

Parkinson disease

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Abstract | Parkinson disease is the second-most common neurodegenerative disorder that affects 2–3% of the population ≥65 years of age. Neuronal loss in the substantia nigra, which causes striatal dopamine deficiency, and intracellular inclusions containing aggregates of α-synuclein are the neuropathological hallmarks of Parkinson disease. Multiple other cell types throughout the central and peripheral autonomic nervous system are also involved, probably from early disease onwards. Although clinical diagnosis relies on the presence of bradykinesia and other cardinal motor features, Parkinson disease is associated with many non-motor symptoms that add to overall disability. The underlying molecular pathogenesis involves multiple pathways and mechanisms: α-synuclein proteostasis, mitochondrial function, oxidative stress, calcium homeostasis, axonal transport and neuroinflammation. Recent research into diagnostic biomarkers has taken advantage of neuroimaging in which several modalities, including PET, single-photon emission CT (SPECT) and novel MRI techniques, have been shown to aid early and differential diagnosis. Treatment of Parkinson disease is anchored on pharmacological substitution of striatal dopamine, in addition to non-dopaminergic approaches to address both motor and non-motor symptoms and deep brain stimulation for those developing intractable L-DOPA-related motor complications. Experimental therapies have tried to restore striatal dopamine by gene-based and cell-based approaches, and most recently, aggregation and cellular transport of α-synuclein have become therapeutic targets. One of the greatest current challenges is to identify markers for prodromal disease stages, which would allow novel disease-modifying therapies to be started earlier.

Two hundred years after James Parkinson's seminal essay on 'the shaking palsy', most of his original clinical observations have stood the test of time. Beyond the perception of Parkinson disease as a disorder of movement, it has since become apparent that a multitude of non-motor features, such as cognitive impairment, autonomic dysfunction, disorders of sleep, depression and hyposmia (impaired smell), are part of the disease and add considerably to overall burden. Tremendous progress has been made in understanding the neuropathology of Parkinson disease and its progression throughout the nervous system, as well as the molecular and neurophysiological mechanisms and perturbations underlying the disease and its symptoms. Above all, highly efficacious therapies have become available, which are focused on pharmacological dopamine substitution (L-DOPA treatment), but with important refinements and ground-breaking expansions, such as the introduction of deep brain stimulation (DBS). These treatment advances have undoubtedly made Parkinson disease the first and still unparalleled example of a neurodegenerative disease that can be effectively managed, leading to sustained symptom control

and quality of life (QOL) up to decades after disease onset. However, none of these treatments is curative and Parkinson disease remains a progressive disorder that eventually causes severe disability — not least by the increasing severity of treatment-resistant motor problems and non-motor symptoms. Thus, modifying disease progression and further delaying disability are the key unmet needs to be addressed by current and future research efforts. Of great future potential is the development of methods to identify individuals at risk and early manifestations that antedate the onset of the defining motor symptoms.

In this Primer, we describe the epidemiology of Parkinson disease, and review our current understanding of the underlying pathology and molecular pathogenesis as well as the perturbations of basal ganglia and cortical connectivity that underlie the cardinal motor features of this illness. We also summarize recent advances in clinical diagnostics, biomarker research and screening, and provide an overview of the natural history of Parkinson disease and the current, as well as future, therapies.

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Epidemiology

Worldwide incidence estimates of Parkinson disease range from 5 to >35 new cases per 100,000 individuals yearly¹, which probably reflects differences in the demographics of the populations studied or in study methods. In a population-based study in Minnesota (USA) with pathological validation of clinical diagnoses, the incidence of Parkinson disease was 21 cases per 100,000 person-years². Parkinson disease is rare before 50 years of age¹, but the incidence increases 5–10-fold from the sixth to the ninth decade of life^{1–3}. The global prevalence, conservatively estimated at 0.3% overall, likewise increases sharply with age to >3% in those >80 years of age⁴ (FIG. 1).

Mortality is not increased in the first decade after disease onset, but increases thereafter, eventually doubling compared with the general population⁵. Improvement in health care has led to longer survival, which is associated with increasing prevalence of Parkinson disease over time in one 20-year study⁶. The number of people with Parkinson disease is expected to double between 2005 and 2030 (REF. 7). Years lived with disability and disability-adjusted life years due to Parkinson disease increased between 1990 and 2010, and a progressive increase in the personal, societal and economic burden associated with the disease is expected in the future as the world population ages^{7–10}.

Parkinson disease is twice as common in men than in women in most populations^{3,11} (FIG. 1), although in a few populations, including one study from Japan, no difference or even a female excess was observed¹². A protective effect of female sex hormones, a sex-associated genetic mechanism or sex-specific differences in exposure to environmental risk factors might explain this male preponderance, although disparities in health care could also contribute.

The incidence seems to vary within subgroups defined by race, ethnicity, genotype or environment. Parkinson disease might be less common in African Americans and Asians in the United States, but systematic race-specific incidence has not been investigated in other multiracial

populations, and societal rather than biological causes might underlie these findings³. Geography and race are often related, and it might be difficult to determine the relative contribution of each to the risk of developing Parkinson disease. In Israel, the prevalence is high, possibly reflecting the higher prevalence of the incompletely penetrant genes associated with Parkinson disease (that is, *LRRK2* (which encodes leucine-rich repeat serine/threonine-protein kinase 2) and *GBA* (which encodes glucocerebrosidase)) in Ashkenazi Jews¹³. The prevalence of Parkinson disease is also high in Inuit, Alaska Native and Native American populations¹⁴. Lifestyle, including dietary exposure to persistent organic pollutants, or shared genetic factors could explain this pattern. The incidence is greater in men of Japanese and Okinawan descent living in Hawaii than in men living in Japan, supporting that environmental factors have a role¹⁵. Gene-environment interactions definitely modify the risk for sporadic Parkinson disease. For example, the incidence of Parkinson disease is significantly greater in individuals exposed to certain environmental factors, such as pesticides and traumatic brain injury, and lower in smokers or caffeine users¹⁶.

Mechanisms/pathophysiology**Neuropathology**

Characteristic features of Parkinson disease include neuronal loss in specific areas of the substantia nigra and widespread intracellular protein (α -synuclein) accumulation. Although neither the loss of pigmented dopaminergic neurons in the substantia nigra^{17,18} nor the deposition of α -synuclein in neurons is specific for Parkinson disease, these two major neuropathologies are specific for a definitive diagnosis of idiopathic Parkinson disease when applied together (FIG. 2).

Gross macroscopic atrophy of the brain is not a feature of Parkinson disease, rather neuronal degeneration occurs in only certain types of neurons within particular brain regions. In early-stage disease, loss of dopaminergic neurons is restricted to the ventrolateral substantia nigra with relative sparing of other midbrain dopaminergic neurons^{19,20} (FIG. 2a–d), but becomes more widespread by end-stage disease. The dramatic loss of these dopaminergic neurons even early in the disease suggests that the degeneration in this region starts before the onset of motor symptoms, which is supported by several recent clinicopathological studies^{21,22}.

The other required neuropathology is the abnormal deposition of α -synuclein in the cytoplasm of certain neurons in several different brain regions²³. Lewy bodies, which are largely made up of aggregated α -synuclein, were the first to be described over a century ago. Following the development of refined histopathological methods, a broader range of α -synuclein aggregates have been described (FIG. 2e–g). The Lewy pathology initially occurs in cholinergic and monoaminergic brainstem neurons and in neurons in the olfactory system, but is also found in limbic and neocortical brain regions with disease progression (FIG. 2h). In patients with Alzheimer pathology, there is a different pattern of α -synuclein pathology that concentrates mainly in limbic brain regions²².

Although heritable forms of Parkinson disease only represent 5–10% of all cases (TABLE 1), they have provided crucial clues to the mechanisms underlying the neuropathology of Parkinson disease. Some of the proteins encoded by genes associated with Parkinson disease are involved in a set of molecular pathways that, when perturbed, can trigger a neuropathology that resembles, or is indistinguishable from, sporadic Parkinson disease. In addition, large genome-wide association studies (GWAS) confirm that some of these genes are also affected in sporadic Parkinson disease²⁴. Examples of these pathways are: α -synuclein proteostasis, mitochondrial function, oxidative stress, calcium homeostasis, axonal transport and neuroinflammation (FIG. 3).

α -Synuclein proteostasis

Intraneuronal protein aggregates that are largely made up of α -synuclein are found in all patients with Parkinson disease. The existence of point mutations and multiplications of SNCA, the gene encoding α -synuclein, that cause heritable forms of Parkinson disease strongly support the notion that α -synuclein is a key player in Parkinson disease (TABLE 1). Similarly, GWAS have revealed a single-nucleotide polymorphism associated with the SNCA locus that alters the risk for sporadic Parkinson disease and is associated with increased expression levels of α -synuclein^{24,25}. A study in human neurons derived from induced pluripotent stem cells and post-mortem frontal cortex samples from patients with Parkinson disease supports the idea that a risk variant associated with Parkinson disease in a non-coding distal enhancer element of SNCA is coupled to increased α -synuclein expression²⁶.

The normal neuronal function of the 140 amino acid α -synuclein protein is not fully understood, but it occurs in the cytosol, possibly also in mitochondria and the nucleus, and probably has a role in synaptic vesicle dynamics, mitochondrial function, intracellular trafficking and might be a potential chaperone^{25,27,28}. α -Synuclein acquires neurotoxic properties during a pathogenetic process in which soluble α -synuclein

monomers initially form oligomers, then progressively combine to form small protofibrils and eventually large, insoluble α -synuclein fibrils (that is, the aggregates that make up Lewy pathology)^{29,30}. The underlying triggers of accumulation and aggregation of α -synuclein can be manifold, for example, a relative overproduction of the protein, the presence of mutations that increase the likelihood for its misfolding and oligomerization or impairments in the molecular pathways that are charged with degrading native or misfolded α -synuclein.

A progressive, age-related decline in proteolytic defence mechanisms in the ageing brain might play an important part in the accumulation of α -synuclein^{31,32} (FIG. 3).

α -Synuclein degradation. Intracellular homeostasis of α -synuclein is maintained by the actions of the ubiquitin–proteasome system and the lysosomal autophagy system (LAS). The relative importance of the ubiquitin–proteasome system and LAS for intracellular α -synuclein proteolysis in neurons is debated and LAS seems to be more important than the ubiquitin–proteasome system to clear oligomeric assemblies³². With respect to LAS, both chaperone-mediated autophagy and macroautophagy are suggested to mediate α -synuclein degradation^{32,33}. Chaperone-mediated autophagy involves specific chaperones that target certain proteins to lysosomes, whereas macroautophagy entails the formation of autophagosomes that are directed to perinuclear lysosomes. Inhibition of either system leads to increased levels of α -synuclein, and evidence for some compensatory crosstalk between the systems exists³⁴. Additional proteases, which are not part of the ubiquitin–proteasome system and LAS, can also cleave α -synuclein in the extracellular space³².

Several lines of evidence suggest that impairment of these degradation systems could contribute to α -synuclein accumulation. Increasing age — the greatest risk factor for Parkinson disease — is associated with reduced LAS and ubiquitin–proteasome system functions³¹, which is consistent with observations of

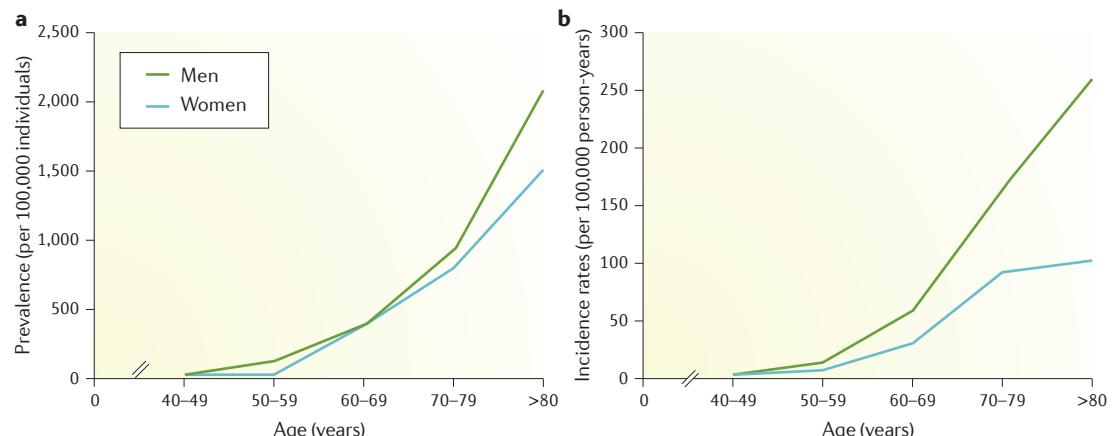


Figure 1 | Incidence and prevalence of Parkinson disease. **a** | Prevalence of Parkinson disease in men and women per 100,000 individuals. **b** | Incidence rate of Parkinson disease per 100,000 person-years. Data are derived from two recent meta-analyses, which used crude rates without adjustments for demographic differences or methodological differences between studies^{4,235}.

increased levels of α -synuclein in nigral dopaminergic neurons during normal ageing³⁵. Pharmacological stimulation of macroautophagy reduces the levels of intracellular α -synuclein in experimental models^{36,37}. In the substantia nigra of patients and experimental models of Parkinson disease, lysosomal enzyme levels are reduced, particularly in neurons containing α -synuclein inclusions³⁸, markers of chaperone-mediated autophagy

are decreased³⁹ and autophagosomes accumulate⁴⁰. Additional observations support the idea that altered proteostasis profoundly influences neuronal accumulation of α -synuclein. For example, α -synuclein oligomers inhibit the ubiquitin–proteasome system⁴¹, accumulating α -synuclein can inhibit macroautophagy^{42,43} and different forms of α -synuclein (wild type, mutant or post-translationally modified) can reduce chaperone-mediated autophagy function^{34,44}. Collectively, these observations suggest a vicious cycle involving the accumulation of α -synuclein due to disrupted proteostasis, which in turn leads to defective α -synuclein degradation.

Several mutations associated with genetic forms of Parkinson disease are associated with reduced LAS function. The G2019S mutation in the gene encoding LRRK2 is associated with impaired LAS and increased aggregation of α -synuclein in dopaminergic neurons that are exposed to α -synuclein fibrils⁴⁵. Heterozygous mutations in the gene encoding the lysosomal enzyme GBA, the most common genetic risk factor for Parkinson disease⁴⁶, are coupled to reduced LAS function⁴⁷. GWAS have revealed two polymorphisms in the GBA locus associated with altered risk of developing Parkinson disease²⁴, and normal ageing is reported to result in a progressive decline in GBA activity⁴⁸. Recent evidence from clinical cohort studies also suggests an increased risk for dementia in individuals with Parkinson disease who carry GBA mutations that, in the homozygotic state, are associated with the neuronopathic type of Gaucher disease^{49,50}. Reduced GBA activity coincides with increased levels of α -synuclein both in cell cultures and animal models^{51,52}. Mutations in the gene encoding vacuolar protein sorting-associated protein 35 (VPS35), which cause autosomal dominant Parkinson disease^{53,54}, also seem to affect α -synuclein handling. VPS35 is part of the retromer complex, which has a key role in sorting lipids and proteins that are newly synthesized or have undergone endocytosis and directs them to either the lysosome, the cell surface or the Golgi apparatus⁵⁵. Notably, Vps35-deficient mice exhibit increased levels of α -synuclein in nigral dopaminergic neurons⁵⁶, whereas overexpression of Vps35 reduces α -synuclein accumulation in transgenic mice who also overexpress α -synuclein and in cultured neurons that are exposed to α -synuclein fibrils⁵⁷. Both VPS35 deficiency and the D620N mutation in VPS35 that causes autosomal dominant Parkinson disease are coupled to reduced cellular levels of lysosome-associated membrane glycoprotein 2 (LAMP2)⁵⁶, suggesting once again that LAS perturbation is key to disease pathogenesis. Finally, mutations in ATP13A2 (also known as PARK9), which encodes a type 5 P-type ATPase that is present in lysosomes and autophagosomes⁵⁸, are associated with a rare juvenile-onset neurological condition (Kufor-Rakeb syndrome) that includes parkinsonian features and responds to dopaminergic therapy⁵⁸. Dysfunction of LAS and vesicular trafficking probably contribute to neurodegeneration in people with mutations in ATP13A2 (REF. 59). Notably, GWAS have revealed that certain ATP13A2 variants are associated with increased

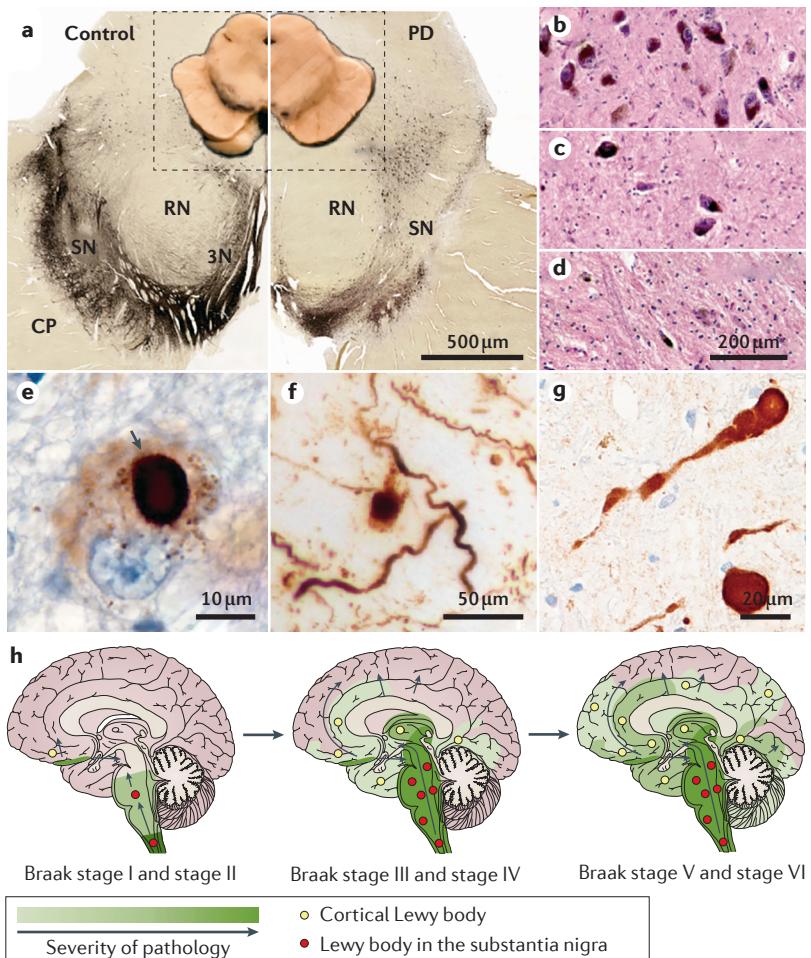


Figure 2 | The main diagnostic neuropathologies for Parkinson disease. **a** | Parkinson disease (PD) is defined by depigmentation of the substantia nigra (SN) (right panel) compared with control (left panel). Macroscopical (inset) and transverse sections of the midbrain upon immunohistochemical staining for tyrosine hydroxylase, the rate limiting enzyme for the synthesis of dopamine, are shown. Selective loss of the ventrolateral parts of the SN with sparing of the more medial and dorsal regions is evident in the histological section. **b–d** | Haematoxylin and eosin staining of the ventrolateral region of the SN showing a normal distribution of pigmented neurons in a healthy control (part **b**) and diagnostically significant moderate (part **c**) or severe (part **d**) pigmented cell loss in PD. **e–g** | Immunohistochemical staining of α -synuclein shows the round, intracytoplasmic Lewy bodies (arrow in part **e**), more diffuse, granular deposits of α -synuclein (part **e** and part **f**), deposits in neuronal cell processes (part **f**), extracellular dot-like α -synuclein structures (part **f**) and α -synuclein spheroids in axons (part **g**). **h** | The theorized progression of α -synuclein aggregation in PD without Alzheimer pathology. α -Synuclein inclusions occur in cholinergic and monoaminergic lower brainstem neurons in asymptomatic cases (Braak stage I and stage II), infiltrate similar neurons in the midbrain and basal forebrain in those with the motor symptoms of PD (Braak stage III and stage IV), and then are found later in limbic and neocortical brain regions with disease progression (Braak stage V and stage VI)²³⁶. 3N, 3rd nerve fibres; CP, cerebral peduncle; RN, red nucleus. Part **h** adapted with permission from REF. 236, Wiley.

Table 1 | Classification of hereditary parkinsonism*

Locus symbol	New designation [‡]	Gene locus	Gene	OMIM (phenotype MIM number; gene/locus MIM number)	Clinical clues
Autosomal dominant Parkinson disease					
PARK1 or PARK4	PARK-SNCA	4q22.1	SNCA	• 168601; 163890 (PARK1) • 605543; 163890 (PARK4)	Missense mutations (PARK1) cause classic Parkinson disease phenotype. Duplication or triplication of this gene (PARK4) causes early-onset Parkinson disease with prominent dementia
PARK8	PARK-LRRK2	12q12	LRRK2	607060; 609007	Classic Parkinson disease phenotype. Variations in LRRK2 include risk-conferring variants and disease-causing mutations
PARK17	PARK-VPS35	16q11.2	VPS35	614203; 601501	Classic Parkinson disease phenotype
Early-onset Parkinson disease (autosomal recessive inheritance)					
PARK2	PARK-Parkin	6q26	PARK2 encoding parkin	600116; 602544	Often presents with lower limb dystonia
PARK6	PARK-PINK1	1p36.12	PINK1	605909; 608309	Psychiatric features are common
PARK7	PARK-DJ1	1p36.23	PARK7 encoding protein deglycase DJ1	606324; 602533	Early-onset Parkinson disease
PARK19B	PARK-DNAJC6	1p31.3	DNAJC6	615528; 608375	Onset of parkinsonism between the third and fifth decades of life
Complex genetic forms (autosomal recessive inheritance)[§]					
PARK9	PARK-ATP13A2	1p36.13	ATP13A2	606693; 610513	Early-onset parkinsonism with a complex phenotype (for example, dystonia, supranuclear gaze palsy, pyramidal signs and cognitive dysfunction); also known as Kufor–Rakeb syndrome
PARK14	PARK-PLA2G6	22q13.1	PLA2G6	256600; 603604	PLAN (or NBIA2) is characterized by a complex clinical phenotype, which does not include parkinsonism in the majority of cases
PARK15	PARK-FBXO7	22q12.3	FBXO7	260300; 605648	Early-onset parkinsonism with pyramidal signs and a variable complex phenotype (for example, supranuclear gaze palsy, early postural instability, chorea and dystonia)
PARK19A	PARK-DNAJC6	1p31.3	DNAJC6	615528; 608375	Juvenile-onset parkinsonism that is occasionally associated with mental retardation and seizures
PARK20	PARK-SYNJ1	21q22.11	SYNJ1	615530; 604297	Patients may have seizures, cognitive decline, abnormal eye movements and dystonia
PARK23	Not yet assigned	15q22.2	VPS13C	616840; 608879	Young-adult-onset parkinsonism associated with progressive cognitive impairment that leads to dementia and dysautonomia

The locus symbols are in accordance with the Online Mendelian Inheritance in Man (OMIM) catalogue (<https://omim.org>). Seven loci, which have been assigned a PARK designation, have a yet unconfirmed relationship to disease (that is, PARK3, unknown gene on 2p13; PARK5, UCHL1 on 4p13; PARK11, GIGYF2S on 2q37.1; PARK13, HTRA2 on 2p13.1; PARK18, ELF4G1 on 3q27.1; PARK21, DNAJC13 on 3q22; and PARK22, CHCHD2 on 7p11.2) and three are classified as risk loci (PARK10 on 1p32; PARK12 on Xq21–q25; and PARK16 on 1q32). Mutations in TMEM230 on 20p12 have also very recently been described to cause monogenic Parkinson disease, but causal relationship to disease is still uncertain^{134,240}. MDS, International Parkinson and Movement Disorder Society; NBIA2, neurodegeneration with brain iron accumulation 2A; PLAN, PLA2G6-associated neurodegeneration. *See REFS 133,241. [‡]On the basis of the recommendations of the MDS Task Force on the nomenclature of genetic movement disorders, which will be regularly updated: MDSGene; available at <http://www.mdsgene.org>^{133,241}. [§]Complex genetic forms that have parkinsonism as a key clinical feature but also present with atypical, multisystem features or other movement disorders.

penetrance of LRRK2 mutations and increased risk of Parkinson disease in GBA mutation carriers, which supports the idea that the proteins encoded by these genes act in shared molecular pathways⁶⁰.

Prion-like propagation of α-synuclein. An additional mechanism for the development of α-synuclein aggregates has recently been proposed. The prion-like hypothesis for α-synuclein posits that once α-synuclein aggregates have formed in a neuron, they can be transported intra-axonally to other brain regions, be released into the extracellular space, be taken up by neighbouring neurons and seed aggregation of endogenous α-synuclein once inside their new cellular host^{61,62}. Cell culture studies have demonstrated that LAS impairment leads to increased secretion of α-synuclein into the extracellular

space through exosomes and that endocytosis is a key mechanism of uptake of extracellular α-synuclein^{63,64}. Thus, initial α-synuclein misfolding in a small number of cells could progressively lead to the spread of α-synuclein aggregates to multiple brain regions over years or decades following the initial insult. This is consistent with the idea that α-synuclein pathology gradually engages more brain regions as the disease progresses, as suggested by Braak *et al.*²³ (FIG. 2h). In addition, this model supports the idea that the first sites of α-synuclein aggregation might be in the gut enteric nerves and the olfactory bulb where they underlie the signs and symptoms associated with prodromal Parkinson disease (for example, anosmia and constipation)^{65,66}, before they spread, eventually leading to motor dysfunction once the substantia nigra becomes involved⁶⁷.

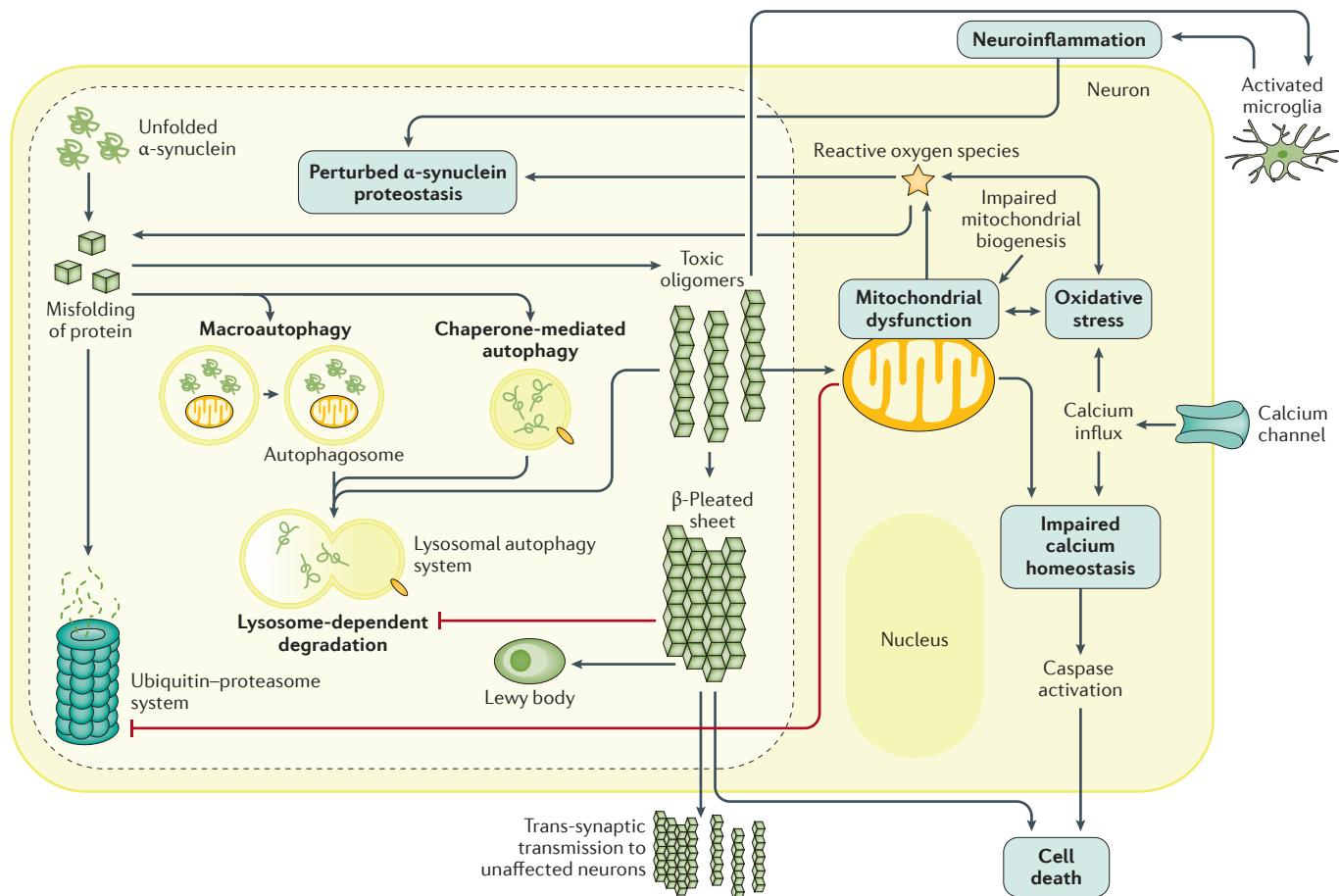


Figure 3 | Molecular mechanisms involved in Parkinson disease. Schematic diagram depicting interactions between major molecular pathways that are implicated in the pathogenesis of Parkinson disease.

Mitochondrial dysfunction

Several lines of evidence have implicated mitochondrial dysfunction as a key element in the pathogenesis of Parkinson disease (reviewed in detail in REFS 68,69; FIG. 3). An emerging picture is one of a vicious cycle in which α-synuclein aggregation and mitochondrial dysfunction exacerbate each other, which could explain why these cellular changes are observed together in degenerating neurons in Parkinson disease.

Activity of mitochondrial complex I, a compound of the electron transport chain, is reduced in several tissues isolated from patients with Parkinson disease^{68,69}. Peroxisome proliferator-activated receptor-γ (PPAR γ) co-activator 1α (PGC1α), a mitochondrial master transcriptional regulator, target genes are generally underexpressed in Parkinson disease⁷⁰. It has been proposed that low levels of α-synuclein are normally present in mitochondria, but that accumulation of the protein inside mitochondria leads to mitochondrial complex I deficits and oxidative stress⁷¹. Activation of PGC1α results in reduced α-synuclein oligomerization and less toxicity *in vitro*, whereas induced PGC1α deficiency by genetic knockdown increases vulnerability to α-synuclein oligomers⁷². Conversely, exposure to α-synuclein oligomers reduces the levels of cellular PGC1α⁷². In animal models, injection of several toxins

that impair mitochondrial function replicates features of Parkinson disease neuropathology^{68,69}. When mitochondrial transcription factor A, which is essential for mitochondrial DNA expression, is selectively depleted in dopaminergic neurons of mice — so-called MitoPark mice — mitochondria in dopaminergic neurons in the substantia nigra develop a defective electron transport chain, leading to neuronal degeneration in adulthood⁷³. Adult mice that lack one allele of *En1* — encoding for engrailed 1, which enhances nuclear translation of the mitochondrial complex proteins NADH-ubiquinone oxidoreductase 75 kDa subunit (NDUFS1) and NDUFS3 — replicate several important features of Parkinson disease neuropathology, such as perturbations of autophagy, neuroinflammation and progressive nigral dopaminergic neuron death following retrograde axonal degeneration⁷⁴. Importantly, axonal degeneration, potentially owing to the energy deficiency, might be an upstream and early neurodegenerative event in Parkinson disease. Human brain imaging studies have demonstrated changes in the striatum in people with Parkinson disease even several years before they are diagnosed^{75,76}, and recent post-mortem studies suggest that nigrostriatal axon terminals are dysfunctional or have degenerated several years before the neuronal cell bodies in the substantia nigra die⁷⁷.

An alternative explanation for the axonal degeneration is that α -synuclein aggregates eventually become obstacles to normal axonal transport⁷⁸.

Recent advances in the understanding of molecular pathways governed by proteins encoded by genes associated with Parkinson disease have provided additional support to the notion that mitochondrial failure is a key event in disease process. For example, *LRRK2* mutations are not only associated with changes in autophagy but also with mitochondrial impairments⁶⁹. Moreover, proteins encoded by *PARK2* and *PINK1*, which are autosomal recessive Parkinson disease genes, cooperate in the clearance of damaged mitochondria through mitophagy⁷⁹. Impaired degradation of MIRO (a protein in the outer mitochondrial membrane that connects the organelle to microtubule motors) seems to have a role in defective clearance of damaged mitochondria. In neurons derived from induced pluripotent stem cells from patients with inherited or sporadic Parkinson disease, degradation of MIRO is reduced, and as a consequence, mitophagy is inefficient, which ultimately could lead to energy failure⁸⁰.

Oxidative stress

Evidence that oxidative stress, as a consequence of mitochondrial dysfunction, is increased in the brain tissue of patients with Parkinson disease is compelling⁸¹ (FIG. 3), but it is debatable whether it occurs early or late during the demise of neurons. Mutations in *DJ1* (also known as *PARK7*), encoding a putative antioxidant, which cause early-onset autosomal recessive Parkinson disease⁸², are associated with increased cellular oxidative stress^{83,84}. Knocking out *Dj1* in mice results in increased protein oxidation in stressed nigral dopaminergic neurons.

Nigral dopaminergic neurons have been suggested to be particularly vulnerable to metabolic and oxidative stress for several reasons. First, they possess particularly long (up to 4.5 metres), unmyelinated axons, with large numbers of synapses (estimated at 1–2.4 million per nigral dopaminergic neuron), which require great energy to be sustained^{85,86}. Second, they (unlike the dopaminergic neurons in the neighbouring ventral tegmental area, which are relatively resilient in Parkinson disease) exhibit autonomous pacemaking activity involving cytosolic calcium oscillations and calcium extrusion at the expense of energy^{87,88}. Third, increased levels of cytosolic dopamine and its metabolites can cause toxic oxidative stress^{89,90}. Last, mitochondrial dysfunction and increased oxidative stress can lead to the depletion of lysosomes⁹¹ and functional impairment of LAS, further demonstrating that several putative pathogenetic pathways in Parkinson disease are intimately linked.

Neuroinflammation

A large number of post-mortem, brain imaging and fluid biomarker studies shows that neuroinflammation is a salient feature of Parkinson disease⁹² (FIG. 3). Although maybe not the initial trigger, neuroinflammation is probably an essential contributor to pathogenesis^{93,94}. Catecholaminergic neurons in the brain tissue of patients

with Parkinson disease and cultured dopaminergic neurons (when exposed to activated microglia or L-DOPA) have been reported to be particularly inclined to express MHC class I proteins, which exposes them to cytotoxic T cell-mediated death if they present antigens⁹⁵.

GWAS indicate that genes associated with the risk of developing Parkinson disease often encode proteins that are expressed in immune cells and that are involved in immune regulation, such as *LRRK2* (which is involved in autophagy by immune cells)^{24,96,97}. Close links between certain genes, protein aggregates and neuroinflammation exist. Evidence from patients and experimental models suggests that α -synuclein aggregation induces both innate and adaptive immunity in Parkinson disease^{93,94} and neuroinflammation can also promote α -synuclein misfolding⁹⁸, suggesting that the two processes participate in a self-aggravating cycle. During prodromal Parkinson disease, tissue inflammation in the olfactory system or gut has been suggested to trigger a sufficient level of α -synuclein misfolding that some α -synuclein aggregates eventually escape the normal degradation mechanisms⁹⁹. Indeed, recent evidence from experiments in *Snca*-overexpressing mice suggests a role of gut microbiota in promoting microglial activation and α -synuclein pathology, as well as motor deficits¹⁰⁰.

However, it would be misleading to suggest that activated immune cells only contribute to the initiation or deterioration of Parkinson disease pathology in the brain. Microglia can phagocytose and degrade extracellular α -synuclein aggregates, and immunotherapies that target α -synuclein, which are currently being developed for clinical trials, rely on the clearance of antibody-bound α -synuclein by activated immune cells¹⁰¹.

Motor circuit pathophysiology

The basal ganglia are part of several parallel, but anatomically segregated thalamo–cortico–basal ganglia circuits, which have important functions in the control of actions and goal-directed behaviour. These circuits are anatomically characterized by a strong convergence of cortical input onto relatively few subcortical output neurons and back to the cortex, suggesting a ‘filter-like’ function. Four circuits with a functionally similar, yet topographically distinct, organization have been identified to subserve limbic, prefrontal-associative, oculomotor and motor functions by linking the corresponding frontal cortical areas and subregions of the thalamus and basal ganglia^{102,103}.

Parkinsonism results from a decreased dopaminergic transmission in the motor region of the striatum with opposing effects on the direct and indirect pathways, which results in increased γ -aminobutyric acid (GABA)-ergic inhibition of thalamocortical projections (FIG. 4). This firing rate model provided a rationale for the renaissance of stereotactic surgery for Parkinson disease in the early 1990s, as akinesia was no longer considered a loss-of-function symptom, but rather the physiological consequence of increased inhibitory output activity of the basal ganglia. Indeed, lesioning of the globus pallidus internus or the subthalamic nucleus proved to be effective in alleviating bradykinesia in animals and

humans^{104,105}. Meanwhile, the model has been amended by additional connections, such as the ‘hyperdirect pathway’, a monosynaptic link between motor cortical areas and the subthalamic nucleus, which changed the perception of the subthalamic nucleus from a passive relay nucleus to a second input structure of the basal ganglia¹⁰⁶. The hyperdirect pathway might have a role in preventing premature responses by reinforcing indirect pathway activity and thereby the ‘breaking’ function of the basal ganglia, thus allowing more time for the selection of the most appropriate response at the cortical level¹⁰⁷. Moreover, recent animal studies have suggested that antidromic activation of the hyperdirect pathway might drive the strong anti-akinetic effect of subthalamic nucleus DBS, which further underlines the functional significance of this second basal ganglia input^{108,109}.

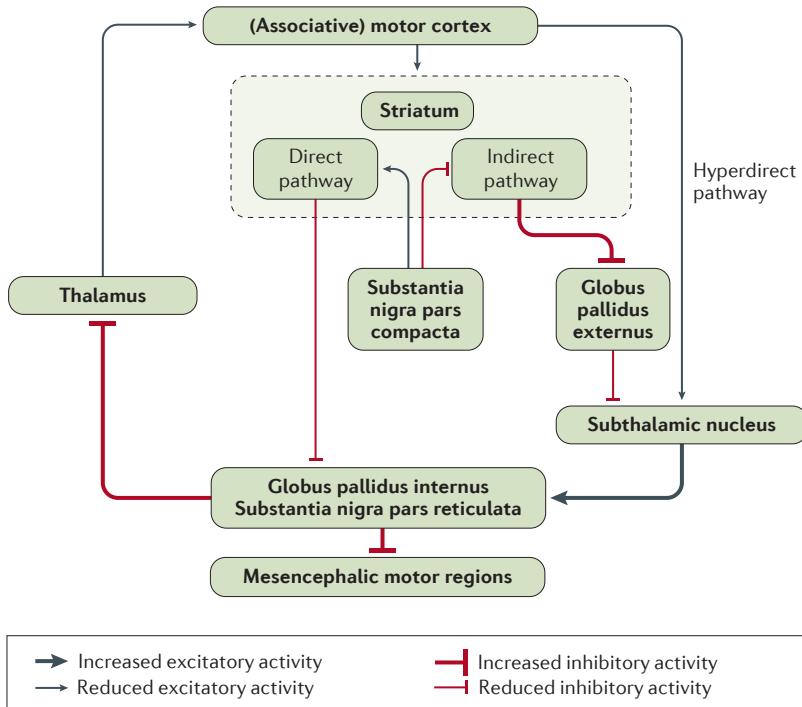


Figure 4 | Motor cortex circuitry activity changes in Parkinson disease.

The motor circuit consists of corticostriatal projections from the primary motor cortex, supplementary motor area, cingulate motor cortex and premotor cortex, terminating on dendrites of striatal medium spiny neurons. The hyperdirect pathway has direct glutamatergic connectivity from the motor cortex to the subthalamic nucleus. The globus pallidus internus and the substantia nigra pars reticulata are the two main output nuclei of the basal ganglia and project to the brainstem and ventrolateral thalamus. The striatal projections to these output nuclei are divided into ‘direct’ and ‘indirect’ pathways. The direct pathway is a monosynaptic connection between medium spiny neurons that express dopamine D1 receptors and GABAergic neurons in the globus pallidus internus and the substantia nigra pars reticulata. The ‘indirect’ pathway originates from medium spiny neurons that express D2 receptors, which project to the globus pallidus externus, and reaches the globus pallidus internus via the subthalamic nucleus as a glutamatergic relay. Through these two pathways, the striatal dopaminergic tone regulates the GABAergic output activity of the basal ganglia. As indicated, parkinsonism is associated with changes in these relays. Indeed, nigrostriatal dopamine deficiency has opposing effects on the direct and indirect pathways. Although D1-mediated direct pathway activity becomes reduced, D2-mediated indirect pathway activity increases, resulting in the net effect of a strong increase in the firing rate of GABAergic basal ganglia output neurons, which over-inhibit downstream thalamocortical and brainstem areas.

However, changes in firing rate are not capable of fully explaining the pathophysiology of hyperkinetic or hypokinetic movement disorders. Growing evidence suggests that movement disorders are characterized by more-complex changes in information processing, such as abnormal neural synchronization and cortico-subcortical coupling in specific frequency bands as indexed by electroencephalogram power density and spectral coherence. The parkinsonian off-state is characterized by enhanced beta-band activity (~20 Hz) in local field potential recordings from the basal ganglia, which is suppressed by dopaminergic medication or DBS in parallel with the clinical improvement of bradykinesia and rigidity^{110,111}. By contrast, hyperkinesia — such as L-DOPA-induced dyskinesia in Parkinson disease — has been associated with increased theta-band activity in the same structures (4–12 Hz)¹¹². High-frequency DBS suppresses either activity and might thus act like a ‘filter’ for abnormally synchronized basal ganglia activity, irrespective of the underlying disorder.

In addition, changes in cerebellar activity and the interaction between the basal ganglia and the cerebellum might be important for the pathophysiology of tremor in Parkinson disease¹¹³, and disorders of balance and gait probably involve abnormal basal ganglia output via projections into the midbrain locomotor region (pedunculopontine and cuneiform nuclei)¹¹⁴. A better understanding of this expanded motor network may help to define alternative targets for DBS in Parkinson disease that target specific symptom profiles.

Diagnosis, screening and prevention

Clinical diagnosis and natural history

Parkinson disease is clinically defined by the presence of bradykinesia and at least one additional cardinal motor feature (rigidity or rest tremor), as well as additional supporting and exclusionary criteria^{115–118} (BOX 1). Onset of motor symptoms is usually unilateral and asymmetry persists throughout the disease. The average age of onset is in the late fifties, with a broad range from <40 to >80 years of age. Young-onset Parkinson disease is commonly defined by an age of onset <45 years and >10% of those individuals have a genetic basis, and the proportion of genetically defined cases rises to >40% of those with disease onset before 30 years of age^{119,120}.

In addition to the cardinal motor features, a majority of patients with Parkinson disease also have non-motor symptoms¹²¹ (FIG. 5). Non-motor symptoms involve a multitude of functions, including disorders of sleep-wake cycle regulation, cognitive impairment (including frontal executive dysfunction, memory retrieval deficits, dementia and hallucinosis), disorders of mood and affect, autonomic dysfunction (mainly orthostatic hypotension, urogenital dysfunction, constipation and hyperhidrosis), as well as sensory symptoms (most prominently hyposmia) and pain¹²¹. Some of these can antedate the onset of classic motor symptoms by years or even decades. Non-motor symptoms become increasingly prevalent over the course of the illness and are a major determinant of QOL, progression of overall disability and of nursing home placement. In one

Box 1 | MDS diagnostic criteria for Parkinson disease**Step 1: diagnosis of parkinsonism (core feature)**

- Presence of bradykinesia as a slowness of movement and a decrement in amplitude or speed (or progressive hesitations or halts) as movements are continued
- In combination with at least one of: rigidity and/or rest tremor

Step 2: determining Parkinson disease as the cause of parkinsonism with two levels of diagnostic certainty

Diagnosis of clinically established Parkinson disease requires all three of the below parameters:

- Absence of absolute exclusion criteria. These criteria include clinical or imaging evidence for alternate diagnoses of parkinsonism, such as atypical parkinsonism, drug-induced parkinsonism or essential tremor.
- Two or more supportive criteria. These include L-DOPA responsiveness, the presence of classic rest tremor, the presence of L-DOPA-induced dyskinésias, the presence of either olfactory loss or cardiac sympathetic denervation on metaiodobenzylguanidine (MIBG) scintigraphy.
- No red flags. This refers to features that are unusual but not absolutely exclusionary for Parkinson disease, for example, the rapid progression of gait impairment that requires wheelchair use or the development of severe autonomic failure within 5 years after onset.

Diagnosis of clinically probable Parkinson disease requires:

- Absence of absolute exclusion criteria (mentioned above)
- Presence of red flags (mentioned above) that are counterbalanced by supportive criteria

For a full listing of absolute exclusion criteria, red flags and supportive criteria see REF. 118.

MDS, International Parkinson and Movement Disorder Society.

long-term study, dementia was present in 83%, hallucinosis in 74%, symptomatic orthostatic hypotension in 48%, constipation in 40% and urinary incontinence in 71% of those patients with Parkinson disease surviving for >20 years. Progressive disability ultimately included treatment-resistant motor symptoms, such as freezing of gait (81%), postural instability and falling (87%, with fractures in 35%) and choking (48%)¹²². Although these milestones of progression are key events in the long-term evolution of Parkinson disease, clinical trials and observational studies so far have focused on the progression of motor impairment as captured by the Unified Parkinson's Disease Rating Scale (UPDRS), which is the most commonly used scale to monitor motor disability associated with Parkinson disease in research settings^{123,124}.

In cases presenting with fully developed classic motor features of Parkinson disease, the clinical diagnosis might seem a straightforward exercise. However, early in the disease, error rates for a clinical diagnosis can be as high as 24% even in specialized centres. The most common misclassifications in clinicopathological series are multiple system atrophy, progressive supranuclear palsy and, less frequently, corticobasal degeneration, and in clinically based studies, common errors relate to essential tremor, drug-induced parkinsonism and vascular parkinsonism¹¹⁶. Accuracy of a clinical diagnosis of Parkinson disease can be improved by the stringent use of standard clinical criteria, such as the UK Parkinson's Disease Society Brain Bank (UKPDSBB) criteria, but even then diagnostic accuracy at first visit is only slightly above 80%, as shown by a recent

meta-analysis of 11 studies assessing a UKPDSBB-based clinical diagnosis against post-mortem pathological examination as the gold standard¹²⁵. Such findings highlight the need for diagnostic tests and biomarkers to enhance diagnostic confidence in early disease, or to eventually diagnose Parkinson disease in its prodromal stages.

Diagnostic tests

Imaging. Visualization of striatal dopamine depletion in patients with Parkinson disease using ¹⁸F-labelled L-DOPA (FIG. 6) and PET was a breakthrough in molecular neuroimaging in the early 1980s¹²⁶. Since then, the field of neuroimaging has seen dramatic advances that are becoming increasingly relevant to Parkinson disease¹²⁷. For example, ¹²³I-ioflupane single-photon emission CT (SPECT) (also known as DaTscan (GE Healthcare)) is approved for clinical routine use and can be used to differentiate between Parkinson disease and clinical mimics that are not associated with presynaptic nigrostriatal terminal dysfunction^{127,128}. Structural MRI helps to identify symptomatic parkinsonism and various MRI techniques can reveal specific changes in the basal ganglia and infratentorial structures in atypical parkinsonism¹²⁹. Advanced MRI techniques and post-processing procedures, including diffusion weighted imaging, volumetric imaging, automated subcortical volume segmentation and multimodal imaging, are being explored to enhance diagnostic accuracy for Parkinson disease versus other types of degenerative parkinsonism^{129–131}. Myocardial sympathetic denervation, assessed with PET or SPECT using noradrenergic tracers, is common in Parkinson disease, but is not seen in patients with atypical parkinsonism or other Parkinson disease mimics¹³². An overview of imaging findings in Parkinson disease is provided in FIG. 6 and Supplementary information S1 (table).

Genetics. The list of mutations causing monogenic types of Parkinson disease continues to grow, as does the number of genes associated with complex phenotypes that include parkinsonism and have been assigned PARK loci (TABLE 1). Several other genes (including GBA, GCH1, ADH1C, TBP, ATXN2, MAPT and GLUD2) have been identified that contribute to an increased risk for the sporadic form of the disease, of which the most prevalent and important are heterozygous mutations in GBA. Large meta-analyses of data sets from GWAS have identified and confirmed many more common low-risk susceptibility variants in other loci in Parkinson disease, which account for additional heritability, each potentially acting in a small but additive manner^{24,133,134}.

Genetic forms only account for a small percentage of Parkinson disease cases in clinical practice, and, as of yet, genetic testing is not part of the routine diagnostic process, except in patients in whom there is a specific suspicion for a possible genetic cause (for example, suggestive family history, early onset (which is typical for several recessive genes) or specific clinical features, such as dystonia as a presenting symptom).

Overall, the implications of genetic testing in clinical routine are limited by reduced penetrance and variable expressivity, and there is currently no impact of genetic findings on practical treatment decisions. This might well change in the future as data from prospective studies on the prognostic implication of genetic mutations emerge and specific therapeutic targets in carriers of mutations associated with Parkinson disease are being pursued. Examples for this are the recently reported increased risk for dementia in patients who have Parkinson disease and carry neuronopathic *GBA* mutations^{49,50} or development programmes for inhibitors of LRRK2 (REF. 135).

Cerebrospinal fluid and blood tests. Although several studies have assessed differential levels of various proteins, most prominently the levels of different α -synuclein species, in the cerebrospinal fluid (CSF) of patients with Parkinson disease versus controls (see Supplementary information S2 (table)), the sensitivities and specificities have been suboptimal and there is currently no clinically useful CSF-based diagnostic test for Parkinson disease¹³⁶. This is also true for blood biomarkers, although associations of different serum or plasma parameters with disease progression have been described, including correlations of lower plasma levels of apolipoprotein A1 with greater severity of motor symptoms¹³⁷.

Screening and prevention

Neuronal dysfunction in Parkinson disease is believed to start long before the defining motor features have become apparent¹³⁸ (FIG. 5). In an attempt to define at-risk individuals or prodromal disease stages, current research efforts have focused on risk factors and biomarkers that have been associated with later diagnosis of Parkinson disease in epidemiological studies^{136,139}. Subtle motor abnormalities (mild parkinsonian signs), such as reduced arm swing, changes in walking pattern, stiffness, tremor or changes in fine motor skills, might be found in as many as 40% of the elderly population, in whom they have been associated with an increased risk for incident Parkinson disease or with other Parkinson disease risk markers⁶⁶. In addition, several studies have demonstrated that subtle motor abnormalities can be detected in individuals a considerable time before the eventual diagnosis, and more-sensitive digital tools to identify these early motor features are being tested¹⁴⁰.

Non-motor symptoms seem to long antedate the onset of classic motor symptoms in individuals with Parkinson disease^{66,139,141} (FIG. 5). Idiopathic rapid eye movement (REM)-sleep behaviour disorder (RBD), in particular, carries a high risk for the development of Parkinson disease or other α -synucleinopathies¹³⁹. Recent prospective studies in large RBD cohorts have found conversion rates to Parkinson disease, dementia associated with Parkinson disease or multiple system

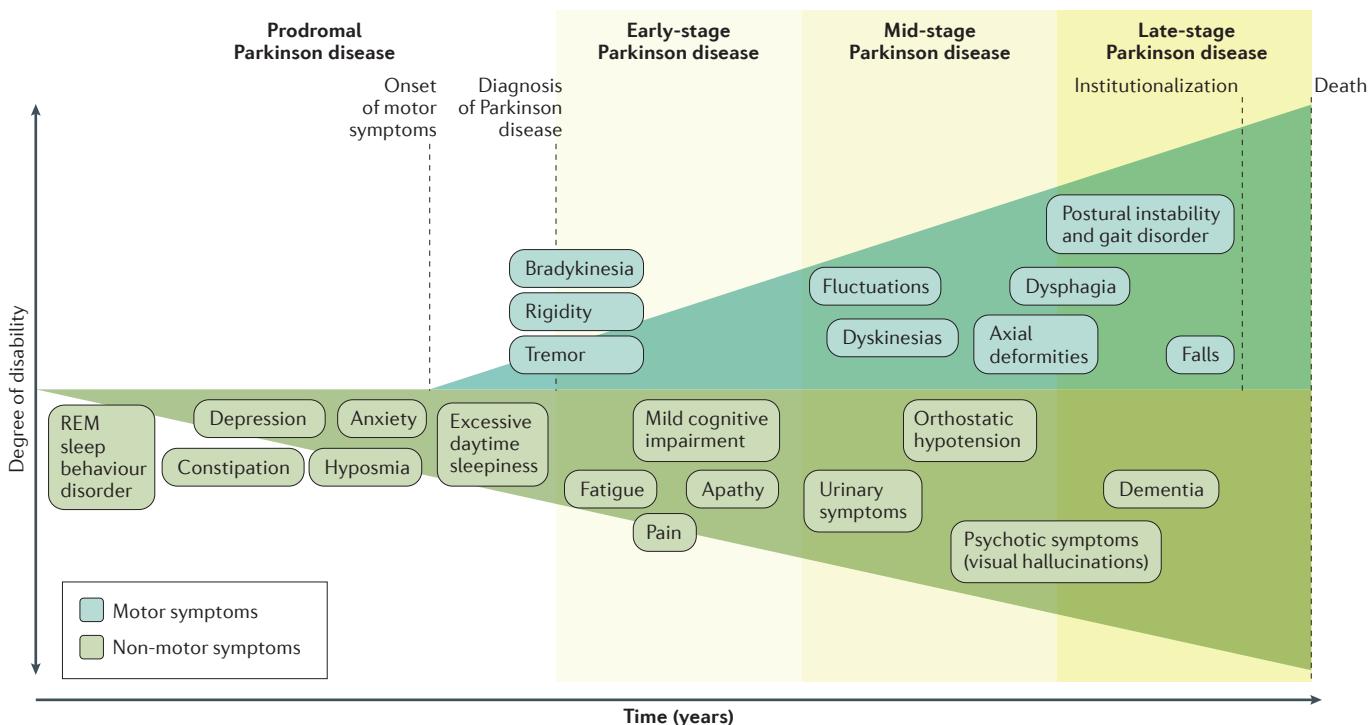


Figure 5 | Clinical symptoms associated with Parkinson disease progression. Diagnosis of Parkinson disease occurs with the onset of motor symptoms (early-stage Parkinson disease) typically in the late fifties, but can be preceded by a prodromal phase of years or even decades, which is characterized by specific non-motor symptoms (prodromal Parkinson disease). Non-motor symptoms become increasingly prevalent and obvious over the course of the illness, but can be present to a variable degree

throughout all stages of Parkinson disease. Progressive disability from Parkinson disease is driven by the combination of these non-motor problems with increasing severity of cardinal motor features, the development of L-DOPA-induced motor complications (mid-stage Parkinson disease) and the evolution of poorly L-DOPA-responsive motor disabilities, such as postural instability, gait problems (including freezing) and dysphagia (late-stage Parkinson disease). REM, rapid eye movement.

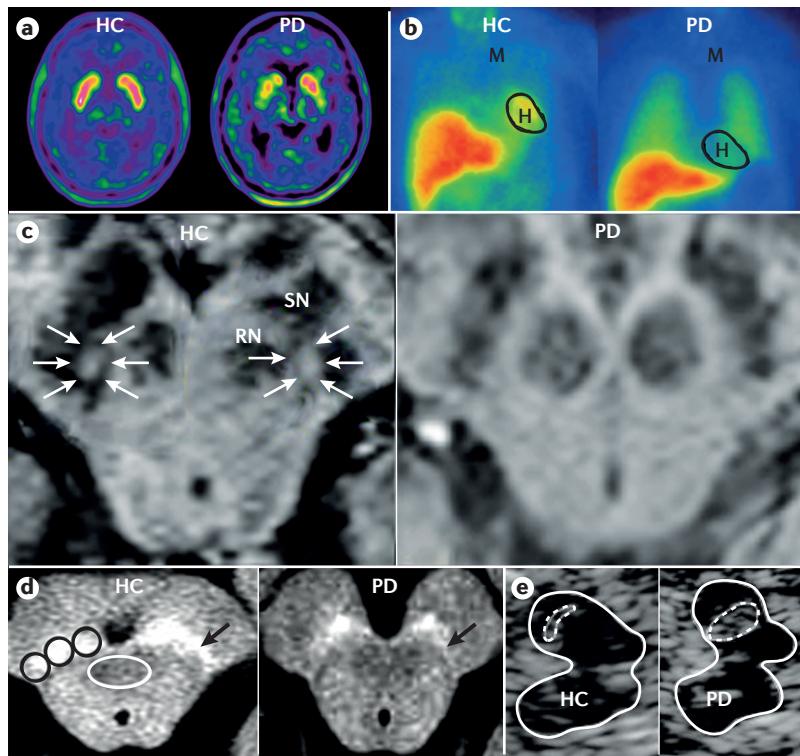


Figure 6 | Imaging methods used to study Parkinson disease. **a** | ^{18}F -DOPA-PET images show normal bilateral tracer uptake in the putamen and caudate nuclei (a typical comma-shaped structure) in a healthy control (HC; left panel), whereas uptake is asymmetric with more marked reduction in the right compared with the left putamen in a patient with early-stage predominantly left-sided Parkinson disease (PD; right panel). **b** | Metaiodobenzylguanidine single-photon emission CT (MIBG SPECT) shows cardiac sympathetic denervation with a profound reduction in MIBG uptake in the heart (H) of a patient with early PD (right panel) compared with a HC (left panel). **c** | MRI of the substantia nigra (SN). Left panel shows a susceptibility weighted imaging (SWI) scan of a HC demonstrating an ovoid area of hyperintensity (arrows) within the dorsolateral SN, which is absent in patients with PD (right panel). **d** | Neuromelanin-sensitive MRI of the SN (black circles in the left panel) shows reduced signal intensity in the lateral part (arrow) in a patient with early PD (right panel) compared with the HC (left panel). No changes are observed in the periaqueductal grey matter (white circle in the left panel). **e** | Transcranial ultrasonography (axial plane) shows an enlarged area of echogenicity in the region of the SN in a patient with PD (right panel) compared with a HC (left panel). The solid lines mark the midbrain area, the dotted lines mark the echogenic area at the anatomical site of the SN on the side of insonation. M, mediastinum; RN, red nucleus. Part **c** is adapted with permission from REF. 237, Wiley. Part **d** is adapted with permission from REF. 238, Elsevier. Part **e** is adapted with permission from REF. 239, Wiley.

atrophy between 15% and 40% over 2–5 years and up to 90% with longer follow-up beyond 10 years¹⁴², but the need for polysomnographic confirmation limits the use of RBD for screening at the population level.

Conversely, hyposmia is an established, albeit less specific risk factor for Parkinson disease, which is easy to screen at relatively low cost and has a relatively high prevalence in the general population⁶⁶. A two-step approach using smell testing as a primary screen and dopamine transporter scan imaging as a secondary screen in hyposmic individuals is being used as a screening approach in the population-based PARS study, in which hyposmia when combined with older age and constipation was associated with deficits in dopamine transporter binding in >40% of such individuals¹⁴³. In a

small cohort of individuals with idiopathic RBD, the presence of hyposmia yielded a predictive value for conversion to Parkinson disease or dementia associated with Parkinson disease of >60% over 5 years¹⁴⁴, which has obvious implications for the calculation of sample sizes for future ‘neuropreventive’ trials targeting prodromal Parkinson disease.

The International Parkinson and Movement Disorder Society (MDS) has published research diagnostic criteria for prodromal Parkinson disease, which are based on epidemiological data about the effects of a large number of risk and prodromal markers⁶⁵. Their predictive validity and, therefore, usefulness for selecting populations for ‘disease prevention’ trials still await prospective testing, but a population-based study has provided first evidence to this effect¹⁴⁵.

Management

Dopaminergic pharmacological targets

Loss of dopaminergic neurons in the substantia nigra pars compacta leading to striatal dopamine depletion is the core mechanism underlying the cardinal motor features of Parkinson disease. Substituting striatal dopamine loss via the systemic administration of the dopamine precursor amino acid L-DOPA represented a revolutionary breakthrough in the treatment of Parkinson disease >50 years ago. Since then, important advances in the understanding of the pharmacological players that regulate nigrostriatal dopaminergic transmission have revealed multiple additional targets for dopaminergic therapies. These can be broadly classified into interventions with primarily presynaptic or postsynaptic activity (FIG. 7).

L-DOPA. L-DOPA has remained the gold standard for Parkinson disease and parkinsonism, and over time, practically all patients with Parkinson disease will require treatment with this agent^{146,147}. However, its use is complicated by the evolution of motor complications, including motor response oscillations and drug-induced dyskinesias (BOX 2). The mechanisms underlying these phenomena, in particular, those responsible for the development of dyskinesias with chronic L-DOPA replacement, are still incompletely understood. Both presynaptic and postsynaptic mechanisms are involved, which eventually lead to non-physiological pulsatile striatal dopamine receptor stimulation and give rise to various maladaptive neuronal responses^{148,149}. The key cause is discontinuous drug delivery due to the short half-life of L-DOPA and the variability in its gastrointestinal absorption and blood–brain barrier transport¹⁵⁰. Novel sustained-release formulations of L-DOPA as well as continuous delivery (either intestinally via percutaneous endoscopic gastro-jejunostomy tubes or subcutaneously via mini-pumps) have been or are being developed to address this problem¹⁵⁰. Clinical observations of reductions of pre-existing dyskinesias with intestinal gel infusions of L-DOPA indeed support the value of the concept of continuous dopaminergic receptor stimulation as a means to prevent the evolution of drug-induced dyskinesias^{150,151}.

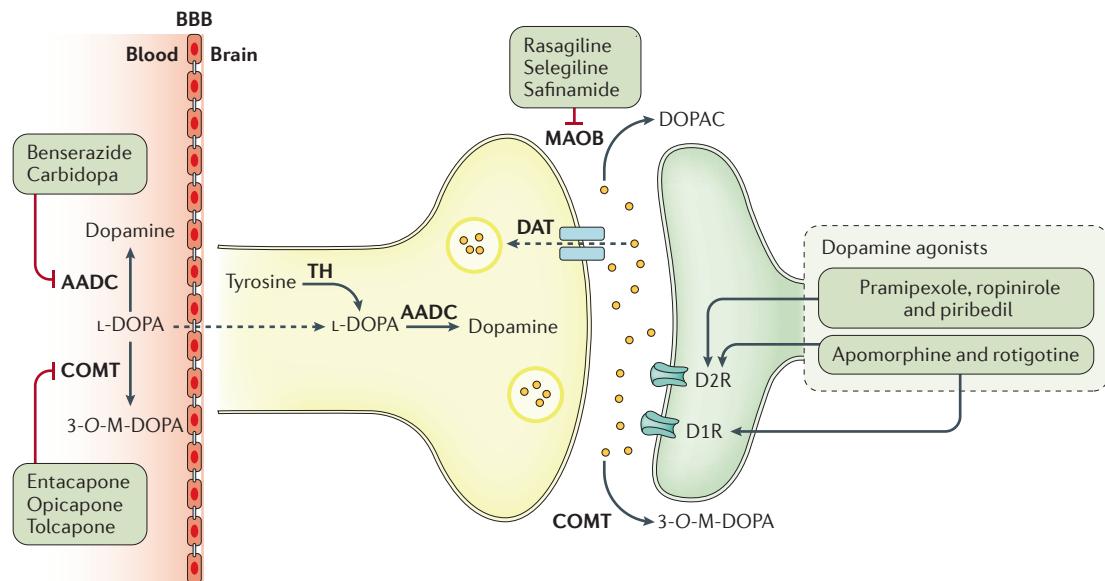


Figure 7 | Dopaminergic drug targets in Parkinson disease. Presynaptic targets include L-DOPA substitution combined with peripherally active inhibitors of aromatic amino acid decarboxylase (AADC) or catechol-O-methyltransferase (COMT). Monoamine oxidase type B (MAOB) inhibitors enhance the synaptic availability of dopamine (both endogenous and exogenous), whereas dopamine agonists act postsynaptically. Dashed arrow from blood to brain designates blood–brain barrier (BBB) transport of L-DOPA. Dashed arrow through the dopamine transporter (DAT) denotes reuptake of dopamine from the synaptic cleft. 3-O-M-DOPA, 3-O-methyl-DOPA; D1R, dopamine D1 receptor; DOPAC, 3,4-dioxy-phenylacetic acid; TH, tyrosine hydroxylase.

Catechol-O-methyltransferase inhibitors. Current L-DOPA preparations include inhibitors of aromatic amino acid decarboxylase (AADC) — for example, carbidopa or benserazide — to prevent peripheral metabolism of dopamine and enhance bioavailability (FIG. 7). As a consequence, the peripheral metabolism of L-DOPA is shifted towards the activity of a secondary metabolic pathway that involves ortho-methylation of L-DOPA via catechol-O-methyltransferase (COMT). Inhibition of this enzyme in the periphery will further enhance bioavailability and the half-life of L-DOPA, which is of particular benefit in patients who have developed motor fluctuations of the wearing-off type¹⁵². Extending the duration of effect of individual L-DOPA doses via COMT inhibitors has become a first-line treatment in these individuals, and currently, three preparations are available for clinical use^{153,154}.

Monoamine oxidase type B inhibitors. Oxidation via monoamine oxidase type B (MAOB) in glial cells is a major clearance mechanism for synaptically released dopamine, next to presynaptic reuptake via the dopamine transporter¹⁵⁵ (FIG. 7). Inhibition of MAOB prolongs and increases synaptic dopamine concentrations, and symptomatic efficacy of MAOB inhibition using the selective inhibitor selegiline as an adjunct to L-DOPA was shown already in the 1970s¹⁵⁶. More recent studies have established the antiparkinsonian efficacy of monotherapy with selegiline and the newer MAOB inhibitor rasagiline, which has also been found to be efficacious when added to L-DOPA in patients with motor fluctuations¹⁵³. Although both selegiline and rasagiline are irreversible ('suicide') inhibitors of MAOB, the most recent marketed agent, safinamide, acts as a reversible MAOB inhibitor¹⁵⁷.

Dopamine agonists. The actions of dopamine on striatal medium spiny neurons are mediated via two classes of dopamine receptors (FIGS 4,7). Dopaminomimetics with direct activity to dopamine receptors (dopamine receptor agonist) mainly target the D2 receptor family and were first introduced into Parkinson disease therapy in the 1970s with the ergot alkaloid bromocriptine and have since become an important medical therapy for motor symptoms^{153,158}. Initial members of this family of drugs were ergoline derivatives, which also activate 5-hydroxytryptamine (5-HT) receptors, including the 5-HT_{2B} subtype, and these became associated with pleuropulmonary and cardiac valvular fibrosis, which introduced important safety concerns. Currently used agents (listed in FIG. 7) are all non-ergoline drugs and are devoid of this activity. An important advantage of dopamine agonists is their longer half-life than L-DOPA, which makes them attractive candidates as adjunct therapies in patients with motor fluctuations^{153,159}. In addition, rotigotine is available as a transdermal patch formulation that affords continuous drug delivery. Overall, dopamine agonists are believed to induce less pulsatile striatal dopamine receptor stimulation than L-DOPA and this is taken as an explanation for the markedly reduced risk to induce motor complications when dopamine agonists are used as initial monotherapy in Parkinson disease^{158,159}. Drawbacks include their reduced overall effect size as compared with L-DOPA and their potential to induce drowsiness and impulse dyscontrol — the latter being possibly associated with their preferential activity at D3 receptors that are located in the ventral striatum, which causes excessive stimulation of the brain reward systems¹⁶⁰. Apomorphine stands out among the other dopamine agonists in terms

of combined activity at both D1 and D2 receptors and equipotency to L-DOPA¹⁶¹. Continuous subcutaneous apomorphine infusions not only smoothen out motor response fluctuations but have also been associated with reductions of pre-existing L-DOPA-induced dyskinésias¹⁶². Currently, new apomorphine formulations, for sublingual use, are in clinical development¹⁶³.

Non-dopaminergic pharmacological targets

Despite the remarkable effect of dopaminergic therapy on the symptoms of Parkinson disease, there is a clear need for therapies that target other pharmacological systems. The symptom categories that need to be addressed by such treatments include the complications of L-DOPA therapy, such as motor fluctuations and L-DOPA-induced dyskinesia as well as L-DOPA-resistant ('non-dopaminergic') motor features including treatment-resistant tremor, freezing of gait, postural instability and falls, swallowing and speech disturbances. Currently, the only available and effective pharmacological treatment for L-DOPA-induced dyskinesia is amantadine, which is thought to work as an *N*-methyl-D-aspartate receptor antagonist^{153,158,159}. TABLE 2 provides a summary of non-dopaminergic pharmacological treatments that are used or are under development to address various motor problems in Parkinson disease.

In some patients, selected non-motor complaints (such as pain, anxiety, panic, depression and restlessness) can fluctuate in response to dopaminergic therapy and these 'non-motor fluctuations' can be equally or more disabling than the motor symptoms^{121,164}. Many non-motor symptoms do not respond to dopamine replacement therapy and some are indeed aggravated or

precipitated by this treatment¹²¹. Anatomical targets for these problems include afferent, efferent and intrinsic basal ganglia connections, various brainstem-originating projections as well as intrinsic cortical connections and, finally, numerous targets outside the central nervous system (for example, in the autonomic nervous system)¹⁶⁵. Non-dopaminergic neurotransmitter and neuro-modulatory systems in these various regions that have been implicated in the symptoms of Parkinson disease include glutamatergic, adenosinergic, noradrenergic, serotonergic, GABAergic, opioidergic, cholinergic and histaminergic pathways¹⁶⁶.

Cognitive dysfunction, depression and autonomic failure are among the most prevalent and troublesome non-motor issues in Parkinson disease. Cholinesterase inhibitors can have striking beneficial effects on the cognitive disturbances of patients with Parkinson disease who also have dementia, an effect that is possibly related to the considerable loss of cholinergic projections from the nucleus basalis of Meynert^{167,168}. The most effective therapy for psychotic symptoms in Parkinson disease is clozapine^{167,168}. All other available atypical neuroleptics, apart from quetiapine, worsen parkinsonism, probably by blocking striatal dopamine D2 receptors. The exact mechanism of action of clozapine in psychosis associated with Parkinson disease is uncertain; a serotonergic effect is strongly supported by the recent positive results using the 5-HT_{2A} inverse agonist pimavanserin¹⁶⁹. It is not known whether depression in Parkinson disease has the same anatomical and pathogenetic bases as depression in the general population¹⁷⁰. In fact, consistent with depression in other circumstances, it is likely that depression in Parkinson disease is not a uniform, homogeneous disorder. Although patients may respond to all types of antidepressant medications, there is limited evidence that tricyclic antidepressants may be more effective than selective serotonin reuptake inhibitors, suggesting a greater role for noradrenergic systems; although this remains to be firmly established¹⁶⁷. Finally, autonomic dysfunction is extremely common, particularly in late-stage Parkinson disease, and pharmacological therapies are largely directed at autonomic nervous system targets. They include the mineralocorticoid fludrocortisone and adrenergic agents (such as midodrine and etilefrine), the noradrenaline precursor (that is, droxidopa) to treat orthostatic hypotension, anti-muscarinics (such as oxybutynin, tolterodine or trospium chloride) for urinary urgency or incontinence, and pro-kinetic drugs (such as macrogol or lubiprostone) to improve constipation^{158,167,171}.

DBS

The breakthrough for DBS as a treatment for Parkinson disease came in 1993 when new concepts of the basal ganglia circuitry led to the identification of the subthalamic nucleus (FIG. 4) as a novel target for DBS¹⁷². DBS is based on the finding that high-frequency (100–200 Hz) electrical stimulation of specific brain targets can mimic the effect of a lesion without the need for destroying brain tissue and involves the implantation of an electrode in brain tissue. Since then, numerous

Box 2 | L-DOPA-induced motor complications

Chronic exposure to L-DOPA is associated with the development of motor complications in ~30% of patients with Parkinson disease after 2–3 years of exposure and >50% after >5 years. Major risk factors are high L-DOPA dose, longer disease duration and younger age. Motor response fluctuations are often referred to as on-off oscillations in analogy to a switch-like action, in which individual doses of L-DOPA produce symptom control ('on' state), which is replaced by the 'off' state when the drug effects have worn off and symptoms recur. They reflect the short half-life of L-DOPA, but variations in gastrointestinal absorption and blood–brain barrier transport as well as striatal pharmacodynamic changes also have a role, thus producing different patterns of variation in motor response.

L-DOPA-induced dyskinésias are adventitious involuntary movements that may be choreic, dystonic or mixed in appearance and occur in different temporal association to the L-DOPA response cycle: dyskinésias associated with the on state are most often choreic in nature and affect the limb, trunk, face and neck, whereas movements associated with the off state are dystonic with painful cramps that chiefly affect the foot and lower limb. Some patients experience dyskinésias (choreic or mixed) at times of transitioning into an on phase or off phase (biphasic dyskinésias). Mechanisms underlying L-DOPA-induced dyskinésias include presynaptic and postsynaptic mechanisms: the loss of dopaminergic nigrostriatal terminals leads to reduced presynaptic dopamine storage capacity and dysregulated dopamine release, whereas discontinuous delivery of L-DOPA results in pulsatile activation of postsynaptic dopamine receptors. Striatal output activity becomes altered owing to supersensitivity of dopamine receptors and structural and molecular changes that lead to altered signal processing in striatal neurons. Serotonergic maladaptive plasticity associated with sprouting of striatal serotonergic terminals, which might result in ectopic dopamine release, and excessive glutamatergic activity in corticostratial and subthalamopallidal projections both contribute to altered activity patterns in basal ganglia–thalamocortical networks.

clinical trials have confirmed the initial observation on the dramatic antiparkinsonian efficacy of DBS of the subthalamic nucleus, which is now an established evidence-based therapy for motor fluctuations and dyskinesia in patients with advanced Parkinson disease¹⁵³. The stimulation-induced improvement is linked to the previous treatment response to L-DOPA, that is, patients with motor symptoms who do not respond to dopaminergic treatment are unlikely to respond to DBS. The only exception to this rule is drug-resistant tremor in patients with an otherwise good response to L-DOPA, in whom DBS can successfully control tremor. In general, ideal candidates have idiopathic Parkinson disease with an excellent L-DOPA response but motor complications

due to long-term medical treatment¹⁷³. Dementia, acute psychosis and major depression are exclusion criteria¹⁷³. Patients with young-onset Parkinson disease fulfil the inclusion criteria for DBS best and are over-represented among the operated group. Although older age is not an absolute exclusion criterion for surgery, surgical adverse events are more often encountered in this group, L-DOPA-resistant symptoms are more frequent, motor rehabilitation is slower and frailty might limit the degree of functional restitution. Bilateral DBS of the subthalamic nucleus reduces (that is, improves) the UPDRS II (activities of daily living) and UPDRS III (motor) scores on average by 50–60% compared with the preoperative medical off-state. Total daily dopaminergic

Table 2 | Non-dopaminergic pharmacological treatments in the motor symptoms of Parkinson disease*

Mechanism of action	Examples	Comments
Motor fluctuations and parkinsonism		
Adenosine A _{2A} receptor antagonists	Istradefylline, preladenant and tozadenant	<ul style="list-style-type: none"> Istradefylline is approved in Japan and is under further evaluation in other countries The development of preladenant has ceased Tozadenant is currently in phase III
Mixed activity that includes inhibition of sodium/calcium channels and monoamine oxidase type B (MAOB) activity	Safinamide and zonisamide	<ul style="list-style-type: none"> Safinamide is approved in Europe and is under review at the US FDA Zonisamide is approved for use in Japan
Tremor		
Anticholinergics	Many available	Often poorly tolerated; newer more-selective muscarinic antagonists are under development
Mixed antagonist: 5-HT _{1A} , 5-HT _{2A} , 5-HT _{2B} , 5-HT _{2C} , muscarinic M1, muscarinic M4, histamine H1, α1-adrenergic, α2-adrenergic, dopamine D2 and dopamine D4 receptors	Clozapine	Not approved for this indication; requires haematological monitoring
L-DOPA-induced dyskinesia		
N-methyl-D-aspartate receptor antagonists	Amantadine and dextromethorphan	<ul style="list-style-type: none"> Amantadine is in routine clinical use; extended-release formulation of amantadine has successfully completed phase III Dextromethorphan/quinidine combination (AVP-923) is under study (quinidine is a CYP2D6 inhibitor)
Mixed antagonist: 5-HT _{1A} , 5-HT _{2A} , 5-HT _{2B} , 5-HT _{2C} , muscarinic M1, muscarinic M4, histamine H1, α1-adrenergic, α2-adrenergic, dopamine D2 and dopamine D4 receptors	Clozapine	Single positive double-blind, placebo-controlled trial, not approved in this indication and requires haematological monitoring
Metabotropic glutamate receptor 5	Mavoglurant (AFQ056) and dipraglurant (ADX48621)	<ul style="list-style-type: none"> Development of mavoglurant stopped after a failed phase IIb trial Dipraglurant is in phase II
α1-Adrenergic and 5-HT _{1A} receptor agonist	Buspirone	Clinically available as an antidepressant; phase III trial in patients with Parkinson disease who have L-DOPA-induced dyskinesia is ongoing
Leviteracetam	Binds to synaptic vesicle glycoprotein 2A and reduces neurotransmitter release	Mixed results in double-blind, placebo-controlled trials
Combined 5-HT _{1A} and 5-HT _{1B} receptor agonist	Eltoprazine	Phase II is ongoing
Selective α7-nicotinic acetylcholine receptor partial agonist	AQW051	Phase II is completed, but results are unavailable
Gait disorders, falls and freezing of gait		
Procholinergic therapy (cholinesterase inhibitors)	Donepezil and rivastigmine; other cholinergic agents (for example, varenicline, a nicotinic agonist) are under study	Variable, mild effects
Noradrenaline reuptake inhibitor	Methylphenidate	Variable effects on freezing of gait using high doses (for example, 80 mg per day)

5-HT, 5-hydroxytryptophan. *See REF. 166.

drug dosage is reduced following surgery by an average of 60%. As a consequence, dyskinetic fluctuations decrease by 60–70% and hypokinetic fluctuations are markedly reduced with a decrease of daily off-time by approximately 70%¹⁷⁴. Several randomized controlled trials have proven that DBS provides a better QOL than best medical management in patients with clinically relevant motor fluctuations and dyskinetic fluctuations¹⁷⁴.

The globus pallidus internus is an alternative surgical target for the treatment of motor complications but does usually not allow reduction of medication¹⁷⁵. Randomized trials comparing the two targets have produced conflicting results of either similar motor benefits¹⁷⁶ or inferior long-term efficacy on motor function and motor fluctuations with surgery in the globus pallidus internus versus the subthalamic nucleus¹⁷⁷.

DBS is a complex therapy that requires a high level of interdisciplinary expertise in the correct surgical placement of the electrode, postoperative programming and the adjustment of neurostimulation and drug therapy¹⁷³. The most relevant adverse events are intracranial bleedings and device complications (such as infections and lead misplacements, among others), which account for a permanent morbidity of >1–3%. The mortality of DBS is <0.5%¹⁷⁸. Thus, the benefit–risk profile of DBS is usually considered to be favourable, in particular, with respect to the large gains in QOL compared with best medical management observed in clinical trials. Psychiatric sequelae of DBS (for example, apathy, depression, impulsiveness or mania) are not uncommon and result from a complex interplay between disease-related psychiatric symptoms, dopaminergic imbalance due to the profound medication changes and stimulation-induced effects on limbic basal ganglia circuits¹⁷⁹. Better devices allowing a finer control over the spatial distribution of current around the electrode, closed-loop neurostimulation systems autoadjusting parameters based on biomarkers and computer-assisted expert systems for surgical planning and postoperative programming are now becoming available and might help to make the procedure less dependent on expert knowledge and provide more consistent outcomes across centres¹⁸⁰.

Exercise-based treatment

Most patients with Parkinson disease have to cope with residual motor disabilities that affect gait and mobility, postural control and balance, as well as speech and swallowing function, which are often poorly responsive to drugs and mostly unresponsive to DBS. In addition to historic use, a steadily increasing number of trials document the effects of various exercise-based strategies on classic motor outcomes (such as the UPDRS), specific parameters (such as gait speed, balance control, freezing, muscle strength and speech) or global measures (for example, QOL)^{153,181,182}. These developments are paralleled by new research evidence for neuroplasticity and the potential neuroprotective effects of activity-enhancing approaches in experimental models¹⁸³ and epidemiological evidence for Parkinson disease risk modulation by physical activity^{184,185}.

Quality of life

Although motor symptoms have been in the foreground of clinical approaches to Parkinson disease, the introduction of patient-reported health-related QOL measures led to the recognition that the multitude of motor and non-motor features of Parkinson disease has a wider effect on the overall health of patients than just motor impairment¹²¹. It is clear that QOL deteriorates with advancing disease and worsening motor disability¹⁸⁶, but in optimally treated patients with Parkinson disease, non-motor features, in particular, autonomic, cognitive and psychiatric aspects, are of greater importance to the QOL of patients¹⁸⁷. Many non-motor symptoms are under-reported and under-recognized, and only in the past years have treatment trials for specific non-motor symptoms been conducted^{167,188}. The most consistently reported non-motor feature of Parkinson disease associated with poorer QOL is depressive mood, including subsyndromal depression¹⁷⁰. Most studies have excluded patients with dementia from QOL studies, but evidence is emerging that the cognitive impairment is also a considerable contributor to the QOL of patients even at early stages¹⁸⁹. The range of other non-motor symptoms seen in Parkinson disease, including constipation, urinary urgency, insomnia, fatigue, pain, other neuropsychiatric presentations and sexual dysfunction, contributes to lower QOL scores, particularly in advanced disease, and are potentially treatable¹⁹⁰.

Clinical trials on the effect of treatments on both motor and non-motor symptoms in Parkinson disease now frequently include QOL measures. QOL measures have been used to combine assessment of efficacy and tolerability of treatments and compare the overall effect of treatments on the overall QOL of patients, rather than on specific disease aspects alone. Examples of treatments that have demonstrated improvement of QOL in patients with Parkinson disease include antiparkinsonian medications, DBS surgery¹⁴⁶, exercise¹⁸², multidisciplinary intervention¹⁹¹ and treatments of non-motor symptoms and comorbid conditions¹⁸⁸.

Well-being of the carer of a patient with Parkinson disease is also affected by the challenges of supporting a person with both physical impairments and mental health issues. The most important contributing factors to carer burden include dementia, falls and non-motor symptoms¹⁹², and the QOL of the patient and mood are closely related to carer burden¹⁹³. Currently, no interventions are validated to improve carer burden, although several trials are under way.

Outlook

Challenges

A major challenge in research on Parkinson disease is that environmental (for example, pollutants) and lifestyle (for example, diet, exercise and smoking) factors probably contribute to lifetime risk, in part by affecting the epigenome in the nervous system. These factors are difficult to accurately measure in large cohorts over the decades that they have a role. Second, Parkinson disease is probably a multifactorial disease. Thus, both upstream pathogenetic molecular mechanisms and some of their

downstream effects probably differ between patients, even if the final outcomes (α -synuclein aggregation and nigral neurodegeneration) are shared. Third, Parkinson disease symptomatology, clinical course and neuropathology vary between patients, which supports the idea that several forms of molecular pathogenesis coexist within the diagnostic realms of the disease. Fourth, disease development might conceivably require the simultaneous activation of more than one pathogenetic pathway, and that certain cellular defence mechanisms fail concomitantly. Last, superimposed on the complex interplay between multiple triggers and failing defences is the process of normal cellular ageing. The greatest risk factor for Parkinson disease is increasing age, and, possibly, the same molecular perturbations that are handled gracefully by a young neuron have catastrophic consequences in an aged counterpart. All this makes the development and testing of putative disease-modifying interventions in clinical trials extremely complex, and the risks to fail the test of controlled trials are high.

Predictive factors

Despite these challenges, there is now also, for the first time, a realistic possibility to define populations at risk or individuals in the earliest stages of Parkinson disease. On the basis of several prospective, population-based studies, various factors that are associated with an increased risk to develop Parkinson disease have been identified. These include behavioural aspects (such as non-smoking or non-use of caffeine), physiological changes (such as hyposmia, constipation or subtle motor abnormalities) and — most strongly of all — the presence of RBD. Genetic, proteomic, metabolomic and tissue biomarkers are also being increasingly characterized, and neuro-imaging might have an important future role in providing information on the risk of developing Parkinson disease. Current prospective cohort studies are trying to assess the sensitivities, specificities and predictive value of various risk markers. Such data will be crucial to define cohorts for future disease modification studies in at-risk populations. The recently proposed MDS research criteria for prodromal Parkinson disease have already been tested retrospectively in one population-based study and hold promise as a first operational step towards future preventive studies¹⁴⁵.

Biomarkers

Specific and sensitive biomarkers that can be used to assess disease risk or progression or can enhance early diagnosis are a major need both in research and clinical care of patients with Parkinson disease. For example, increased levels of serum or plasma uric acid have been found to be associated with a decreased risk of developing Parkinson disease; a meta-analysis calculated a pooled rate ratio of 0.80 for Parkinson disease in people with a one standard deviation increase in the levels of uric acid¹⁹⁴. Another recent study used data from six independent cohorts of patients with Parkinson disease and healthy controls to develop a classification model from 30 genetic risk factors, family history, olfactory function, sex and age, and found excellent separation between the two diagnostic groups¹⁹⁵.

Alterations of gut microbiota have only recently moved into the focus of Parkinson disease research, with several studies showing differences in the composition of the gut microbiota in patients with Parkinson disease compared with controls¹⁹⁶ (see *Supplementary information S2* (table)). The application of ‘omics’ techniques — such as proteomics, metabolomics and transcriptomics — are powerful tools that are capable of mass analyses to identify small changes in protein, metabolite or RNA profiles in fluids or even tissue from healthy and diseased individuals, and they have been started to be used in Parkinson disease¹⁹⁷. The recent initiation of large multi-site consortia for biomarker development in Parkinson disease holds promise for the discovery of new powerful biomarkers¹³⁶.

In addition, recent studies have tried to detect α -synuclein-related pathology in the autonomic nervous system using skin punch biopsies, biopsies of the salivary glands as well as gastrointestinal biopsies with the ultimate goal to define diagnostic markers for the earliest stages of Parkinson disease⁶⁶. Two studies found increased aggregation of α -synuclein and fibre loss in autonomic sudomotor and pilomotor fibres in skin punch biopsies, and these characteristics separated patients with Parkinson disease from controls with a high diagnostic accuracy^{198,199}. Intriguingly, studies have also found immunostaining for phosphorylated α -synuclein in colonic and submandibular gland biopsies in individuals with idiopathic RBD, suggesting that this marker might be used to detect prodromal Parkinson disease (see below)^{200,201}.

Experimental therapies

Two highly experimental techniques that are focused on achieving structural or neurochemical brain repair in Parkinson disease have generated great interest during recent decades: gene therapy and cell transplantation. Although progress towards successful clinical translation has not been rapid in either case, both remain conceptually important approaches that, if successful, can have great clinical impact. In addition, recent advances in our understanding of the molecular pathogenesis of Parkinson disease have revealed novel therapeutic targets for disease-modifying pharmacological therapies.

Gene therapy. The two main strategies for gene therapy in Parkinson disease are viral vector-mediated expression of growth factors or neurotransmitter-synthesizing enzymes. A vast body of experimental evidence suggests that members of the glial cell line-derived neurotrophic factor (GDNF) family protect nigral dopaminergic neurons from death and promote regeneration of their axons following damage²⁰². A small open-label trial with GDNF injections into the putamen in patients with Parkinson disease generated optimism by suggesting that GDNF might reduce symptoms²⁰³, but a larger placebo-controlled trial with injections of smaller doses of GDNF failed to show clinical benefit²⁰⁴. This disappointing outcome of the controlled GDNF trial did not deter the company Ceregen from testing gene therapy that induced expression of neurturin, a less potent

member of the GDNF family of growth factors that had demonstrated efficacy in animal models²⁰⁵. In a series of trials targeting the putamen or both the putamen and the substantia nigra, it was found that adeno-associated virus (AAV)-mediated expression of neurturin was safe, but again neither approach stood the test of randomized clinical trials^{206,207}. Post-mortem findings suggested that neurturin was only expressed in relatively few cells surrounding the injection tracts, limiting its neurorestorative potential, and it has been suggested to target earlier stages of Parkinson disease, when more of the nigrostriatal axons still remain functional^{206,207}. An ongoing safety trial (predicted to be completed in 2018) is testing the effects of AAV-mediated expression of GDNF in Parkinson disease, and with improved vector technology and better understanding of the effects of growth factors, it is likely that there will be additional attempts at inducing neurorestoration in the future.

Clinical trials are also underway using viral vector-mediated expression of key enzymes in the dopamine synthesis pathway. Thus, in different studies, lentiviral and AAV vectors that express tyrosine hydroxylase with cofactors and AADC have been injected into the striatum, with initial safety reports already published^{208,209}. The strategy is to genetically modify cells in the striatum so that they can produce and release dopamine locally, either from tyrosine or from peripherally administered L-DOPA or dopamine. Animal studies have demonstrated that this approach is feasible and that it might not just provide relief of dopamine-dependent motor symptoms, but by providing constant dopamine receptor stimulation, it could also reduce the risk of motor fluctuations developing later on^{210,211}. Another approach has targeted the subthalamic nucleus with AAV2 vector-mediated delivery of glutamate decarboxylase to induce GABAergic inhibition of subthalamic nucleus firing, with promising results from a sham-surgery-controlled phase II trial²¹².

Fetal cell transplantation. In the 1990s, cell transplantation was considered a promising approach to brain repair in Parkinson disease. Open-label trials suggested that immature dopaminergic neurons obtained from aborted embryos or fetuses not only could restore striatal dopamine transmission and connectivity (as evidenced by *in vivo* PET and morphological findings upon autopsy) but also could reduce the motor symptoms^{213,214}. However, the publication of two US NIH-sponsored, double-blind, placebo-controlled trials in 2001 and 2003 brought the clinical programmes to a standstill^{215,216}. Not only was there no evidence of clinical benefit in these trials but it was also reported that some patients developed uncontrollable graft-induced dyskinesias^{215,216}, which was also confirmed in a retrospective analysis of patients in the open-label trials²¹⁷. Although the clinical trials were halted, laboratory research into the mechanisms underlying these adverse effects started at the same time as the modern era of research into regenerative stem cell therapies was born. On the basis of animal experiments and clinical observations, today, it is believed that graft-induced dyskinesias

are unlikely to develop if the number of serotonergic neurons included in the dissected graft tissue is minimized²¹⁸ and if the selected patients do not already exhibit L-DOPA-induced dyskinesias before surgery²¹⁹. In 2008, concerns were raised about the future of the cell transplantation approach when it was reported that Lewy pathology appears inside grafted neurons over 10 years after surgery. Today, the consensus is that there are progressive signs of degeneration (loss of dopamine transporters and tyrosine hydroxylase, as well as the appearance of Lewy pathology) in grafted neurons²²⁰, but that it takes well over a decade before these pathological changes might impair the function of the transplants. In the open-label setting, quantitative evidence of beneficial graft effects was shown in at least two patients up to 15–18 years after surgery²²¹. In 2015, an open-label study with fetal dopaminergic neuron implants was initiated by the European Union-funded team TRANSEURO; this group plans to have operated on 20 patients, all at a relatively early stage of disease, before the end of 2017 (REFS 222,223). This trial will help to clarify whether graft-induced dyskinesias can be avoided and will also help to lay the foundations for future trials that use stem cell-derived neurons as the source of transplantable dopaminergic neurons.

Stem cells as donor tissue. Over the past 10–15 years, experimental stem cell therapy in animal models of Parkinson disease has developed dramatically²²². Today, it is possible to generate dopaminergic neurons with midbrain characteristics from two forms of human pluripotent stem cells: human embryonic stem cells and human induced pluripotent stem cells²²². Recent studies have shown that they survive grafting into animals, grow axons that innervate the brain and support functional recovery from lesion-induced deficits²²². Current research is focused on resolving issues related to scaling up production of cells, guaranteeing safety and meeting the increasing regulatory demands that are placed on biological products²²². Although several commercial entities already advertise different types of stem cell therapies for Parkinson disease, the underlying scientific rationale for using the proposed cell type is frequently not strong²²⁴. The development of stem cell-based trials should follow the guidelines set forth by the International Society for Stem Cell Research²²⁵. One can expect that cell transplantation clinical trials in Parkinson disease that use stem cell-derived products, which are well-validated scientifically, will commence 2–3 years from now. Progress in this area is stimulated by the existence of an international consortium of scientists called G-Force-PD, which is devoted to clinical translation of stem cell therapy in Parkinson disease and holds regular meetings in which technical advances and protocols are shared²²².

Novel targets for disease modification. The growing evidence for oligomerization and fibrillar aggregation of pathological α-synuclein species and their possible cell-to-cell transmission as a key event in the molecular pathogenesis of Parkinson disease has moved

multimerization and extracellular and intracellular handling of this protein into focus as targets for novel therapies. Two immunological approaches are currently in active clinical development. First, active immunization with a novel vaccine containing short peptides that are homologous to α -synuclein conjugated to a carrier (AFFITOPE, AFFiRiS) induced the formation of antibodies specifically directed against the carboxy terminus of human α -synuclein, cleared α -synuclein aggregates and reduced neuropathology in a transgenic mouse model of Parkinson disease²²⁶. A phase I/II safety trial in 28 individuals with Parkinson disease also provided evidence for induction antibody formation against α -synuclein without safety concerns, as well as immunological efficacy of booster vaccinations up to 3 years after initial immunization²²⁷. A placebo-controlled phase II trial of AFF03, a second AFFITOPE technology vaccine against α -synuclein, is currently ongoing.

A second approach is passive immunization via monoclonal antibodies against α -synuclein and, to date, three candidate compounds have reached early phases of clinical development²²⁸. Following the recent publication of a first-in-human placebo-controlled phase I study of one of these compounds (PRX002)²²⁹, this field will almost certainly also gain momentum over the next 5 years in Parkinson disease.

Other α -synuclein targeting approaches focus on extracellular α -synuclein-binding sites, inhibitors of α -synuclein aggregation or enhancers of α -synuclein clearance via the LAS^{235,230}, but have not yet progressed into clinical development. However, other experimental approaches pursue different targets, including the agonist of the glucagon-like peptide 1 receptor (exenatide)²³¹, the urate precursor (inosine)²³², the GBA chaperone (ambroxol)²³³ or the calcium channel antagonist (isradipine)²³⁴, and some of these are currently being tested in clinical trials in Parkinson disease.

Clinical trials

To date, all clinical disease modification trials in Parkinson disease have been performed in individuals with early disease as defined by the presence of classic motor features, and they have all failed to show unequivocal positive effects. Studying interventions in the prodromal or preclinical phase of disease should offer greater promise of success, assuming less-advanced pathology and greater potential to intervene at crucial points of molecular pathogenesis. These could have a greater likelihood of success if patients who are recruited into clinical trials can be shown to share some form of biomarker that relates to the therapeutically targeted pathogenetic mechanism.

1. Twelves, D., Perkins, K. S. & Counsell, C. Systematic review of incidence studies of Parkinson's disease. *Mov. Disord.* **18**, 19–31 (2003).
2. Savica, R., Grossardt, B. R., Bower, J. H., Ahlskog, J. E. & Rocca, W. A. Incidence and pathology of synucleinopathies and tauopathies related to parkinsonism. *JAMA Neurol.* **70**, 859–866 (2013).
3. Van Den Eeden, S. K. *et al.* Incidence of Parkinson's disease: variation by age, gender, and race/ethnicity. *Am. J. Epidemiol.* **157**, 1015–1022 (2003).
4. Pringsheim, T., Jette, N., Frolkis, A. & Steeves, T. D. The prevalence of Parkinson's disease: a systematic review and meta-analysis. *Mov. Disord.* **29**, 1583–1590 (2014).
5. Pinter, B. *et al.* Mortality in Parkinson's disease: a 38-year follow-up study. *Mov. Disord.* **30**, 266–269 (2015).
6. Lix, L. M. *et al.* Socioeconomic variations in the prevalence and incidence of Parkinson's disease: a population-based analysis. *J. Epidemiol. Community Health* **64**, 335–340 (2010).
7. Dorsey, E. R. *et al.* Projected number of people with Parkinson disease in the most populous nations, 2005 through 2030. *Neurology* **68**, 384–386 (2007).
8. Vos, T. *et al.* Years lived with disability (YLDs) for 1160 sequelae of 289 diseases and injuries 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* **380**, 2163–2196 (2012).
9. Murray, C. J. *et al.* Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* **380**, 2197–2223 (2012).
10. Leibson, C. L. *et al.* Direct medical costs associated with Parkinson's disease: a population-based study. *Mov. Disord.* **21**, 1864–1871 (2006).
11. Baldereschi, M. *et al.* Parkinson's disease and parkinsonism in a longitudinal study: two-fold higher incidence in men. ILSA Working Group. Italian Longitudinal Study on Aging. *Neurology* **55**, 1358–1363 (2000).
12. Kusumi, M., Nakashima, K., Harada, H., Nakayama, H. & Takahashi, K. Epidemiology of Parkinson's disease in Yonago City, Japan: comparison with a study carried out 12 years ago. *Neuroepidemiology* **15**, 201–207 (1996).
13. Chillag-Talmor, O. *et al.* Use of a refined drug tracer algorithm to estimate prevalence and incidence of Parkinson's disease in a large Israeli population. *J. Parkinsons Dis.* **1**, 35–47 (2011).
14. Gordon, P. H., Mehal, J. M., Holman, R. C., Rowland, A. S. & Cheek, J. E. Parkinson's disease among American Indians and Alaska natives: a nationwide prevalence study. *Mov. Disord.* **27**, 1456–1459 (2012).
15. Morens, D. M. *et al.* Epidemiologic observations on Parkinson's disease: incidence and mortality in a prospective study of middle-aged men. *Neurology* **46**, 1044–1050 (1996).
16. Ascherio, A. & Schwarzschild, M. A. The epidemiology of Parkinson's disease: risk factors and prevention. *Lancet Neurol.* **15**, 1257–1272 (2016). **This is a comprehensive and up-to-date review of epidemiological data on risk factors for Parkinson disease.**
17. Dickson, D. W. *et al.* Neuropathological assessment of Parkinson's disease: refining the diagnostic criteria. *Lancet Neurol.* **8**, 1150–1157 (2009). **This is a joint review of diagnostic criteria for a neuropathological diagnosis of Parkinson disease by leading neuropathologists.**
18. Halliday, G. M., Holton, J. L., Revesz, T. & Dickson, D. W. Neuropathology underlying clinical variability in patients with synucleinopathies. *Acta Neuropathol.* **122**, 187–204 (2011).
19. Fearnley, J. M. & Lees, A. J. Ageing and Parkinson's disease: substantia nigra regional selectivity. *Brain* **114**, 2283–2301 (1991).
20. Damier, P., Hirsch, E. C., Agid, Y. & Graybiel, A. M. The substantia nigra of the human brain. II. Patterns of loss of dopamine-containing neurons in Parkinson's disease. *Brain* **122**, 1437–1448 (1999).
21. Dijkstra, A. A. *et al.* Stage-dependent nigral neuronal loss in incidental Lewy body and Parkinson's disease. *Mov. Disord.* **29**, 1244–1251 (2014).
22. Iacono, D. *et al.* Parkinson disease and incidental Lewy body disease: just a question of time? *Neurology* **85**, 1670–1679 (2015).
23. Braak, H. *et al.* Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol. Aging* **24**, 197–211 (2003). **In this landmark paper, Braak and colleagues first introduced the concept of spreading of pathology in the parkinsonian brain.**
24. Nalls, M. A. *et al.* Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. *Nat. Genet.* **46**, 989–993 (2014). **This paper is a large meta-analysis of GWAS that describes the genetic variants that alter the risk for Parkinson disease.**
25. Vekrellis, K., Xilouri, M., Emmanouilidou, E., Rideout, H. J. & Stefanis, L. Pathological roles of alpha-synuclein in neurological disorders. *Lancet Neurol.* **10**, 1015–1025 (2011).
26. Soldner, F. *et al.* Parkinson-associated risk variant in distal enhancer of alpha-synuclein modulates target gene expression. *Nature* **533**, 95–99 (2016).
27. Wales, P., Pinho, R., Lazaro, D. F. & Outeiro, T. F. Limelight on alpha-synuclein: pathological and mechanistic implications in neurodegeneration. *J. Parkinsons Dis.* **3**, 415–459 (2013).
28. Burre, J. The synaptic function of alpha-synuclein. *J. Parkinsons Dis.* **5**, 699–713 (2015).
29. Kim, C. & Lee, S. J. Controlling the mass action of alpha-synuclein in Parkinson's disease. *J. Neurochem.* **107**, 303–316 (2008).
30. Melki, R. Role of different alpha-synuclein strains in synucleinopathies, similarities with other neurodegenerative diseases. *J. Parkinsons Dis.* **5**, 217–227 (2015).
31. Kaushik, S. & Cuervo, A. M. Proteostasis and aging. *Nat. Med.* **21**, 1406–1415 (2015).
32. Xilouri, M., Brekk, O. R. & Stefanis, L. Alpha-synuclein and protein degradation systems: a reciprocal relationship. *Mol. Neurobiol.* **47**, 537–551 (2013).
33. Brundin, P., Li, J. Y., Holton, J. L., Lindvall, O. & Revesz, T. Research in motion: the enigma of Parkinson's disease pathology spread. *Nat. Rev. Neurosci.* **9**, 741–745 (2008). **This review marks the start of a new research area that is focused on the cell-to-cell propagation of α -synuclein aggregates, based on observations of Lewy bodies in grafted neurons in Parkinson disease.**
34. Xilouri, M., Vogiatzi, T., Vekrellis, K., Park, D. & Stefanis, L. Aberrant alpha-synuclein confers toxicity to neurons in part through inhibition of chaperone-mediated autophagy. *PLoS ONE* **4**, e5515 (2009).
35. Chu, Y. & Kordower, J. H. Age-associated increases of alpha-synuclein in monkeys and humans are associated with nigrostriatal dopamine depletion: is this the target for Parkinson's disease? *Neurobiol. Dis.* **25**, 134–149 (2007).
36. Sarkar, S., Davies, J. E., Huang, Z., Tunnacliffe, A. & Rubinsztein, D. C. Trehalose, a novel mTOR-independent autophagy enhancer, accelerates the clearance of mutant huntingtin and alpha-synuclein. *J. Biol. Chem.* **282**, 5641–5652 (2007).

37. Steele, J. W. *et al.* Latrepirdine stimulates autophagy and reduces accumulation of alpha-synuclein in cells and in mouse brain. *Mol. Psychiatry* **18**, 882–888 (2013).
38. Chu, Y., Dodiya, H., Aebischer, P., Olanow, C. W. & Kordower, J. H. Alterations in lysosomal and proteasomal markers in Parkinson's disease: relationship to alpha-synuclein inclusions. *Neurobiol. Dis.* **35**, 385–398 (2009).
39. Alvarez-Erviti, L. *et al.* Chaperone-mediated autophagy markers in Parkinson disease brains. *Arch. Neurol.* **67**, 1464–1472 (2010).
40. Anglade, P. *et al.* Apoptosis and autophagy in nigral neurons of patients with Parkinson's disease. *Histol. Histopathol.* **12**, 25–31 (1997).
41. Emmanouilidou, E., Stefanis, L. & Vekrellis, K. Cell-produced alpha-synuclein oligomers are targeted to, and impair, the 26S proteasome. *Neurobiol. Aging* **31**, 953–968 (2010).
42. Winslow, A. R. *et al.* Alpha-synuclein impairs macroautophagy: implications for Parkinson's disease. *J. Cell Biol.* **190**, 1023–1037 (2010).
43. Tanik, S. A., Schultheiss, C. E., Volpicelli-Daley, L. A., Brundin, K. R. & Lee, V. M. Lewy body-like alpha-synuclein aggregates resist degradation and impair macroautophagy. *J. Biol. Chem.* **288**, 15194–15210 (2013).
44. Martinez-Vicente, M. *et al.* Dopamine-modified alpha-synuclein blocks chaperone-mediated autophagy. *J. Clin. Invest.* **118**, 777–788 (2008).
45. Volpicelli-Daley, L. A. *et al.* G2019S-LRRK2 expression augments alpha-synuclein sequestration into inclusions in neurons. *J. Neurosci.* **36**, 7415–7427 (2016).
46. Sidransky, E. *et al.* Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. *N. Engl. J. Med.* **361**, 1651–1661 (2009).
47. Fernandes, H. J. *et al.* ER stress and autophagic perturbations lead to elevated extracellular alpha-synuclein in GBA-N370S Parkinson's iPSC-derived dopamine neurons. *Stem Cell Reports* **6**, 342–356 (2016).
48. Rocha, E. M. *et al.* Progressive decline of glucocerebrosidase in aging and Parkinson's disease. *Ann. Clin. Transl. Neurol.* **2**, 433–438 (2015).
49. Cilia, R. *et al.* Survival and dementia in GBA-associated Parkinson's disease: the mutation matters. *Ann. Neurol.* **80**, 662–673 (2016).
50. Liu, G. *et al.* Specifically neuropathic Gaucher's mutations accelerate cognitive decline in Parkinson's. *Ann. Neurol.* **80**, 674–685 (2016).
51. Mazzulli, J. R. *et al.* Gaucher disease glucocerebrosidase and alpha-synuclein form a bidirectional pathogenic loop in synucleinopathies. *Cell* **146**, 37–52 (2011).
52. Sardi, S. P. *et al.* Augmenting CNS glucocerebrosidase activity as a therapeutic strategy for parkinsonism and other Gaucher-related synucleinopathies. *Proc. Natl. Acad. Sci. USA* **110**, 3537–3542 (2013).
53. Vilarino-Guell, C. *et al.* VPS35 mutations in Parkinson disease. *Am. J. Hum. Genet.* **89**, 162–167 (2011).
54. Zimprich, A. *et al.* A mutation in VPS35, encoding a subunit of the retromer complex, causes late-onset Parkinson disease. *Am. J. Hum. Genet.* **89**, 168–175 (2011).
55. Seaman, M. & Freeman, C. L. Analysis of the retromer complex-WASH complex interaction illuminates new avenues to explore in Parkinson disease. *Commun. Integr. Biol.* **7**, e29483 (2014).
56. Tang, F. L. *et al.* VPS35 in dopamine neurons is required for endosome-to-golgi retrieval of Lamp2a, a receptor of chaperone-mediated autophagy that is critical for alpha-synuclein degradation and prevention of pathogenesis of Parkinson's disease. *J. Neurosci.* **35**, 10613–10628 (2015).
57. Dhungel, N. *et al.* Parkinson's disease genes VPS35 and EIF4G1 interact genetically and converge on alpha-synuclein. *Neuron* **85**, 76–87 (2015).
58. Ramirez, A. *et al.* Hereditary parkinsonism with dementia is caused by mutations in ATP13A2, encoding a lysosomal type 5P-type ATPase. *Nat. Genet.* **38**, 1184–1191 (2006).
59. Dehay, B. *et al.* Lysosomal dysfunction in Parkinson disease: ATP13A2 gets into the groove. *Autophagy* **8**, 1389–1391 (2012).
60. Lubbe, S. J. *et al.* Additional rare variant analysis in Parkinson's disease cases with and without known pathogenic mutations: evidence for oligogenic inheritance. *Hum. Mol. Genet.* **25**, 5483–5489 (2016).
61. Angot, E., Steiner, J. A., Hansen, C., Li, J. Y. & Brundin, P. Are synucleinopathies prion-like disorders? *Lancet Neurol.* **9**, 1128–1138 (2010).
62. Brundin, P., Melki, R. & Kopito, R. Prion-like transmission of protein aggregates in neurodegenerative diseases. *Nat. Rev. Mol. Cell Biol.* **11**, 301–307 (2010).
63. Tyson, T., Steiner, J. A. & Brundin, P. Sorting out release, uptake and processing of alpha-synuclein during prion-like spread of pathology. *J. Neurochem.* **139** (Suppl. 1), 275–289 (2016).
64. Mao, X. *et al.* Pathological alpha-synuclein transmission initiated by binding lymphocyte-activation gene 3. *Science* **353**, aah3374 (2016).
65. Berg, D. *et al.* MDS research criteria for prodromal Parkinson's disease. *Mov. Disord.* **30**, 1600–1611 (2015). **This is the first paper that provides a research framework to operationalize the diagnosis of the prodromal stages of Parkinson disease.**
66. Mahlknecht, P., Seppi, K. & Poewe, W. The concept of prodromal Parkinson's disease. *J. Parkinsons Dis.* **5**, 681–697 (2015).
67. George, S., Rey, N. L., Reichenbach, N., Steiner, J. A. & Brundin, P. Alpha-synuclein: the long distance runner. *Brain Pathol.* **23**, 350–357 (2013).
68. Schapira, A. H. Mitochondrial dysfunction in Parkinson's disease. *Cell Death Differ.* **14**, 1261–1266 (2007).
69. Bose, A. & Beal, M. F. Mitochondrial dysfunction in Parkinson's disease. *J. Neurochem.* **139** (Suppl. 1), 216–231 (2016).
70. Zheng, B. *et al.* PGC-1alpha, a potential therapeutic target for early intervention in Parkinson's disease. *Sci. Transl. Med.* **2**, 52ra73 (2010).
71. Devi, L., Raghavendran, V., Prabhu, B. M., Avadhani, N. G. & Anandatheerthavarada, H. K. Mitochondrial import and accumulation of alpha-synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. *J. Biol. Chem.* **283**, 9089–9100 (2008).
72. Eschbach, J. *et al.* Mutual exacerbation of peroxisome proliferator-activated receptor gamma coactivator 1alpha deregulation and alpha-synuclein oligomerization. *Ann. Neurol.* **77**, 15–32 (2015).
73. Ekstrand, M. I. *et al.* Progressive parkinsonism in mice with respiratory-chain-deficient dopamine neurons. *Proc. Natl. Acad. Sci. USA* **104**, 1325–1330 (2007).
74. Nordstrom, U. *et al.* Progressive nigrostriatal terminal dysfunction and degeneration in the engrailed1 heterozygous mouse model of Parkinson's disease. *Neurobiol. Dis.* **73**, 70–82 (2015).
75. Sossi, V. *et al.* Changes of dopamine turnover in the progression of Parkinson's disease as measured by positron emission tomography: their relation to disease-compensatory mechanisms. *J. Cereb. Blood Flow Metab.* **24**, 869–876 (2004).
76. Sossi, V. *et al.* Dopamine turnover increases in asymptomatic LRRK2 mutations carriers. *Mov. Disord.* **25**, 2717–2723 (2010).
77. Kordower, J. H. *et al.* Disease duration and the integrity of the nigrostriatal system in Parkinson's disease. *Brain* **136**, 2419–2431 (2013).
78. Lamberts, J. T., Hildebrandt, E. N. & Brundin, P. Spreading of alpha-synuclein in the face of axonal transport deficits in Parkinson's disease: a speculative synthesis. *Neurobiol. Dis.* **77**, 276–283 (2015).
79. Pickrell, A. M. & Youle, R. J. The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron* **85**, 257–273 (2015).
80. Hsieh, C. H. *et al.* Functional impairment in vitro degradation and mitophagy is a shared feature in familial and sporadic Parkinson's disease. *Cell Stem Cell* **19**, 709–724 (2016).
81. Dias, V., Junn, E. & Mouradian, M. M. The role of oxidative stress in Parkinson's disease. *J. Parkinsons Dis.* **3**, 461–491 (2013).
82. Bonifati, V. *et al.* Mutations in the DJ-1 gene associated with autosomal recessive early-onset parkinsonism. *Science* **299**, 256–259 (2003).
83. Di Nottia, M. *et al.* DJ-1 modulates mitochondrial response to oxidative stress: clues from a novel diagnosis of PARK7. *Clin. Genet.* <http://dx.doi.org/10.1111/cge.12841> (2016).
84. Guzman, J. N. *et al.* Oxidant stress evoked by pacemaking in dopaminergic neurons is attenuated by DJ-1. *Nature* **468**, 696–700 (2010).
85. Bolam, J. P. & Pissadaki, E. K. Living on the edge with too many mouths to feed: why dopamine neurons die. *Mov. Disord.* **27**, 1478–1483 (2012).
86. Pissadaki, E. K. & Bolam, J. P. The energy cost of action potential propagation in dopamine neurons: clues to susceptibility in Parkinson's disease. *Front. Comput. Neurosci.* **7**, 13 (2013).
87. Surmeier, D. J., Guzman, J. N., Sanchez-Padilla, J. & Schumacker, P. T. The role of calcium and mitochondrial oxidant stress in the loss of substantia nigra pars compacta dopaminergic neurons in Parkinson's disease. *Neuroscience* **198**, 221–231 (2011).
88. Surmeier, D. J. *et al.* Calcium and Parkinson's disease. *Biochem. Biophys. Res. Commun.* **483**, 1013–1019 (2017).
89. Mosharov, E. V. *et al.* Interplay between cytosolic dopamine, calcium, and alpha-synuclein causes selective death of substantia nigra neurons. *Neuron* **62**, 218–229 (2009).
90. Lotharius, J. & Brundin, P. Pathogenesis of Parkinson's disease: dopamine, vesicles and alpha-synuclein. *Nat. Rev. Neurosci.* **3**, 932–942 (2002).
91. Dehay, B. *et al.* Pathogenic lysosomal depletion in Parkinson's disease. *J. Neurosci.* **30**, 12535–12544 (2010).
92. Moehle, M. S. & West, A. B. M1 and M2 immune activation in Parkinson's disease: foe and ally? *Neuroscience* **302**, 59–73 (2015).
93. Ransohoff, R. M. How neuroinflammation contributes to neurodegeneration. *Science* **353**, 777–783 (2016).
94. Hirsch, E. C. & Hunot, S. Neuroinflammation in Parkinson's disease: a target for neuroprotection? *Lancet Neurol.* **8**, 382–397 (2009).
95. Cebrian, C. *et al.* MHC-I expression renders catecholaminergic neurons susceptible to T-cell-mediated degeneration. *Nat. Commun.* **5**, 3633 (2014).
96. Schapansky, J., Nardozzi, J. D., Felizia, F. & LaVoie, M. J. Membrane recruitment of endogenous LRRK2 precedes its potent regulation of autophagy. *Hum. Mol. Genet.* **23**, 4201–4214 (2014).
97. Ma, B. *et al.* LRRK2 modulates microglial activity through regulation of chemokine (C-X3-C) receptor 1-mediated signalling pathways. *Hum. Mol. Genet.* **25**, 3515–3523 (2016).
98. Gao, H. M. *et al.* Neuroinflammation and oxidation/nitration of alpha-synuclein linked to dopaminergic neurodegeneration. *J. Neurosci.* **28**, 7687–7698 (2008).
99. Lema Tome, C. M. *et al.* Inflammation and alpha-synuclein's prion-like behavior in Parkinson's disease — is there a link? *Mol. Neurobiol.* **47**, 561–574 (2013).
100. Sampson, T. R. *et al.* Gut microbiota regulate motor deficits and neuroinflammation in a model of Parkinson's disease. *Cell* **167**, 1469–1480.e12 (2016).
101. George, S. & Brundin, P. Immunotherapy in Parkinson's disease: micromanaging alpha-synuclein aggregation. *J. Parkinsons Dis.* **5**, 413–424 (2015).
102. Alexander, G., Crutcher, M. D. & DeLong, M. R. Basal ganglia-thalamocortical circuits: parallel substrates for motor, oculomotor, "prefrontal" and "limbic" functions. *Prog. Brain Res.* **85**, 119–146 (1990).
103. Alexander, G. D., DeLong, M. R. & Strick, P. L. Parallel organization of functionally segregated circuits linking basal ganglia and cortex. *Annu. Rev. Neurosci.* **9**, 357–381 (1986).
104. Bergman, H., Wichmann, T. & DeLong, M. R. Reversal of experimental parkinsonism by lesions of the subthalamic nucleus. *Science* **249**, 1436–1438 (1990).
105. Laitinen, L. V., Bergenheim, A. T. & Hariz, M. I. Leksell's posteroventral pallidotomy in the treatment of Parkinson's disease. *J. Neurosurg.* **76**, 53–61 (1992).
106. Nambu, A. *et al.* Excitatory cortical inputs to pallidal neurons via the subthalamic nucleus in the monkey. *J. Neurophysiol.* **84**, 289–300 (2000).
107. Frank, M. J., Samanta, J., Moustafa, A. A. & Sherman, S. J. Hold your horses: impulsivity, deep brain stimulation, and medication in parkinsonism. *Science* **318**, 1309–1312 (2007).
108. Gradinaru, V., Mogri, M., Thompson, K. R., Henderson, J. M. & Deisseroth, K. Optical deconstruction of parkinsonian neural circuitry. *Science* **324**, 354–359 (2009).
109. Li, Q. *et al.* Therapeutic deep brain stimulation in parkinsonian rats directly influences motor cortex. *Neuron* **76**, 1030–1041 (2012).
110. Kuhn, A. A. *et al.* High-frequency stimulation of the subthalamic nucleus suppresses oscillatory beta activity in patients with Parkinson's disease in parallel with improvement in motor performance. *J. Neurosci.* **28**, 6165–6173 (2008).

111. Kuhn, A. A., Kupsch, A., Schneider, G. H. & Brown, P. Reduction in subthalamic 8–35 Hz oscillatory activity correlates with clinical improvement in Parkinson's disease. *Eur. J. Neurosci.* **23**, 1956–1960 (2006).
112. Chen, C. C. *et al.* Oscillatory pallidal local field potential activity correlates with involuntary EMG in dystonia. *Neurology* **66**, 418–420 (2006).
113. Dirkx, M. F. *et al.* The cerebral network of Parkinson's tremor: an effective connectivity fMRI study. *J. Neurosci.* **36**, 5362–5372 (2016).
114. Windels, F., Thevathasan, W., Silburn, P. & Sah, P. Where and what is the PPN and what is its role in locomotion? *Brain* **138**, 1135–1134 (2015).
115. Kalia, L. V. & Lang, A. E. Parkinson's disease. *Lancet* **386**, 896–912 (2015).
116. Tolosa, E., Wenning, G. & Poewe, W. The diagnosis of Parkinson's disease. *Lancet Neurol.* **5**, 75–86 (2006).
117. Gibb, W. R. & Lees, A. J. The relevance of the Lewy body to the pathogenesis of idiopathic Parkinson's disease. *J. Neurol. Neurosurg. Psychiatry* **51**, 745–752 (1988).
118. Postuma, R. B. *et al.* MDS clinical diagnostic criteria for Parkinson's disease. *Mov. Disord.* **30**, 1591–1601 (2015).
119. Alcalay, R. N. *et al.* Frequency of known mutations in early-onset Parkinson disease: implication for genetic counseling: the consortium on risk for early onset Parkinson disease study. *Arch. Neurol.* **67**, 1116–1122 (2010).
120. Marder, K. S. *et al.* Predictors of parkin mutations in early-onset Parkinson disease: the consortium on risk for early-onset Parkinson disease study. *Arch. Neurol.* **67**, 731–738 (2010).
121. Chaudhuri, K. R. & Schapira, A. H. Non-motor symptoms of Parkinson's disease: dopaminergic pathophysiology and treatment. *Lancet Neurol.* **8**, 464–474 (2009).
122. Hely, M. A., Reid, W. G., Adena, M. A., Halliday, G. M. & Morris, J. G. The Sydney multicenter study of Parkinson's disease: the inevitability of dementia at 20 years. *Mov. Disord.* **23**, 857–844 (2008).
123. Goetz, C. G. *et al.* Movement Disorder Society-sponsored revision of the Unified Parkinson's Disease Rating Scale (MDS-UPDRS): scale presentation and clinimetric testing results. *Mov. Disord.* **23**, 2129–2170 (2008).
124. Venuto, C. S., Potter, N. B., Ray Dorsey, E. & Kieburz, K. A review of disease progression models of Parkinson's disease and applications in clinical trials. *Mov. Disord.* **31**, 947–956 (2016).
125. Rizzo, G. *et al.* Accuracy of clinical diagnosis of Parkinson disease: a systematic review and meta-analysis. *Neurology* **86**, 566–576 (2016).
126. Garnett, E. S., Firnau, G. & Nahmias, C. Dopamine visualized in the basal ganglia of living man. *Nature* **305**, 137–138 (1983).
127. Stoessl, A. J., Lehericy, S. & Strafella, A. P. Imaging insights into basal ganglia function, Parkinson's disease, and dystonia. *Lancet* **384**, 532–544 (2014). **This paper provides a state-of-the-art review of the recent advances in neuroimaging of Parkinson disease.**
128. Politis, M. Neuroimaging in Parkinson disease: from research setting to clinical practice. *Nat. Rev. Neurol.* **10**, 708–722 (2014).
129. Mahlknecht, P. *et al.* Significance of MRI in diagnosis and differential diagnosis of Parkinson's disease. *Neurodegener. Dis.* **7**, 300–318 (2010).
130. Scherfler, C. *et al.* Diagnostic potential of automated subcortical volume segmentation in atypical parkinsonism. *Neurology* **86**, 1242–1249 (2016).
131. Tuite, P. Magnetic resonance imaging as a potential biomarker for Parkinson's disease. *Transl. Res.* **175**, 4–16 (2016).
132. Treglia, G. *et al.* MIBG scintigraphy in differential diagnosis of Parkinsonism: a meta-analysis. *Clin. Auton. Res.* **22**, 43–55 (2012).
133. Marras, C. *et al.* Nomenclature of genetic movement disorders: recommendations of the International Parkinson and Movement Disorder Society Task Force. *Mov. Disord.* **31**, 436–457 (2016).
134. Lill, C. M. Genetics of Parkinson's disease. *Mol. Cell. Probes* **30**, 386–396 (2016).
135. Brundin, P., Atkin, G. & Lamberts, J. T. Basic science breakthroughs: new therapeutic advances in Parkinson's disease. *Mov. Disord.* **30**, 1521–1527 (2015). **This review provides an outlook on emerging targets for future therapies of Parkinson disease.**
136. Chen-Plotkin, A. S. Unbiased approaches to biomarker discovery in neurodegenerative diseases. *Neuron* **84**, 594–607 (2014).
137. Swanson, C. R. *et al.* Plasma apolipoprotein A1 associates with age at onset and motor severity in early Parkinson's disease patients. *Mov. Disord.* **30**, 1648–1656 (2015).
138. Stern, M. B., Lang, A. & Poewe, W. Toward a redefinition of Parkinson's disease. *Mov. Disord.* **27**, 54–60 (2012).
139. Salat, D., Noyce, A. J., Schrag, A. & Tolosa, E. Challenges of modifying disease progression in prediagnostic Parkinson's disease. *Lancet Neurol.* **15**, 637–648 (2016).
140. Espay, A. J. *et al.* Technology in Parkinson's disease: challenges and opportunities. *Mov. Disord.* **31**, 1272–1282 (2016).
141. Pont-Sunyer, C. *et al.* The onset of nonmotor symptoms in Parkinson's disease (the ONSET PD study). *Mov. Disord.* **30**, 229–237 (2015).
142. Iraizoz, A., Santamaría, J. & Tolosa, E. Idiopathic rapid eye movement sleep behaviour disorder: diagnosis, management, and the need for neuroprotective interventions. *Lancet Neurol.* **15**, 405–419 (2016).
143. Jennings, D. *et al.* Imaging prodromal Parkinson disease: the Parkinson Associated Risk Syndrome Study. *Neurology* **83**, 1739–1746 (2014).
144. Mahlknecht, P. *et al.* Olfactory dysfunction predicts early transition to a Lewy body disease in idiopathic RBD. *Neurology* **84**, 654–658 (2015). **This paper provides important evidence on the effects of combining Parkinson disease risk markers to enhance predictivity for conversion in at-risk populations.**
145. Mahlknecht, P. *et al.* Prodromal Parkinson's disease as defined per MDS research criteria in the general elderly community. *Mov. Disord.* **31**, 1405–1408 (2016).
146. PD Med Collaborative Group *et al.* Long-term effectiveness of dopamine agonists and monoamine oxidase B inhibitors compared with levodopa as initial treatment for Parkinson's disease (PD MED): a large, open-label, pragmatic randomised trial. *Lancet* **384**, 1196–1205 (2014).
147. LeWitt, P. A. & Fahn, S. Levodopa therapy for Parkinson disease: a look backward and forward. *Neurology* **86**, S3–S12 (2016).
148. Olanow, C. W., Obeso, J. A. & Stocchi, F. Continuous dopamine-receptor treatment of Parkinson's disease: scientific rationale and clinical implications. *Lancet Neurol.* **5**, 677–687 (2006).
149. Cenci, M. A. Presynaptic mechanisms of L-DOPA-induced dyskinesia: the findings, the debate, and the therapeutic implications. *Front. Neurol.* **5**, 242 (2014).
150. Poewe, W. & Antonini, A. Novel formulations and modes of delivery of levodopa. *Mov. Disord.* **30**, 114–120 (2015).
151. Antonini, A. *et al.* Effect of levodopa–carbidopa intestinal gel on dyskinesia in advanced Parkinson's disease patients. *Mov. Disord.* **31**, 530–537 (2016).
152. Muller, T. Catechol-O-methyltransferase inhibitors in Parkinson's disease. *Drugs* **75**, 157–174 (2015).
153. Fox, S. H. *et al.* The Movement Disorder Society evidence-based medicine review update: treatments for the motor symptoms of Parkinson's disease. *Mov. Disord.* **26**, S2–S41 (2011).
154. Ferreira, J. J. *et al.* Opicapone as an adjunct to levodopa in patients with Parkinson's disease and end-of-dose motor fluctuations: a randomised, double-blind, controlled trial. *Lancet Neurol.* **15**, 154–165 (2016).
155. Schapira, A. H. Monoamine oxidase B inhibitors for the treatment of Parkinson's disease: a review of symptomatic and potential disease-modifying effects. *CNS Drugs* **25**, 1061–1071 (2011).
156. Birkmayer, W., Riederer, P., Ambrozi, L. & Youdim, M. B. Implications of combined treatment with 'Madopar' and L-doprenil in Parkinson's disease. A long-term study. *Lancet* **1**, 439–443 (1977).
157. Schapira, A. H. *et al.* Assessment of safety and efficacy of safinamide as a levodopa adjunct in patients with Parkinson disease and motor fluctuations: a randomized clinical trial. *JAMA Neurol.* **74**, 216–224 (2017).
158. Connolly, B. S. & Lang, A. E. Pharmacological treatment of Parkinson disease: a review. *JAMA* **311**, 1670–1683 (2014). **This paper provides a comprehensive review on the current options for the pharmacological management of Parkinson disease.**
159. Jankovic, J. & Poewe, W. Therapies in Parkinson's disease. *Curr. Opin. Neurol.* **25**, 433–447 (2012).
160. Voon, V., Mehta, A. R. & Hallett, M. Impulse control disorders in Parkinson's disease: recent advances. *Curr. Opin. Neurol.* **24**, 324–330 (2011).
161. Frankel, J. P., Lees, A. J., Kempster, P. A. & Stern, G. M. Subcutaneous apomorphine in the treatment of Parkinson's disease. *J. Neurol. Neurosurg. Psychiatry* **53**, 96–101 (1990).
162. Katzenschlager, R. *et al.* Continuous subcutaneous apomorphine therapy improves dyskinetics in Parkinson's disease: a prospective study using single-dose challenges. *Mov. Disord.* **20**, 151–157 (2005).
163. Hauser, R. A. *et al.* Sublingual apomorphine (APL-13027) for the acute conversion of OFF to ON in Parkinson's disease. *Mov. Disord.* **31**, 1366–1372 (2016).
164. Storch, A. *et al.* Nonmotor fluctuations in Parkinson disease: severity and correlation with motor complications. *Neurology* **80**, 800–809 (2013).
165. Lim, S. Y., Fox, S. H. & Lang, A. E. Overview of the extranigral aspects of Parkinson disease. *Arch. Neurol.* **66**, 167–172 (2009).
166. Kalia, L. V., Brotchie, J. M. & Fox, S. H. Novel nondopaminergic targets for motor features of Parkinson's disease: review of recent trials. *Mov. Disord.* **28**, 131–144 (2013).
167. Seppi, K. *et al.* The Movement Disorder Society evidence-based medicine review update: treatments for the non-motor symptoms of Parkinson's disease. *Mov. Disord.* **26**, S42–S80 (2011).
168. Connolly, B. & Fox, S. H. Treatment of cognitive, psychiatric, and affective disorders associated with Parkinson's disease. *Neurotherapeutics* **11**, 78–91 (2014).
169. Cummings, J. *et al.* Pimavanserin for patients with Parkinson's disease psychosis: a randomised, placebo-controlled phase 3 trial. *Lancet* **383**, 533–540 (2014).
170. Weintraub, D. & Burn, D. J. Parkinson's disease: the quintessential neuropsychiatric disorder. *Mov. Disord.* **26**, 1022–1031 (2011).
171. Perez-Lloret, S., Rey, M. V., Pavé-Le Traon, A. & Rascol, O. Emerging drugs for autonomic dysfunction in Parkinson's disease. *Expert Opin. Emerg. Drugs* **18**, 39–53 (2013).
172. Limousin, P. *et al.* Effect of parkinsonian signs and symptoms of bilateral subthalamic nucleus stimulation. *Lancet* **345**, 91–95 (1995).
173. Bronstein, J. M. *et al.* Deep brain stimulation for Parkinson disease: an expert consensus and review of key issues. *Arch. Neurol.* **68**, 165 (2011).
174. Deuschl, G. & Agid, Y. Subthalamic neurostimulation for Parkinson's disease with early fluctuations: balancing the risks and benefits. *Lancet Neurol.* **12**, 1025–1034 (2013). **This review summarizes the evidence base for the efficacy and safety of DBS in Parkinson disease.**
175. Odekerken, V. J. *et al.* Subthalamic nucleus versus globus pallidus bilateral deep brain stimulation for advanced Parkinson's disease (NSTAPS study): a randomised controlled trial. *Lancet Neurol.* **12**, 37–44 (2013).
176. Stern, M. B., Follett, K. A. & Weaver, F. M. Randomized trial of deep brain stimulation for Parkinson disease: thirty-six-month outcomes; turning tables: should GPi become the preferred DBS target for Parkinson disease? Author response. *Neurology* **80**, 225 (2013).
177. Odekerken, V. J. *et al.* GPi versus STN deep brain stimulation for Parkinson disease: three-year follow-up. *Neurology* **86**, 755–761 (2016).
178. Voges, J. *et al.* Thirty days complication rate following surgery performed for deep-brain-stimulation. *Mov. Disord.* **22**, 1486–1489 (2007).
179. Volkmann, J., Daniels, C. & Witt, K. Neuropsychiatric effects of subthalamic neurostimulation in Parkinson disease. *Nat. Rev. Neurol.* **6**, 487–498 (2010).
180. Kuhn, A. A. & Volkmann, J. Innovations in deep brain stimulation methodology. *Mov. Disord.* **32**, 11–19 (2017).
181. Keus, S. H., Munneke, M., Nijkraak, M. J., Kwakkel, G. & Bloem, B. R. Physical therapy in Parkinson's disease: evolution and future challenges. *Mov. Disord.* **24**, 1–14 (2009).
182. Bloem, B. R., de Vries, N. M. & Ebersbach, G. Nonpharmacological treatments for patients with Parkinson's disease. *Mov. Disord.* **30**, 1504–1520 (2015).
183. Ahlskog, J. E. Does vigorous exercise have a neuroprotective effect in Parkinson disease? *Neurology* **77**, 288–294 (2011).

184. Yang, F. *et al.* Physical activity and risk of Parkinson's disease in the Swedish National March Cohort. *Brain* **138**, 269–275 (2015).
185. Xu, Q. *et al.* Physical activities and future risk of Parkinson disease. *Neurology* **75**, 341–348 (2010).
186. Hinnell, C. *et al.* Nonmotor versus motor symptoms: how much do they matter to health status in Parkinson's disease? *Mov. Disord.* **27**, 236–241 (2012).
187. Schrag, A., Jahanshahi, M. & Quinn, N. What contributes to quality of life in patients with Parkinson's disease? *J. Neurol. Neurosurg. Psychiatry* **69**, 308–312 (2000).
188. Schrag, A., Sauerbier, A. & Chaudhuri, K. R. New clinical trials for nonmotor manifestations of Parkinson's disease. *Mov. Disord.* **30**, 1490–1504 (2015).
189. Lawson, R. A. *et al.* Cognitive decline and quality of life in incident Parkinson's disease: the role of attention. *Parkinsonism Relat. Disord.* **27**, 47–53 (2016).
190. Weerkamp, N. J. *et al.* Nonmotor symptoms in nursing home residents with Parkinson's disease: prevalence and effect on quality of life. *J. Am. Geriatr. Soc.* **61**, 1714–1721 (2013).
191. van der Mark, M. A. *et al.* Effectiveness of multidisciplinary care for Parkinson's disease: a randomized, controlled trial. *Mov. Disord.* **28**, 605–611 (2013).
192. Greenwell, K., Gray, W. K., van Wersch, A., van Schaik, P. & Walker, R. Predictors of the psychosocial impact of being a carer of people living with Parkinson's disease: a systematic review. *Parkinsonism Relat. Disord.* **21**, 1–11 (2015).
193. Schrag, A., Horvis, A., Morley, D., Quinn, N. & Jahanshahi, M. Caregiver burden in parkinson's disease is closely associated with psychiatric symptoms, falls, and disability. *Parkinsonism Relat. Disord.* **12**, 35–41 (2006).
194. Weisskopf, M. G., O'Reilly, E., Chen, H., Schwarzschild, M. A. & Ascherio, A. Plasma urate and risk of Parkinson's disease. *Am. J. Epidemiol.* **166**, 561–567 (2007).
195. Nalls, M. A. *et al.* Diagnosis of Parkinson's disease on the basis of clinical and genetic classification: a population-based modelling study. *Lancet Neurol.* **14**, 1002–1009 (2015).
196. Schepers, F. Can microbiota research change our understanding of neurodegenerative diseases? *Neurodegener. Dis. Manag.* **6**, 81–85 (2016).
197. Delenclos, M., Jones, D. R., McLean, P. J. & Uitti, R. J. Biomarkers in Parkinson's disease: advances and strategies. *Parkinsonism Relat. Disord.* **22**, S106–S110 (2016).
198. Gibbons, C. H., Garcia, J., Wang, N., Shih, L. C. & Freeman, R. The diagnostic discrimination of cutaneous alpha-synuclein deposition in Parkinson disease. *Neurology* **87**, 505–512 (2016).
199. Doppler, K. *et al.* Cutaneous neuropathy in Parkinson's disease: a window into brain pathology. *Acta Neuropathol.* **128**, 99–109 (2014).
200. Sprenger, F. S. *et al.* Enteric nervous system alpha-synuclein immunoreactivity in idiopathic REM sleep behavior disorder. *Neurology* **85**, 1761–1768 (2015).
201. Vilas, D. *et al.* Assessment of alpha-synuclein in submandibular glands of patients with idiopathic rapid-eye-movement sleep behaviour disorder: a case-control study. *Lancet Neurol.* **15**, 708–718 (2016).
202. Kordower, J. H. & Björklund, A. Trophic factor gene therapy for Parkinson's disease. *Mov. Disord.* **28**, 96–109 (2013).
203. Gill, S. S. *et al.* Direct brain infusion of glial cell line-derived neurotrophic factor in Parkinson disease. *Nat. Med.* **9**, 589–595 (2003).
204. Lang, A. E. *et al.* Randomized controlled trial of intraputamenal glial cell line-derived neurotrophic factor infusion in Parkinson disease. *Ann. Neurol.* **59**, 459–466 (2006).
205. Kordower, J. H. *et al.* Delivery of neurturin by AAV2 (CERE-120)-mediated gene transfer provides structural and functional neuroprotection and neurorestoration in MPTP-treated monkeys. *Ann. Neurol.* **60**, 706–715 (2006).
206. Bartus, R. T. & Johnson, E. M. Jr. Clinical tests of neurotrophic factors for human neurodegenerative diseases, part 1: where have we been and what have we learned? *Neurobiol. Dis.* **97**, 156–168 (2017).
207. Bartus, R. T. & Johnson, E. M. Jr. Clinical tests of neurotrophic factors for human neurodegenerative diseases, part 2: where do we stand and where must we go next? *Neurobiol. Dis.* **97**, 169–178 (2017).
208. Mittermeyer, G. *et al.* Long-term evaluation of a phase 1 study of AADC gene therapy for Parkinson's disease. *Hum. Gene Ther.* **23**, 377–381 (2012).
209. Palfi, S. *et al.* Long-term safety and tolerability of ProSavin, a lentiviral vector-based gene therapy for Parkinson's disease: a dose escalation, open-label, phase 1/2 trial. *Lancet* **383**, 1138–1146 (2014).
210. Björklund, A., Björklund, T. & Kirik, D. Gene therapy for dopamine replacement in Parkinson's disease. *Sci. Transl. Med.* **1**, 2ps2 (2009).
211. Carlsson, T. *et al.* Reversal of dyskinesias in an animal model of Parkinson's disease by continuous L-DOPA delivery using rAAV vectors. *Brain* **128**, 559–569 (2005).
212. LeWitt, P. A. *et al.* AAV2-GAD gene therapy for advanced Parkinson's disease: a double-blind, sham-surgery controlled, randomised trial. *Lancet Neurol.* **10**, 309–319 (2011).
213. Lindvall, O. *et al.* Grafts of fetal dopamine neurons survive and improve motor function in Parkinson's disease. *Science* **247**, 574–577 (1990).
214. Piccini, P. *et al.* Dopamine release from nigral transplants visualized *in vivo* in a Parkinson's patient. *Nat. Neurosci.* **2**, 1137–1140 (1999).
215. Freed, C. R. *et al.* Transplantation of embryonic dopamine neurons for severe Parkinson's disease. *N. Engl. J. Med.* **344**, 710–719 (2001).
216. Olanow, C. W. *et al.* A double-blind controlled trial of bilateral fetal nigral transplantation in Parkinson's disease. *Ann. Neurol.* **54**, 403–414 (2003).
217. Hagell, P. *et al.* Dyskinesias following neural transplantation in Parkinson's disease. *Nat. Neurosci.* **5**, 627–628 (2002).
218. Carta, M., Carlsson, T., Munoz, A., Kirik, D. & Björklund, A. Role of serotonin neurons in the induction of levodopa- and graft-induced dyskinesias in Parkinson's disease. *Mov. Disord.* **25**, S174–S179 (2010).
219. Lane, E. L., Vercammen, L., Cenci, M. A. & Brundin, P. Priming for L-DOPA-induced abnormal involuntary movements increases the severity of amphetamine-induced dyskinesia in grafted rats. *Exp. Neurol.* **219**, 355–358 (2009).
220. Brundin, P. & Kordower, J. H. Neuropathology in transplants in Parkinson's disease: implications for disease pathogenesis and the future of cell therapy. *Prog. Brain Res.* **200**, 221–241 (2012).
221. Kefalopoulou, Z. *et al.* Long-term clinical outcome of fetal cell transplantation for Parkinson disease: two case reports. *JAMA Neurol.* **71**, 83–87 (2014).
222. Barker, R. A., Drouin-Ouellet, J. & Parmar, M. Cell-based therapies for Parkinson disease—past insights and future potential. *Nat. Rev. Neurol.* **11**, 492–503 (2015).
- This paper provides a thoughtful and critical review on the limitations and the potential of cell-based therapies for Parkinson disease.**
223. Petit, G. H., Olsson, T. T. & Brundin, P. The future of cell therapies and brain repair: Parkinson's disease leads the way. *Neuropathol. Appl. Neurobiol.* **40**, 60–70 (2014).
224. Barker, R. A. *et al.* Are stem cell-based therapies for Parkinson's disease ready for the clinic in 2016? *J. Parkinsons Dis.* **6**, 57–63 (2016).
225. Kimmelman, J. *et al.* New ISSCR guidelines: clinical translation of stem cell research. *Lancet* **387**, 1979–1981 (2016).
226. Mandler, M. *et al.* Next-generation active immunotherapy for synucleinopathies: implications for Parkinson's disease clinical trials. *Acta Neuropathol.* **127**, 861–879 (2014).
227. McGuire, Kuhl, M. Foxbed blog: vaccine for Parkinson's reports positive results from boost study. *MichaelJFox.org* https://www.michaeljfox.org/foundation/news-detail.php?#vaccine-for-parkinson-reports-positive-results-from-boost-study&et_cid=663719&et_rid=81507667&et_lid=Read+More&mem_cid=12016.
228. Bergstrom, A. L., Kallunki, P. & Fog, K. Development of passive immunotherapies for synucleinopathies. *Mov. Disord.* **31**, 203–213 (2016).
229. Schenk, D. B. *et al.* First-in-human assessment of PRX002, an anti- α -synuclein monoclonal antibody, in healthy volunteers. *Mov. Disord.* **32**, 211–218 (2017).
230. Wrasidlo, W. M. *et al.* A de novo compound targeting α -synuclein improves deficits in models of Parkinson's disease. *Brain* **139**, 3217–3236 (2016).
231. Aviles-Olmos, I. *et al.* Motor and cognitive advantages persist 12 months after exenatide exposure in Parkinson's disease. *J. Parkinsons Dis.* **4**, 337–344 (2014).
232. Parkinson Study Group SURE-PD Investigators *et al.* Inosine to increase serum and cerebrospinal fluid urate in Parkinson disease: a randomized clinical trial. *JAMA Neurol.* **71**, 141–150 (2014).
233. Migdalas-Richards, A., Daly, L., Bezard, E. & Schapira, A. H. Ambroxol effects in glucocerebrosidase and alpha-synuclein transgenic mice. *Ann. Neurol.* **80**, 766–775 (2016).
234. Parkinson Study Group. Phase II safety, tolerability, and dose selection study of isradipine as a potential disease-modifying intervention in early Parkinson's disease (STEADY-PD). *Mov. Disord.* **28**, 1823–1831 (2013).
235. Hirsch, L., Jette, N., Frolikis, A., Steeves, T. & Pringsheim, T. The incidence of Parkinson's disease: a systematic review and meta-analysis. *Neuroepidemiology* **46**, 292–300 (2016).
236. Halliday, G. M. & McCann, H. The progression of pathology in Parkinson's disease. *Ann. NY Acad. Sci.* **1184**, 188–195 (2010).
237. Reiter, E. *et al.* Dorsolateral nigral hyperintensity on 3.0T susceptibility-weighted imaging in neurodegenerative Parkinsonism. *Mov. Disord.* **30**, 1068–1076 (2015).
238. Ohtsuka, C. *et al.* Changes in substantia nigra and locus caeruleus in patients with early-stage Parkinson's disease using neuromelanin-sensitive MR imaging. *Neurosci. Lett.* **541**, 93–98 (2013).
239. Schmidauer, C. *et al.* Transcranial ultrasound shows mild hypoechogenicity in restless legs syndrome. *Ann. Neurol.* **58**, 630–634 (2005).
240. Deng, H. X. *et al.* Identification of TMEM230 mutations in familial Parkinson's disease. *Nat. Genet.* **48**, 733–739 (2016).
241. Lill, C. M. *et al.* Launching the Movement Disorders Society Genetic Mutation Database (MDSGene). *Mov. Disord.* **31**, 607–609 (2016).

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Competing interests

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