

# Explant Culture Analysis

*Sean Nguyen*

## Overall Objective

### Load Libraries

```
library(tidyverse)
library(cowplot)
library(broom)
library(plotly)
```

### Import data

#### Convert data from 'wide' to 'long' format

```
# data
data1 <- data %>%
  gather(Sample, Count, 2:37)

# Separate samples by identifiers
data2 <- data1 %>%
  separate(Sample, into=c("Sample_ID", "Dilution_factor",
                        "Injection", "Tech_rep", sep = "_")) %>%
  select(-`_`)

# Standards
standards1 <- standards %>%
  gather(Sample, Count, 2:13)

standards2 <- standards1 %>%
  separate(Sample, into=c("Sample_ID", "When", "Dilution_factor",
                        "Nano_day", "Injection", "Tech_Rep", sep = "_")) %>%
  select(-`_`)
```

#### Factor the data into categorical variables

```
# Refactoring Columns for samples
data2$Sample_ID <- as.factor(data2$Sample_ID)
data2$Dilution_factor <- as.numeric(data2$Dilution_factor)
data2$Injection <- as.factor(data2$Injection)
data2$Tech_rep <- as.numeric(data2$Tech_rep)

data2
```

```
## # A tibble: 36,000 × 6
```

```
##      particle_size Sample_ID Dilution_factor Injection Tech_rep Count
## *          <dbl>      <fctr>          <dbl>      <fctr>      <dbl> <int>
## 1             0.5          1             125          1          0      0
## 2             1.5          1             125          1          0      0
## 3             2.5          1             125          1          0      0
## 4             3.5          1             125          1          0      0
## 5             4.5          1             125          1          0      0
## 6             5.5          1             125          1          0      0
## 7             6.5          1             125          1          0      0
## 8             7.5          1             125          1          0      0
## 9             8.5          1             125          1          0      0
## 10            9.5          1             125          1          0      0
## # ... with 35,990 more rows
```

*# Refactoring COlumnns for key*

```
key$Sample_ID <- as.factor(key$Sample_ID)
key$Animal <- as.factor(key$Animal)
key$Condition <- as.factor(key$Condition)
```

key

```
## # A tibble: 6 × 3
##   Sample_ID Animal Condition
##     <fctr> <fctr>    <fctr>
## 1         1  1373   Normal
## 2         2  1371 lowOxygen
## 3         3  1370   Normal
## 4         4  1370 lowOxygen
## 5         5  1373 lowOxygen
## 6         6  1371   Normal
```

*# Refactoring columns for standards*

```
standards2$Sample_ID <- as.factor(standards2$Sample_ID)
standards2$When <- as.factor(standards2$When)
standards2$Dilution_factor <- as.numeric(standards2$Dilution_factor)
standards2$Injection <- as.factor(standards2$Injection)
standards2$Nano_day <- as.numeric(standards2$Nano_day)
```

standards2

```
## # A tibble: 12,000 × 8
##   particle_size Sample_ID When Dilution_factor Nano_day Injection
## *          <dbl>      <fctr> <fctr>          <dbl>      <dbl>      <fctr>
## 1             0.5      std after             125          1          1
## 2             1.5      std after             125          1          1
## 3             2.5      std after             125          1          1
## 4             3.5      std after             125          1          1
## 5             4.5      std after             125          1          1
## 6             5.5      std after             125          1          1
## 7             6.5      std after             125          1          1
## 8             7.5      std after             125          1          1
## 9             8.5      std after             125          1          1
## 10            9.5      std after             125          1          1
## # ... with 11,990 more rows, and 2 more variables: Tech_Rep <chr>,
## #   Count <int>
```

## Back calculate standards

```
standards2 <- standards2 %>%
  mutate(True_Count=Dilution_factor*Count)

# Set the correct order of 'categorical factors'
standards2$Nano_day <- factor(standards2$Nano_day, levels=c('1'))
standards2$When <- factor(standards2$When, levels=c('before','after'))

standards2
```

```
## # A tibble: 12,000 × 9
##   particle_size Sample_ID   When Dilution_factor Nano_day Injection
##         <dbl>    <fctr> <fctr>         <dbl>    <fctr>    <fctr>
## 1         0.5      std  after          125      1        1
## 2         1.5      std  after          125      1        1
## 3         2.5      std  after          125      1        1
## 4         3.5      std  after          125      1        1
## 5         4.5      std  after          125      1        1
## 6         5.5      std  after          125      1        1
## 7         6.5      std  after          125      1        1
## 8         7.5      std  after          125      1        1
## 9         8.5      std  after          125      1        1
## 10        9.5      std  after          125      1        1
## # ... with 11,990 more rows, and 3 more variables: Tech_Rep <chr>,
## #   Count <int>, True_Count <dbl>
```

## Summarize three technical standard replicates

```
standards3 <- standards2 %>%
  group_by(particle_size,Sample_ID,When,Dilution_factor,Nano_day,Injection) %>%
  summarise( tech_N = length(True_Count),
             tech_mean = mean(True_Count),
             tech_sd = sd(True_Count),
             tech_se = tech_sd/sqrt(tech_N))

standards3
```

```
## Source: local data frame [4,000 x 10]
## Groups: particle_size, Sample_ID, When, Dilution_factor, Nano_day [?]
##
##   particle_size Sample_ID   When Dilution_factor Nano_day Injection
##         <dbl>    <fctr> <fctr>         <dbl>    <fctr>    <fctr>
## 1         0.5      std  before          125      1        1
## 2         0.5      std  before          125      1        2
## 3         0.5      std  after          125      1        1
## 4         0.5      std  after          125      1        2
## 5         1.5      std  before          125      1        1
## 6         1.5      std  before          125      1        2
## 7         1.5      std  after          125      1        1
## 8         1.5      std  after          125      1        2
## 9         2.5      std  before          125      1        1
## 10        2.5      std  before          125      1        2
## # ... with 3,990 more rows, and 4 more variables: tech_N <int>,
```

```
## # tech_mean <dbl>, tech_sd <dbl>, tech_se <dbl>
```

### Summarize standards by injection

```
standards4 <- standards3 %>%
  group_by(Nano_day, When, particle_size) %>%
  summarise( inj_N = length(tech_mean),
             inj_mean = mean(tech_mean),
             inj_sd = sd(tech_mean),
             inj_se = inj_sd/sqrt(inj_N))
standards4
```

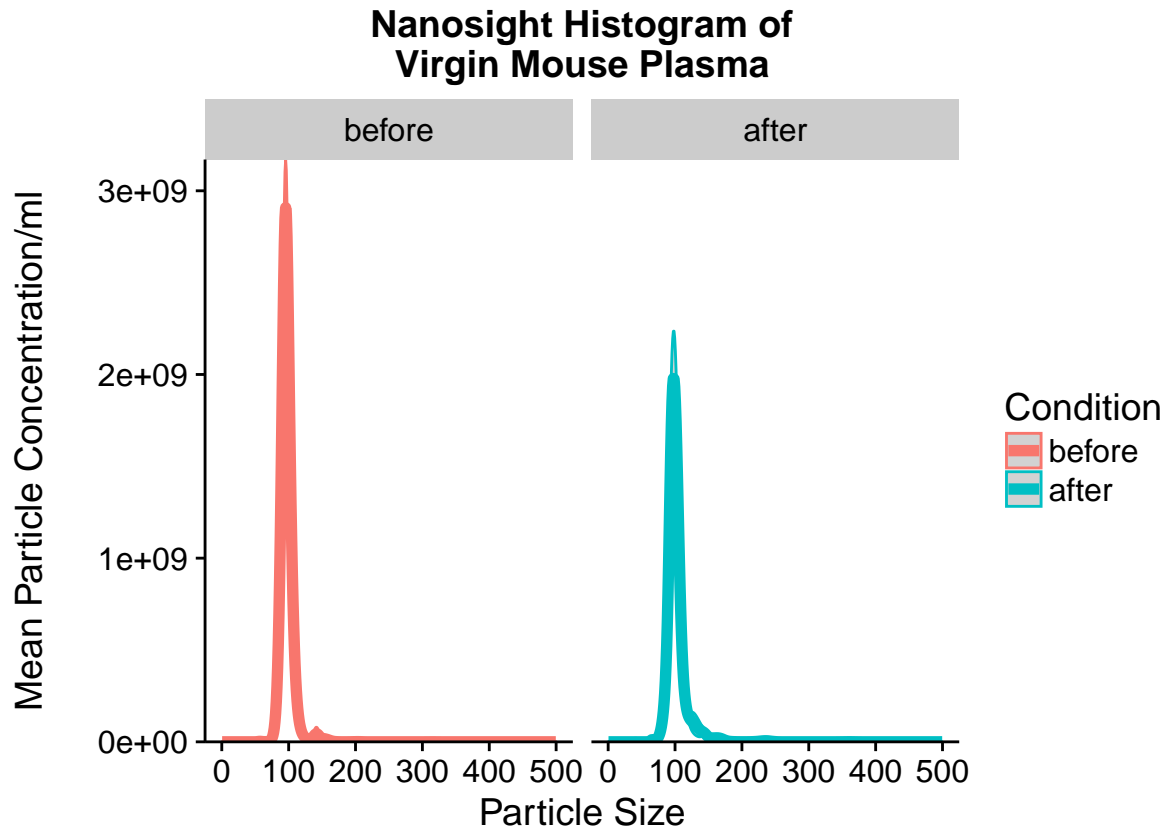
```
## Source: local data frame [2,000 x 7]
## Groups: Nano_day, When [?]
##
##   Nano_day  When particle_size inj_N inj_mean inj_sd inj_se
##   <fctr> <fctr>      <dbl> <int>   <dbl>  <dbl>  <dbl>
## 1      1 before         0.5     2      0      0      0
## 2      1 before         1.5     2      0      0      0
## 3      1 before         2.5     2      0      0      0
## 4      1 before         3.5     2      0      0      0
## 5      1 before         4.5     2      0      0      0
## 6      1 before         5.5     2      0      0      0
## 7      1 before         6.5     2      0      0      0
## 8      1 before         7.5     2      0      0      0
## 9      1 before         8.5     2      0      0      0
## 10     1 before         9.5     2      0      0      0
## # ... with 1,990 more rows
```

### Plot before and after plots, facet by experimental day

```
std_plot <- standards4 %>%
  ggplot(aes(x=particle_size, y=inj_mean, color=When))+
  geom_ribbon(aes(ymin=inj_mean-inj_se, ymax=inj_mean+inj_se),
            alpha=0.2, fill = alpha('grey12', 0.2)) + #error bars
  geom_line(size=2) + xlim(0,500)+ #line size, x-axis scale
  scale_y_continuous(expand=c(0,0))+ #set bottom of graph
  xlab("Particle Size") + # X axis label
  ylab("\nMean Particle Concentration/ml\n") + # Y axis label
  ggtitle("Nanosight Histogram of\nVirgin Mouse Plasma")+ #title
  labs(color="Condition")+ #Label table title
  facet_grid(. ~ When)

std_plot
```

```
## Warning: Removed 1000 rows containing missing values (geom_path).
```



Standards particle concentrations from each experimental day

```
standards_df <- standards4 %>%
  group_by(Nano_day,When) %>%
  summarise(total=sum(inj_mean))

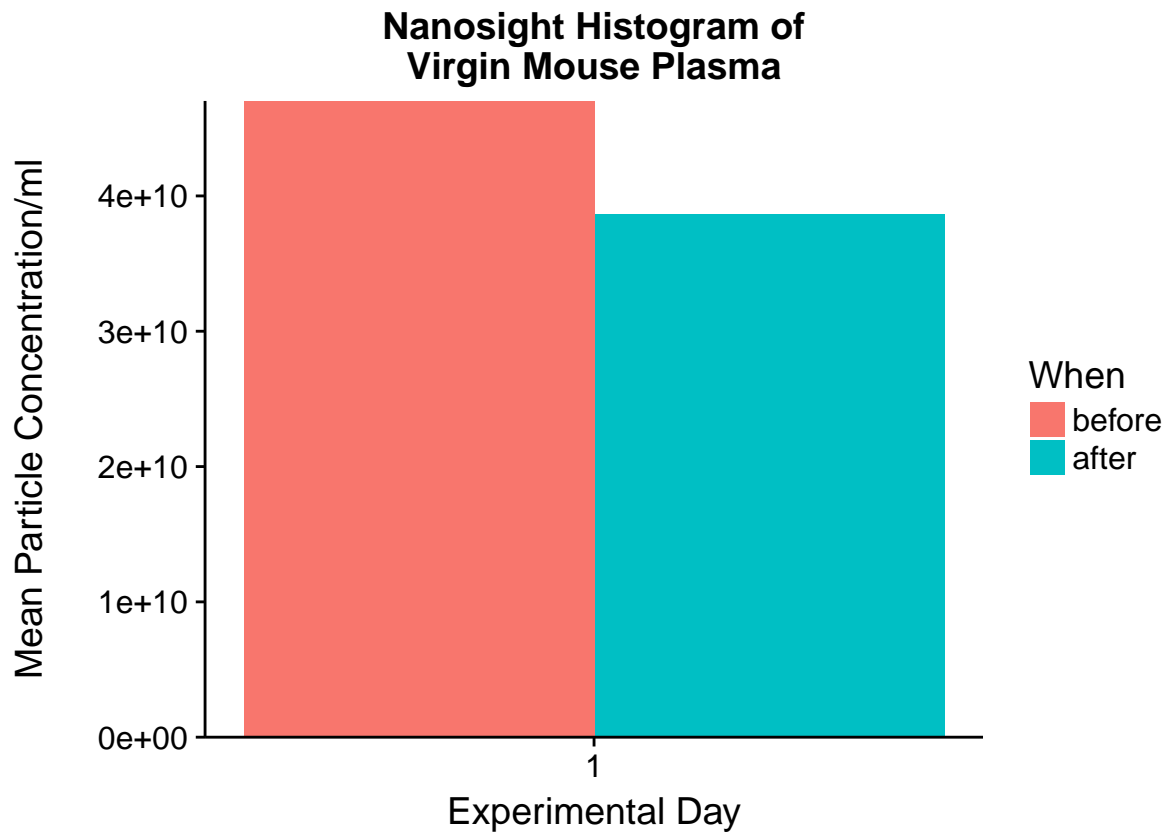
standards_df

## Source: local data frame [2 x 3]
## Groups: Nano_day [?]
##
##   Nano_day  When      total
##   <fctr> <fctr>    <dbl>
## 1       1 before 47020454229
## 2       1 after 38623334583
```

Bar graph of standards particle concentrations

```
standards_df %>%
  ggplot(aes(x=Nano_day,y=total,fill=When))+
  geom_col(position="dodge")+
  scale_y_continuous(expand=c(0,0))+ #set bottom of graph
  xlab("Experimental Day") + # X axis label
```

```
ylab("\nMean Particle Concentration/ml\n") + # Y axis label
ggtitle("Nanosight Histogram of\nVirgin Mouse Plasma")+ #title
labs(color="When") #Label table title
```



#### Intraassay variability

```
Intra.assay_cv <- standards_df %>%
  group_by(Nano_day) %>%
  summarise(Day_N = length(total),
            Day_mean = mean(total),
            Day_sd = sd(total),
            Day_se = Day_sd/sqrt(Day_N),
            Day_cv = Day_sd/Day_mean )
Intra.assay_cv
```

```
## # A tibble: 1 × 6
##   Nano_day Day_N   Day_mean   Day_sd   Day_se   Day_cv
##   <fctr> <int>     <dbl>     <dbl>     <dbl>     <dbl>
## 1       1     2 42821894406 5937660244 4198559823 0.1386594
```

## Sample analysis

Back calculate the original concentration of the sample

```
data2 <- data2 %>%
  mutate(True_Count = Dilution_factor*Count)
data2

## # A tibble: 36,000 × 7
##   particle_size Sample_ID Dilution_factor Injection Tech_rep Count
##   <dbl>         <fctr>         <dbl>      <fctr>    <dbl> <int>
## 1         0.5         1         125         1         0     0
## 2         1.5         1         125         1         0     0
## 3         2.5         1         125         1         0     0
## 4         3.5         1         125         1         0     0
## 5         4.5         1         125         1         0     0
## 6         5.5         1         125         1         0     0
## 7         6.5         1         125         1         0     0
## 8         7.5         1         125         1         0     0
## 9         8.5         1         125         1         0     0
## 10        9.5         1         125         1         0     0
## # ... with 35,990 more rows, and 1 more variables: True_Count <dbl>
```

Average three technical readings

```
data3 <- data2 %>%
  group_by(particle_size, Sample_ID, Dilution_factor, Injection) %>%
  summarise( tech_N = length(True_Count),
             tech_mean = mean(True_Count),
             tech_sd = sd(True_Count),
             tech_se = tech_sd/sqrt(tech_N))
data3

## Source: local data frame [12,000 x 8]
## Groups: particle_size, Sample_ID, Dilution_factor [?]
##
##   particle_size Sample_ID Dilution_factor Injection tech_N tech_mean
##   <dbl>         <fctr>         <dbl>      <fctr>    <int>    <dbl>
## 1         0.5         1         125         1         3         0
## 2         0.5         1         125         2         3         0
## 3         0.5         2         125         1         3         0
## 4         0.5         2         125         2         3         0
## 5         0.5         3         125         1         3         0
## 6         0.5         3         125         2         3         0
## 7         0.5         4         125         1         3         0
## 8         0.5         4         125         2         3         0
## 9         0.5         5         125         1         3         0
## 10        0.5         5         125         2         3         0
## # ... with 11,990 more rows, and 2 more variables: tech_sd <dbl>,
## #   tech_se <dbl>
```

## Summarize samples by injection (average both injections)

```
data4 <- data3 %>%
  group_by(particle_size, Sample_ID, Dilution_factor) %>%
  summarise( inj_N = length(tech_mean),
             inj_mean = mean(tech_mean),
             inj_sd = sd(tech_mean),
             inj_se = inj_sd/sqrt(inj_N))

data4

## Source: local data frame [6,000 x 7]
## Groups: particle_size, Sample_ID [?]
##
##   particle_size Sample_ID Dilution_factor inj_N inj_mean inj_sd inj_se
##   <dbl>         <fctr>         <dbl> <int>    <dbl>  <dbl>  <dbl>
## 1         0.5         1           125     2      0      0      0
## 2         0.5         2           125     2      0      0      0
## 3         0.5         3           125     2      0      0      0
## 4         0.5         4           125     2      0      0      0
## 5         0.5         5           125     2      0      0      0
## 6         0.5         6           125     2      0      0      0
## 7         1.5         1           125     2      0      0      0
## 8         1.5         2           125     2      0      0      0
## 9         1.5         3           125     2      0      0      0
## 10        1.5         4           125     2      0      0      0
## # ... with 5,990 more rows

# Average technical replicates and merge with key
merge <- left_join(key, data3, by= "Sample_ID")

merge

## # A tibble: 12,000 × 10
##   Sample_ID Animal Condition particle_size Dilution_factor Injection
##   <fctr> <fctr>    <fctr>         <dbl>         <dbl>    <fctr>
## 1         1  1373   Normal         0.5           125         1
## 2         1  1373   Normal         0.5           125         2
## 3         1  1373   Normal         1.5           125         1
## 4         1  1373   Normal         1.5           125         2
## 5         1  1373   Normal         2.5           125         1
## 6         1  1373   Normal         2.5           125         2
## 7         1  1373   Normal         3.5           125         1
## 8         1  1373   Normal         3.5           125         2
## 9         1  1373   Normal         4.5           125         1
## 10        1  1373   Normal         4.5           125         2
## # ... with 11,990 more rows, and 4 more variables: tech_N <int>,
## #   tech_mean <dbl>, tech_sd <dbl>, tech_se <dbl>

# Average injection replicates and merge with key
merge1 <- left_join(key, data4, by= "Sample_ID")

merge1

## # A tibble: 6,000 × 9
##   Sample_ID Animal Condition particle_size Dilution_factor inj_N inj_mean
```



```
##      <fctr> <fctr>      <fctr>      <dbl>      <dbl> <int>      <dbl>
## 1      1    1373    Normal      0.5      125     2        0
## 2      1    1373    Normal      1.5      125     2        0
## 3      1    1373    Normal      2.5      125     2        0
## 4      1    1373    Normal      3.5      125     2        0
## 5      1    1373    Normal      4.5      125     2        0
## 6      1    1373    Normal      5.5      125     2        0
## 7      1    1373    Normal      6.5      125     2        0
## 8      1    1373    Normal      7.5      125     2        0
## 9      1    1373    Normal      8.5      125     2        0
## 10     1    1373    Normal      9.5      125     2        0
## # ... with 5,990 more rows, and 2 more variables: inj_sd <dbl>,
## #   inj_se <dbl>
```

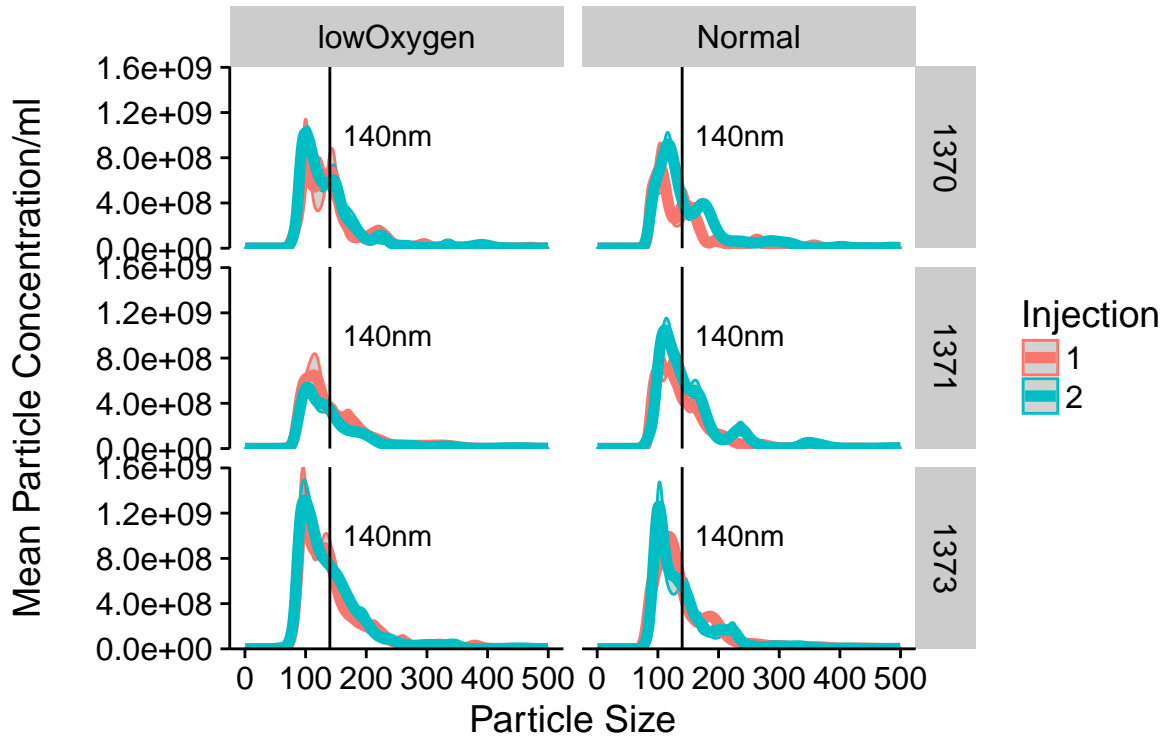
## Quick visualizations

### Graphing all samples

```
sample_plot <- merge %>%
  ggplot(aes(x=particle_size, y=tech_mean,color=Injection ))+ #plot
  geom_ribbon(aes(ymin=tech_mean-tech_se,
                 ymax=tech_mean+tech_se),
             alpha=0.2,fill = alpha('grey12', 0.2)) + #error bars
  geom_line(size=2.0) + xlim(0,500)+ #line size, x-axis scale
  scale_y_continuous(expand=c(0,0))+ #set bottom of graph
  xlab("Particle Size") + # X axis label
  ylab("\nMean Particle Concentration/ml\n") + # Y axis label
  ggtitle("Nanosight Histogram of\nVirgin Mouse Plasma")+ #title
  labs(color="Injection")+ #Label table title
  facet_grid(Animal ~ Condition)+
  geom_vline(xintercept = 140)+
  annotate("text", x= 235, y = 1E9, label= "140nm")

sample_plot
```

## Nanosight Histogram of Virgin Mouse Plasma



Particle concentration values for each of the samples

```
merge2 <- merge1 %>%
  group_by(Animal,Condition) %>%
  summarise(particle_conc=sum(inj_mean))
merge2
```

```
## Source: local data frame [6 x 3]
## Groups: Animal [?]
##
##   Animal Condition particle_conc
##   <fctr>    <fctr>         <dbl>
## 1   1370 lowOxygen  62711510438
## 2   1370   Normal  54048424354
## 3   1371 lowOxygen  46143447083
## 4   1371   Normal  65833232625
## 5   1373 lowOxygen  93505022625
## 6   1373   Normal  70609587271
```

Summary statistics of particle concentration (averaging n=6 for each time point)

```
merge3 <- merge2 %>%
  group_by(Condition) %>%
```

```

    summarise(Condition_N=length(particle_conc),
              Condition_mean = mean(particle_conc),
              Condition_sd = sd(particle_conc),
              Condition_se = Condition_sd/sqrt(Condition_N))
merge3

```

```

## # A tibble: 2 × 5
##   Condition Condition_N Condition_mean Condition_sd Condition_se
##   <fctr>      <int>      <dbl>      <dbl>      <dbl>
## 1 lowOxygen      3    67453326715  24034211567  13876158518
## 2 Normal         3    63497081417   8524155743   4921423613

```

## Boxplot

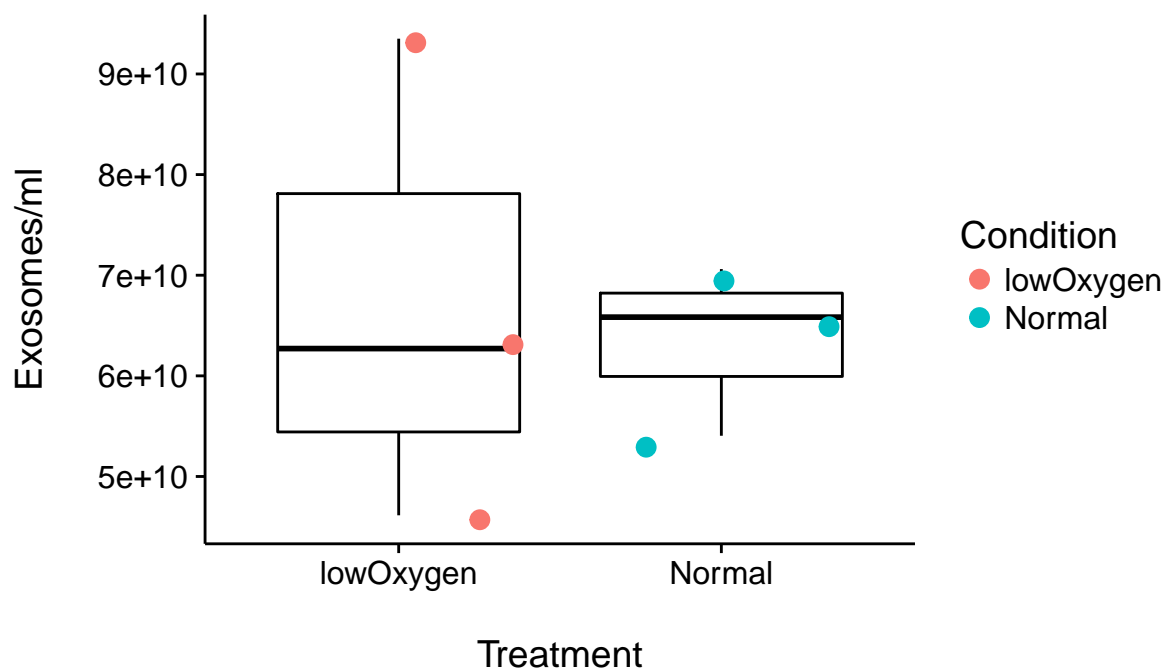
```

plot1 <- merge2 %>%
  #filter(!Animal== '1371' | !Condition == 'lowOxygen') %>%
  group_by(Condition) %>%
  ggplot(aes(x= Condition, y = particle_conc, color=Condition)) +
  geom_boxplot(colour="black", fill=NA) +
  geom_point(aes(text = paste("Animal:", Animal)),
             position='jitter', size=3)+
  xlab("\nTreatment\n") + # X axis label
  ylab("\nExosomes/ml\n") + # Y axis label
  ggtitle("GD 17.5 Placental Exosome \nExplant Culture (Ultracentrifugation)\n")+ #title
  labs(color="Condition") # Label table title

plot1

```

## GD 17.5 Placental Exosome Explant Culture (Ultracentrifugation)



##Interactive Plot

```
# ggplotly(plot1)
```

## Statistics

```
fit <- t.test(particle_conc ~ Condition,data=merge2)
```

```
tidy(fit)
```

```
##      estimate estimate1 estimate2 statistic  p.value parameter
## 1 3956245299 67453326715 63497081417  0.268711 0.8087912  2.495319
##      conf.low  conf.high                method alternative
## 1 -48744278892 56656769489 Welch Two Sample t-test    two.sided
```