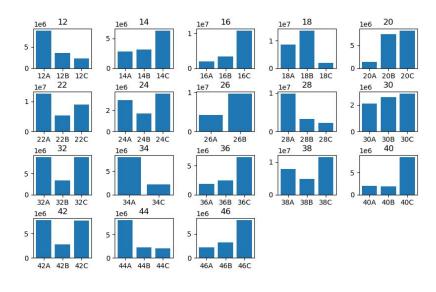
RNAseq - Internal Standard

RNAseq data is collected in a relative framework:

 Gene abundance is calculate as percent of sequence library

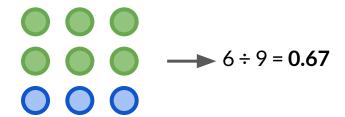
Limitation: cannot provide information on extent/direction of changes in any particular gene

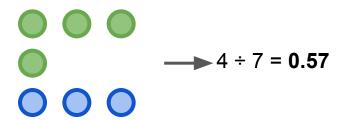
Counts of total reads mapped



Limitation: cannot provide information on extent/direction of changes in any particular gene

Example: a <u>decrease in</u> <u>expression</u> of a transcript can be due to either:

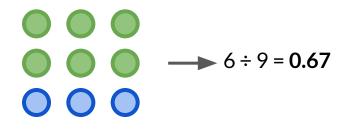




Limitation: cannot provide information on extent/direction of changes in any particular gene

Example: a <u>decrease in</u> <u>expression</u> of a transcript can be due to either:

1. A decrease in abundance of that transcript

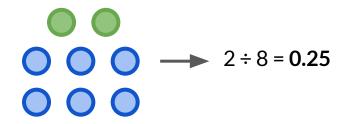


Limitation: cannot provide information on extent/direction of changes in any particular gene

Example: a decrease in expression of a transcript can be due to either:

- 1. A decrease in abundance of that transcript
- 2. An increase in the abundance of unrelated transcript



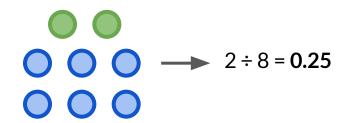


Limitation: cannot provide information on extent/direction of changes in any particular gene

Example: a decrease in expression of a transcript can be due to either:

- 1. A decrease in abundance of that transcript
- 2. An increase in the abundance of unrelated transcript
- → Hard to differentiate between the 2

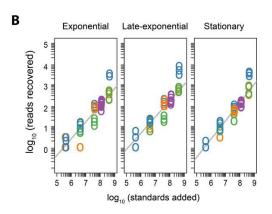




Methods Summary:

- Add many different standard molecules at different <u>known</u> concentrations
- Obtain <u>sequencing</u>
 <u>efficiency</u> from: standards
 added vs. standard
 recovered

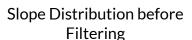
Group	ID	Length (nt)	Standards added	Symbol
A	Std. 5	979	441,334,667	0
	Std. 6	964	40,179,884	0
	Std. 1	1068	4,205,482	0
	Std. 12	991	428,059	0
В	Std. 11	996	410,023,533	0
	Std. 4	1028	39,362,526	0
	Std. 10	969	4,178,282	0
С	Std. 3	1000	408,554,669	0
	Std. 7	991	41,496,261	0
	Std. 8	1000	4,106,141	0
	Std. 2	1077	378,467	0
D	Std. 14	301	123,756,534	0
	Std. 13	613	119,426,056	0
	Std. 15	1504	115,793,038	0

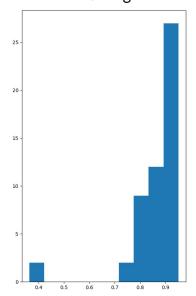


Gifford et al., 2016

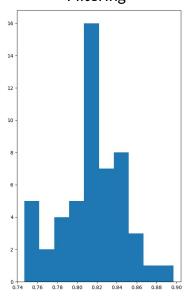
Methods Summary:

- Obtain median across all standards
- Fit linear line to median (global) standard data
- Filter standards to remove outliers
- Fit linear line to local standard data to obtain slope and intercept
- Use y=mx+b to correct counts of gene transcript
- Divide transcript count by cell count to obtain transcript/cell



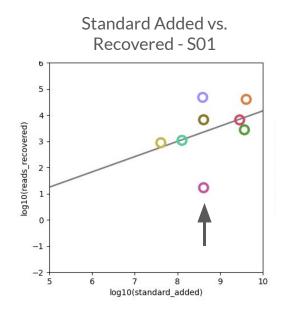


Slope Distribution after Filtering

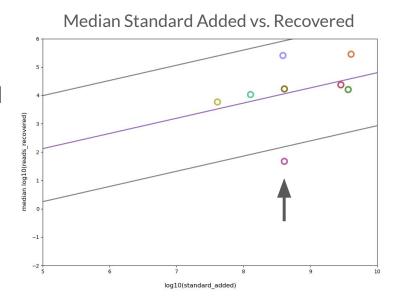


Standard Filtering:

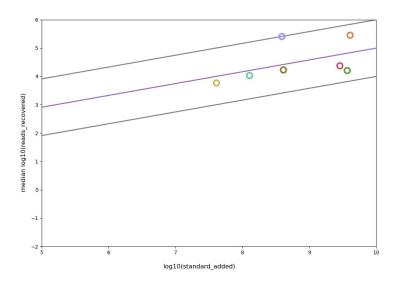
 Some standards are outliers → need to be removed



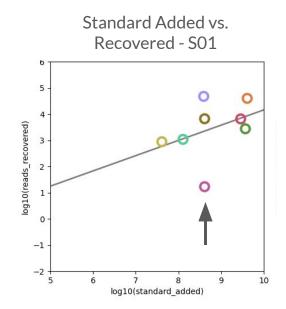
- Some standards are outliers → need to be removed
- Obtain median across all standards and filter standards <u>globally</u>
- Outliers are determined based on median of standards across samples



- Some standards are outliers → need to be removed
- Obtain median across all standards and filter standards <u>globally</u>
- Outliers are determined based on median of standards across samples



- Some standards are outliers → need to be removed
- Obtain median across all standards and filter standards <u>globally</u>
- Outliers are determined based on median of standards across samples
- Standards are then removed per sample



- Some standards are outliers → need to be removed
- Obtain median across all standards and filter standards <u>globally</u>
- Outliers are determined based on median of standards across samples
- Standards are then removed per sample
- Fit linear line to obtain slope and intercept for back-calculating transcript counts

