamess .gamess
user@hostname:~$ vim .bashrc
```

<APPEND THE NEXT LINE>

```
export PATH=$HOME/.gamess:$PATH
```

Installing M3C

```
user@hostname:~$ tar xzf M3C-1.0-intelc-14.0.3-i686.tar.gz
user@hostname:~$ ls
gamess-i686.tar.gz  M3C  M3C-1.0-intelc-14.0.3-i686.tar.gz
user@hostname:~$ mv M3C .M3C
user@hostname:~$ vim .bashrc
```

<APPEND THE NEXT TWO LINES>

```
export M3C_HOME=$HOME/.M3C/
export PATH=$M3C_HOME/bin:$PATH
```

Work space Setup.

<OPEN A NEW TERMINAL WINDOW>

```
user@hostname:~$ cp -r .M3C/examples/ M3C-examples
user@hostname:~$ ls
gamess-i686.tar.gz  M3C-1.0-intelc-14.0.3-i686.tar.gz  M3C-examples

user@hostname:~$ cd M3C-examples/

user@hostname:~/M3C-examples$ ls
b3lyp.optg-GAMESS.inp  diMethylFluoride  b3lyp.freqs-GAMESS.inp  methylFluoride  pm3.optg-GAMESS.inp  reactorT.m3c
```

The tutorial itself.

```
user@hostname:~/M3C-examples$ cd methylFluoride/
user@hostname:~/M3C-examples/methylFluoride$ ls
```

```
CH3F+.m3c fragments.inp init
```

```
user@hostname$ M3C.fragments H3,C,F
```

```
H, C, F, H2, HC, HF, CF, H3, H2C, H2F, HCF, H3C, H3F, H2CF, H3CF
```

```
user@hostname:~/M3C-examples/methylFluoride$ cd init
```

```
user@hostname:~/M3C-examples/methylFluoride/init$ M3C.viewXYZ
```

```
CF.xyz ... OK
C.xyz ... OK
F.xyz ... OK
H2CF.xyz ... OK
H2C.xyz ... OK
H2F.xyz ... OK
H2.xyz ... OK
H3CF.xyz ... OK
H3C.xyz ... OK
H3F.xyz ... OK
H3.xyz ... OK
HCF.xyz ... OK
HC.xyz ... OK
HF.xyz ... OK
H.xyz ... OK
```

```
user@hostname:~/M3C-examples/methylFluoride/init$ ls
```

```
CF.gif C.gif F.gif H2CF.gif H2C.gif H2F.gif H2.gif H3CF.gif H3C.gif H3F.gif H3.gif HCF.gif HC.gif HF.gif H.gif
CF.xyz C.xyz F.xyz H2CF.xyz H2C.xyz H2F.xyz H2.xyz H3CF.xyz H3C.xyz H3F.xyz H3.xyz HCF.xyz HC.xyz HF.xyz H.xyz
```

```
user@hostname:~/M3C-examples/methylFluoride/init$ gwenview . &
```

```
user@hostname:~/M3C-examples/methylFluoride/init$ cd ..
```

```
user@hostname:~/M3C-examples/methylFluoride$ M3C-games.geniso fragments.inp ../pm3.optg-GAMESS.inp ../reactorT.m3c 5 init results
```

```
Running: F C ... OK Time elapsed: 0h 0m 0s
Running: CF H ... OK Time elapsed: 0h 0m 23s
Running: HF HC ... OK Time elapsed: 0h 0m 37s
Running: HCF H2 ... OK Time elapsed: 0h 0m 48s
Running: H2F H2C ... OK Time elapsed: 0h 2m 33s
Running: H2CF H3 ... OK Time elapsed: 0h 1m 6s
Running: H3F H3C ... OK Time elapsed: 0h 7m 55s
Running: F C ... OK Time elapsed: 0h 0m 1s
Running: CF H ... OK Time elapsed: 0h 0m 45s
Running: HF HC ... OK Time elapsed: 0h 1m 47s
Running: HCF H2 ... OK Time elapsed: 0h 1m 9s
Running: H2F H2C ... OK Time elapsed: 0h 2m 17s
Running: H2CF H3 ... OK Time elapsed: 0h 3m 1s
Running: H3F H3C ... OK Time elapsed: 0h 3m 43s
Running: H3CF ... OK Time elapsed: 0h 1m 37s
Total: 0h 27m 42s
```

```
user@hostname:~/M3C-examples/methylFluoride$ ls
```

```
CH3F+.m3c fragments.inp init results
```

```
user@hostname:~/M3C-examples/methylFluoride$ cd results/
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ ls
```

```
CF.q0.m2-1.xyz H2CF.q1.m1-1.xyz H2F.q1.m1-1.xyz H3C.q1.m1-1.xyz H3.q0.m2-2.xyz HC.q1.m1-1.xyz history-F.q0.m2 history-H2F.q1.m1 history-H3F.q1.m2 history-HF.q0.m1
CF.q1.m1-1.xyz H2CF.q1.m1-2.xyz H2.q0.m1-1.xyz H3C.q1.m1-4.xyz H3.q0.m2-3.xyz HF.q0.m1-1.xyz history-F.q1.m1 history-H2.q0.m1 history-H3.q0.m2 history-HF.q1.m2
C.q0.m1-1.xyz H2CF.q1.m1-3.xyz H2.q1.m2-1.xyz H3C.q1.m1-5.xyz H3.q0.m2-5.xyz HF.q1.m2-1.xyz history-H2CF.q0.m2 history-H2.q1.m2 history-H3.q1.m1 history-H.q0.m2
C.q1.m2-1.xyz H2C.q0.m1-1.xyz H3CF.q1.m2-1.xyz H3F.q0.m1-1.xyz H3.q1.m1-1.xyz history-CF.q0.m2 history-H3CF.q1.m1 history-HCF.q0.m1 history-H.q1.m0
C.q0.m2-1.xyz H2C.q1.m2-1.xyz H3CF.q1.m2-2.xyz H3F.q1.m2-1.xyz HCF.q0.m1-1.xyz history-CF.q1.m1 history-H2C.q0.m1 history-H3C.q0.m2 history-HCF.q1.m2 history-H.q0.m2-1.xyz
F.q1.m1-1.xyz H2F.q0.m2-1.xyz H3C.q0.m2-1.xyz H3F.q1.m2-2.xyz HCF.q1.m2-1.xyz history-C.q0.m1 history-H2C.q1.m1 history-H3C.q1.m1 history-HC.q0.m2 H.q1.m0-1.xyz
H2CF.q0.m2-1.xyz H2F.q0.m2-2.xyz H3C.q0.m2-3.xyz H3.q0.m2-1.xyz HC.q0.m2-1.xyz history-C.q1.m2 history-H2F.q0.m2 history-H3F.q0.m1 history-HC.q1.m1
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ rm -rf history-*
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ ls
```

```
CF.q0.m2-1.xyz F.q0.m2-1.xyz H2CF.q1.m1-2.xyz H2F.q0.m2-1.xyz H2.q1.m2-1.xyz H3C.q0.m2-3.xyz H3F.q0.m1-1.xyz H3.q0.m2-2.xyz HCF.q0.m1-1.xyz HF.q0.m1-1.xyz
CF.q1.m1-1.xyz F.q1.m1-1.xyz H2CF.q1.m1-3.xyz H2F.q0.m2-2.xyz H3CF.q1.m2-1.xyz H3C.q1.m1-1.xyz H3F.q1.m2-1.xyz H3.q0.m2-3.xyz HCF.q1.m2-1.xyz HF.q1.m2-1.xyz
C.q0.m1-1.xyz H2CF.q0.m2-1.xyz H2C.q0.m1-1.xyz H2F.q1.m1-1.xyz H3CF.q1.m2-2.xyz H3C.q1.m1-4.xyz H3F.q1.m2-2.xyz H3.q0.m2-5.xyz HC.q0.m2-1.xyz H.q0.m2-1.xyz
C.q1.m2-1.xyz H2CF.q1.m1-1.xyz H2C.q1.m2-1.xyz H2.q0.m1-1.xyz H3C.q0.m2-1.xyz H3C.q1.m1-5.xyz H3.q0.m2-1.xyz H3.q1.m1-1.xyz HC.q1.m1-1.xyz H.q1.m0-1.xyz
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ M3C.viewXYZ
```

```
CF.q0.m2-1.xyz ... OK
CF.q1.m1-1.xyz ... OK
C.q0.m1-1.xyz ... OK
C.q1.m2-1.xyz ... OK
F.q0.m2-1.xyz ... OK
F.q1.m1-1.xyz ... OK
H2CF.q0.m2-1.xyz ... OK
H2CF.q1.m1-1.xyz ... OK
H2CF.q1.m1-2.xyz ... OK
H2CF.q1.m1-3.xyz ... OK
H2CF.q1.m1-3.xyz ... OK
H2C.q0.m1-1.xyz ... OK
H2C.q1.m2-1.xyz ... OK
H2F.q0.m2-1.xyz ... OK
H2F.q0.m2-2.xyz ... OK
H2F.q1.m1-1.xyz ... OK
H2.q0.m1-1.xyz ... OK
```



```

H2.q1.m2-1.xyz ... OK
H3CF.q1.m2-1.xyz ... OK
H3CF.q1.m2-2.xyz ... OK
H3C.q0.m2-1.xyz ... OK
H3C.q0.m2-3.xyz ... OK
H3C.q1.m1-1.xyz ... OK
H3C.q1.m1-4.xyz ... OK
H3C.q1.m1-5.xyz ... OK
H3F.q0.m1-1.xyz ... OK
H3F.q1.m2-1.xyz ... OK
H3F.q1.m2-2.xyz ... OK
H3.q0.m2-1.xyz ... OK
H3.q0.m2-2.xyz ... OK
H3.q0.m2-3.xyz ... OK
H3.q0.m2-5.xyz ... OK
H3.q1.m1-1.xyz ... OK
HCF.q0.m1-1.xyz ... OK
HCF.q1.m2-1.xyz ... OK
HC.q0.m2-1.xyz ... OK
HC.q1.m1-1.xyz ... OK
HF.q0.m1-1.xyz ... OK
HF.q1.m2-1.xyz ... OK
H.q0.m2-1.xyz ... OK
H.q1.m0-1.xyz ... OK

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ ls

```

```

CF.q0.m2-1.gif F.q0.m2-1.gif H2CF.q1.m1-2.gif H2F.q0.m2-1.gif H2.q1.m2-1.gif H3C.q0.m2-3.gif H3F.q0.m1-1.gif H3.q0.m2-2.gif HCF.q0.m1-1.gif HF.q0.m1-1.gif
CF.q0.m2-1.xyz F.q0.m2-1.xyz H2CF.q1.m1-2.xyz H2F.q0.m2-1.xyz H2.q1.m2-1.gif H3C.q0.m2-3.xyz H3F.q0.m1-1.xyz H3.q0.m2-2.gif HCF.q0.m1-1.xyz HF.q0.m1-1.xyz
CF.q1.m1-1.gif F.q1.m1-1.gif H2CF.q1.m1-3.gif H2F.q0.m2-2.gif H3CF.q1.m2-1.gif H3C.q1.m1-1.gif H3F.q1.m2-1.gif H3.q0.m2-3.gif HCF.q1.m2-1.gif HF.q1.m2-1.gif
CF.q1.m1-1.xyz F.q1.m1-1.xyz H2CF.q1.m1-3.xyz H2F.q0.m2-2.gif H3CF.q1.m2-1.gif H3C.q1.m1-1.gif H3F.q1.m2-1.gif H3.q0.m2-3.gif HCF.q1.m2-1.gif HF.q1.m2-1.gif
C.q0.m1-1.gif H2CF.q0.m2-1.gif H2C.q0.m1-1.gif H2F.q1.m1-1.gif H3CF.q1.m2-2.gif H3C.q1.m1-4.gif H3F.q1.m2-2.gif H3.q0.m2-5.gif HC.q0.m2-1.gif H.q0.m2-1.gif
C.q0.m1-1.xyz H2CF.q0.m2-1.xyz H2C.q0.m1-1.gif H2F.q1.m1-1.gif H3CF.q1.m2-2.gif H3C.q1.m1-4.gif H3F.q1.m2-2.gif H3.q0.m2-5.gif HC.q0.m2-1.gif H.q0.m2-1.gif
C.q1.m2-1.gif H2CF.q1.m1-1.gif H2C.q1.m2-1.gif H2.q0.m1-1.gif H3CF.q1.m2-2.gif H3C.q1.m1-5.gif H3.q0.m2-1.gif H3.q1.m1-1.gif HC.q1.m1-1.gif H.q1.m0-1.gif
C.q1.m2-1.xyz H2CF.q1.m1-1.xyz H2C.q1.m2-1.xyz H2.q0.m1-1.gif H3C.q0.m2-1.gif H3C.q1.m1-5.gif H3.q0.m2-1.gif H3.q1.m1-1.gif HC.q1.m1-1.gif H.q1.m0-1.gif

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ gwenview . &

```

```

<REMOVE MOLECULES WHICH ARE SEPARATED IN TWO OR MORE PIECES>

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ rm H2CF.q1.m1-2.* H2CF.q1.m1-3.* H2F.q0.m2-1.* H2F.q0.m2-2.* H3.q0.m2-2.*

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ rm H3.q0.m2-3.* H3C.q0.m2-3.* H3C.q1.m1-4.* H3C.q1.m1-5.* H3F.q1.*

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ rm *.gif

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ ls

```

```

CF.q0.m2-1.xyz C.q1.m2-1.xyz H2CF.q0.m2-1.xyz H2C.q1.m2-1.xyz H2.q1.m2-1.xyz H3C.q0.m2-1.xyz H3.q0.m2-5.xyz HCF.q1.m2-1.xyz HF.q0.m1-1.xyz H.q1.m0-1.xyz
CF.q1.m1-1.xyz F.q0.m2-1.xyz H2CF.q1.m1-1.xyz H2F.q1.m1-1.xyz H3CF.q1.m2-1.xyz H3C.q1.m1-1.xyz H3.q1.m1-1.xyz HC.q0.m2-1.xyz HF.q1.m2-1.xyz
C.q0.m1-1.xyz F.q1.m1-1.xyz H2C.q0.m1-1.xyz H2.q0.m1-1.xyz H3CF.q1.m2-2.xyz H3.q0.m2-1.xyz HCF.q0.m1-1.xyz HC.q1.m1-1.xyz H.q0.m2-1.xyz

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ cp -r ../results ../results.backup

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ M3C-gamess.optg ././b3lyp.optg-GAMESS.inp

```

```

Running: CF.q0.m2-1, CF.q1.m1-1 ... OK Time elapsed: 0h 1m 7s
Running: C.q0.m1-1, C.q1.m2-1 ... OK Time elapsed: 0h 0m 0s
Running: F.q0.m2-1, F.q1.m1-1 ... OK Time elapsed: 0h 0m 0s
Running: H2CF.q0.m2-1, H2CF.q1.m1-1 ... OK Time elapsed: 0h 3m 36s
Running: H2C.q0.m1-1, H2C.q1.m2-1 ... OK Time elapsed: 0h 0m 47s
Running: H2F.q1.m1-1, H2.q0.m1-1 ... OK Time elapsed: 0h 0m 19s
Running: H2.q1.m2-1, H3CF.q1.m2-1 ... OK Time elapsed: 0h 8m 31s
Running: H3CF.q1.m2-2, H3C.q0.m2-1 ... OK Time elapsed: 0h 3m 59s
Running: H3C.q1.m1-1, H3.q0.m2-1 ... OK Time elapsed: 0h 1m 51s
Running: H3.q0.m2-5, H3.q1.m1-1 ... OK Time elapsed: 0h 0m 23s
Running: HCF.q0.m1-1, HCF.q1.m2-1 ... OK Time elapsed: 0h 2m 2s
Running: HC.q0.m2-1, HC.q1.m1-1 ... OK Time elapsed: 0h 0m 29s
Running: HF.q0.m1-1, HF.q1.m2-1 ... OK Time elapsed: 0h 0m 19s
Running: H.q0.m2-1, H.q1.m0-1 ... OK Time elapsed: 0h 0m 0s
Running: ... OK Time elapsed: 0h 0m 0s
Total: 0h 23m 23s

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ ls

```

```

CF.q0.m2-1.xyz C.q1.m2-1.xyz H2CF.q0.m2-1.xyz H2C.q1.m2-1.xyz H2.q1.m2-1.xyz H3C.q0.m2-1.xyz H3.q0.m2-5.xyz HCF.q1.m2-1.xyz HF.q0.m1-1.xyz H.q1.m0-1.xyz
CF.q0.m2-1.xyz0 C.q1.m2-1.xyz0 H2CF.q0.m2-1.xyz0 H2C.q1.m2-1.xyz0 H2.q1.m2-1.xyz0 H3C.q0.m2-1.xyz0 H3.q0.m2-5.xyz0 HCF.q1.m2-1.xyz0 HF.q0.m1-1.xyz0 H.q1.m0-1.xyz0
CF.q1.m1-1.xyz F.q0.m2-1.xyz H2CF.q1.m1-1.xyz H2F.q1.m1-1.xyz H3CF.q1.m2-1.xyz H3C.q1.m1-1.xyz H3.q1.m1-1.xyz HC.q0.m2-1.xyz HF.q1.m2-1.xyz
CF.q1.m1-1.xyz0 F.q0.m2-1.xyz0 H2CF.q1.m1-1.xyz0 H2F.q1.m1-1.xyz0 H3CF.q1.m2-1.xyz0 H3C.q1.m1-1.xyz0 H3.q1.m1-1.xyz0 HC.q0.m2-1.xyz0 HF.q1.m2-1.xyz0
C.q0.m1-1.xyz F.q1.m1-1.xyz H2C.q0.m1-1.xyz H2.q0.m1-1.xyz H3CF.q1.m2-2.xyz H3.q0.m2-1.xyz HCF.q0.m1-1.xyz HC.q1.m1-1.xyz H.q0.m2-1.xyz
C.q0.m1-1.xyz0 F.q1.m1-1.xyz0 H2C.q0.m1-1.xyz0 H2.q0.m1-1.xyz0 H3CF.q1.m2-2.xyz0 H3.q0.m2-1.xyz0 HCF.q0.m1-1.xyz0 HC.q1.m1-1.xyz0 H.q0.m2-1.xyz0

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ M3C.viewXYZ

```

```

CF.q0.m2-1.xyz ... OK
CF.q1.m1-1.xyz ... OK
C.q0.m1-1.xyz ... OK
C.q1.m2-1.xyz ... OK
F.q0.m2-1.xyz ... OK
F.q1.m1-1.xyz ... OK

```

```
H2CF.q0.m2-1.xyz ... OK
H2CF.q1.m1-1.xyz ... OK
H2C.q0.m1-1.xyz ... OK
H2C.q1.m2-1.xyz ... OK
H2F.q1.m1-1.xyz ... OK
H2.q0.m1-1.xyz ... OK
H2.q1.m2-1.xyz ... OK
H3CF.q1.m2-1.xyz ... OK
H3CF.q1.m2-2.xyz ... OK
H3C.q0.m2-1.xyz ... OK
H3C.q1.m1-1.xyz ... OK
H3.q0.m2-1.xyz ... OK
H3.q0.m2-5.xyz ... OK
H3.q1.m1-1.xyz ... OK
HCF.q0.m1-1.xyz ... OK
HCF.q1.m2-1.xyz ... OK
HC.q0.m2-1.xyz ... OK
HC.q1.m1-1.xyz ... OK
HF.q0.m1-1.xyz ... OK
HF.q1.m2-1.xyz ... OK
H.q0.m2-1.xyz ... OK
H.q1.m0-1.xyz ... OK
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ gwenview . &
```

```
<REMOVE MOLECULES WHICH ARE SEPARATED IN TWO OR MORE PIECES>
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ rm H3.q0.m2-1.*
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ ls
```

CF.q0.m2-1.gif	C.q0.m1-1.xyz0	F.q1.m1-1.xyz	H2C.q0.m1-1.gif	H2F.q1.m1-1.xyz0	H3CF.q1.m2-1.xyz	H3C.q1.m1-1.gif	HCF.q0.m1-1.gif	HC.q0.m2-1.xyz0	HF.q1.m2-1.xyz
CF.q0.m2-1.xyz	C.q1.m2-1.gif	F.q1.m1-1.xyz0	H2C.q0.m1-1.xyz	H2.q0.m1-1.gif	H3CF.q1.m2-1.xyz0	H3C.q1.m1-1.xyz	HCF.q0.m1-1.xyz	HC.q1.m1-1.gif	HF.q1.m2-1.xyz0
CF.q0.m2-1.xyz0	C.q1.m2-1.xyz	H2CF.q0.m2-1.gif	H2C.q0.m1-1.xyz0	H2.q0.m1-1.xyz	H3CF.q1.m2-2.gif	H3C.q1.m1-1.xyz0	HCF.q0.m1-1.xyz0	HC.q1.m1-1.xyz	H.q0.m2-1.gif
CF.q1.m1-1.gif	C.q1.m2-1.xyz0	H2CF.q0.m2-1.xyz	H2C.q1.m2-1.gif	H2.q0.m1-1.xyz0	H3CF.q1.m2-2.xyz	H3.q0.m2-5.xyz	HCF.q1.m2-1.gif	HC.q1.m1-1.xyz0	H.q0.m2-1.xyz
CF.q1.m1-1.xyz	F.q0.m2-1.gif	H2CF.q0.m2-1.xyz0	H2C.q1.m2-1.gif	H2.q1.m2-1.gif	H3CF.q1.m2-2.xyz0	H3.q0.m2-5.xyz0	HCF.q1.m2-1.xyz	HF.q0.m1-1.gif	H.q0.m2-1.xyz0
CF.q1.m1-1.xyz0	F.q0.m2-1.xyz	H2CF.q1.m1-1.gif	H2C.q1.m2-1.xyz	H2.q1.m2-1.xyz0	H3C.q0.m2-1.gif	H3.q1.m1-1.gif	HCF.q1.m2-1.xyz0	HF.q0.m1-1.xyz	H.q1.m0-1.gif
C.q0.m1-1.gif	F.q0.m2-1.xyz0	H2CF.q1.m1-1.xyz	H2F.q1.m1-1.gif	H2.q1.m2-1.xyz0	H3C.q0.m2-1.xyz	H3.q1.m1-1.xyz	HC.q0.m2-1.gif	HF.q0.m1-1.xyz0	H.q1.m0-1.xyz
C.q0.m1-1.xyz	F.q1.m1-1.gif	H2CF.q1.m1-1.xyz0	H2F.q1.m1-1.xyz	H3CF.q1.m2-1.gif	H3C.q0.m2-1.xyz0	H3.q1.m1-1.xyz0	HC.q0.m2-1.xyz	HF.q1.m2-1.gif	H.q1.m0-1.xyz0

```
user@hostname:~/M3C-examples/methylFluoride/results$ cp -r ../results ../results.backup2
```

```
user@hostname:~/M3C-examples/methylFluoride/results$ M3C-gamess.freqs ../.b3lyp.freqs-GAMESS.inp
```

Running:	CF.q0.m2-1,	CF.q1.m1-1 ...	OK	Time elapsed:	0h 2m 1s
Running:	C.q0.m1-1,	C.q1.m2-1 ...	OK	Time elapsed:	0h 0m 13s
Running:	F.q0.m2-1,	F.q1.m1-1 ...	OK	Time elapsed:	0h 0m 14s
Running:	H2CF.q0.m2-1,	H2CF.q1.m1-1 ...	OK	Time elapsed:	0h 8m 3s
Running:	H2C.q0.m1-1,	H2C.q1.m2-1 ...	OK	Time elapsed:	0h 2m 23s
Running:	H2F.q1.m1-1,	H2.q0.m1-1 ...	OK	Time elapsed:	0h 1m 19s
Running:	H2.q1.m2-1,	H3CF.q1.m2-1 ...	OK	Time elapsed:	0h 9m 4s
Running:	H3CF.q1.m2-2,	H3C.q0.m2-1 ...	OK	Time elapsed:	0h 11m 11s
Running:	H3C.q1.m1-1,	H3.q0.m2-5 ...	OK	Time elapsed:	0h 2m 11s
Running:	H3.q1.m1-1,	HCF.q0.m1-1 ...	OK	Time elapsed:	0h 2m 24s
Running:	HCF.q1.m2-1,	HC.q0.m2-1 ...	OK	Time elapsed:	0h 3m 31s
Running:	HC.q1.m1-1,	HF.q0.m1-1 ...	OK	Time elapsed:	0h 0m 52s
Running:	HF.q1.m2-1,	H.q0.m2-1 ...	OK	Time elapsed:	0h 0m 49s
Running:	H.q1.m0-1	...	OK	Time elapsed:	0h 0m 3s
				Total:	0h 44m 18s

```
user@hostname:~/M3C-examples/methylFluoride/results$ ls
```

CF.q0.m2-1.gif	C.q0.m1-1.xyz0	F.q1.m1-1.xyz	H2C.q0.m1-1.rxyz	H2.q0.m1-1.gif	H3CF.q1.m2-1.xyz0	H3C.q1.m1-1.xyz	HCF.q0.m1-1.xyz	HC.q1.m1-1.rxyz	H.q0.m2-1.gif
CF.q0.m2-1.rxyz	C.q1.m2-1.gif	F.q1.m1-1.xyz0	H2C.q0.m1-1.rxyz	H2.q0.m1-1.rxyz	H3CF.q1.m2-2.gif	H3C.q1.m1-1.xyz0	HCF.q0.m1-1.rxyz	HC.q1.m1-1.rxyz	H.q0.m2-1.rxyz
CF.q0.m2-1.rxyz0	C.q1.m2-1.rxyz	H2CF.q0.m2-1.gif	H2C.q0.m1-1.rxyz0	H2.q0.m1-1.rxyz	H3CF.q1.m2-2.rxyz	H3.q0.m2-5.rxyz	HCF.q1.m2-1.gif	HC.q1.m1-1.rxyz0	H.q0.m2-1.rxyz0
CF.q1.m1-1.gif	C.q1.m2-1.rxyz0	H2CF.q0.m2-1.rxyz	H2C.q1.m2-1.gif	H2.q0.m1-1.rxyz0	H3CF.q1.m2-2.xyz	H3.q0.m2-5.rxyz	HCF.q1.m2-1.rxyz	HF.q0.m1-1.rxyz	H.q0.m2-1.rxyz0
CF.q1.m1-1.rxyz	F.q0.m2-1.gif	H2CF.q0.m2-1.rxyz0	H2C.q1.m2-1.rxyz	H2.q1.m2-1.gif	H3CF.q1.m2-2.xyz0	H3.q0.m2-5.rxyz0	HCF.q1.m2-1.rxyz0	HF.q0.m1-1.rxyz	H.q1.m0-1.gif
CF.q1.m1-1.rxyz0	F.q0.m2-1.rxyz	H2CF.q1.m1-1.gif	H2C.q1.m2-1.rxyz0	H2.q1.m2-1.rxyz	H3C.q0.m2-1.gif	H3.q1.m1-1.rxyz	HCF.q1.m2-1.rxyz0	HF.q0.m1-1.rxyz0	H.q1.m0-1.rxyz
C.q0.m1-1.gif	F.q0.m2-1.rxyz0	H2CF.q1.m1-1.rxyz	H2F.q1.m1-1.rxyz	H2.q1.m2-1.rxyz0	H3C.q0.m2-1.rxyz	H3.q1.m1-1.rxyz	HC.q0.m2-1.rxyz	HF.q1.m2-1.rxyz	H.q1.m0-1.rxyz0
C.q0.m1-1.rxyz	F.q1.m1-1.gif	H2CF.q1.m1-1.rxyz0	H2F.q1.m1-1.rxyz	H3CF.q1.m2-1.gif	H3C.q0.m2-1.rxyz0	H3.q1.m1-1.rxyz0	HC.q0.m2-1.rxyz	HF.q1.m2-1.rxyz	H.q1.m0-1.rxyz0
C.q0.m1-1.rxyz0	F.q1.m1-1.rxyz	H2CF.q1.m1-1.rxyz	H2F.q1.m1-1.rxyz0	H3CF.q1.m2-1.rxyz	H3C.q1.m1-1.gif	HCF.q0.m1-1.rxyz	HC.q0.m2-1.rxyz0	HF.q1.m2-1.rxyz	H.q1.m0-1.rxyz0
C.q1.m1-1.rxyz	F.q1.m1-1.rxyz	H2C.q0.m1-1.gif	H2F.q1.m1-1.rxyz	H3CF.q1.m2-1.rxyz	H3C.q1.m1-1.rxyz	HCF.q0.m1-1.rxyz	HC.q1.m1-1.gif	HF.q1.m2-1.rxyz0	

```
user@hostname:~/M3C-examples/methylFluoride/results$ cat H3CF.q1.m2-2.rxyz
```

```
5
Energy = -139.2622142579
C 0.5063172279 -0.2100414032 -0.1685057135
F -0.2546710826 0.7995943914 0.0969352537
H 0.1557351925 -1.1353725153 0.4524849110
H 0.7554782499 -0.3594035325 -1.2172639004
H 1.3649855324 -0.2706254204 0.6195076992
```

```
FREQUENCIES 9
3202.02
2555.74
```

2212.72
1461.16
1285.72
1092.85
1056.26
966.02
733.31

user@hostname:~/M3C-examples/methylFluoride/results\$ grep -H "^I" *.xyz

H2F.q1.m1-1.xyz:I

H2F.q1.m1-1.xyz:I

<THIS MOLECULE HAS TWO IMAGINARY FREQUENCIES, THEN YOU HAVE TO DELETE THEIR RELATED FILES>

user@hostname:~/M3C-examples/methylFluoride/results\$ rm H2F.q1.m1-1.*

user@hostname:~/M3C-examples/methylFluoride/results\$ cp -r ./results ../results.backup3

user@hostname:~/M3C-examples/methylFluoride/results\$ cp ../CH3F+.m3c .

user@hostname:~/M3C-examples/methylFluoride/results\$ M3C.makeDB CH3F+.m3c

BEGIN FRAGMENTS_DATABASE

#	Label	Z	M	L	SYM	geomFile	Eelec	maxVib
#	H(d1)	0	2	0	1	H.q0.m2-1.xyz	-13.572100	
	C(s1)	0	1	0	1	C.q0.m1-1.xyz	-1027.790000	
	F(d1)	0	2	0	1	F.q0.m2-1.xyz	-2713.690000	
	H2(s1)	0	1	0	1	H2.q0.m1-1.xyz	-31.833900	H(d1)+H(d1)
	HC(d1)	0	2	0	1	HC.q0.m2-1.xyz	-1046.730000	H(d1)+C(s1)
	HF(s1)	0	1	0	1	HF.q0.m1-1.xyz	-2732.950000	H(d1)+F(d1)
	CF(d1)	0	2	0	1	CF.q0.m2-1.xyz	-3748.880000	C(s1)+F(d1)
	H2C(s1)	0	1	0	1	H2C.q0.m1-1.xyz	-1064.370000	H(d1)+HC(d1)
	HCF(s1)	0	1	0	1	HCF.q0.m1-1.xyz	-3765.840000	H(d1)+CF(d1)
	H3C(d1)	0	2	0	1	H3C.q0.m2-1.xyz	-1083.470000	H2(s1)+HC(d1)
	H2CF(d1)	0	2	0	1	H2CF.q0.m2-1.xyz	-3783.830000	H2(s1)+CF(d1)
	Hp(1)	1	0	0	1	H.q1.m0-1.xyz	0.000000	
	Cp(d1)	1	2	0	1	C.q1.m2-1.xyz	-1018.080000	
	Fp(s1)	1	1	0	1	F.q1.m1-1.xyz	-2692.420000	
	H2p(s1)	1	2	0	1	H2.q1.m2-1.xyz	-16.298600	Hp(1)+H(d1)
	HCp(s1)	1	1	0	1	HC.q1.m1-1.xyz	-1035.860000	H(d1)+Cp(d1)
	HFp(s1)	1	2	0	1	HF.q1.m2-1.xyz	-2716.870000	Hp(1)+F(d1)
	CFp(s1)	1	1	0	1	CF.q1.m1-1.xyz	-3739.460000	Cp(d1)+F(d1)
	H3p(s1)	1	1	0	1	H3.q1.m1-1.xyz	-35.977800	Hp(1)+H2(s1)
	H2Cp(d1)	1	2	0	1	H2C.q1.m2-1.xyz	-1054.540000	H2(s1)+Cp(d1)
	HCFp(d1)	1	2	0	1	HCF.q1.m2-1.xyz	-3753.040000	Cp(d1)+HF(s1)
	H3Cp(s1)	1	1	0	1	H3C.q1.m1-1.xyz	-1073.720000	H(d1)+H2Cp(d1)
	H2CFp(s1)	1	1	0	1	H2CF.q1.m1-1.xyz	-3774.770000	H2(s1)+CFp(s1)
	H3CFp(d1)	1	2	0	1	H3CF.q1.m2-1.xyz	-3789.280000	H(d1)+H2CFp(s1)
	H3CFp(d2)	1	2	0	1	H3CF.q1.m2-2.xyz	-3789.520000	H(d1)+H2CFp(s1)

END FRAGMENTS_DATABASE

<THE NEXT STEP IS TO REPLACE THE PREVIOUSLY GENERATED BLOCK BY THE EQUIVALENT ONE IN THE CH3F+.m3c FILE>

<THE M3C FILE IS READY TO BE EXECUTED>

user@hostname:~/M3C-examples/methylFluoride/results\$ M3C -i CH3F+.m3c

Input file = CH3F+.m3c

Blocks input file

> BEGIN GOPTIONS
> systemRadius = 8.0
> overlappingRadius = 0.4
> useRandomWalkers = FALSE

...

START TIME: Wed Jan 14 16:10:45 2015

GOPTIONS:zero = 0.10E-11
GOPTIONS:systemRadius = 8.00000 A
GOPTIONS:randomWalkStepRadius = 1.00000 A
GOPTIONS:printLevel = 1
GOPTIONS:debugLevel = 1

+-----+
| FRAGMENTS DATABASE INITIALIZATION |
+-----+

```

file name = H.g0.m2-1.rxyz
name = H(d1)
Moments of inertia = [ 0.00000 0.00000 0.00000 ] amu*angs**2
Moments of inertia = [ 0.00000 0.00000 0.00000 ] a.u.
Radius = 0.37000 A
Eelec = -13.5721000 eV
Eelec = -0.4987653 a.u.
Mass = 1.0080000 amu
(fr, fv) = ( 0 0 )
maxEvib = 0.0000000 eV

```

...

```

+-----+
| END FRAGMENTS DATABASE INITIALIZATION |
+-----+

```

```

reactives = H3Cfp(d1)
excitationEnergy = 10.00000 eV
numberOfEvents = 10000
numberOfExperiments = 2
task = V,T,S:0,V,T,S:1:-1
geometryHistoryFilePrefix = geom
freqBlockingCheck = 4
track = energy

```

```

#-----#
# ENERGY HISTORY
#-----#
#
```

	trans eV	intermol eV	elec eV	vib eV	rot eV	tot eV	formula
aV	4.03427	0.00000	-3787.41000	3.01840	1.07732	-3779.28000	H3Cp(s1)+F(d1)
pT	1.70911	0.00000	-3787.41000	5.34357	1.07732	-3779.28000	H3Cp(s1)+F(d1)
rS	2.11221	0.00000	-3787.41000	4.94047	1.07732	-3779.28000	H3Cp(s1)+F(d1)
rV	1.57511	0.00000	-3787.41000	4.87930	1.67559	-3779.28000	H3Cp(s1)+F(d1)

...

```

#-----#
# Channels histogram
#-----#
#
```

item	1	2	aver	desv
H3Cp+F	0.121	0.095	0.108	0.013
H+H2Cp+F	0.606	0.549	0.578	0.029
H2+HCp+F	0.273	0.356	0.315	0.042

item	1	2	aver	desv
H2(s1)+HCp(s1)+F(d1)	0.273	0.356	0.315	0.042
H(d1)+H2Cp(d1)+F(d1)	0.606	0.549	0.578	0.029
H3Cp(s1)+F(d1)	0.121	0.095	0.108	0.013

...

```

-----
ELAPSED TIME: 0 h 0 m 16 s
END TIME: Wed Jan 14 16:11:01 2015
-----

```

```

user@hostname$ M3C.analysis CH3F+.m3c energy
user@hostname$ M3C.analysis CH3F+.m3c ecorr Et.vs.Ev
user@hostname$ M3C.analysis CH3F+.m3c ecorr Er.vs.Ev
user@hostname$ M3C.analysis CH3F+.m3c ecorr Er.vs.Et
user@hostname$ M3C.analysis CH3F+.m3c species
user@hostname$ M3C.analysis CH3F+.m3c channels

```

<CHANGE "tracking = energy" BY "tracking = none" IN CH3F+.m3c FILE IN ORDER TO SAVE EXECUTION TIME>

```

user@hostname:~/results$ M3C.p -i CH3F+.m3c
Running: 0.00000, 1.00000 ... OK Time elapsed: 0h 0m 12s
Running: 2.00000, 3.00000 ... OK Time elapsed: 0h 0m 22s
Running: 4.00000, 5.00000 ... OK Time elapsed: 0h 0m 21s
Running: 6.00000, 7.00000 ... OK Time elapsed: 0h 0m 23s
Running: 8.00000, 9.00000 ... OK Time elapsed: 0h 0m 25s
Running: 10.00000, 11.00000 ... OK Time elapsed: 0h 0m 40s
Running: 12.00000, 13.00000 ... OK Time elapsed: 0h 0m 35s
Running: 14.00000, 15.00000 ... OK Time elapsed: 0h 0m 40s
Running: 16.00000, 17.00000 ... OK Time elapsed: 0h 0m 45s
Running: 18.00000, 19.00000 ... OK Time elapsed: 0h 0m 50s

```

```

Running: 20.00000, 21.00000 ... OK    Time elapsed: 0h 0m 50s
Running: 22.00000, 23.00000 ... OK    Time elapsed: 0h 0m 57s
Running: 24.00000, 25.00000 ... OK    Time elapsed: 0h 0m 57s
Running: 26.00000, 27.00000 ... OK    Time elapsed: 0h 0m 54s
Running: 28.00000, 29.00000 ... OK    Time elapsed: 0h 0m 57s
Running: 30.00000 ... OK              Time elapsed: 0h 0m 32s
                                         Total: 0h 10m 20s

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ M3C.analysis CH3F+.m3c fit_sBR

```

```
...
```

```

### ERROR ### M3CfitBR: HCFp has not been mapped.
                  Remove it from the input file

```

```
...
```

```

<THIS ERROR MEANS THAT HCF+ MOLECULE IS NOT INTO THE CALCULATED SET OF FRAGMENTS>
<IN ORDER TO FIX IT, COMMENT OUT THE LINE "HCFp 4.4 0.0 32.0" AND/OR MORE LINES AS REQUIRED BY USING THE CHARACTER #>

```

```

user@hostname:~/M3C-examples/methylFluoride/results$ M3C.analysis CH3F+.m3c fit_sBR
user@hostname:~/M3C-examples/methylFluoride/results$ M3C.analysis CH3F+.m3c fit_sfE
user@hostname:~/M3C-examples/methylFluoride/results$ M3C.analysis CH3F+.m3c S.vs.E "p"

```

RESULT

