Agar tray preparation

1. Up to 1 week in advance, wash all agar trays, lids, and 1L bottles. Do a final rinse with dI H2O 1 day in advance.
2. Prepare 1L bottles: 750 mL dI H2O and 7.5g Phytoagar (1%).
3. Autoclave flasks 30 minutes, liquid setting, 121°C.
4. If needed, place flasks in 55°C water bath until cool enough to hold with a gloved hand.
5. Perform the following steps in laminar flow hood:
   1. Spray trays with 70% ethanol to sterilize. Wipe with paper towel to partially dry.
   2. Pour 1.5L (2 bottles) of agar per tray.
   3. Using a sterile glass pipette or p1000 with sterile tips, remove all bubbles.
   4. Cool 2 hours uncovered in hood. Then cover with ethanol-sterilized lids and store in lab until inoculation.
      1. If rushed, can cool uncovered in hood until solidified, then add ethanol-sterilized lids and gently move to shelf or cart.
6. Store lidded trays in lab until inoculation (up to 1 day later).