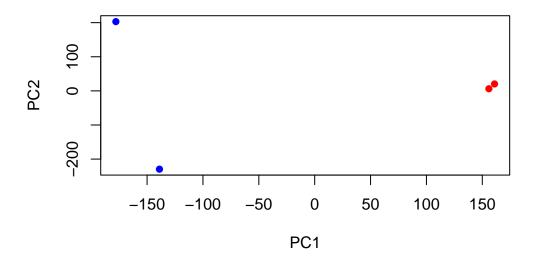
Lab 17: Analyzing Sequencing Data in the Cloud

Nicholas Chiu

Downstream Analysis

```
# Install Packages
  #BiocManager::install("tximport")
  #BiocManager::install("rhdf5")
  library(tximport)
  # setup the folder and filenames to read
  folders <- dir(pattern="SRR21568*")</pre>
  samples <- sub("_quant", "", folders)</pre>
  files <- file.path( folders, "abundance.h5" )</pre>
  names(files) <- samples</pre>
  txi.kallisto <- tximport(files, type = "kallisto", txOut = TRUE)</pre>
1 2 3 4
  head(txi.kallisto$counts)
                SRR2156848 SRR2156849 SRR2156850 SRR2156851
ENST00000539570
                                     0
                                          0.00000
ENST00000576455
                                          2.62037
                                                            0
                          0
                                     0.00000
ENST00000510508
ENST00000474471
                                     1 1.00000
ENST00000381700
                                     0.00000
ENST00000445946
                                          0.00000
```

```
colSums(txi.kallisto$counts)
SRR2156848 SRR2156849 SRR2156850 SRR2156851
  2563611
              2600800
                         2372309
                                     2111474
  sum(rowSums(txi.kallisto$counts)>0)
[1] 94561
  # Filtering
  to.keep <- rowSums(txi.kallisto$counts) > 0
  kset.nonzero <- txi.kallisto$counts[to.keep,]</pre>
  keep2 <- apply(kset.nonzero,1,sd)>0
  x <- kset.nonzero[keep2,]</pre>
PCA
  pca <- prcomp(t(x), scale=TRUE)</pre>
  summary(pca)
Importance of components:
                            PC1
                                      PC2
                                               PC3
                                                     PC4
Standard deviation
                       183.6379 177.3605 171.3020 1e+00
Proportion of Variance
                         0.3568
                                  0.3328
                                            0.3104 1e-05
Cumulative Proportion
                         0.3568
                                  0.6895
                                            1.0000 1e+00
  # Plot PCAs
  plot(pca$x[,1], pca$x[,2],
       col=c("blue","blue","red","red"),
       xlab="PC1", ylab="PC2", pch=16)
  ## Plot using ggplot
  library(ggplot2)
```

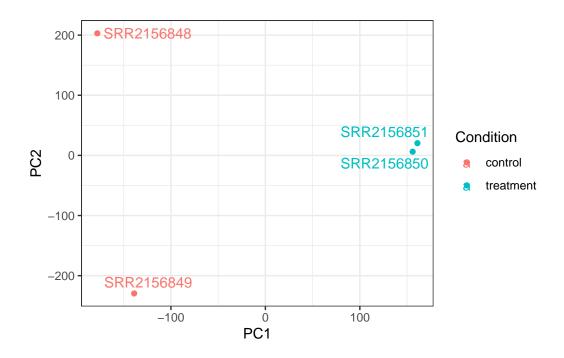


```
library(ggrepel)

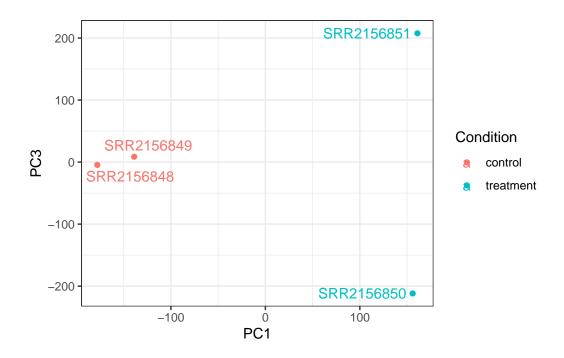
# Make metadata object for the samples
colData <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))
rownames(colData) <- colnames(txi.kallisto$counts)

# Make the data.frame for ggplot
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)

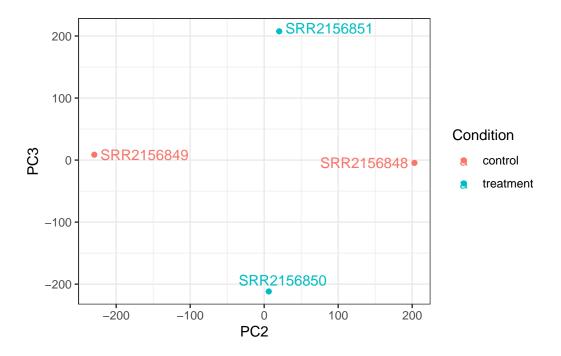
# PC1 vs PC2
ggplot(y) +
   aes(PC1, PC2, col=Condition) +
   geom_point() +
   geom_text_repel(label=rownames(y)) +
   theme_bw()</pre>
```



```
# PC1 vs PC3
ggplot(y) +
  aes(PC1, PC3, col=Condition) +
  geom_point() +
  geom_text_repel(label=rownames(y)) +
  theme_bw()
```



```
# PC2 vs PC3
ggplot(y) +
  aes(PC2, PC3, col=Condition) +
  geom_point() +
  geom_text_repel(label=rownames(y)) +
  theme_bw()
```



Differential-expression analysis

library(DESeq2)

Warning: package 'DESeq2' was built under R version 4.3.3

Loading required package: S4Vectors

Loading required package: stats4

Loading required package: BiocGenerics

Attaching package: 'BiocGenerics'

The following objects are masked from 'package:stats':

IQR, mad, sd, var, xtabs

The following objects are masked from 'package:base':

anyDuplicated, aperm, append, as.data.frame, basename, cbind, colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget, order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply, union, unique, unsplit, which.max, which.min

Attaching package: 'S4Vectors'

The following object is masked from 'package:utils':

findMatches

The following objects are masked from 'package:base':

expand.grid, I, unname

Loading required package: IRanges

Attaching package: 'IRanges'

The following object is masked from 'package:grDevices':

windows

Loading required package: GenomicRanges

Loading required package: GenomeInfoDb

Loading required package: SummarizedExperiment

Loading required package: MatrixGenerics

Loading required package: matrixStats

Attaching package: 'MatrixGenerics'

The following objects are masked from 'package:matrixStats':

colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse, colCounts, colCummaxs, colCummins, colCumprods, colCumsums, colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs, colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats, colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds, colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads, colWeightedMeans, colWeightedMedians, colWeightedSds, colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet, rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods, rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps, rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins, rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks, rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars, rowWeightedMads, rowWeightedMeans, rowWeightedMedians, rowWeightedSds, rowWeightedVars

Loading required package: Biobase

Welcome to Bioconductor

Vignettes contain introductory material; view with 'browseVignettes()'. To cite Bioconductor, see 'citation("Biobase")', and for packages 'citation("pkgname")'.

Attaching package: 'Biobase'

rowMedians

The following object is masked from 'package:MatrixGenerics':

The following objects are masked from 'package:matrixStats': anyMissing, rowMedians

```
sampleTable <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))</pre>
  rownames(sampleTable) <- colnames(txi.kallisto$counts)</pre>
  dds <- DESeqDataSetFromTximport(txi.kallisto,</pre>
                                    sampleTable,
                                    ~condition)
using counts and average transcript lengths from tximport
  dds <- DESeq(dds)
estimating size factors
using 'avgTxLength' from assays(dds), correcting for library size
estimating dispersions
gene-wise dispersion estimates
mean-dispersion relationship
-- note: fitType='parametric', but the dispersion trend was not well captured by the
   function: y = a/x + b, and a local regression fit was automatically substituted.
   specify fitType='local' or 'mean' to avoid this message next time.
final dispersion estimates
fitting model and testing
  res <- results(dds)</pre>
  head(res)
```

log2 fold change (MLE): condition treatment vs control

Wald test p-value: condition treatment vs control

NA

DataFrame with 6 rows and 6 columns

ENST00000445946

| Datarramo wrom o | TOWN GILL | O OOLUMIID | | | |
|------------------|---------------------|------------------------|---------------------|---------------------|---------------------|
| | baseMean | ${\tt log2FoldChange}$ | lfcSE | stat | pvalue |
| | <numeric></numeric> | <numeric></numeric> | <numeric></numeric> | <numeric></numeric> | <numeric></numeric> |
| ENST00000539570 | 0.000000 | NA | NA | NA | NA |
| ENST00000576455 | 0.761453 | 3.155061 | 4.86052 | 0.6491203 | 0.516261 |
| ENST00000510508 | 0.000000 | NA | NA | NA | NA |
| ENST00000474471 | 0.484938 | 0.181923 | 4.24871 | 0.0428185 | 0.965846 |
| ENST00000381700 | 0.000000 | NA | NA | NA | NA |
| ENST00000445946 | 0.000000 | NA | NA | NA | NA |
| | padj | | | | |
| | <numeric></numeric> | | | | |
| ENST00000539570 | NA | | | | |
| ENST00000576455 | NA | | | | |
| ENST00000510508 | NA | | | | |
| ENST00000474471 | NA | | | | |
| ENST00000381700 | NA | | | | |