

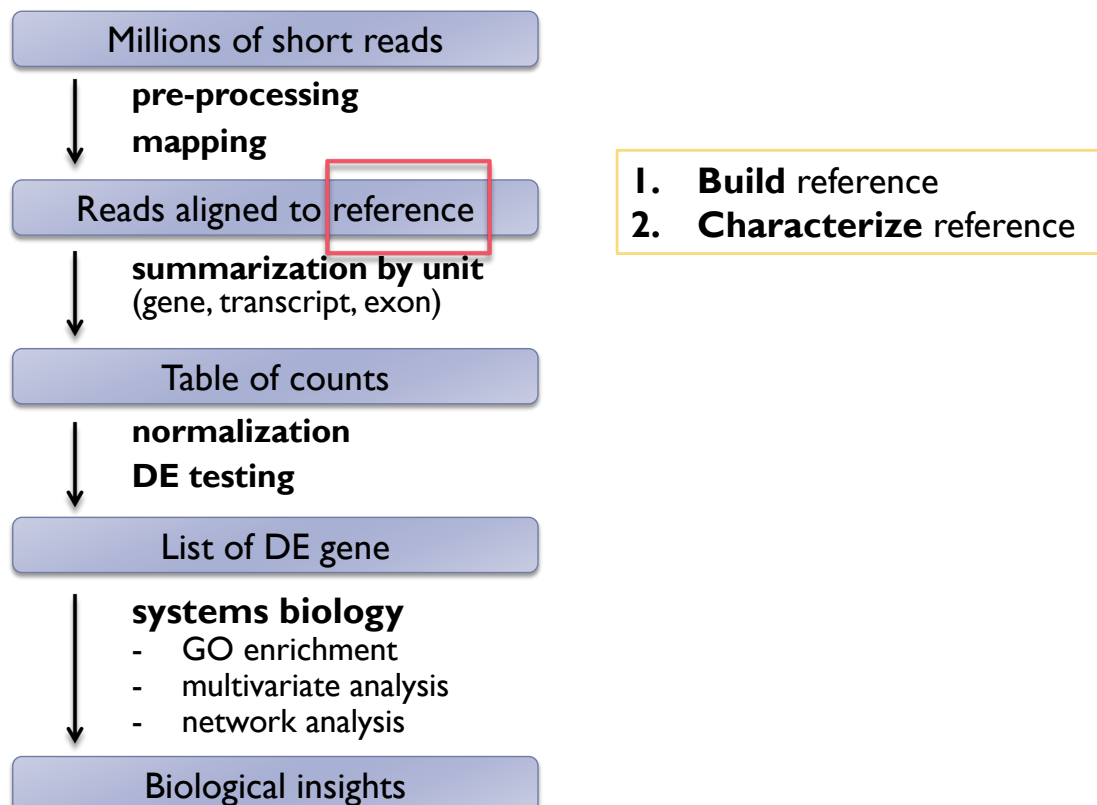
RNA-seq解析パイプライン： *de novo*

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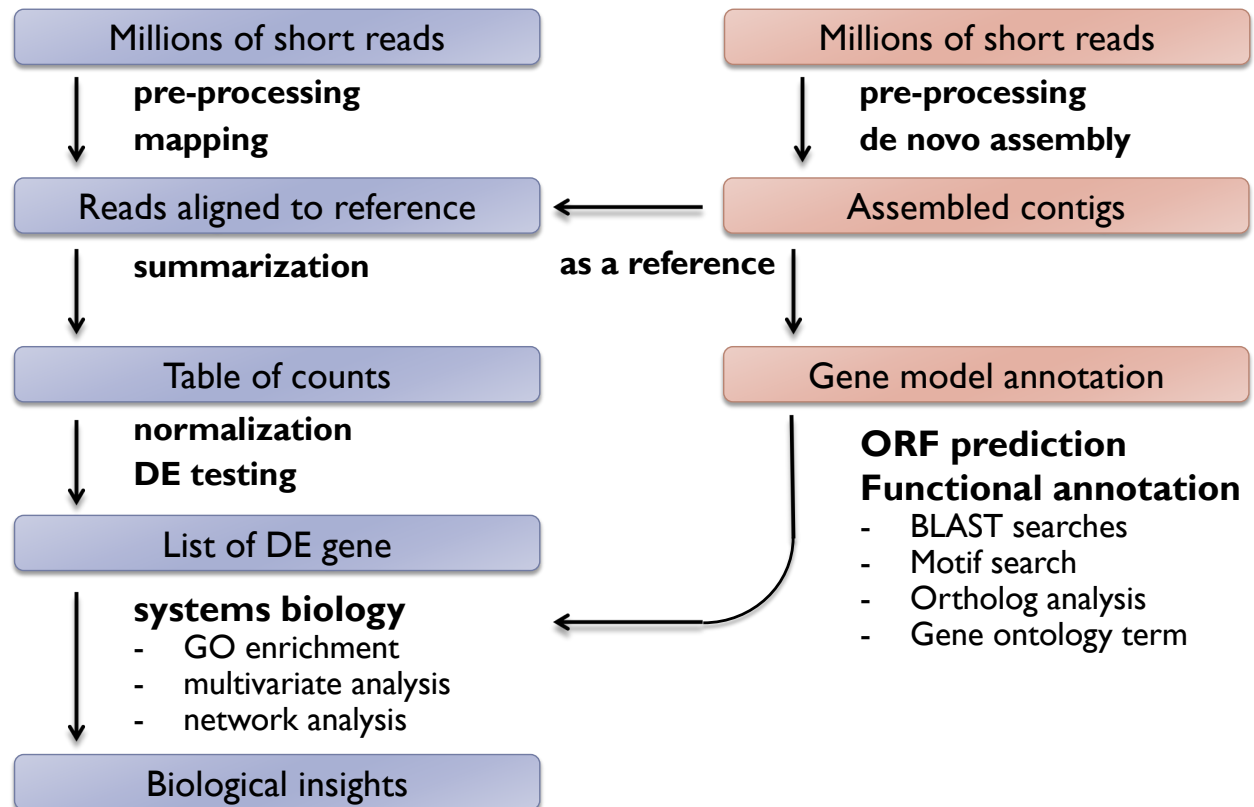
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de novo RNA-seq



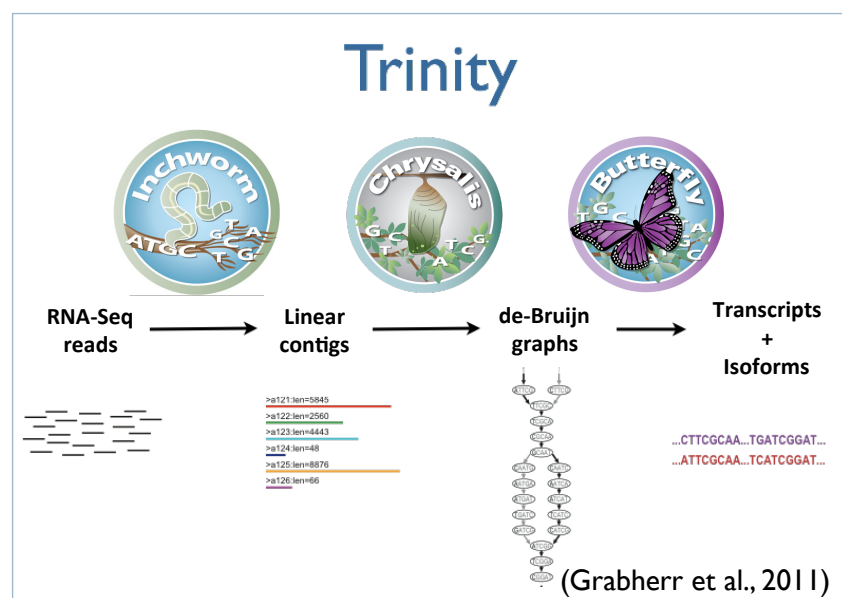
RNA-seq analysis pipeline (*de novo* strategy)



de novo assemblers of RNA-seq

De novo assemblers use reads to assemble transcripts directly, which does not depend on a reference genome.

- ▶ Trinity
- ▶ Oases
- ▶ TransAbyss
- ▶ EBARDenovo
- ▶ ...



<https://github.com/trinityrnaseq/trinityrnaseq/wiki>

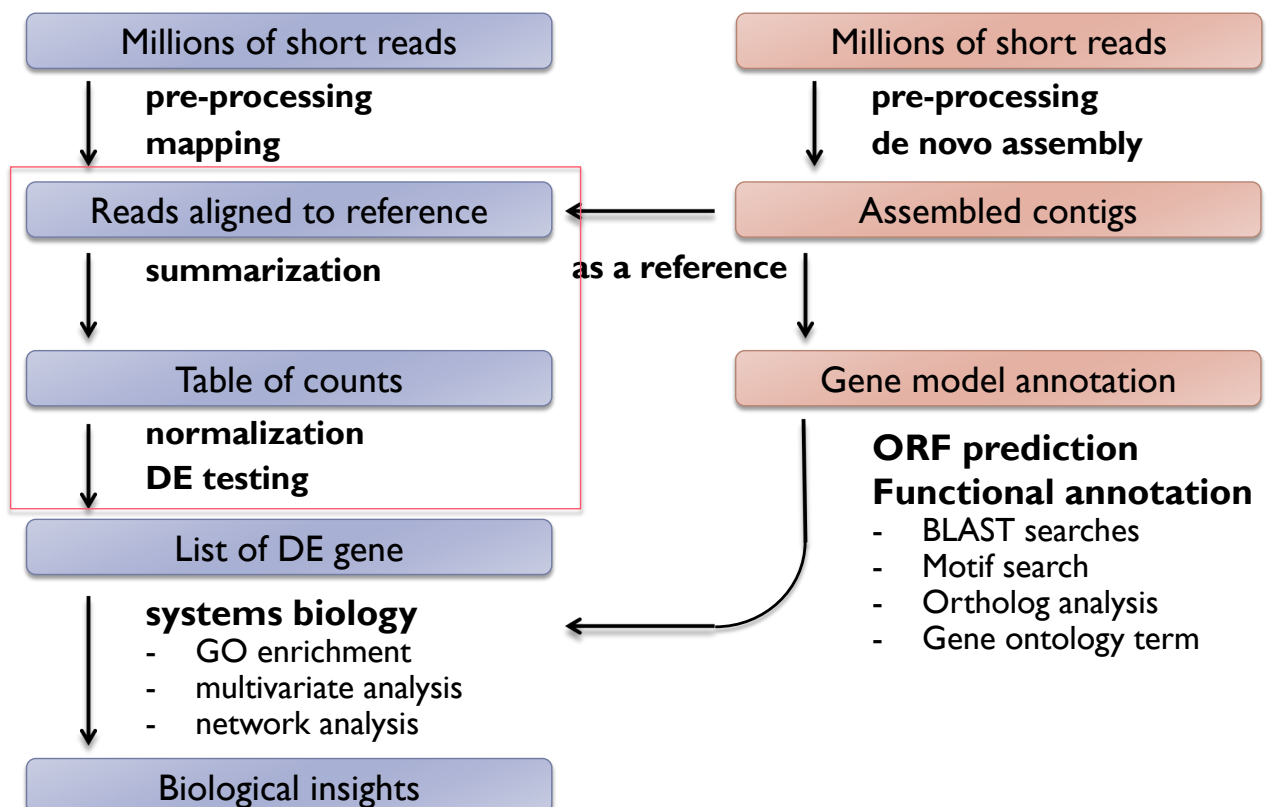
Trinity example

- ▶ Input: Illumina short reads in FASTQ | FASTA format
- ▶ Output: assembled contigs in FASTA format

```
# Run Trinity
$ Trinity --seqType fq --left left_all.fq --right right_all.fq \
  --CPU 8 --max_memory 20G
```

(Trinity is supported on only Linux)

RNA-seq analysis pipeline (*de novo* strategy)



DEG analysis

- ▶ Follow transcript-based RNA-seq pipeline

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Advanced

Clean up reference sequences

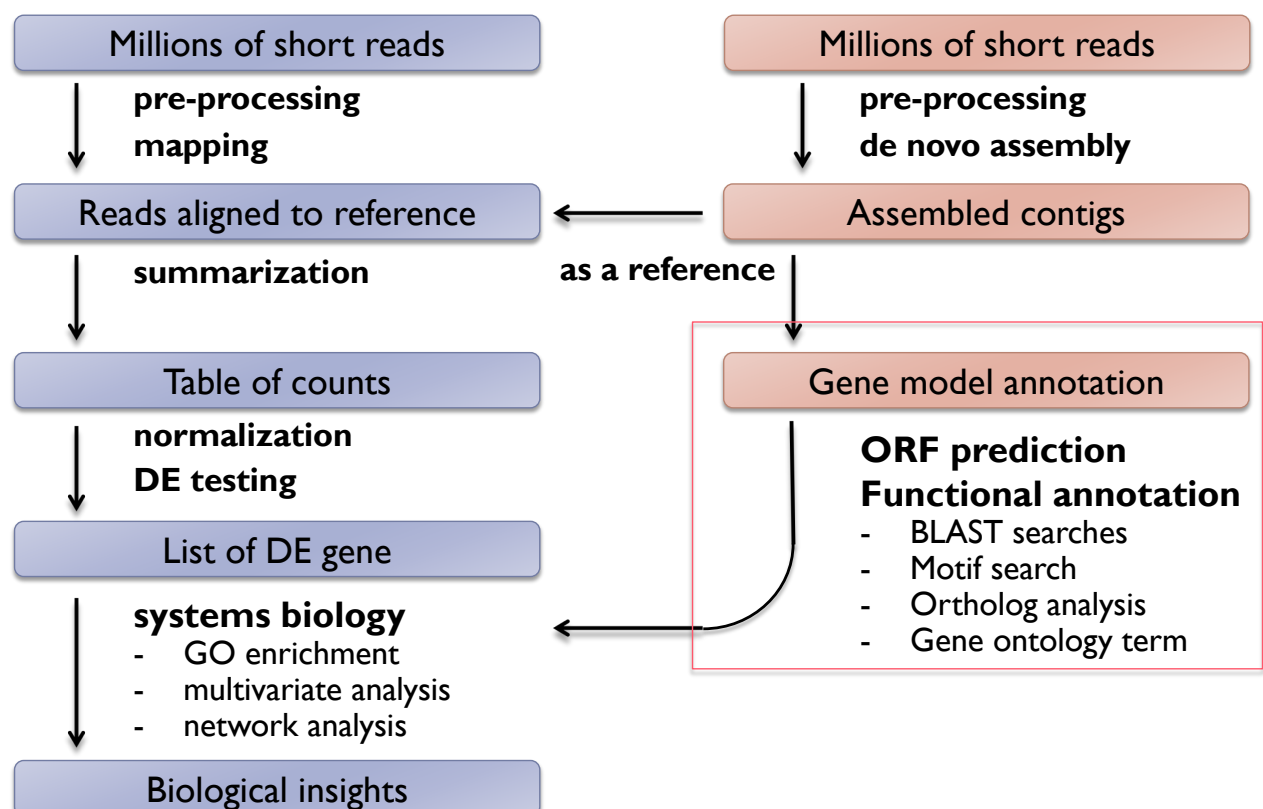
- ▶ Issues: Inflation of the number of Trinity contigs is often observed.
 - ▶ Trinity outputs splicing variants separately
 - ▶ Contaminations
 - ▶ Artifacts (bad contigs)
 - ▶ Incomplete contigs with very low expression.
- ▶ Solution
 - ▶ Filter out unwanted contigs.
 - ▶ Filter out very lowly expressed transcripts.
 - ▶ Clustering similar sequences.

Remove redundancy in reference sequences

► Strategy and Tools

- Choose one representative transcript from each cluster based on Trinity component information. (longest or highest expression)
- Clustering
 - CDHIT-EST (<http://weizhongli-lab.org/cd-hit/>)
 - Corset (Davidson et al., 2014).
 - RapClust (<https://github.com/COMBINE-lab/RapClust>)

RNA-seq analysis pipeline (*de novo* strategy)



ORF prediction

- ▶ Special consideration in ORF prediction after de novo RNA-seq assembly
 - ▶ Sometimes partial: Start Met or terminal codon may be missing.
 - ▶ Ideally one ORF is present per contig, but erroneously joined contigs may include multiple ORFs.
 - ▶ Possible frame shifts.
 - ▶ Frame shifts do not occur so often in Illumina, while it happens very frequently in 454 and IonProton.
- ▶ Recommended software: TransDecoder

Functional Annotation of Predicted ORFs

- ▶ **BLAST**
 - ▶ NCBI NR (or UniProt)
 - ▶ species of interest (model organisms, close relatives etc)
 - ▶ specific DB (SwissProt, rRNA DB, CEGMA etc)
 - ▶ self (assembly v.s. assembly)
- ▶ **Motif search**
 - ▶ Pfam, SignalP etc.
- ▶ **Ortholog analysis**
 - ▶ vs model organism
 - ▶ ortholog database (OrthoDB, eggNOG, OrthoMCL etc)
 - ▶ close relatives
- ▶ **Gene Ontology term assignment**

Quick annotation by BLASTX

- ▶ Query: assembled contigs
(nucleotide sequences in multi-fasta format)
- ▶ DB: Protein sequences of a model organism

Format DB

```
$ makeblastdb -in protein.fa -dbtype prot
```

Search

```
$ blastx -query trinity_contigs -db protein.fa \  
-num_threads 8 -evalue 1.0e-8 -outfmt 0 > blastxout.txt
```

Protein motif search using InterProScan

- ▶ Query: Translated ORF sequences
- ▶ Software: InterProScan
 - ▶ <https://github.com/ebi-pf-team/interproscan/wiki>

Search

```
$ interproscan.sh -I proteins.fasta -f XML,TSV --goterms  
--pathways
```

Assign Gene Ontology terms

- ▶ Tools

- ▶ InterProScan
- ▶ BLAST2GO
- ▶ Transfer model organisms GO terms based on orthology.

Let's try Trinity assembly

- ▶ ex9: de novo RNA-seq assembly using Trinity