

Nanosight Analysis

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Overall Objective

Load Libraries

```
library(tidyverse)
library(cowplot)
library(broom)
library(pwr)
```

Import data

```
setwd("~/GitHub/time-course/data")

rawdata <- "revised_MASTER-ExperimentSummary.csv"
timecourse <- "timecourse2017.csv"

data <- read_csv(rawdata)
tc <- read_csv(timecourse, na = c("", "NA"))
```

Convert data from ‘wide’ to ‘long’ format

```
data1 <- data %>%
  gather(Sample, Count, 2:250)

# Separate samples by identifiers
data2 <- data1 %>%
  separate(Sample, into=c("Sample_ID", "Dilution_factor", "Injection", "Tech_rep", sep = "_")) %>%
  select(-`_`)
```

Factor the data into categorical variables

```
# Refactoring Columns for samples
data2$Sample_ID <- as.factor(data2$Sample_ID)
data2$Dilution_factor <- as.numeric(data2$Dilution_factor)
data2$Injection <- as.factor(data2$Injection)
data2$Tech_rep <- as.numeric(data2$Tech_rep)

# Refactoring Columns for timecourse
tc$Sample_ID <- as.factor(tc$Sample_ID)
tc$Day <- as.factor(tc$Day)
```

```
tc$Weight <- as.numeric(tc$Weight)
tc$TEI_Day <- as.factor(tc$TEI_Day)
```

```
tc1 <- tc %>%
  select(Day:Pups)
tc1
```

```
## # A tibble: 40 × 5
##   Day Sample_ID TEI_Day Weight Pups
##   <fctr>   <fctr>   <fctr>   <dbl> <int>
## 1     1         1       6  19.60     0
## 2     1         2       2  18.60     0
## 3     1         3       1  19.30     0
## 4     1         4       3  20.50     0
## 5     1         5       4  18.74     0
## 6     1         6       5  18.30     0
## 7     5         7       6  20.83    10
## 8     5         8       3  19.88    10
## 9     5         9       2  24.41     9
## 10    5        10       4  20.01     6
## # ... with 30 more rows
```

Back calculate the original concentration of the sample

```
data2 <- data2 %>%
  mutate(True_Count=Dilution_factor*Count)
data2
```

```
## # A tibble: 249,000 × 7
##   particle_size Sample_ID Dilution_factor Injection Tech_rep Count
##           <dbl>   <fctr>           <dbl>   <fctr>     <dbl> <int>
## 1           0.5         2           500         1         0     0
## 2           1.5         2           500         1         0     0
## 3           2.5         2           500         1         0     0
## 4           3.5         2           500         1         0     0
## 5           4.5         2           500         1         0     0
## 6           5.5         2           500         1         0     0
## 7           6.5         2           500         1         0     0
## 8           7.5         2           500         1         0     0
## 9           8.5         2           500         1         0     0
## 10          9.5         2           500         1         0     0
## # ... with 248,990 more rows, and 1 more variables: True_Count <dbl>
```

Average the three technical replicate readings

```
data3 <- data2 %>%
  group_by(particle_size,Sample_ID,Dilution_factor,Injection) %>%
  summarise( tech_N = length(True_Count),
             tech_mean = mean(True_Count),
             tech_sd = sd(True_Count),
             tech_se = tech_sd/sqrt(tech_N))
data3
```

```
## Source: local data frame [82,000 x 8]
## Groups: particle_size, Sample_ID, Dilution_factor [?]
##
##   particle_size Sample_ID Dilution_factor Injection tech_N tech_mean
##           <dbl>    <fctr>           <dbl>    <fctr>  <int>    <dbl>
## 1             0.5         1             500         1         3         0
## 2             0.5         1             500         2         3         0
## 3             0.5        10             500         1         3         0
## 4             0.5        10             500         2         3         0
## 5             0.5        11             500         1         3         0
## 6             0.5        11             500         2         3         0
## 7             0.5        12             500         1         6         0
## 8             0.5        12             500         2         3         0
## 9             0.5        13             500         1         3         0
## 10            0.5        13             500         2         3         0
## # ... with 81,990 more rows, and 2 more variables: tech_sd <dbl>,
## #   tech_se <dbl>
test1 <- left_join(tc1,data3, by= "Sample_ID")
```

Summarize samples by injection (average both injections)

```
data4 <- data3 %>%
  group_by(particle_size,Sample_ID,Dilution_factor) %>%
  summarise( inj_N = length(tech_mean),
             inj_mean = mean(tech_mean),
             inj_sd = sd(tech_mean),
             inj_se = inj_sd/sqrt(inj_N))
data4
```

```
## Source: local data frame [41,000 x 7]
## Groups: particle_size, Sample_ID [?]
##
##   particle_size Sample_ID Dilution_factor inj_N inj_mean inj_sd inj_se
##           <dbl>    <fctr>           <dbl> <int>    <dbl>  <dbl>  <dbl>
## 1             0.5         1             500     2         0         0         0
## 2             0.5        10             500     2         0         0         0
## 3             0.5        11             500     2         0         0         0
## 4             0.5        12             500     2         0         0         0
## 5             0.5        13             500     2         0         0         0
## 6             0.5        14             500     2         0         0         0
## 7             0.5        15             500     2         0         0         0
## 8             0.5        16             500     2         0         0         0
## 9             0.5        17             500     2         0         0         0
## 10            0.5        18             500     2         0         0         0
## # ... with 40,990 more rows
test2 <- left_join(tc1,data4, by= "Sample_ID")
test2
```

```
## # A tibble: 41,000 × 11
##   Day Sample_ID TEI_Day Weight  Pups particle_size Dilution_factor
##   <fctr>    <chr>  <fctr>  <dbl> <int>    <dbl>          <dbl>
```

```
## 1      1      1      6  19.6    0      0.5      500
## 2      1      1      6  19.6    0      1.5      500
## 3      1      1      6  19.6    0      2.5      500
## 4      1      1      6  19.6    0      3.5      500
## 5      1      1      6  19.6    0      4.5      500
## 6      1      1      6  19.6    0      5.5      500
## 7      1      1      6  19.6    0      6.5      500
## 8      1      1      6  19.6    0      7.5      500
## 9      1      1      6  19.6    0      8.5      500
## 10     1      1      6  19.6    0      9.5      500
## # ... with 40,990 more rows, and 4 more variables: inj_N <int>,
## #   inj_mean <dbl>, inj_sd <dbl>, inj_se <dbl>
```

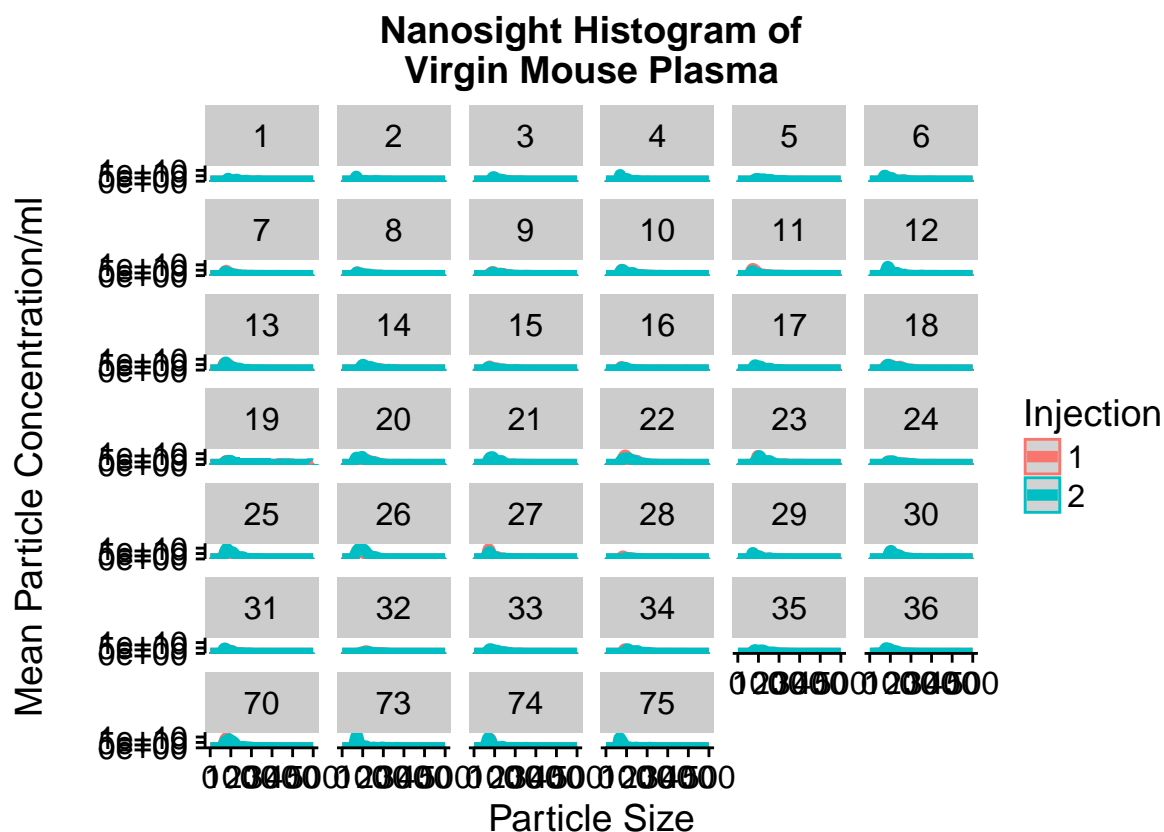
Quick visualizations

Graphing all samples

```
test1$Sample_ID_correct = factor(test1$Sample_ID, levels=c('1','2','3','4','5','6','7','8','9','10','11'))

graph1 <- test1 %>%
  ggplot(aes(x=particle_size, y=tech_mean,color=Injection ))+ #plot
  geom_ribbon(aes(ymin=tech_mean-tech_se, ymax=tech_mean+tech_se),alpha=0.2,fill = alpha('grey12', 0.2))
  geom_line(size=2.0) + xlim(0,500)+ #line size, x-axis scale
  scale_y_continuous(expand=c(0,0))+ #set bottom of graph
  xlab("Particle Size") + # X axis label
  ylab("\nMean Particle Concentration/ml\n") + # Y axis label
  ggtitle("Nanosight Histogram of\nVirgin Mouse Plasma")+ #title
  labs(color="Injection")+ #Label table title
  facet_wrap( ~ Sample_ID_correct, nrow=7)

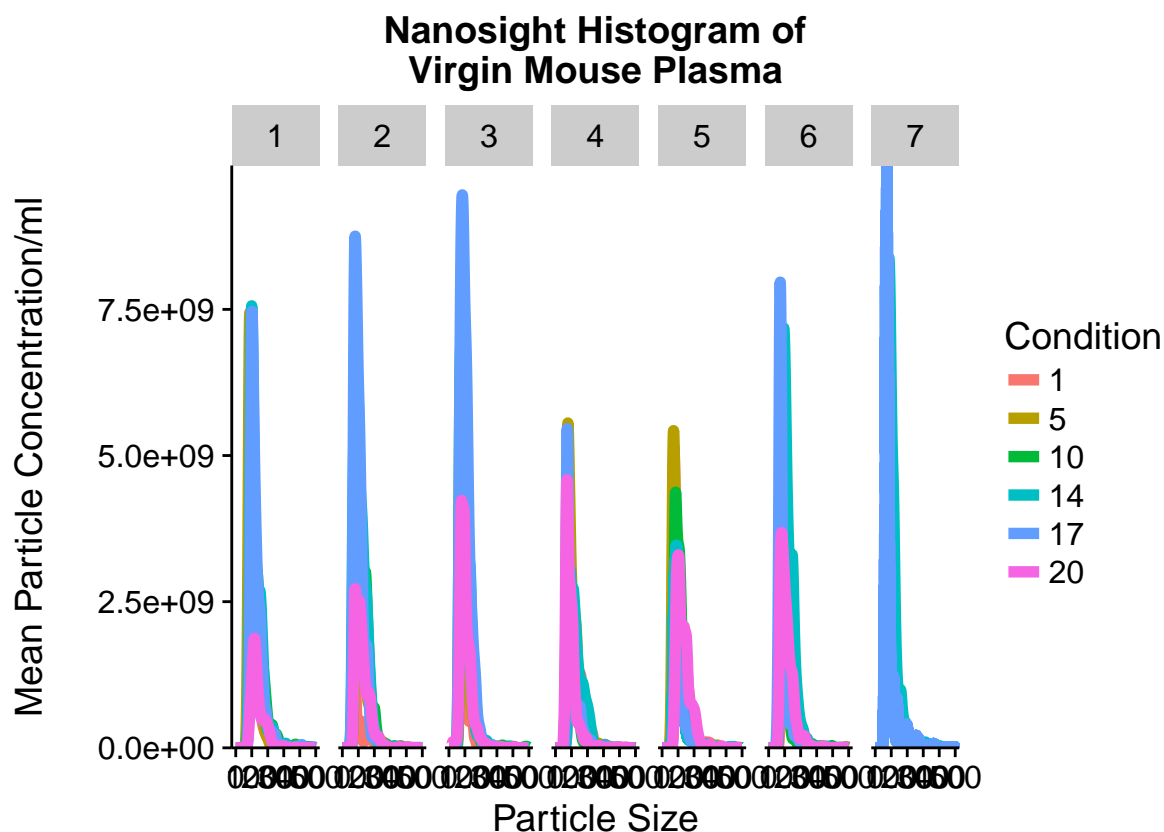
graph1
```



Graphing averaged samples by experimental day

```
graph2 <- test2 %>%
  group_by(TEI_Day) %>%
  ggplot(aes(x=particle_size, y=inj_mean,color=Day ))+ #plot
  #geom_ribbon(aes(ymin=inj_mean-inj_se, ymax=inj_mean+inj_se),alpha=0.2,fill = alpha('grey12', 0.2)) +
  geom_line(size=2) + xlim(0,500)+ #line size, x-axis scale
  scale_y_continuous(expand=c(0,0))+ #set bottom of graph
  xlab("Particle Size") + # X axis label
  ylab("\nMean Particle Concentration/ml\n") + # Y axis label
  ggtitle("Nanosight Histogram of\nVirgin Mouse Plasma")+ #title
  labs(color="Condition")+ #Label table title
  facet_wrap(~ TEI_Day, ncol=7)

graph2
```



Particle concentration values for each of the 36 samples

```
test3 <- test2 %>%
  group_by(Day, Sample_ID) %>%
  summarise(particle_conc = sum(inj_mean))
test3
```

```
## Source: local data frame [40 x 3]
## Groups: Day [?]
##
##      Day Sample_ID particle_conc
##    <fctr>    <chr>         <dbl>
## 1      1         1      124235076333
## 2      1         2      89249062500
## 3      1         3      175595167167
## 4      1         4      153741474667
## 5      1         5      182057049833
## 6      1         6      282458569250
## 7      5        10      306075805167
## 8      5        11      241497990000
## 9      5        12      336677212208
## 10     5         7      167230730583
## # ... with 30 more rows
```

Summary statistics of particle concentration (averaging n=6 for each time point)

```
test4 <- test3 %>%
  group_by(Day) %>%
  summarise(Day_N=length(particle_conc),
            Day_mean = mean(particle_conc),
            Day_sd = sd(particle_conc),
            Day_se = Day_sd/sqrt(Day_N))
test4
```

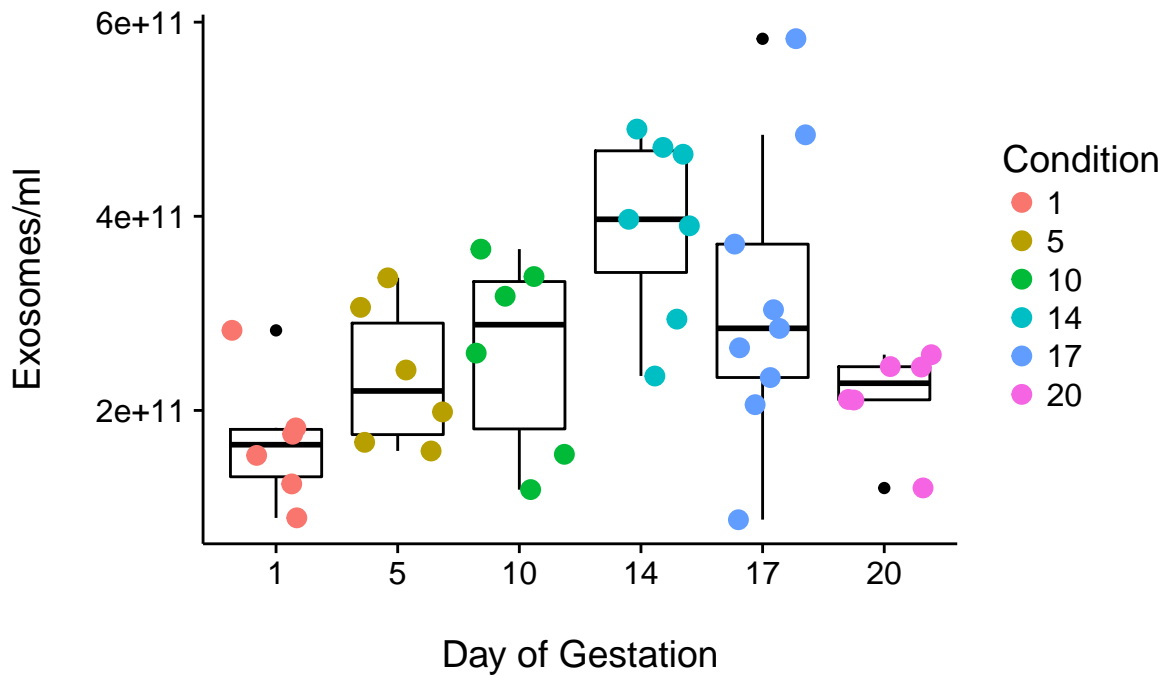
A tibble: 6 × 5

##	Day	Day_N	Day_mean	Day_sd	Day_se
##	<fctr>	<int>	<dbl>	<dbl>	<dbl>
## 1	1	6	167889399958	65842785695	26880204699
## 2	5	6	234722109938	73806691215	30131455513
## 3	10	6	258974135458	101749179329	41538928517
## 4	14	7	391625801212	95846005651	36226385016
## 5	17	9	313145842646	148840936571	49613645524
## 6	20	6	214896696056	50294400099	20532602860

Boxplot

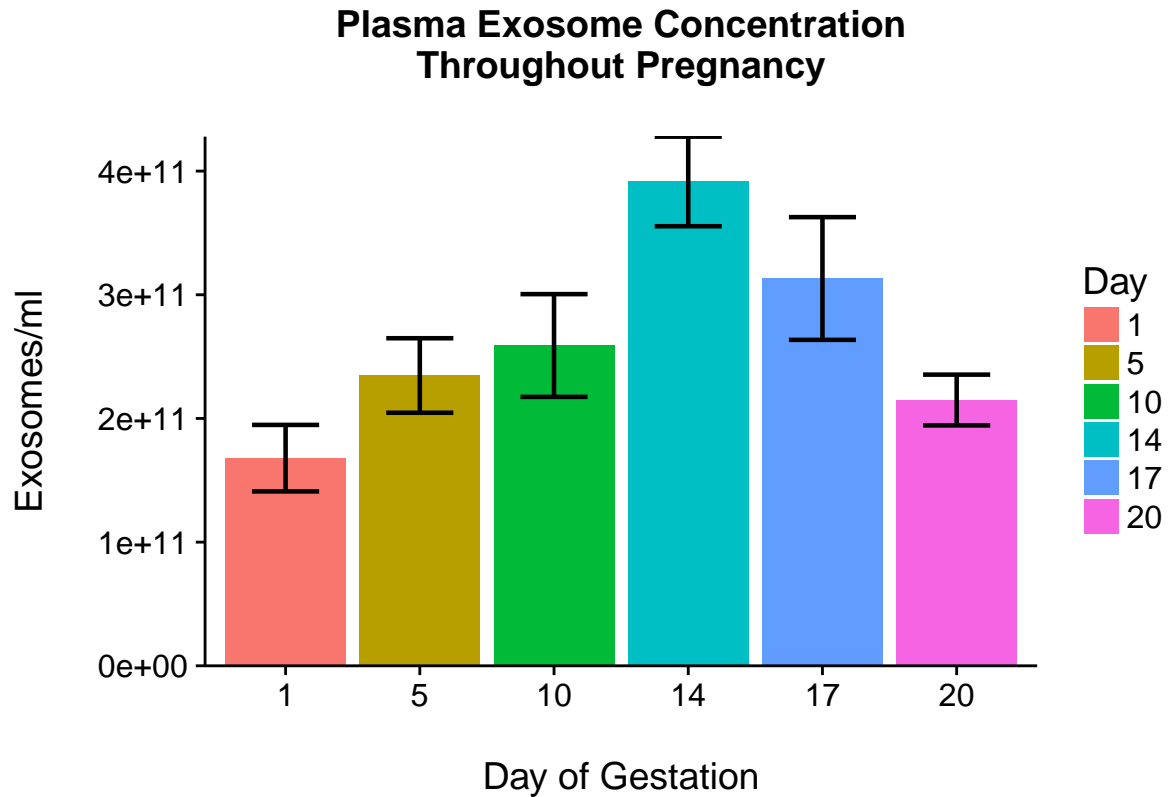
```
plot1 <- test3 %>%
  group_by(Day) %>%
  ggplot(aes(factor(Day),particle_conc, color=Day)) +
  geom_boxplot(colour="black",fill=NA) +
  geom_point(position='jitter',size=3)+
  xlab("\nDay of Gestation\n") + # X axis label
  ylab("\nExosomes/ml\n") + # Y axis label
  ggtitle("Plasma Exosome Concentration\nThroughout Pregnancy\n")+ #title
  labs(color="Condition")#Label table title
plot1
```

Plasma Exosome Concentration Throughout Pregnancy



Bar plot

```
plot <- test4 %>%
  ggplot(aes(x=Day, y=Day_mean, fill=Day ))+ #plot
  geom_col()+
  geom_errorbar(aes(ymin=Day_mean-Day_se, ymax=Day_mean+Day_se), width=.5,
                size=0.8, colour="black", position=position_dodge(.9)) + #error bars
  scale_y_continuous(expand=c(0,0))+ #set bottom of graph
  xlab("\nDay of Gestation\n") + # X axis label
  ylab("\nExosomes/ml\n") + # Y axis label
  ggtitle("Plasma Exosome Concentration\nThroughout Pregnancy\n")+ #title
  labs(color="Condition")#Label table title
plot
```

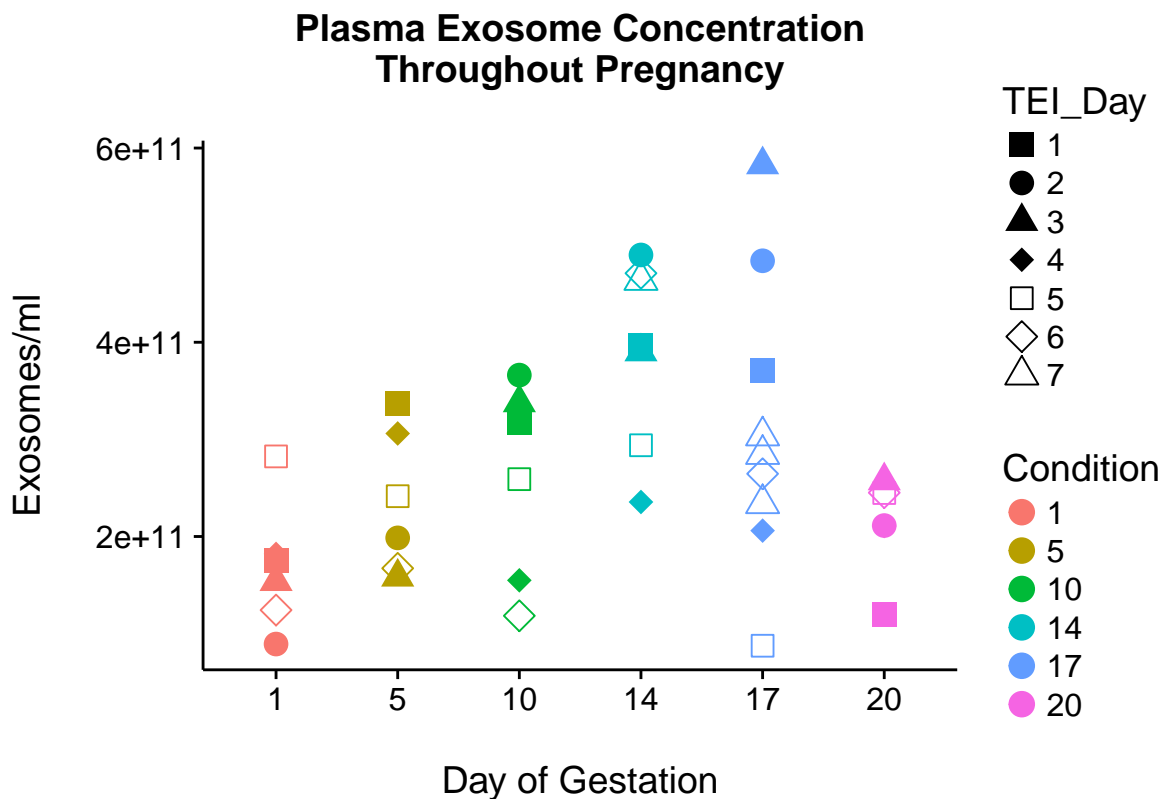
Looking at Variation between the days the samples were run

```
test7 <- test3 %>%
  left_join(tc1)

## Joining, by = c("Day", "Sample_ID")

plot2 <- test7 %>%
  ggplot(aes(x = Day, y = particle_conc, color = Day, shape=TEI_Day))+
  geom_point(position= 'dodge',size=4)+
  scale_shape_manual(values=c(15,16,17,18,22,23,24))+
  xlab("\nDay of Gestation\n") + # X axis label
  ylab("\nExosomes/ml\n") + # Y axis label
  ggtitle("Plasma Exosome Concentration\nThroughout Pregnancy\n")+ #title
  labs(color="Condition")#Label table title

plot2
```



Statistics

Shapiro test

```
tidy(shapiro.test(test3$particle_conc))
```

```
##      statistic    p.value          method
## 1 0.9577149 0.1398459 Shapiro-Wilk normality test
```

p value > 0.05 therefore conclude data is normally distributed

ANOVA

```
fit <- aov(particle_conc ~ Day, data=test3)
stats <- tidy(fit)
stats
```

```
##      term df      sumsq      meansq statistic    p.value
## 1      Day  5 2.092696e+23 4.185391e+22  4.116699 0.004967575
## 2 Residuals 34 3.456733e+23 1.016686e+22      NA      NA
```

Statistically significant, thus Tukey's HSD post hoc analysis can determine significant differences.

Tukey Post Hoc Test

```
HSD <- TukeyHSD(fit)
tukey <- tidy(HSD)
tukey
```

##	term	comparison	estimate	conf.low	conf.high	adj.p.value
## 1	Day	5-1	66832709979	-108873429601	242538849559	0.857604669
## 2	Day	10-1	91084735500	-84621404080	266790875080	0.626373082
## 3	Day	14-1	223736401254	54421728588	393051073919	0.004172081
## 4	Day	17-1	145256442688	-15140584212	305653469588	0.094520772
## 5	Day	20-1	47007296097	-128698843483	222713435677	0.964178159
## 6	Day	10-5	24252025521	-151454114059	199958165101	0.998248763
## 7	Day	14-5	156903691274	-12410981391	326218363940	0.082369473
## 8	Day	17-5	78423732709	-81973294191	238820759609	0.681433255
## 9	Day	20-5	-19825413882	-195531553462	155880725698	0.999337197
## 10	Day	14-10	132651665754	-36663006912	301966338419	0.197356245
## 11	Day	17-10	54171707188	-106225319712	214568734088	0.908193136
## 12	Day	20-10	-44077439403	-219783578983	131628700177	0.972790824
## 13	Day	17-14	-78479958566	-231848850888	74888933757	0.638991899
## 14	Day	20-14	-176729105156	-346043777822	-7414432491	0.036582117
## 15	Day	20-17	-98249146591	-258646173491	62147880309	0.449695482

Significant Tukey Post Hoc Test Values

```
tukey %>%
  filter(adj.p.value<0.05) %>%
  arrange(adj.p.value)
```

##	term	comparison	estimate	conf.low	conf.high	adj.p.value
## 1	Day	14-1	223736401254	54421728588	393051073919	0.004172081
## 2	Day	20-14	-176729105156	-346043777822	-7414432491	0.036582117

Looking at nanoparticle range

```
nano_100 <- data4 %>%
  filter(particle_size<120.5)

nano_100_data <- left_join(tc1,nano_100, by= "Sample_ID")
nano_100_data %>%
  group_by(Day,Sample_ID) %>%
  summarise(particle_conc=sum(inj_mean)) %>%
  ggplot(aes(factor(Day),particle_conc, color=Day)) +
  geom_boxplot(colour="black",fill=NA) +
  geom_point(position='jitter',size=3)+
  xlab("\nDay of Gestation\n") + # X axis label
  ylab("\nExosomes/ml\n") + # Y axis label
  ggtitle("Plasma Exosome Concentration\nThroughout Pregnancy\n")+ #title
  labs(color="Condition")#Label table title
```

Plasma Exosome Concentration Throughout Pregnancy

