

A tutorial for implementing Step Selection Function in R

P. Antkowiak*

H. Tripke†

C. Wilhelm‡

November 27, 2014

Contents

1	Introduction	2
1.1	Purpose and applications of SSFs	2
1.2	Our SSF workflow in R	2
1.3	Installing and loading Packages	2
2	Processing the Waypoint Data	4
2.1	Loading Waypoint Data (*.csv, ESRI)	4
2.2	Creating a “Spatial Points Data Frame”	5
2.3	Creating an “ltraj” object	5
2.4	Creating Bursts	9
2.5	Distinguish different behaviors: Broken stick model and autocorrelation	11
2.6	Creating Random Steps	12
2.7	Different distributions as an option for rdSteps	14
3	Processing Spatial Covariates	14
3.1	Load Raster Data (ESRI, *.tif, (*.shp))	14
3.2	Raster Extraction	15
3.3	Checking for Multicollinearity	17
4	Final SSF Model	17
4.1	generalized linear mixed model	18
4.2	mixed conditional logistic regression	19
4.3	Predictions	20
4.3.1	Predictions for GLMM	20
4.3.2	Predictions for MCLOGIT	23
5	Acknowledgements	26
6	Appendix	27

*M.Sc. programme “GIS und Umweltmodellierung” at University of Freiburg

†M.Sc. programme “Wildlife, Biodiversity and Vegetation” at University of Freiburg

‡M.Sc. programme “Wildlife, Biodiversity and Vegetation” at University of Freiburg

1 Introduction

1.1 Purpose and applications of SSFs

In addition to Resources Selection Functions (RSF) another powerful tool for evaluating data on animal movements and habitat selection are Step Selection Functions (SSF). The latter are used to estimate resource selection by comparing observed habitat use with available structures. Given GPS locations of a collared individual are connected by a linear segment. These segments are considered as steps. The time intervals influencing the step length should be chosen carefully (i.e. by conducting a pilot study) to meet the requirements of the study questions, the target species and its behaviour. The SSF then calculates random steps by taking measured angle and distance along steps and using the observed positions as starting points. These alternative steps represent the available habitat, which could have been chosen, within a realistic step length of the observed positions. Finally, we can compare spatial attributes on both and test for effects that explain habitat selection by animals [2].

So far, SSF models were mainly done using Geospatial Modelling Environment (GME) that works with a GIS¹. However, more and more packages for analyzing animal movements are provided in R. None of these packages is designed for doing a SSF only but quite a number provide already helpful functions to perform single steps of the Selection Function. Therefore, the aim of this tutorial is to collect all functions necessary to conduct a SSF and order them in a way that intuitively makes you understand how to run a SSF with your own data. Each step will be explained using an exemplary dataset of GPS locations collected from seven Cougars (*Puma concolor*) in the year 2010 (in the following addressed as “xmpl”).

1.2 Our SSF workflow in R

Figure 1 provides an overview of all necessary steps and potential options to conduct a SSF. This tutorial will guide you through each step and gives brief instructions on how to implement the functions and what to consider beforehand. To conduct a SSF using this tutorial we need you to store your initial data in two independent datasets:

1. A raster file of your spatial attributes (*Raster data*) and
2. GPS locations of your individuals assigned with a time stamp (*Waypoint data*).

We will start with the *Waypoint data* because these need to be transformed a couple of times to be able to work with them. You can find the single steps on the right side of Figure 1. While there are many options to adjust your *Waypoint data* the *Raster data* describing your spatial attributes needs not much of change. Once you created random steps for your observed positions you can extract the spatial attributes for each of those positions by using the function `extract`. At this point, *Waypoint* and *Raster data* will be combined and your final model can be written.

1.3 Installing and loading Packages

Before you can actually start using this tutorial for conducting SSF you need to install a bunch of packages in R. Some of them require others so that you have to add all these to your library:

```
## for implementing SSF
install.packages("hab")
install.packages("hab", repos = "http://ase-research.org/R/") # regular
install.packages("hab", repos = "http://ase-research.org/R/",
                 type = "source") # for self-compiling
```

¹www.spatial ecology.com/gme/

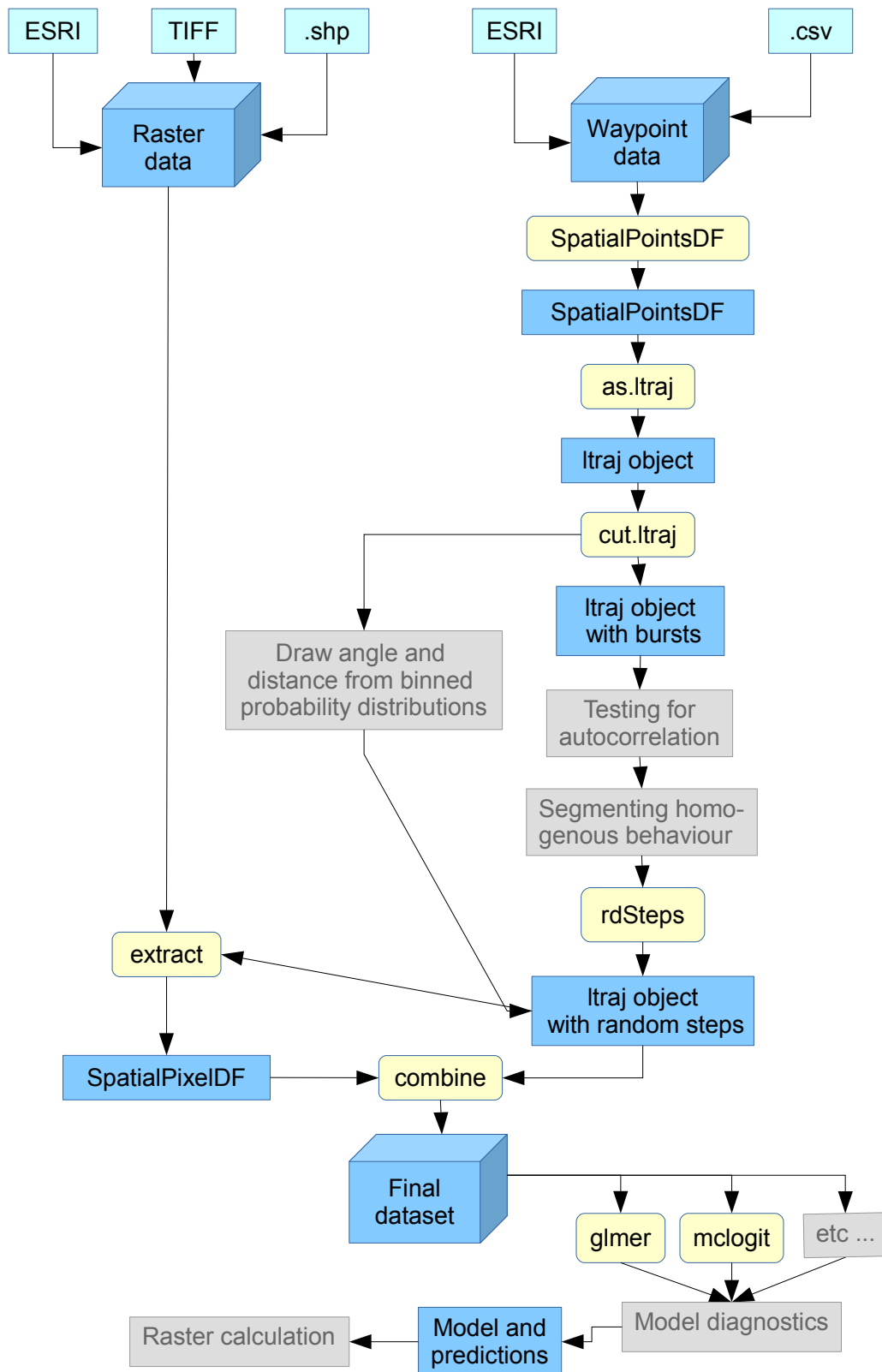


Figure 1: Stepwise conducting a Step Selection Function using existing R-packages. The yellow boxes show the name of the function applied while the blue boxes provide the type of object or data. In light grey optional steps are highlighted that are not implemented in this tutorial.

Keep fingers off the `adehabitat` package! It is outdated and replaced by four different packages designed for different analyses:

```
install.packages("adehabitatHR") # dealing with home ranges
install.packages("adehabitatHS") # habitat selection
install.packages("adehabitatLT") # trajectories
install.packages("adehabitatMA") # maps

install.packages("tkrplot")

# for handling raster data
install.packages("move")
install.packages("raster")
install.packages("rgdal")

# for analyzing the data
install.packages("mclogit")
install.packages("lme4")
install.packages("effects")
```

For faster processing the `install.packages` function is deactivated.

Loading packages:

```
library(hab)
library(adehabitatMA)
library(adehabitatHR)
library(adehabitatHS)
library(adehabitatLT)

library(sp)
library(raster)

library(mclogit)

## Error in library(mclogit): there is no package called 'mclogit'

library(lme4)
library(effects)

## Error in library(effects): there is no package called 'effects'
```

2 Processing the Waypoint Data

2.1 Loading Waypoint Data (*.csv, ESRI)

The data for the analysis should be saved in a simple *.csv file format. The table should have headings for each column in the first row and each observation should include at least the four following values:

- animal ID
- x-coordinate (easting)

- y-coordinate (northing)
- date and/or time

Remember that the coordinates need to be provided in the same coordinate system and spatial projection as the raster data.

Depending on your analysis you can include further values such as:

- ID for each record
- GPS precision
- other recording parameters such as season or month
- temperature / elevation at the moment of record
- other values that might be of interest in the further analysis

Use the following commands to set your working directory and read the data:

```
setwd("P:/Henriette/BestPracticeR/SSF-workflow/Code")
xmpl = read.csv("UTMsREDUCED.csv", head=T)
```

You can execute `head(xmpl)` and `str(xmpl)` to check, whether the data was successfully read.

2.2 Creating a “Spatial Points Data Frame”

The functions used along the rest of the toolchain can only process data which is stored as an object of class “SpatialPointsDataFrame”. This object class stores the coordinates separately and can be created using the according function from package `sp`.

```
library(sp)

xmpl.spdf = SpatialPointsDataFrame(coords = xmpl[,c("easting", "northing")],
                                   data = xmpl)
```

In the `coords` argument of the function you should specify the “x” and “y” values for your dataset. In our example, we simply assign the two coordinate columns of the example dataset, but you can read the coordinates from a separate file if you want.

To see, what information is stored in the data frame, use the `names` command:

```
names(xmpl)

## [1] "cat"      "LINE_NO"  "GMT_DATE" "GMT_TIME" "LMT_DATE" "LMT_TIME"
## [7] "easting"  "northing" "summer1"  "day1"     "hour"     "month"
```

2.3 Creating an “ltraj” object

After storing the data in a “Spatial Points Data Frame”, you now need to connect the single points and turn them into a set of trajectories. This operation is carried out by the function `as.ltraj` from the “hab” package and produces objects of class “ltraj”. The function `as.ltraj` requires at least three arguments to work:

1. “xy” (x- and y- coordinates for each point)

2. “date” (timestamp for each point, given as POSIXct class)
3. “id” (the animal id)

Both, coordinates and animal id can easily be adopted from the “Spatial Points Data Frame”. The timestamp however, needs to be stored as a “POSIXct” value with date and time in the same cell. If it is not stored in the required format yet, you therefore need to convert it first:

```
date <- as.POSIXct(strptime(paste(xmpl.spdf$LMT_DATE, xmpl.spdf$LMT_TIME),
                                "%d/%m/%Y %H:%M:%S"))
```

If your dataset already features a “POSIXct” timestamp, you can skip this step. Now you can proceed and actually create the “ltraj” object by executing the following command:

```
xmpl.ltr <- hab:::as.ltraj(xy = xmpl.spdf@coords, date = date,
                          id = xmpl.spdf$cat)
```

Two comments to the function used: By typing `hab:::as.ltraj` you tell R to use the `as.ltraj` function from the “hab” package which is speed optimized against its `adehabitatLT` sibling. Unlike the `xmpl.spdf@coords` prompt which works for any “SPDF” object, the `xmpl.spdf$cat` prompt is specific to your dataset. In the example dataset, animal ID’s are stored as an integer vector called `cat`. If this differs in your dataset and you should change the prompt accordingly. The `id` creates subsets in your data set which you can see in the structure below.

You now may want to have a closer look at the created “ltraj” object. Display its structure with by executing `str(xmpl.ltr)`. The “ltraj” object is a list containing seven different dataframes, each for one individual, which we defined with the `id`. In each dataframe all information is stored for every single observation of the individual:

- “x” and “y” (x- and y- coordinates for each point)
- “date” (timestamp for each point, given as POSIXct class)
- “dx” and “dy” (changes in x- and y- values)
- “dist” (length of trajectory)
- “dt” (time between relocations in s)
- “R2n” (???????)
- “abs.angle” (absolute angle of trajectory)
- “rel.angle” (angle between previous and actual trajectory)

```
str(xmpl.ltr, width=80)
```

```
## List of 7
## $ : 'data.frame': 1111 obs. of 10 variables:
## ..$ x : num [1:1111] 692362 692304 691745 691738 691743 ...
## ..$ y : num [1:1111] 5476589 5476529 5476031 5476042 5476035 ...
## ..$ date : POSIXct[1:1111], format: "2010-01-26 15:00:56" "2010-01-26 21:02:53" ..
## ..$ dx : num [1:1111] -57.79 -559.05 -7.28 4.68 2.45 ...
## ..$ dy : num [1:1111] -60.13 -497.63 10.99 -7.18 2.54 ...
## ..$ dist : num [1:1111] 83.4 748.45 13.18 8.57 3.53 ...
## ..$ dt : num [1:1111] 21717 21532 10767 10776 10775 ...
```

```

## ..$ R2n      : num [1:1111] 0 6955 691594 688499 690582 ...
## ..$ abs.angle: num [1:1111] -2.336 -2.414 2.156 -0.993 0.802 ...
## ..$ rel.angle: num [1:1111] NA -0.0778 -1.7126 3.1338 1.795 ...
## ..- attr(*, "id")= chr "10286"
## ..- attr(*, "burst")= chr "10286"
## ..- attr(*, "infolocs")='data.frame': 1111 obs. of  1 variable:
## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 1 2 3 4 5 6 7 8 9 10
## $ :'data.frame': 933 obs. of  10 variables:
## ..$ x      : num [1:933] 693628 693722 693638 693782 693789 ...
## ..$ y      : num [1:933] 5484250 5484205 5484182 5484248 5484243 ...
## ..$ date   : POSIXct[1:933], format: "2010-02-21 12:03:14" "2010-02-21 15:01:51" ...
## ..$ dx     : num [1:933] 93.91 -84.48 144.27 6.78 -357.94 ...
## ..$ dy     : num [1:933] -45.08 -22.48 66.34 -4.99 345.67 ...
## ..$ dist   : num [1:933] 104.17 87.42 158.79 8.42 497.6 ...
## ..$ dt     : num [1:933] 10717 21660 21482 10794 10839 ...
## ..$ R2n    : num [1:933] 0 10852 4653 23627 25793 ...
## ..$ abs.angle: num [1:933] -0.448 -2.882 0.431 -0.635 2.374 ...
## ..$ rel.angle: num [1:933] NA -2.43 -2.97 -1.07 3.01 ...
## ..- attr(*, "id")= chr "10287"
## ..- attr(*, "burst")= chr "10287"
## ..- attr(*, "infolocs")='data.frame': 933 obs. of  1 variable:
## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 1112 1113 1114 1115
## $ :'data.frame': 812 obs. of  10 variables:
## ..$ x      : num [1:812] 698168 698171 698172 698180 698002 ...
## ..$ y      : num [1:812] 5488180 5488171 5488169 5488189 5489527 ...
## ..$ date   : POSIXct[1:812], format: "2010-02-01 12:01:18" "2010-02-01 15:00:54" ...
## ..$ dx     : num [1:812] 3.214 0.945 7.374 -177.826 4.936 ...
## ..$ dy     : num [1:812] -8.676 -2.081 20.414 1337.288 -0.265 ...
## ..$ dist   : num [1:812] 9.25 2.29 21.71 1349.06 4.94 ...
## ..$ dt     : num [1:812] 10776 10802 10828 21653 10758 ...
## ..$ R2n    : num [1:812] 0.00 8.56e+01 1.33e+02 2.26e+02 1.84e+06 ...
## ..$ abs.angle: num [1:812] -1.216 -1.1444 1.2242 1.703 -0.0536 ...
## ..$ rel.angle: num [1:812] NA 0.0716 2.3686 0.4788 -1.7566 ...
## ..- attr(*, "id")= chr "10288"
## ..- attr(*, "burst")= chr "10288"
## ..- attr(*, "infolocs")='data.frame': 812 obs. of  1 variable:
## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 2045 2046 2047 2048
## $ :'data.frame': 775 obs. of  10 variables:
## ..$ x      : num [1:775] 725277 725254 724811 726203 730378 ...
## ..$ y      : num [1:775] 5462266 5462033 5463762 5464355 5464921 ...
## ..$ date   : POSIXct[1:775], format: "2010-03-09 18:01:53" "2010-03-09 21:00:47" ...
## ..$ dx     : num [1:775] -23.4 -442.6 1392 4174.6 5401.3 ...
## ..$ dy     : num [1:775] -233 1730 592 567 148 ...
## ..$ dist   : num [1:775] 234 1785 1513 4213 5403 ...
## ..$ dt     : num [1:775] 10734 54036 118772 21618 10805 ...
## ..$ R2n    : num [1:775] 0 54825 2457311 5220574 33066643 ...
## ..$ abs.angle: num [1:775] -1.6708 1.8213 0.4022 0.1349 0.0275 ...
## ..$ rel.angle: num [1:775] NA -2.791 -1.419 -0.267 -0.107 ...
## ..- attr(*, "id")= chr "10289"
## ..- attr(*, "burst")= chr "10289"
## ..- attr(*, "infolocs")='data.frame': 775 obs. of  1 variable:

```

```

## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 2857 2858 2859 2860
## $ : 'data.frame': 1638 obs. of 10 variables:
## ..$ x : num [1:1638] 727631 727789 727793 726560 726284 ...
## ..$ y : num [1:1638] 5447646 5447722 5447708 5445560 5445322 ...
## ..$ date : POSIXct[1:1638], format: "2010-03-01 15:01:20" "2010-03-01 18:00:58" ..
## ..$ dx : num [1:1638] 157.94 4.15 -1232.31 -276.79 112.8 ...
## ..$ dy : num [1:1638] 75.9 -13.9 -2148.2 -237.5 -134.8 ...
## ..$ dist : num [1:1638] 175.2 14.5 2476.6 364.7 175.7 ...
## ..$ dt : num [1:1638] 10778 10796 21694 10753 43289 ...
## ..$ R2n : num [1:1638] 0 30701 30119 5497537 7213751 ...
## ..$ abs.angle: num [1:1638] 0.448 -1.28 -2.092 -2.433 -0.874 ...
## ..$ rel.angle: num [1:1638] NA -1.728 -0.812 -0.341 1.559 ...
## ..- attr(*, "id")= chr "10290"
## ..- attr(*, "burst")= chr "10290"
## ..- attr(*, "infolocs")='data.frame': 1638 obs. of 1 variable:
## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 3632 3633 3634 3635
## $ : 'data.frame': 1026 obs. of 10 variables:
## ..$ x : num [1:1026] 732538 732540 733902 733793 733834 ...
## ..$ y : num [1:1026] 5454593 5454585 5455217 5456784 5457345 ...
## ..$ date : POSIXct[1:1026], format: "2010-03-10 18:01:49" "2010-03-10 21:01:48" ..
## ..$ dx : num [1:1026] 1.39 1362.2 -109.5 41.85 -4.68 ...
## ..$ dy : num [1:1026] -8.63 632.89 1566.31 561.62 -10.11 ...
## ..$ dist : num [1:1026] 8.74 1502.05 1570.13 563.18 11.14 ...
## ..$ dt : num [1:1026] 10799 21576 10746 10914 10697 ...
## ..$ R2n : num [1:1026] 0.00 7.63e+01 2.25e+06 6.37e+06 9.25e+06 ...
## ..$ abs.angle: num [1:1026] -1.412 0.435 1.641 1.496 -2.004 ...
## ..$ rel.angle: num [1:1026] NA 1.846 1.206 -0.144 2.783 ...
## ..- attr(*, "id")= chr "10291"
## ..- attr(*, "burst")= chr "10291"
## ..- attr(*, "infolocs")='data.frame': 1026 obs. of 1 variable:
## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 5270 5271 5272 5273
## $ : 'data.frame': 1550 obs. of 10 variables:
## ..$ x : num [1:1550] 703848 704820 703803 704148 703890 ...
## ..$ y : num [1:1550] 5470822 5466573 5467903 5468040 5468453 ...
## ..$ date : POSIXct[1:1550], format: "2010-03-19 15:03:05" "2010-03-20 18:03:08" ..
## ..$ dx : num [1:1550] 971.8 -1017.4 345.5 -258.2 -27.5 ...
## ..$ dy : num [1:1550] -4248 1330 137 412 204 ...
## ..$ dist : num [1:1550] 4358 1674 372 487 206 ...
## ..$ dt : num [1:1550] 97203 43191 32334 10859 10675 ...
## ..$ R2n : num [1:1550] 0 18993292 8521412 7826708 5614599 ...
## ..$ abs.angle: num [1:1550] -1.346 2.224 0.378 2.13 1.705 ...
## ..$ rel.angle: num [1:1550] NA -2.713 -1.846 1.752 -0.426 ...
## ..- attr(*, "id")= chr "10293"
## ..- attr(*, "burst")= chr "10293"
## ..- attr(*, "infolocs")='data.frame': 1550 obs. of 1 variable:
## .. ..$ pkey: Factor w/ 7845 levels "10286.2010-01-26 15:00:56",...: 6296 6297 6298 6299
## - attr(*, "class")= chr [1:2] "ltraj" "list"
## - attr(*, "typeII")= logi TRUE
## - attr(*, "regular")= logi FALSE

```

To get a visual impression of your data you can plot the trajectory for all or for one particular animal:


```
plot(xmpl.ltr)
```

The result of this code chunk is shown in Figure 2.

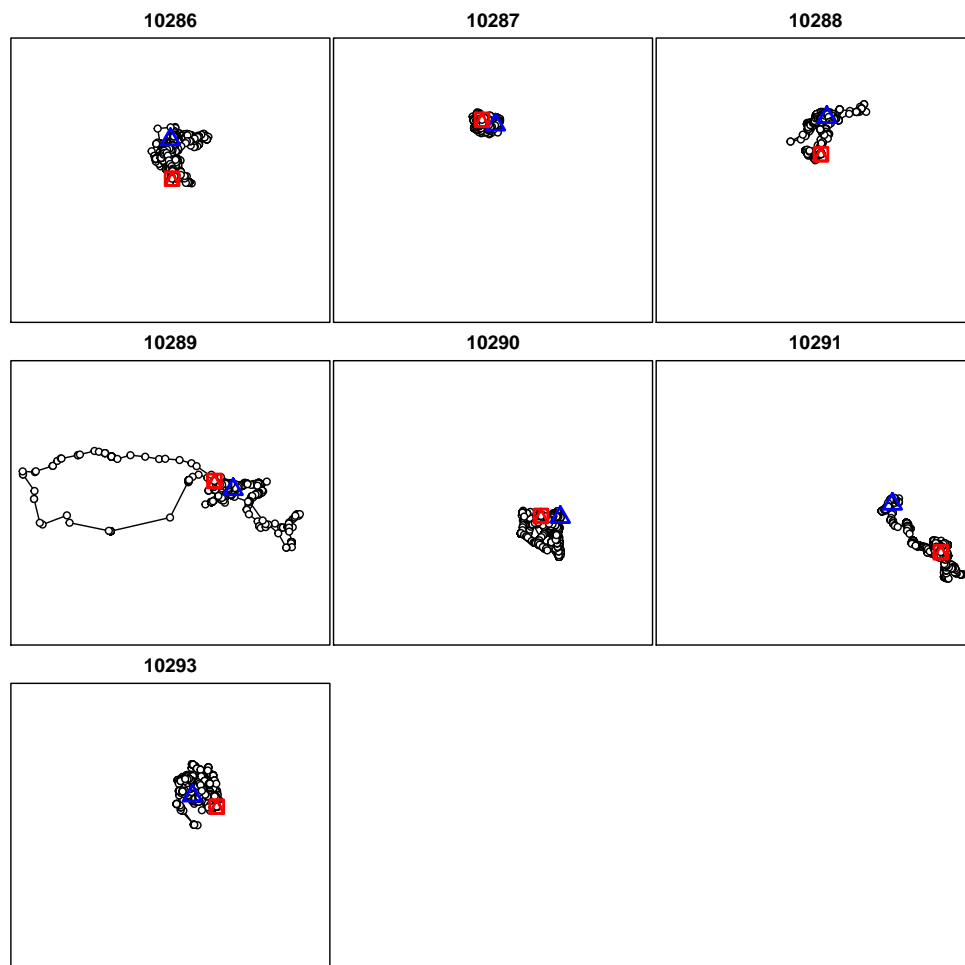


Figure 2: Visualization of the observed positions and the path from seven collared cougars.

To get a list of all cat IDs, use the `unique` command. Choose one that you are interested in!

```
unique(xmpl.spdf$cat)
```

```
## [1] 10286 10287 10288 10289 10290 10291 10293
```

You can then look at the trajectory of a single individual only.

```
plot(xmpl.ltr, id=10289)
```

The result of this code chunk is shown in Figure 3.

2.4 Creating Bursts

As described in the previous section, the relocations stored in the “ltraj” object are already divided into the different individuals. This partition is called a “burst”. For analysing the data, there might be the need to create “sub-bursts” for each animal within your trajectory. For example, if the animals were only recorded during the day, the monitoring took place over two

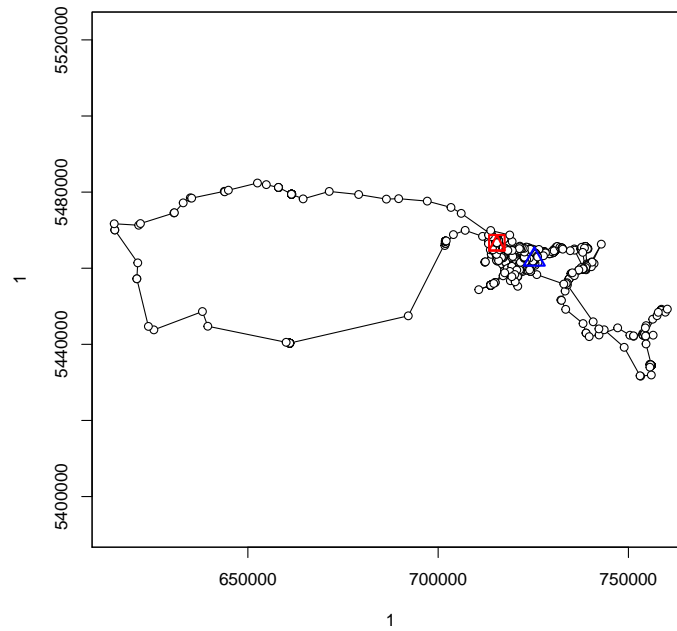


Figure 3: Visualization of the observed positions and the path from only one individual.

consecutive years or the time lag between the relocations differs remarkably, each accumulation of relocations can be defined as a different burst. Looking at those different parts separately might be necessary for different reasons. It is also possible to split your data into homogenous behaviour. Suggestions how to do that are explained in one of the following sections.

The function `cutltraj` splits the given bursts of your “ltraj” object into smaller bursts according to a specified criterion. In contrast, the function `bindltraj` combines the bursts of an object of class “ltraj” with the same attribute `id` to one unique burst.

To find out if there are missing values and to get an overview of the time lags between the relocations, you can plot `dt` of your “ltraj” object. For that, you need to define the time interval you are looking at [1].

In our example, the locations of the cougars were recorded every 3 hours, starting at 3 AM. The location at midnight is always missing. As `dt`, the time between successive relocations, is measured in seconds, we need to convert it into 3 hours.

```
plotltr(xmpl.ltr, "dt/3600/3")
```

The result of this code chunk is shown in Figure 4. The relocations which are 3 hours apart, are plotted at value 1 on the y-axis, if one relocation is missing (time lag of 6 hours) the relocation gets the value 2. As you can see, there are a lot of relocations missing. To keep the time lag constant, we now want to split the existing bursts (individuals) into “sub-bursts” where the time lag is bigger than 3 hours. A constant time lag is necessary to have comparable trajectories for further analyses.

To cut our data at our desired interval, we need a function which defines `dt`. Because we want to keep relocations which are only a few minutes “wrong”, we added 10 extra minutes.

Note: We still keep data with different time lags (i 3:10 h). As this is an irregular type II trajectory, it might cause problems with different functions (e.g. checking for autocorrelation). There are functions to convert it into a regular one, look at the `adehabitatLT` tutorial by C. Calenge [1], the help section to get more information or define an explicit time lag in the function

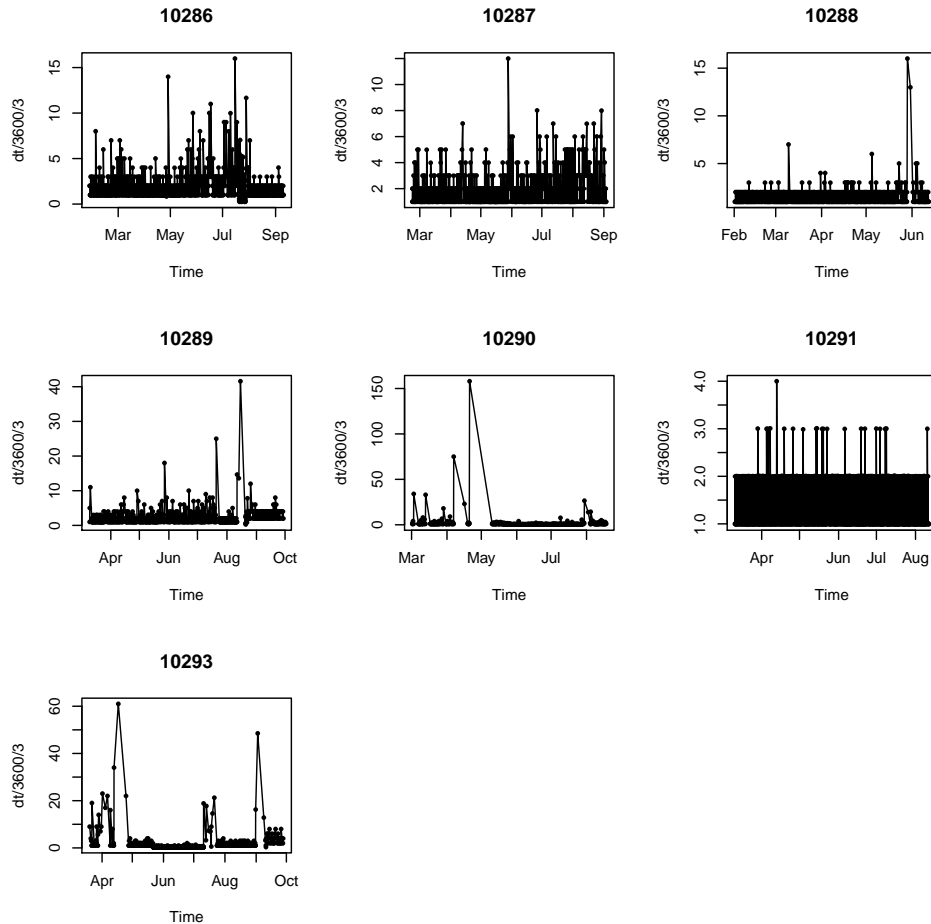


Figure 4: For each individual the observations are plotted. The y-axis shows the time interval from one observation to the next. Observations at 1 means that they are not more than 3 hours apart.

below. Because this however might lead to a massive loss of data, we didn't do that.

```
foo = function(dt) { return(dt > (3800*3)) }
```

Then we split the object of class `ltraj` into smaller bursts using `cutltraj` and the function above. The bursts we had before applying this function still remain.

```
xmpl.cut <- cutltraj(xmpl.ltr, "foo(dt)", nexttr = TRUE)

## Warning in cutltraj(xmpl.ltr, "foo(dt)", nexttr = TRUE): At least 3
relocations are needed for a burst
## 1249 relocations have been deleted
```

There are two options of cutting the trajectory depending on `nexttr`. If it is set as `FALSE`, the burst stops before the first relocation matching the criterion. if it is set as `TRUE`, it stops after. [1]

2.5 Distinguish different behaviors: Broken stick model and autocorrelation

NEEDS EXPLANATION! !!! Autocorrelation between dist or angle means homogenous behaviour

2.6 Creating Random Steps

Given your final bursts we now randomly draw angles and distances from your observed data to get random steps. The angle is taken from the previous trajectory to the actual one while the distance is taken from the starting point to the next observed position. This means that each burst should at least contain three observed positions to create random steps for one relocation. The first and the last relocations only contribute to the calculation but don't have their "own" random steps as there is no previous trajectory an angle could be calculated from. Neither there is an endpoint to draw a distance. This means that the number of points with random steps is lower as your actual recorded relocations. Before applying the function `rdSteps` you might want to check for correlation between turning angle and distance. In case your individuals tend to move long distances by turning only in small angles (e.g. a species migrating) but stop for several days for feeding you want to pick the distance and the angle as pairs. If no correlation is found you can pick both variables independently. To check for this we use the `plot` function but first have to convert our "ltraj" object back to a data frame by using `ld`.

```
# ld is a quick way to create a data frame from an ltraj object  
xmpl.cut.df <- ld(xmpl.cut)
```

```
plot(xmpl.cut.df$dist, xmpl.cut.df$rel.angle)
```

The result of this code chunk is shown in Figure 5.

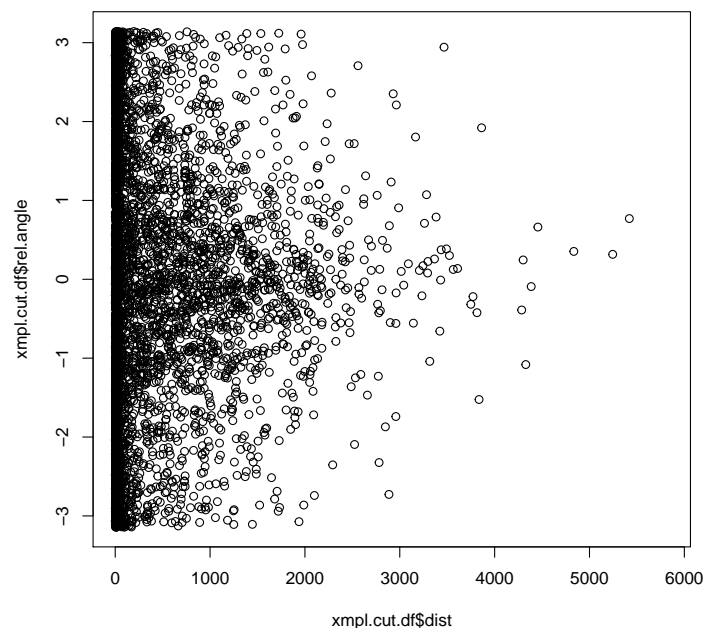


Figure 5: Testing for correlation of the observed turning angle and step length. The y-axis shows the difference in the turning angle. The shape indicates a correlation for longer steps and doing only small turning angles (around 1).

The plot shows a correlation of step length and turning angle and therefore the random steps should be taken as pairs (`simult = T`). Per default the angle and distance for each random step is drawn from the observed values you provide with `x`. If your random steps shall be taken from a different dataset you can do so by writing it in `rand.dist = YourDataSet`. Hereby, you can

also specify a distribution for estimating angle and distance. In our case we stick to the same dataset and apply `rdSteps`.

```
xmpl.steps <- rdSteps(x = xmpl.cut, nrs = 10, simult = T, rand.dis = NULL,
                      distMax = Inf, reproducible = TRUE, only.others = FALSE)
# use simult = FALSE if your data is not correlated
```

The function `rdSteps` uses some default settings which offer you options to modify your random steps. You can for example, easily change the number of steps taken from the observed data by defining `nrs` (default is 10) or if you only need steps shorter than a certain value specify `distMax` to that value (per default all steps are taken). By setting `reproducible = TRUE` a seed is used to get reproducible random steps. If you want to exclude your current individual to draw angle and distance from this set include `only.others = TRUE`. All in all, `rdSteps` is very straight forward and computes a lot of useful things for you:

```
head(xmpl.steps)
```

##		x	y	date	dx	dy		
##	10286.2	691738.1	5476042	2010-01-27 06:01:12	4.681700	-7.18200		
##	10286.2.1	691738.1	5476042	2010-01-27 06:01:12	105.850281	-134.39670		
##	10286.2.2	691738.1	5476042	2010-01-27 06:01:12	-2.562687	-86.12695		
##	10286.2.3	691738.1	5476042	2010-01-27 06:01:12	-124.866698	675.96404		
##	10286.2.4	691738.1	5476042	2010-01-27 06:01:12	85.786965	69.73609		
##	10286.2.5	691738.1	5476042	2010-01-27 06:01:12	-280.215683	-448.38129		
##		dist	dt	R2n	abs.angle	rel.angle	id	burst
##	10286.2	8.573181	10776	173.7518	-0.9931106	3.1337881	10286	10286.003
##	10286.2.1	171.075294	10776	173.7518	-7.1868501	-3.0599515	10286	10286.003
##	10286.2.2	86.165064	10776	173.7518	-1.6005423	2.5263563	10286	10286.003
##	10286.2.3	687.400232	10776	173.7518	-4.5297242	-0.4028256	10286	10286.003
##	10286.2.4	110.555529	10776	173.7518	-5.6006288	-1.4737302	10286	10286.003
##	10286.2.5	528.740587	10776	173.7518	-2.1293594	1.9975393	10286	10286.003
##		pkey	case	strata				
##	10286.2	10286.2010-01-27	06:01:12	1	1			
##	10286.2.1	10286.2010-01-27	06:01:12	0	1			
##	10286.2.2	10286.2010-01-27	06:01:12	0	1			
##	10286.2.3	10286.2010-01-27	06:01:12	0	1			
##	10286.2.4	10286.2010-01-27	06:01:12	0	1			
##	10286.2.5	10286.2010-01-27	06:01:12	0	1			

The table still includes your “cat id”, “burst id”, the “rel.angle” and “dist” of your observed positions. Furthermore, the “case” is provided as categories of 0 and 1 for available and used. The “strata” marks all 10 random steps and the visited location so you can later tell your function what to compare. Depending on your analysis you might want to compare only your observed positions with the endpoints of your random steps or you want to investigate in the selection of spatial attributes along the path. For latter you need to implement more code because we did not use this option in our tutorial (see the grey box in Figure 1).

Now only new coordinates for your random points are missing. Instead two columns provide the differences of your x- and y- coordinates for each random step (“dx” and “dy”). To get new coordinates for your random steps we simply add these differences to your initial coordinates and create two new columns.

```
xmpl.steps$new_x <- xmpl.steps$x + xmpl.steps$dx
xmpl.steps$new_y <- xmpl.steps$y + xmpl.steps$dy
```

After running this chapter you get your final “SpatialPointDataFrame” to use with the selection function.

```
head(xmpl.steps)
```

##		x	y	date	dx	dy		
##	10286.2	691738.1	5476042	2010-01-27 06:01:12	4.681700	-7.18200		
##	10286.2.1	691738.1	5476042	2010-01-27 06:01:12	105.850281	-134.39670		
##	10286.2.2	691738.1	5476042	2010-01-27 06:01:12	-2.562687	-86.12695		
##	10286.2.3	691738.1	5476042	2010-01-27 06:01:12	-124.866698	675.96404		
##	10286.2.4	691738.1	5476042	2010-01-27 06:01:12	85.786965	69.73609		
##	10286.2.5	691738.1	5476042	2010-01-27 06:01:12	-280.215683	-448.38129		
##		dist	dt	R2n	abs.angle	rel.angle	id	burst
##	10286.2	8.573181	10776	173.7518	-0.9931106	3.1337881	10286	10286.003
##	10286.2.1	171.075294	10776	173.7518	-7.1868501	-3.0599515	10286	10286.003
##	10286.2.2	86.165064	10776	173.7518	-1.6005423	2.5263563	10286	10286.003
##	10286.2.3	687.400232	10776	173.7518	-4.5297242	-0.4028256	10286	10286.003
##	10286.2.4	110.555529	10776	173.7518	-5.6006288	-1.4737302	10286	10286.003
##	10286.2.5	528.740587	10776	173.7518	-2.1293594	1.9975393	10286	10286.003
##		pkey	case	strata	new_x	new_y		
##	10286.2	10286.2010-01-27	06:01:12	1	1	691742.7 5476035		
##	10286.2.1	10286.2010-01-27	06:01:12	0	1	691843.9 5475908		
##	10286.2.2	10286.2010-01-27	06:01:12	0	1	691735.5 5475956		
##	10286.2.3	10286.2010-01-27	06:01:12	0	1	691613.2 5476718		
##	10286.2.4	10286.2010-01-27	06:01:12	0	1	691823.8 5476112		
##	10286.2.5	10286.2010-01-27	06:01:12	0	1	691457.8 5475594		

2.7 Different distributions as an option for rdSteps

!!!!!!!!!!!!!!!!NEEDS FURTHER EXPLANATION!!!!!!!!!!!!!!!!!!!!!!!!!!!!

3 Processing Spatial Covariates

This section explains the handling of spatial parameters that will be tested for selection by the target species. You should store these data in raster files (ESRI *.adf or georeferenced *.tif). These should have the same coordinate system as your telemetry data and should (for time reasons) already be clipped to your study area. For instructions how to do this in R, please read the GIS instructions from the other group ;)

3.1 Load Raster Data (ESRI, *.tif, (*.shp))

With a simple function stored in the package **raster** you are able to upload any raster file into R. Exemplarily we use raster data on the following parameters for the study area:

- ruggedness of the terrain
- land cover
- canopy cover

- distance to the nearest highway
- distance to the nearest road

For reading the raster data, three packages are required:

The source files for the raster data can be stored in the working directory or loaded by specifying the exact path. The `raster()` function is a universal and very powerful tool for loading all kinds of raster data. For reading shapefiles, use the `readOGR()` function. Below is an example of how to read a set of raster layers.

```
# First, make sure that your working directory is still
# the one specified earlier:
# getwd()

# Now read the layers (in our case the layers are stored in a different place):
ruggedness <- raster("P:/SSF PROJECT/NEW GIS LAYERS/tri1")
landcover <- raster("P:/SSF PROJECT/NEW GIS LAYERS/lc_30")
canopycover <- raster("P:/SSF PROJECT/NEW GIS LAYERS/cc_abmt")
disthighway <- raster("P:/SSF PROJECT/NEW GIS LAYERS/disthwy")
distrdoad <- raster("P:/SSF PROJECT/NEW GIS LAYERS/distsmrd")

# It is enough to load the whole folder were your *.adf files are
# stored in. The function raster() finds the needed files itself.
```

You can plot the data for a first overview. As this can take a while with large datasets, we outcommented the following chunk.

```
plot(ruggedness)
plot(landcover)
plot(canopycover)
plot(disthighway)
plot(distrdoad)
```

3.2 Raster Extraction

Now that you have generated the random steps and loaded the raster data, you can take the next step and actually connect the observed and potential points with the spatial covariates. There are different functions that can do this. When choosing one, you need to consider that raster files are large and juggling with them occupies lots of memory and computing power. For this reason we suggest using the `extract()` function that allows for querying single pixel values without loading the whole source file into working memory. The code for compiling the final dataset involves three steps:

- Converting the `xmpl.steps` data frame into a Spatial Points Data frame
- extracting the raster values
- combining them to the final dataset

Converting `xmpl.steps` into a “SpatialPointsDataFrame”:

```
xmpl.steps.spdf <- SpatialPointsDataFrame(coords =
                                           xmpl.steps[,c("new_x", "new_y")],
                                           data = xmpl.steps)
```

Extracting the values from each raster layer:

```
ruggedness.extr <- extract(ruggedness, xmpl.steps.spdf,
                           method='simple', sp=F, df=T)
canopycover.extr <- extract(canopycover, xmpl.steps.spdf,
                           method='simple', sp=F, df=T)
disthighway.extr <- extract(disthighway, xmpl.steps.spdf,
                           method='simple', sp=F, df=T)
distroad.extr <- extract(distroad, xmpl.steps.spdf,
                        method='simple', sp=F, df=T)
landcover.extr <- extract(landcover, xmpl.steps.spdf,
                        method='simple', sp=F, df=T)
```

The extraction is done separately for each layer. The option `method = 'simple'` extracts value from nearest cell whereas `method = 'bilinear'` interpolates from the four nearest cells. You can adjust this option according to the resolution of your dataset and ecological considerations. `df=T` returns the result as a data frame and `sp=F` ensures that the output is not added to the original dataset right away.

Automatically adding the extracted values to the original dataset sounds like a handy option. For two reasons we do not use it here: Firstly, we want to set the column names manually for not ending up with several columns called “w001001”. Secondly, our data include a categorical covariate (landcover) that we want to reclassify and flag as a factor.

This is the code for compiling the final dataset:

```
xmpl.steps.spdf$ruggedness <- ruggedness.extr[,2]
xmpl.steps.spdf$canopycover <- canopycover.extr[,2]
xmpl.steps.spdf$disthighway <- disthighway.extr[,2]
xmpl.steps.spdf$distroad <- distroad.extr[,2]

# The landcover covariate comes coded in integers between 0 and 10
# and is by default (mis)interpreted as an integer string.

unique(landcover.extr[,2])

## [1] 3 2 1 6 8 7 0 9 4 10 5

# Re-classifying landcover:
xmpl.steps.spdf$landcover <- as.factor(
  ifelse(landcover.extr[,2] == 0, NA,
  ifelse(landcover.extr[,2] < 5, "forest",
  ifelse(landcover.extr[,2] < 8, "open", NA))))
```

Now your final dataset should be ready for analysis. Examine it:

```
head(xmpl.steps.spdf)

##           x           y           date           dx           dy
## 10286.2    691738.1 5476042 2010-01-27 06:01:12    4.681700   -7.18200
## 10286.2.1 691738.1 5476042 2010-01-27 06:01:12  105.850281 -134.39670
## 10286.2.2 691738.1 5476042 2010-01-27 06:01:12   -2.562687  -86.12695
## 10286.2.3 691738.1 5476042 2010-01-27 06:01:12 -124.866698  675.96404
## 10286.2.4 691738.1 5476042 2010-01-27 06:01:12   85.786965   69.73609
## 10286.2.5 691738.1 5476042 2010-01-27 06:01:12 -280.215683 -448.38129
```



```
##           dist      dt      R2n  abs.angle  rel.angle      id      burst
## 10286.2      8.573181 10776 173.7518 -0.9931106  3.1337881 10286 10286.003
## 10286.2.1 171.075294 10776 173.7518 -7.1868501 -3.0599515 10286 10286.003
## 10286.2.2  86.165064 10776 173.7518 -1.6005423  2.5263563 10286 10286.003
## 10286.2.3 687.400232 10776 173.7518 -4.5297242 -0.4028256 10286 10286.003
## 10286.2.4 110.555529 10776 173.7518 -5.6006288 -1.4737302 10286 10286.003
## 10286.2.5 528.740587 10776 173.7518 -2.1293594  1.9975393 10286 10286.003
##
##                pkey case strata      new_x      new_y ruggedness
## 10286.2      10286.2010-01-27 06:01:12      1      1 691742.7 5476035 11.704700
## 10286.2.1 10286.2010-01-27 06:01:12      0      1 691843.9 5475908 15.198684
## 10286.2.2 10286.2010-01-27 06:01:12      0      1 691735.5 5475956 18.027756
## 10286.2.3 10286.2010-01-27 06:01:12      0      1 691613.2 5476718 16.522711
## 10286.2.4 10286.2010-01-27 06:01:12      0      1 691823.8 5476112 11.832160
## 10286.2.5 10286.2010-01-27 06:01:12      0      1 691457.8 5475594  5.744563
##
##      canopycover disthighway distroad landcover
## 10286.2           63      1738.189 210.0000      forest
## 10286.2.1          72      1612.762 330.0000      forest
## 10286.2.2          37      1745.680 271.6616      forest
## 10286.2.3          76      2204.949 361.2478      forest
## 10286.2.4          63      1718.139 120.0000      forest
## 10286.2.5          76      1749.286 180.0000      forest
```

3.3 Checking for Multicollinearity

Before including all environmental factors in your analysis, you should check if two or even more variables are exact or highly correlated. The threshold for the correlation coefficient is 0.7 or higher. To create a correlation matrix, we first need to convert the Spatial Points Data Frame into a data frame.

```
xmpl.steps.df = as.data.frame(xmpl.steps.spdf)

attach(xmpl.steps.df)

## The following objects are masked _by_ .GlobalEnv:
##
##      canopycover, date, disthighway, distroad, landcover, ruggedness

Z = cbind(ruggedness,canopycover,disthighway,distroad,landcover)

detach(xmpl.steps.df)

cor(Z)

## Error in cor(Z): 'x' must be numeric
```

As you can see, the canopycover and the landcover are highly correlated. The correlation coefficient for the distance to roads and the distance to highways is very high as well. For our further analysis we therefore only use landcover and the distance to roads.

4 Final SSF Model

There are several options to analyze the generated data. As [2] describe in their review of SSFs, conditional logistic regression has been the most commonly used procedure. Recently,

researchers have tried to account for among-individual heterogeneity in their dataset. Thurfjell et al. [2] recommend two R packages providing the necessary functionality: The `lme4` package and the `mclogit` package. For giving two simple examples, we will demonstrate data analysis with a mixed conditional logistic regression (`mclogit`) and a generalized linear mixed model (`lme4`). We will test for effects of ruggedness, canopy cover, land cover, distance to road and distance to highway. The geographic covariates will be included as quadratic terms, resulting in the equation:

case ruggedness+ruggedness²+disthighway+disthighway²+distroad+distroad²+landcover

Both models should produce rather similar outputs. Certainly, you are free to implement any type of model you find more elaborate or more suitable.

In a second step, we will use one of the fitted models to generate predictions and plot them.

4.1 generalized linear mixed model

family binomial (with a binomial distribution of error) nested random effect for ID and strata ...where the random effect takes the form of (1—id/strata).

structure of pseudo replication: id, stratum

```
xmpl.glmm.fit <- glmer(case ~
  landcover +
  ruggedness + I(ruggedness^2) +
  distroad + I(distroad^2) +
  (1|id/strata),
  family = binomial, data=xmpl.steps.df)

summary(xmpl.glmm.fit)
```

The model does not run and we get error messages suggesting us to scale the variables. Using the function `scale()` we center and scale the data so we get comparable values for each predictor. The according equation is:

$$f(x) = (x - \text{mean})/SD(x)$$

```
library(lme4)
library(effects)

## Error in library(effects): there is no package called 'effects'

xmpl.glmm.fit.sc <- glmer(case ~
  landcover +
  scale(ruggedness) + I(scale(ruggedness)^2) +
  scale(distroad) + I(scale(distroad)^2) +
  (1|id/strata),
  family = binomial, data=xmpl.steps.df)

summary(xmpl.glmm.fit.sc)

## Generalized linear mixed model fit by maximum likelihood (Laplace
## Approximation) [glmerMod]
## Family: binomial ( logit )
```

```

## Formula: case ~ landcover + scale(ruggedness) + I(scale(ruggedness)^2) +
##       scale(distroad) + I(scale(distroad)^2) + (1 | id/strata)
## Data: xmpl.steps.df
##
##      AIC      BIC   logLik deviance df.resid
## 32715.2 32786.3 -16349.6 32699.2    53401
##
## Scaled residuals:
##      Min       1Q   Median       3Q      Max
## -0.3346 -0.3296 -0.3255 -0.2955  6.8536
##
## Random effects:
## Groups      Name      Variance Std.Dev.
## strata:id (Intercept) 0.00e+00 0.000e+00
## id         (Intercept) 1.59e-13 3.988e-07
## Number of obs: 53409, groups: strata:id, 4940; id, 7
##
## Fixed effects:
##
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)    -2.215426   0.022127 -100.12 < 2e-16 ***
## landcoveropen   -0.216703   0.035502  -6.10 1.03e-09 ***
## scale(ruggedness)  0.011641   0.021362   0.54 0.58580
## I(scale(ruggedness)^2) -0.029234   0.010682  -2.74 0.00621 **
## scale(distroad)   -0.025928   0.030423  -0.85 0.39408
## I(scale(distroad)^2)  0.005637   0.011432   0.49 0.62191
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) lndcvr scl(r) I(scl(r)^2) scl(d)
## landcoverpn -0.338
## scl(rggdns)  0.179  0.012
## I(scl(r)^2) -0.365 -0.082 -0.633
## scal(dstrd)  0.397  0.043 -0.252  0.092
## I(scl(d)^2) -0.486 -0.012  0.187 -0.073      -0.861

plot(allEffects(xmpl.glmm.fit.sc))

## Error in plot(allEffects(xmpl.glmm.fit.sc)): error in evaluating the
## argument 'x' in selecting a method for function 'plot': Error: could not find
## function "allEffects"

```

4.2 mixed conditional logistic regression

The conventional logistic regression models do not take into account that in wildlife telemetry data observations are not independent but rather linked. If for example the selection differs among individual animals, the model may be flawed by pseudo replication. The mixed conditional logistic regression allows for specifying a random effects structure and thereby is capable of handling matched observations from different animals. Here is how to implement it in R:

```

library(mclgfit)

## Error in library(mclgfit): there is no package called 'mclgfit'

# Transform dataset to a data frame
xmpl.steps.df <- as.data.frame(xmpl.steps.spdf)

# Recall the column names of the dataset:
#head(xmpl.steps.df)

#xmpl.steps.spdf$id <- as.integer(xmpl.steps.spdf$id)

# column "case" indicates whether a site was visited or not. "strata" indicates the burst number

# The actual logistic model:
xmpl.logit.fit <-
  mclgfit(cbind(case, strata) ~
    landcover +
    ruggedness + I(ruggedness^2) +
    distroad + I(distroad^2), random=~1|id,
    data = xmpl.steps.df, start.theta=1000)

## Error in eval(expr, envir, enclos): could not find function "mclgfit"

## when including random effects we run into problems. We get the error message that the data is not
#random=~1|id,
#disthighway + I(disthighway^2),

summary(xmpl.logit.fit)

## Error in summary(xmpl.logit.fit): error in evaluating the argument 'object'
in selecting a method for function 'summary': Error: object 'xmpl.logit.fit'
not found

```

4.3 Predictions

Formula

$$w(x) = \exp(\beta_1 * x_1 + \beta_2 * x_2 + \dots + \beta_p * x_p)$$

4.3.1 Predictions for GLMM

—————PREDICTIONS FOR GLMM—————

```

# predictions for glmm

#plot(predict(xmpl.glmm.fit.sc, type="response"))
summary(xmpl.glmm.fit.sc)

## Generalized linear mixed model fit by maximum likelihood (Laplace
## Approximation) [glmerMod]
## Family: binomial ( logit )
## Formula: case ~ landcover + scale(ruggedness) + I(scale(ruggedness)^2) +

```

```

##      scale(distroad) + I(scale(distroad)^2) + (1 | id/strata)
##      Data: xmpl.steps.df
##
##      AIC      BIC    logLik deviance df.resid
## 32715.2 32786.3 -16349.6 32699.2    53401
##
## Scaled residuals:
##      Min      1Q  Median      3Q      Max
## -0.3346 -0.3296 -0.3255 -0.2955  6.8536
##
## Random effects:
## Groups      Name      Variance Std.Dev.
## strata:id (Intercept) 0.00e+00 0.000e+00
## id          (Intercept) 1.59e-13 3.988e-07
## Number of obs: 53409, groups: strata:id, 4940; id, 7
##
## Fixed effects:
##
##              Estimate Std. Error z value Pr(>|z|)
## (Intercept)      -2.215426    0.022127 -100.12 < 2e-16 ***
## landcoveropen      -0.216703    0.035502  -6.10 1.03e-09 ***
## scale(ruggedness)    0.011641    0.021362   0.54 0.58580
## I(scale(ruggedness)^2) -0.029234    0.010682  -2.74 0.00621 **
## scale(distroad)      -0.025928    0.030423  -0.85 0.39408
## I(scale(distroad)^2)   0.005637    0.011432   0.49 0.62191
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) lndcvr scl(r) I(scl(r)^2) scl(d)
## landcoverpn -0.338
## scl(rggdns)  0.179  0.012
## I(scl(r)^2) -0.365 -0.082 -0.633
## scal(dstrd)  0.397  0.043 -0.252  0.092
## I(scl(d)^2) -0.486 -0.012  0.187 -0.073      -0.861

mean(xmpl.steps.df$ruggedness)

## [1] 21.39603

sd(xmpl.steps.df$ruggedness)

## [1] 15.22119

summary(scale(xmpl.steps.df$ruggedness))

##      V1
## Min.   :-1.4057
## 1st Qu.: -0.7389
## Median :-0.2735
## Mean    : 0.0000
## 3rd Qu.: 0.4773
## Max.    :12.0550

```

```

rm(mydata)

## Warning in rm(mydata): object 'mydata' not found

mydata = data.frame(ruggedness=seq(-1.5,15,0.1))
#mydata = data.frame(ruggedness=scale(xmpl.steps.df$ruggedness))

mydata$wruggforest = exp(
  xmpl.glmm.fit.sc@beta[3] * mydata$ruggedness +
  xmpl.glmm.fit.sc@beta[4] * mydata$ruggedness^2 +
  xmpl.glmm.fit.sc@beta[5] * median(scale(xmpl.steps.df$distroad)) +
  xmpl.glmm.fit.sc@beta[6] * median((scale(xmpl.steps.df$distroad))^2))

mydata$wruggopen = exp(
  xmpl.glmm.fit.sc@beta[2] +
  xmpl.glmm.fit.sc@beta[3] * mydata$ruggedness +
  xmpl.glmm.fit.sc@beta[4] * mydata$ruggedness^2 +
  xmpl.glmm.fit.sc@beta[5] * median(scale(xmpl.steps.df$distroad)) +
  xmpl.glmm.fit.sc@beta[6] * median((scale(xmpl.steps.df$distroad))^2))

#plot(mydata$ruggedness, mydata$wruggforest, type="l", col="darkgreen", lwd=2)
#lines(mydata$ruggedness, mydata$wruggopen, type="l", col="green", lwd=2)

## rescaling

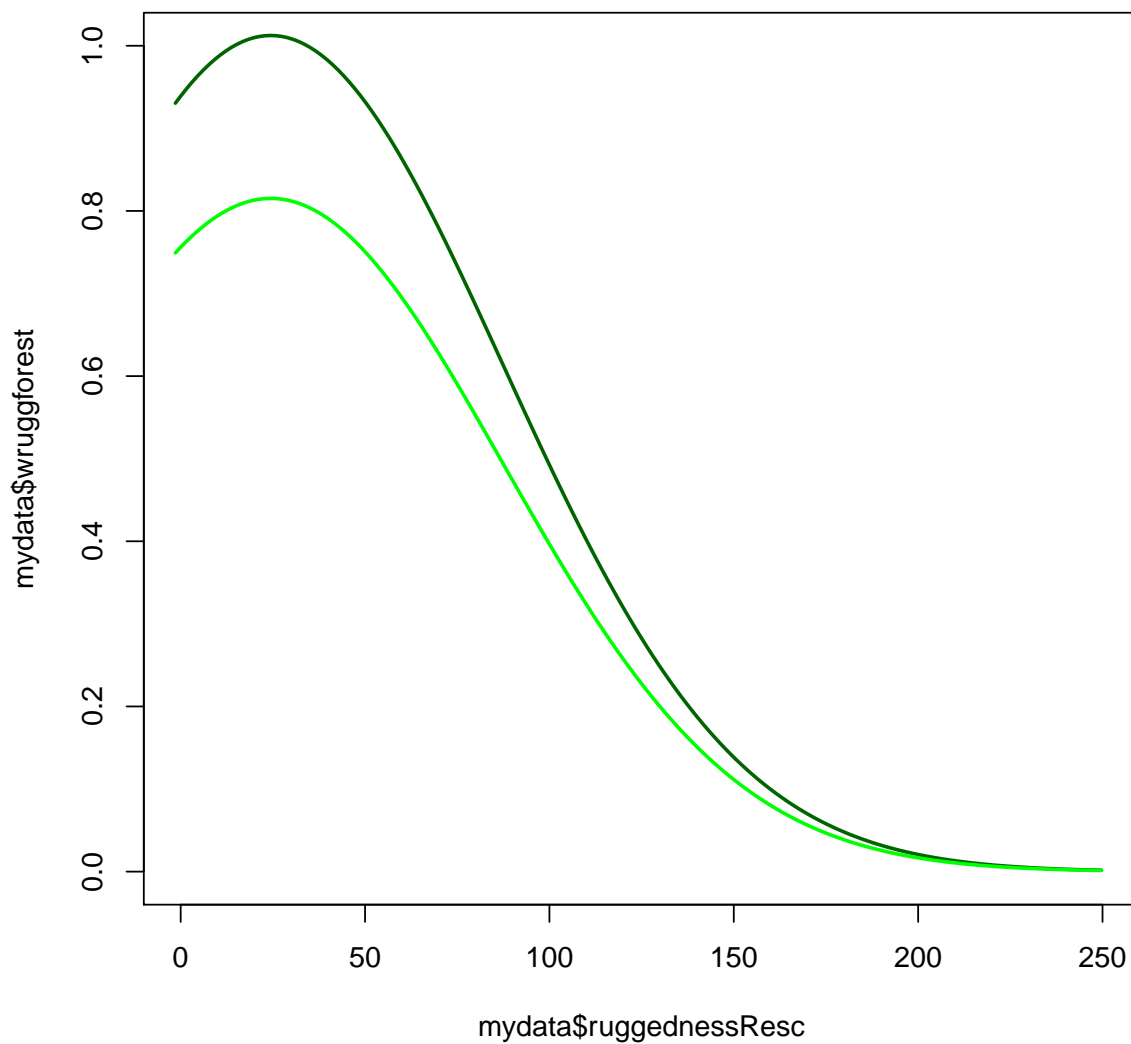
mydata$ruggednessResc <- (mydata$ruggedness * sd(xmpl.steps.df$ruggedness) + mean(xmpl.steps

#head(mydata)
#str(mydata)
#mydata$ruggednessResc

plot(mydata$ruggednessResc, mydata$wruggforest, type="l", col="darkgreen", lwd=2, ylim=c(0,
lines(mydata$ruggednessResc, mydata$wruggopen, type="l", col="green", lwd=2)

#abline(h=1,lty=2,col="wheat4")

```



4.3.2 Predictions for MCLOGIT

—————PREDICTIONS FOR MCLOGIT—————

```
#predictions for mclogit

summary(xmpl.steps.df$ruggedness)

##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##      0.00  10.15   17.23   21.40  28.66   204.90

summary(xmpl.logit.fit)

## Error in summary(xmpl.logit.fit): error in evaluating the argument 'object'
## in selecting a method for function 'summary': Error: object 'xmpl.logit.fit'
## not found

#plot(predict(xmpl.logit.fit))
```

```

mydata = data.frame(ruggedness=seq(0,250,1))

mydata$wruggforest = exp(xmpl.logit.fit$coefficients[2] * mydata$ruggedness +
                        xmpl.logit.fit$coefficients[3] * mydata$ruggedness^2 +
                        xmpl.logit.fit$coefficients[4] * median(xmpl.steps.df$distroad) +
                        xmpl.logit.fit$coefficients[5] * (median(xmpl.steps.df$distroad))^2)

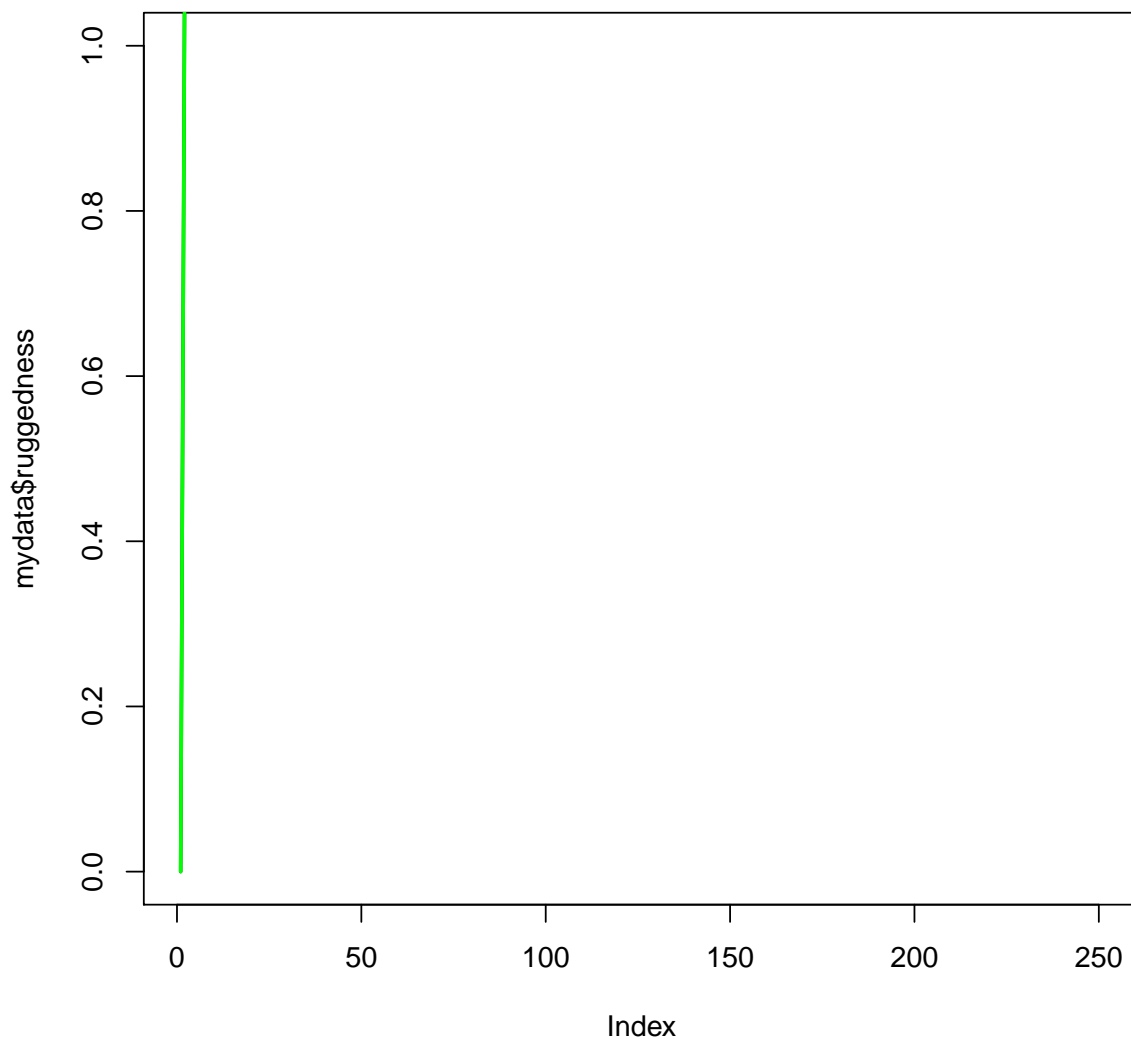
## Error in eval(expr, envir, enclos): object 'xmpl.logit.fit' not found

mydata$wruggopen = exp(xmpl.logit.fit$coefficients[1] +
                      xmpl.logit.fit$coefficients[2] * mydata$ruggedness +
                      xmpl.logit.fit$coefficients[3] * mydata$ruggedness^2 +
                      xmpl.logit.fit$coefficients[4] * median(xmpl.steps.df$distroad) +
                      xmpl.logit.fit$coefficients[5] * (median(xmpl.steps.df$distroad))^2)

## Error in eval(expr, envir, enclos): object 'xmpl.logit.fit' not found

plot(mydata$ruggedness,mydata$wruggforest, type="l", col="darkgreen", lwd=2, ylim=c(0,1))
lines(mydata$ruggedness,mydata$wruggopen,type="l", col="green", lwd=2)
#abline(h=1,lty=2,col="wheat4")

```

save models —————

```
save(pippo, file="pippo.Rdata")  
  
## Error in save(pippo, file = "pippo.Rdata"): object 'pippo' not found  
  
pippo <- load("pippo.Rdata")  
  
## Warning in readChar(con, 5L, useBytes = TRUE): cannot open compressed file  
'pippo.Rdata', probable reason 'No such file or directory'  
## Error in readChar(con, 5L, useBytes = TRUE): cannot open the connection
```

5 Acknowledgements

Don't forget to thank TeX and R and other opensource communities if you use their products! The correct way to cite R is shown when typing `"citation()"`, and `"citation("mgcv")"` for packages.

Special thanks to ♥ ♥ Simone ♥♥♥ ♥?❀! You were our best team member! ❀
Save Models!

6 Appendix

Session Info:

```
## R version 3.1.1 (2014-07-10)
## Platform: i386-w64-mingw32/i386 (32-bit)
##
## locale:
## [1] LC_COLLATE=English_United Kingdom.1252
## [2] LC_CTYPE=English_United Kingdom.1252
## [3] LC_MONETARY=English_United Kingdom.1252
## [4] LC_NUMERIC=C
## [5] LC_TIME=English_United Kingdom.1252
##
## attached base packages:
## [1] stats      graphics  grDevices  utils      datasets  methods    base
##
## other attached packages:
## [1] rgdal_0.9-1      lme4_1.1-7      Rcpp_0.11.3
## [4] Matrix_1.1-4     raster_2.3-12   adehabitatHS_0.3.9
## [7] hab_1.17         adehabitatHR_0.4.11 deldir_0.1-6
## [10] adehabitatLT_0.3.16 CircStats_0.2-4  boot_1.3-11
## [13] MASS_7.3-33      ade4_1.6-2      adehabitatMA_0.3.8
## [16] sp_1.0-15        knitr_1.7
##
## loaded via a namespace (and not attached):
## [1] evaluate_0.5.5  formatR_1.0     grid_3.1.1      highr_0.4
## [5] lattice_0.20-29 minqa_1.2.4     nlme_3.1-117    nloptr_1.0.4
## [9] splines_3.1.1   stringr_0.6.2   tools_3.1.1
```

References

- [1] C. Calenge. Package adehabitatlt. 2014.
- [2] Henrik Thurfjell, Simone Ciuti, and Mark S Boyce. Applications of step-selection functions in ecology and conservation. *Movement Ecology*, 2(1):4, 2014.