

RNAhybrid_complete

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setup

dir

```
# home
setwd("D:/Krueger_Lab/Publications/miR181_paper/Figure2/RNAhybrid")

# work
#setwd("Z:/Personen/Nikita/Publications/miR181_paper/Figure2/RNAhybrid")
```

packages

```
#home
source("D:/Krueger_Lab/Publications/miR181_paper_v21022023/Figure_theme/theme_paper.R")
#work
#source("Z:/Personen/Nikita/Publications/miR181_paper_v21022023/Figure_theme/theme_paper.R")

library(BSgenome.Mmusculus.UCSC.mm10)

## Loading required package: BSgenome
## Loading required package: BiocGenerics
##
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:stats':
##
##     IQR, mad, sd, var, xtabs
## The following objects are masked from 'package:base':
##
##     anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##     colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
##     get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,
##     match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,
##     Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort,
##     table, tapply, union, unique, unsplit, which.max, which.min
## Loading required package: S4Vectors
## Loading required package: stats4
##
## Attaching package: 'S4Vectors'
```

```

## The following objects are masked from 'package:base':
##
##     expand.grid, I, unname
## Loading required package: IRanges
##
## Attaching package: 'IRanges'
## The following object is masked from 'package:grDevices':
##
##     windows
## Loading required package: GenomeInfoDb
## Loading required package: GenomicRanges
## Loading required package: Biostrings
## Loading required package: XVector
##
## Attaching package: 'Biostrings'
## The following object is masked from 'package:base':
##
##     strsplit
## Loading required package: rtracklayer
library(dplyr)

##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:Biostrings':
##
##     collapse, intersect, setdiff, setequal, union
## The following object is masked from 'package:XVector':
##
##     slice
## The following objects are masked from 'package:GenomicRanges':
##
##     intersect, setdiff, union
## The following object is masked from 'package:GenomeInfoDb':
##
##     intersect
## The following objects are masked from 'package:IRanges':
##
##     collapse, desc, intersect, setdiff, slice, union
## The following objects are masked from 'package:S4Vectors':
##
##     first, intersect, rename, setdiff, setequal, union
## The following objects are masked from 'package:BiocGenerics':
##
##     combine, intersect, setdiff, union

```

```

## The following objects are masked from 'package:stats':
##
##   filter, lag
## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union
library(ggplot2)
library(circlize)

## =====
## circlize version 0.4.15
## CRAN page: https://cran.r-project.org/package=circlize
## Github page: https://github.com/jokergoo/circlize
## Documentation: https://jokergoo.github.io/circlize\_book/book/
##
## If you use it in published research, please cite:
## Gu, Z. circlize implements and enhances circular visualization
##   in R. Bioinformatics 2014.
##
## This message can be suppressed by:
##   suppressPackageStartupMessages(library(circlize))
## =====
library(ComplexHeatmap)

## Loading required package: grid

##
## Attaching package: 'grid'

## The following object is masked from 'package:Biostrings':
##
##   pattern
## =====
## ComplexHeatmap version 2.15.2
## Bioconductor page: http://bioconductor.org/packages/ComplexHeatmap/
## Github page: https://github.com/jokergoo/ComplexHeatmap
## Documentation: http://jokergoo.github.io/ComplexHeatmap-reference
##
## If you use it in published research, please cite either one:
## - Gu, Z. Complex Heatmap Visualization. iMeta 2022.
## - Gu, Z. Complex heatmaps reveal patterns and correlations in multidimensional
##   genomic data. Bioinformatics 2016.
##
##
## The new InteractiveComplexHeatmap package can directly export static
## complex heatmaps into an interactive Shiny app with zero effort. Have a try!
##
## This message can be suppressed by:
##   suppressPackageStartupMessages(library(ComplexHeatmap))
## =====
library(seqinr)

##

```

```
## Attaching package: 'seqinr'

## The following object is masked from 'package:dplyr':
##
##      count

## The following object is masked from 'package:Biostrings':
##
##      translate

library(GenomicRanges)
library(stringr)
```

data

```
#home
f2bs <- readRDS("D:/Krueger_Lab/Publications/miR181_paper/Figure2/MRE_bound_gene_and_bound_region/mir181")
#work
#f2bs <- readRDS("Z:/Personen/Nikita/Publications/miR181_paper/Figure2/MRE_bound_gene_and_bound_region/")

head(f2bs)
```

```
##      seqnames      start      end width strand scoreSum scoreMean scoreMax
## 1      chr1 6245651 6245657      7      + 9.52553 4.762765 6.00678
## 2      chr1 6248341 6248347      7      + 92.68921 23.172303 48.76900
## 3      chr1 6248857 6248863      7      + 14.07133 7.035665 7.04425
## 4      chr1 6248918 6248924      7      + 38.91451 12.971503 20.65080
## 5      chr1 7170481 7170487      7      + 66.92218 13.384436 25.84490
## 6      chr1 9899605 9899611      7      + 25.15963 6.289907 8.61019
##      geneType geneName      geneID region BS_ID      mir_IP
## 1 protein_coding Rb1cc1 ENSMUSG00000025907 cds      5 mmu-miR-181a-5p
## 2 protein_coding Rb1cc1 ENSMUSG00000025907 cds      8 mmu-miR-181a-5p
## 3 protein_coding Rb1cc1 ENSMUSG00000025907 cds     10 mmu-miR-181a-5p
## 4 protein_coding Rb1cc1 ENSMUSG00000025907 cds     11 mmu-miR-181a-5p
## 5 protein_coding Pcmt1 ENSMUSG00000051285 utr3     19 mmu-miR-181a-5p
## 6 protein_coding Sgk3 ENSMUSG00000025915 utr3     23 mmu-miR-181a-5p
##      n_mir181 n_mir181a n_mir181b n_mir181c n_mir181d      set WT KO
## 1      1      1      0      0      0 ago_bs_mir181_chi 1 1
## 2      5      5      0      0      0 ago_bs_mir181_chi 1 1
## 3      6      6      0      0      0 ago_bs_mir181_chi 1 0
## 4      6      6      0      0      0 ago_bs_mir181_chi 1 1
## 5      4      4      0      0      0 ago_bs_mir181_chi 1 1
## 6      1      1      0      0      0 ago_bs_mir181_chi NA NA
##      geneID.2 geneName.1 region.1 counts.bs.1_KO counts.bs.2_KO
## 1 ENSMUSG00000025907 Rb1cc1 cds      4      3
## 2 ENSMUSG00000025907 Rb1cc1 cds     28     32
## 3 ENSMUSG00000025907 Rb1cc1 cds     13     11
## 4 ENSMUSG00000025907 Rb1cc1 cds     15     15
## 5 ENSMUSG00000051285 Pcmt1 utr3     12     22
## 6      <NA>      <NA>      <NA>      NA      NA
##      counts.bs.3_KO counts.bs.4_WT counts.bs.5_WT counts.bs.6_WT
## 1      3      3      10      3
## 2     27     46     41     20
## 3      4     22     13     12
## 4     10     33     20     18
```

```

## 5          14          16          20          9
## 6          NA          NA          NA          NA
##          geneID.1 counts.bg.1_K0 counts.bg.2_K0 counts.bg.3_K0
## 1 ENSMUSG00000025907          1609          1973          1250
## 2 ENSMUSG00000025907          1609          1973          1250
## 3 ENSMUSG00000025907          1609          1973          1250
## 4 ENSMUSG00000025907          1609          1973          1250
## 5 ENSMUSG00000051285          1355          1706          1064
## 6          <NA>          NA          NA          NA
## counts.bg.4_WT counts.bg.5_WT counts.bg.6_WT resBs.baseMean
## 1          2638          2231          1352          92.10645
## 2          2638          2231          1352          281.53271
## 3          2638          2231          1352          145.51107
## 4          2638          2231          1352          186.74162
## 5          1654          1348          755          151.36245
## 6          NA          NA          NA          NA
## resBs.log2FoldChange resBs.lfcSE resBs.stat resBs.pvalue resBs.padj
## 1          -0.1093039  0.5923673  0.03419066  0.8533018  0.9652601
## 2           0.2749428  0.2351157  1.35874137  0.2437557  0.6729889
## 3          -0.1805519  0.3623758  0.25017050  0.6169550  0.8961239
## 4          -0.2606282  0.3062717  0.73169661  0.3923338  0.7868678
## 5           0.1466485  0.3122905  0.22052922  0.6386370  0.9013566
## 6          NA          NA          NA          NA          NA
## resBg.baseMean resBg.log2FoldChange resBg.lfcSE resBg.stat resBg.pvalue
## 1          NA          NA          NA          NA          NA
## 2          NA          NA          NA          NA          NA
## 3          NA          NA          NA          NA          NA
## 4          NA          NA          NA          NA          NA
## 5          NA          NA          NA          NA          NA
## 6          NA          NA          NA          NA          NA
## resBg.padj tpm.counts.bg.1_K0 tpm.counts.bg.2_K0 tpm.counts.bg.3_K0
## 1          NA          133.7259          117.9980          129.8669
## 2          NA          133.7259          117.9980          129.8669
## 3          NA          133.7259          117.9980          129.8669
## 4          NA          133.7259          117.9980          129.8669
## 5          NA          248.6210          225.2505          244.0445
## 6          NA          NA          NA          NA
## tpm.counts.bg.4_WT tpm.counts.bg.5_WT tpm.counts.bg.6_WT
## 1          139.8635          146.2855          163.5360
## 2          139.8635          146.2855          163.5360
## 3          139.8635          146.2855          163.5360
## 4          139.8635          146.2855          163.5360
## 5          193.5994          195.1330          201.6149
## 6          NA          NA          NA
##          BS_ID.1 tpm_support_KO tpm_support_WT tpm_supported down
## 1 ENSMUSG00000025907.bs5          3          3          TRUE FALSE
## 2 ENSMUSG00000025907.bs8          3          3          TRUE FALSE
## 3 ENSMUSG00000025907.bs10          3          3          TRUE FALSE
## 4 ENSMUSG00000025907.bs11          3          3          TRUE FALSE
## 5 ENSMUSG00000051285.bs4          3          3          TRUE FALSE
## 6          <NA>          NA          NA          NA    NA

```

```

#colours
farbeneg <- "#b4b4b4"

```

```

farbe1 <- "#0073C2FF"
farbe2 <- "#EFC000FF"
farbe3 <- "#CD534CFF"
farbe4 <- "#7AA6DCFF"
farbe5 <- "#868686FF"
farbe6 <- "#003C67FF"
farbe7 <- "#8F7700FF"
farbe8 <- "#3B3B3BFF"
farbe9 <- "#A73030FF"
farbe10 <- "#4A6990FF"
farbe11 <- "#FF6F00FF"
farbe12 <- "#C71000FF"
farbe13 <- "#008EAOFF"
farbe14 <- "#8A4198FF"
farbe15 <- "#5A9599FF"
farbe16 <- "#FF6348FF"

RNApcol <- "#b56504"
RNAncol <- "#027d73"
RPFpcol <- "#c4c404"
RPFncol <- "#8d0391"

```

Get DNA sequences

#resize ranges

```

f2bsLA <- f2bs
f2bsLA$start <- f2bsLA$start -25
f2bsLA$end <- f2bsLA$end + 50
f2bsLA$n_mir181a <- as.numeric(f2bsLA$n_mir181a)

```

```

df181A <- mutate(f2bsLA, Sequence = as.character(getSeq(BSgenome.Mmusculus.UCSC.mm10, seqnames, start, end)))
df181A$rownum <- rownames(df181A)
df181A <- df181A[as.numeric(df181A$n_mir181a) > 0,]

```

#and turn T into Us

```
df181A$Sequence <- gsub('T', 'U', df181A$Sequence)
```

```
head(df181A)
```

```

##   seqnames   start     end width strand scoreSum scoreMean scoreMax
## 1   chr1 6245626 6245707     7    + 9.52553 4.762765 6.00678
## 2   chr1 6248316 6248397     7    + 92.68921 23.172303 48.76900
## 3   chr1 6248832 6248913     7    + 14.07133 7.035665 7.04425
## 4   chr1 6248893 6248974     7    + 38.91451 12.971503 20.65080
## 5   chr1 7170456 7170537     7    + 66.92218 13.384436 25.84490
## 6   chr1 9899580 9899661     7    + 25.15963 6.289907 8.61019
##      geneType geneName      geneID region BS_ID      mir_IP
## 1 protein_coding Rb1cc1 ENSMUSG00000025907  cds      5 mmu-miR-181a-5p
## 2 protein_coding Rb1cc1 ENSMUSG00000025907  cds      8 mmu-miR-181a-5p
## 3 protein_coding Rb1cc1 ENSMUSG00000025907  cds     10 mmu-miR-181a-5p
## 4 protein_coding Rb1cc1 ENSMUSG00000025907  cds     11 mmu-miR-181a-5p

```

```

## 5 protein_coding Pcmdt1 ENSMUSG00000051285 utr3 19 mmu-miR-181a-5p
## 6 protein_coding Sgk3 ENSMUSG00000025915 utr3 23 mmu-miR-181a-5p
## n_mir181 n_mir181a n_mir181b n_mir181c n_mir181d set WT KO
## 1 1 1 0 0 0 ago_bs_mir181_chi 1 1
## 2 5 5 0 0 0 ago_bs_mir181_chi 1 1
## 3 6 6 0 0 0 ago_bs_mir181_chi 1 0
## 4 6 6 0 0 0 ago_bs_mir181_chi 1 1
## 5 4 4 0 0 0 ago_bs_mir181_chi 1 1
## 6 1 1 0 0 0 ago_bs_mir181_chi NA NA
## geneID.2 geneName.1 region.1 counts.bs.1_KO counts.bs.2_KO
## 1 ENSMUSG00000025907 Rb1cc1 cds 4 3
## 2 ENSMUSG00000025907 Rb1cc1 cds 28 32
## 3 ENSMUSG00000025907 Rb1cc1 cds 13 11
## 4 ENSMUSG00000025907 Rb1cc1 cds 15 15
## 5 ENSMUSG00000051285 Pcmdt1 utr3 12 22
## 6 <NA> <NA> <NA> NA NA
## counts.bs.3_KO counts.bs.4_WT counts.bs.5_WT counts.bs.6_WT
## 1 3 3 10 3
## 2 27 46 41 20
## 3 4 22 13 12
## 4 10 33 20 18
## 5 14 16 20 9
## 6 NA NA NA NA
## geneID.1 counts.bg.1_KO counts.bg.2_KO counts.bg.3_KO
## 1 ENSMUSG00000025907 1609 1973 1250
## 2 ENSMUSG00000025907 1609 1973 1250
## 3 ENSMUSG00000025907 1609 1973 1250
## 4 ENSMUSG00000025907 1609 1973 1250
## 5 ENSMUSG00000051285 1355 1706 1064
## 6 <NA> NA NA NA
## counts.bg.4_WT counts.bg.5_WT counts.bg.6_WT resBs.baseMean
## 1 2638 2231 1352 92.10645
## 2 2638 2231 1352 281.53271
## 3 2638 2231 1352 145.51107
## 4 2638 2231 1352 186.74162
## 5 1654 1348 755 151.36245
## 6 NA NA NA NA
## resBs.log2FoldChange resBs.lfcSE resBs.stat resBs.pvalue resBs.padj
## 1 -0.1093039 0.5923673 0.03419066 0.8533018 0.9652601
## 2 0.2749428 0.2351157 1.35874137 0.2437557 0.6729889
## 3 -0.1805519 0.3623758 0.25017050 0.6169550 0.8961239
## 4 -0.2606282 0.3062717 0.73169661 0.3923338 0.7868678
## 5 0.1466485 0.3122905 0.22052922 0.6386370 0.9013566
## 6 NA NA NA NA NA
## resBg.baseMean resBg.log2FoldChange resBg.lfcSE resBg.stat resBg.pvalue
## 1 NA NA NA NA NA
## 2 NA NA NA NA NA
## 3 NA NA NA NA NA
## 4 NA NA NA NA NA
## 5 NA NA NA NA NA
## 6 NA NA NA NA NA
## resBg.padj tpm.counts.bg.1_KO tpm.counts.bg.2_KO tpm.counts.bg.3_KO
## 1 NA 133.7259 117.9980 129.8669
## 2 NA 133.7259 117.9980 129.8669

```

```
## 3      NA      133.7259      117.9980      129.8669
## 4      NA      133.7259      117.9980      129.8669
## 5      NA      248.6210      225.2505      244.0445
## 6      NA      NA      NA      NA
##      tpm.counts.bg.4_WT tpm.counts.bg.5_WT tpm.counts.bg.6_WT
## 1      139.8635      146.2855      163.5360
## 2      139.8635      146.2855      163.5360
## 3      139.8635      146.2855      163.5360
## 4      139.8635      146.2855      163.5360
## 5      193.5994      195.1330      201.6149
## 6      NA      NA      NA
##      BS_ID.1 tpm_support_KO tpm_support_WT tpm_supported down
## 1 ENSMUSG00000025907.bs5      3      3      TRUE FALSE
## 2 ENSMUSG00000025907.bs8      3      3      TRUE FALSE
## 3 ENSMUSG00000025907.bs10     3      3      TRUE FALSE
## 4 ENSMUSG00000025907.bs11     3      3      TRUE FALSE
## 5 ENSMUSG00000051285.bs4      3      3      TRUE FALSE
## 6      <NA>      NA      NA      NA      NA
##
##                                     Sequence
## 1 UAAAGGACUGGACUCCUGGCCUCCUCAUUUUUGUGUAUGUAUUUUUUUUUUUUAACUAGGACUAAAUUUCUUUUUUUUUUU
## 2 CAAGAAUAGAAAGUACAACAGGCAUUACAACCACUACCUCACCAAAAACUCCUCCUCCACUACUGUUCAGGACACCUUAUG
## 3 UACAAAAAGAACAGUGUGACUUAGCAAAUUAUUUAAAAUGUACAGCUGUAGAAAUAAGAAAUUUUUGAAAAAGUAAAAUG
## 4 UAUUUAUUGAAAAAGUAAAAUGUUCUCUAGAAAUAACACUAAAGGAAAAGCAUCAGCAAGAACUCCAUCUUUAAAAUUGAG
## 5 GGAAAAUUUCUGCUUCUCUCAUAGAGAUUUUUUAAAGAGCUAGUGAAUGUUAAAAGUAGGAAGUGGCUACUUGACACAACUAGUU
## 6 GAAGUGUAAUAAAAUGCUACCAGAUGUUUUUUUAAAGUGGUACCCACCAUAAUGUCUCUGUCACAUUUUAUUUACAAAUG
##      rownum
## 1      1
## 2      2
## 3      3
## 4      4
## 5      5
## 6      6
```

find seed

```
#find both seeds
seed1 <- df181A %>% filter(str_detect(Sequence, "GAAUGU"))
seed2 <- df181A %>% filter(str_detect(Sequence, "GAUUGU"))

#combine
seedm <- rbind(seed1, seed2)
#remove duplicates
seedm <- seedm[!duplicated(seedm$rownum),]
#remove NAs in gene name
seedm <- seedm[!is.na(seedm$geneName),]

head(seedm)
```

```
##      seqnames      start      end width strand scoreSum scoreMean scoreMax
## 5      chr1      7170456      7170537      7      + 66.92218 13.384436 25.8449
## 7      chr1      9899846      9899927      7      + 20.67430  6.891433 11.2987
## 22     chr1      43570279      43570360      7      + 55.09962 13.774905 21.7788
## 31     chr1      58754246      58754327      7      + 21.46580 10.732900 15.0257
```



```

## 51 chr1 85849941 85850022 7 + 56.80659 18.935530 31.2761
## 86 chr1 119528130 119528211 7 + 19.78365 9.891825 12.0423
## geneType geneName geneID region BS_ID mir_IP
## 5 protein_coding Pcmt1 ENSMUSG00000051285 utr3 19 mmu-miR-181a-5p
## 7 protein_coding Sgk3 ENSMUSG00000025915 utr3 24 mmu-miR-181a-5p
## 22 protein_coding Nck2 ENSMUSG00000066877 utr3 97 mmu-miR-181b-5p
## 31 protein_coding Cflar ENSMUSG00000026031 utr3 130 mmu-miR-181a-5p
## 51 protein_coding Cab39 ENSMUSG00000036707 utr3 209 mmu-miR-181a-5p
## 86 protein_coding Tmem185b ENSMUSG00000098923 utr3 320 mmu-miR-181a-5p
## n_mir181 n_mir181a n_mir181b n_mir181c n_mir181d set WT KO
## 5 4 4 0 0 0 ago_bs_mir181_chi 1 1
## 7 1 1 0 0 0 ago_bs_mir181_chi NA NA
## 22 2 1 1 0 0 ago_bs_mir181_chi 1 1
## 31 170 163 5 2 0 ago_bs_mir181_chi 0 1
## 51 326 297 29 0 0 ago_bs_mir181_chi 1 0
## 86 7 7 0 0 0 ago_bs_mir181_chi NA NA
## geneID.2 geneName.1 region.1 counts.bs.1_KO counts.bs.2_KO
## 5 ENSMUSG00000051285 Pcmt1 utr3 12 22
## 7 <NA> <NA> <NA> NA NA
## 22 ENSMUSG00000066877 Nck2 utr3 11 15
## 31 ENSMUSG00000026031 Cflar utr3 6 14
## 51 ENSMUSG00000036707 Cab39 utr3 2 2
## 86 <NA> <NA> <NA> NA NA
## counts.bs.3_KO counts.bs.4_WT counts.bs.5_WT counts.bs.6_WT
## 5 14 16 20 9
## 7 NA NA NA NA
## 22 10 20 21 12
## 31 11 24 23 10
## 51 3 85 50 32
## 86 NA NA NA NA
## geneID.1 counts.bg.1_KO counts.bg.2_KO counts.bg.3_KO
## 5 ENSMUSG00000051285 1355 1706 1064
## 7 <NA> NA NA NA
## 22 ENSMUSG00000066877 5026 7988 4386
## 31 ENSMUSG00000026031 1371 1785 1002
## 51 ENSMUSG00000036707 1606 2091 1226
## 86 <NA> NA NA NA
## counts.bg.4_WT counts.bg.5_WT counts.bg.6_WT resBs.baseMean
## 5 1654 1348 755 151.3625
## 7 NA NA NA NA
## 22 8478 6584 3537 300.8802
## 31 2308 1960 1040 148.2842
## 51 2802 2422 1406 234.7085
## 86 NA NA NA NA
## resBs.log2FoldChange resBs.lfcSE resBs.stat resBs.pvalue resBs.padj
## 5 0.1466485 0.3122905 0.2205292 6.386370e-01 9.013566e-01
## 7 NA NA NA NA NA
## 22 -0.2719827 0.3328411 0.6723828 4.122221e-01 8.000076e-01
## 31 -0.3288789 0.3315817 1.0018062 3.168738e-01 7.352453e-01
## 51 -3.9344782 0.5635927 107.5785787 3.324592e-25 1.174121e-21
## 86 NA NA NA NA NA
## resBg.baseMean resBg.log2FoldChange resBg.lfcSE resBg.stat resBg.pvalue
## 5 NA NA NA NA NA
## 7 NA NA NA NA NA

```

```

## 22          NA          NA          NA          NA          NA
## 31          NA          NA          NA          NA          NA
## 51          NA          NA          NA          NA          NA
## 86          NA          NA          NA          NA          NA
##      resBg.padj tpm.counts.bg.1_K0 tpm.counts.bg.2_K0 tpm.counts.bg.3_K0
## 5          NA          248.6210          225.2505          244.0445
## 7          NA          NA          NA          NA
## 22          NA          1377.3876          1575.2874          1502.5578
## 31          NA          132.1224          123.7842          120.7079
## 51          NA          277.0625          259.5818          264.3940
## 86          NA          NA          NA          NA
##      tpm.counts.bg.4_WT tpm.counts.bg.5_WT tpm.counts.bg.6_WT
## 5          193.5994          195.1330          201.6149
## 7          NA          NA          NA
## 22          1482.1666          1423.5269          1410.7365
## 31          141.8877          149.0175          145.8645
## 51          308.3687          329.6465          353.0162
## 86          NA          NA          NA
##      BS_ID.1 tpm_support_K0 tpm_support_WT tpm_supported down
## 5  ENSMUSG000000051285.bs4          3          3          TRUE FALSE
## 7          <NA>          NA          NA          NA    NA
## 22 ENSMUSG000000066877.bs31          3          3          TRUE FALSE
## 31 ENSMUSG000000026031.bs4          3          3          TRUE FALSE
## 51 ENSMUSG000000036707.bs4          3          3          TRUE  TRUE
## 86          <NA>          NA          NA          NA    NA
##
##                                     Sequence
## 5  GGAAAAUUUCUGCUUCUCUCAUAGAGAUUUUUAAGAGCUAGUGAAUGUUAAGUAGGAAGUGGCUACUUGACACAACUAGUU
## 7  GGCAAGUCUGGGUUGGUGUGAAUGUGUGUCACCUACACAUUCUAAACAGAAGGUAACAUAAGUUAGCAGUGACAUUUCAGU
## 22 AUUAUUAUUUUGCUUUACAGGGAUUUUUUCAGGGUUUACAAAAGAAUAUGUGAUUAGUAGUAACAGAAUGUUUAUGAAGAA
## 31 UGGGUGUAUAGUGUAUAGUGGUUCAAGAUUUGACACUGAAUGUAACUUGAGACUUACCUGAGUUUGUCAUGCGACUGGGUAA
## 51 UGUUAUUAUUUCUUAAGAAUGCUCAUUUCUUUUAUUCGUUUAAUUGUACAGCAGAGGAAUGUUUUGUAGUAGUAUGUAAC
## 86 UGCAUAUAUUAGUAUUUAUAUGAAUGUUUAGCAGUGUUAUCUGUGUUGAUUGUAGUUCUUGGCAGUAAUGUAUUGUGUAA
##      rownum
## 5          5
## 7          7
## 22         22
## 31         31
## 51         51
## 86         86

```

Write to .fasta

this is deactivated for now because we only need it once right now just remove the eval if needed

```

candgeneNameA <- as.list(seedm$geneName)
candnameA <- as.list(seedm$rownum)
condgeneSeqA <- as.list(seedm$Sequence)

#change to output directory

setwd("D:/Krueger_Lab/Publications/miR181_paper_nongithub/Figure2/RNAhybrid/fastafiles_complete/A")

for (i in 1:length(candgeneNameA)) {

```

```

write.fasta(condgeneSeqA[i], candnameA[i], paste(candnameA[i], candgeneNameA[i], "miR_181a", 'fasta',
})

Personalized_Reader <- function(lambda){
  read.table(lambda, sep = ":") %>% select(V1, V5, V6, V7, V10, V11)}

#remove NA file...I just dont get it....where is it coming from?

#File lists
reslistA <- list.files(path = "D:/Krueger_Lab/Publications/miR181_paper_nongithub/Figure2/RNAhybrid/res")

#import
myfilelistA <- lapply(reslistA, Personalized_Reader)

resframeA <- bind_rows(myfilelistA)

#colnames
colnames(resframeA) <- c("rownumber", "mfs", "pvalue", "start_position", "binding_bases", "non_binding_bases")

resframeA[is.na(resframeA$non_binding_bases), "non_binding_bases"] <- " "

head(resframeA)

```

```

##   rownumber   mfs   pvalue start_position binding_bases
## 1      1004 -24.7 0.024036          66      UGGCUGUC ACUUACA
## 2      1005 -25.4 0.015632          46      UGGCUGUC ACUUACA
## 3      1035 -11.0 1.000000           1          CUUACA
## 4      1043 -18.4 0.703212          46      UG C   UCG AA   CUUACA
## 5      1050 -14.6 0.999997          60      GCUG CG AAC UACAA
## 6      1054 -17.8 0.828455          25      GUGG UGUCG AACU  ACA
##               non_binding_bases
## 1      UGAG      GCA      A
## 2      UGAG      GCA      A
## 3      UGAGUGGCUGUCGCAA      A
## 4      UGAG  G UG  C      A
## 5      UGAGUG      U  C  U
## 6      UGA      C      C  U  A

```

merge with original df

```

# make seperate objects for each mature mirna just to see if they are much different

seedm$rownumber <- as.character(seedm$rownum)
resframeA$rownumber <- as.character(resframeA$rownumber)

```

```
bsseqHA <- left_join(seedm, resframeA, by="rownumber")
```

```
head(bsseqHA)
```

```
##      seqnames      start      end width strand scoreSum scoreMean scoreMax
## 1      chr1    7170456    7170537     7      + 66.92218 13.384436 25.8449
## 2      chr1    9899846    9899927     7      + 20.67430  6.891433 11.2987
## 3      chr1   43570279   43570360     7      + 55.09962 13.774905 21.7788
## 4      chr1   58754246   58754327     7      + 21.46580 10.732900 15.0257
## 5      chr1   85849941   85850022     7      + 56.80659 18.935530 31.2761
## 6      chr1 119528130 119528211     7      + 19.78365  9.891825 12.0423
##      geneType geneName      geneID region BS_ID      mir_IP
## 1 protein_coding Pcmt1d1 ENSMUSG000000051285 utr3    19 mmu-miR-181a-5p
## 2 protein_coding Sgk3 ENSMUSG000000025915 utr3    24 mmu-miR-181a-5p
## 3 protein_coding Nck2 ENSMUSG000000066877 utr3    97 mmu-miR-181b-5p
## 4 protein_coding Cflar ENSMUSG000000026031 utr3   130 mmu-miR-181a-5p
## 5 protein_coding Cab39 ENSMUSG000000036707 utr3   209 mmu-miR-181a-5p
## 6 protein_coding Tmem185b ENSMUSG000000098923 utr3   320 mmu-miR-181a-5p
##      n_mir181 n_mir181a n_mir181b n_mir181c n_mir181d      set WT KO
## 1          4          4          0          0          0 ago_bs_mir181_chi 1 1
## 2          1          1          0          0          0 ago_bs_mir181_chi NA NA
## 3          2          1          1          0          0 ago_bs_mir181_chi 1 1
## 4         170         163          5          2          0 ago_bs_mir181_chi 0 1
## 5         326         297         29          0          0 ago_bs_mir181_chi 1 0
## 6          7          7          0          0          0 ago_bs_mir181_chi NA NA
##      geneID.2 geneName.1 region.1 counts.bs.1_KO counts.bs.2_KO
## 1 ENSMUSG000000051285 Pcmt1d1 utr3          12          22
## 2          <NA>          <NA>          <NA>          NA          NA
## 3 ENSMUSG000000066877 Nck2 utr3          11          15
## 4 ENSMUSG000000026031 Cflar utr3           6          14
## 5 ENSMUSG000000036707 Cab39 utr3           2           2
## 6          <NA>          <NA>          <NA>          NA          NA
##      counts.bs.3_KO counts.bs.4_WT counts.bs.5_WT counts.bs.6_WT
## 1          14          16          20           9
## 2          NA          NA          NA          NA
## 3          10          20          21          12
## 4          11          24          23          10
## 5           3          85          50          32
## 6          NA          NA          NA          NA
##      geneID.1 counts.bg.1_KO counts.bg.2_KO counts.bg.3_KO
## 1 ENSMUSG000000051285      1355      1706      1064
## 2          <NA>          NA          NA          NA
## 3 ENSMUSG000000066877      5026      7988      4386
## 4 ENSMUSG000000026031      1371      1785      1002
## 5 ENSMUSG000000036707      1606      2091      1226
## 6          <NA>          NA          NA          NA
##      counts.bg.4_WT counts.bg.5_WT counts.bg.6_WT resBs.baseMean
## 1          1654          1348           755       151.3625
## 2          NA          NA          NA          NA
## 3          8478          6584          3537       300.8802
## 4          2308          1960          1040       148.2842
## 5          2802          2422          1406       234.7085
## 6          NA          NA          NA          NA
##      resBs.log2FoldChange resBs.lfcSE resBs.stat resBs.pvalue resBs.padj
```

```

## 1      0.1466485    0.3122905    0.2205292 6.386370e-01 9.013566e-01
## 2      NA          NA          NA          NA          NA
## 3     -0.2719827    0.3328411    0.6723828 4.122221e-01 8.000076e-01
## 4     -0.3288789    0.3315817    1.0018062 3.168738e-01 7.352453e-01
## 5     -3.9344782    0.5635927 107.5785787 3.324592e-25 1.174121e-21
## 6      NA          NA          NA          NA          NA
##      resBg.baseMean resBg.log2FoldChange resBg.lfcSE resBg.stat resBg.pvalue
## 1      NA          NA          NA          NA          NA
## 2      NA          NA          NA          NA          NA
## 3      NA          NA          NA          NA          NA
## 4      NA          NA          NA          NA          NA
## 5      NA          NA          NA          NA          NA
## 6      NA          NA          NA          NA          NA
##      resBg.padj tpm.counts.bg.1_KO tpm.counts.bg.2_KO tpm.counts.bg.3_KO
## 1      NA          248.6210          225.2505          244.0445
## 2      NA          NA          NA          NA
## 3      NA          1377.3876          1575.2874          1502.5578
## 4      NA          132.1224          123.7842          120.7079
## 5      NA          277.0625          259.5818          264.3940
## 6      NA          NA          NA          NA
##      tpm.counts.bg.4_WT tpm.counts.bg.5_WT tpm.counts.bg.6_WT
## 1      193.5994          195.1330          201.6149
## 2      NA          NA          NA
## 3      1482.1666          1423.5269          1410.7365
## 4      141.8877          149.0175          145.8645
## 5      308.3687          329.6465          353.0162
## 6      NA          NA          NA
##      BS_ID.1 tpm_support_KO tpm_support_WT tpm_supported down
## 1 ENSMUSG00000051285.bs4      3      3      TRUE FALSE
## 2      <NA>      NA      NA      NA      NA
## 3 ENSMUSG00000066877.bs31      3      3      TRUE FALSE
## 4 ENSMUSG00000026031.bs4      3      3      TRUE FALSE
## 5 ENSMUSG00000036707.bs4      3      3      TRUE  TRUE
## 6      <NA>      NA      NA      NA      NA
##
##                                     Sequence
## 1 GGAAAAUUUCUGCUUCUCUCAUAGAGAUUUUUAAGAGCUAGUGAAUGUUAAGUAGGAAGUGGCUACUUGACACAACUAGUU
## 2 GGCAAGUCUGGGUUGGUGUGAAUGUGUGUACCUACACAUCUAAACAGAAGGUAACAAUAAGUUAGCAGUGACAUAUUCAGU
## 3 AUUAUUAUUUGCUUUACAGGGAAAAUUUUCAGGGUUUACAAAAGAAUAUGUGAUUAGUAGUAAACAGAAUGUUUAUGAAGAA
## 4 UGGGUGUAUAGUGUAUAGUGGUUCAAGAUUUGACACUGAAUGUAACUUGAGACUUAACUGAGUUUGUCAUGCGACUGGGUAA
## 5 UGUUAUAAAUUCUUAAGAAUGCUCAUUUCUUUAAAUCGUUUAAUUUGUACAGCAGAGGAAUGUUAAUUGUAGUAGUAUGUAAAC
## 6 UGCAUAUAUUAUUAUUAUGAAUGUUUAGCAGUGUUAUCUGUGUUGAUUGUAGUUCUUGGCAGUAAUGUAUUGUGUUAA
##
##      rownum rownumber   mfs   pvalue start_position
## 1      5         5 -23.3 0.056025          17
## 2      7         7 -18.8 0.612351           7
## 3     22        22 -13.2 1.000000          13
## 4     31        31 -18.3 0.725425          21
## 5     51        51 -21.9 0.129199          19
## 6     86        86 -18.4 0.703212          56
##
##                                     binding_bases
## 1      GAGUG   CUG      UCG  ACUUACAA
## 2      GAGUG      GGCU  GUCGC ACUUACA
## 3      GAGUG   CU      GUC CAA      CU  UACA
## 4      GAGUG      GAGU  GGCUGU G  ACUUACA
## 5      GAGUG      GGC      UGUCG C  CUUACAA

```

```
## 6          GAG GGCUGUCG AC  ACA
##          non_binding_bases
## 1          U      G          CA
## 2          UGAGU          A      A
## 3          U      G          G          A
## 4          U          U          C CA      A
## 5          AA
## 6          U      U          CA UU      A
```

-continue here

Process data (remove gaps)

Due to the loops in the mRNA there are additional spaces in the mirna. We only want the binding and non binding bases of the mirna in the correct order. For that we will remove all gaps that origin in the mRNA loops.

```
#binding and non binding bases as characters in a list
Alistbb <- strsplit(resframeA$binding_bases,"")
Alistnb <- strsplit(resframeA$non_binding_bases,"")

#combine the two lists
Alist <- Map(cbind, Alistbb, Alistnb)

## Warning in cbind(...): number of rows of result is not a multiple of vector
## length (arg 2)

Alist <- lapply(Alist, as.data.frame)

#remove all empty rows (mRNA loops)
Alist0 <- lapply(Alist, function(x){
  x[!(x[,1]== " " & x[,2] == " "),]
})

#rewrite as characters
AlistF <- lapply(Alist0, function(x){
  paste(x[,1], collapse = '')
})

#Attach lists back onto original data.frame as new column
resframeA$binding_nospace <-unlist(AlistF)
head(resframeA$binding_nospace)
```

```
## [1] "      UGGCUGUC  ACUUACA " "      UGGCUGUC  ACUUACA "
## [3] "              CUUACA " "      UG C  UCG AACUUACA "
## [5] "      GCUG CG AAC UACAA" "      GUGG UGUCG AACU ACA "
```

Transform into Numbers

add 0s

replace all gaps with 0 and all letters with 1

```
#0
resframeA$binding_nospace <- chartr(" ", "0", resframeA$binding_nospace)

#1
resframeA$binding_nospace <- mgsub::mgsub(resframeA$binding_nospace, c("A", "U", "C", "G"), c(rep("1", 4), rep("0", 16)))

head(resframeA)
```

```
##   rownumber   mfs   pvalue start_position      binding_bases
## 1      1004 -24.7 0.024036           66      UGGCUGUC ACUUACA
## 2      1005 -25.4 0.015632           46      UGGCUGUC ACUUACA
## 3      1035 -11.0 1.000000            1      CUUACA
## 4      1043 -18.4 0.703212           46      UG C  UCG AA  CUUACA
## 5      1050 -14.6 0.999997           60      GCUG CG AAC UACAA
## 6      1054 -17.8 0.828455           25      GUGG UGUCG AACU  ACA
##           non_binding_bases      binding_nospace
## 1      UGAG      GCA      A 0000111111110001111110
## 2      UGAG      GCA      A 00001111111110001111110
## 3      UGAGUGGCUGUCGCAA      A 0000000000000000001111110
## 4  UGAG  G  UG  C      A 000011010011101111111110
## 5      UGAGUG  U  C  U      00000011110110111011111
## 6      UGA  C  C  U      A 000111101111101111101110
```

seperate into columns

for each base make 1 column so it can be added and also put into a heatmap

```
#for the heatmap with every binding site
heatframeA <- do.call(rbind.data.frame, strsplit(resframeA$binding_nospace, ""))
heatframeA <- sapply( heatframeA, as.numeric )
colnames(heatframeA) <- c(23:1)
rownames(heatframeA) <- resframeA[,1]
head(heatframeA)
```

```
##      23 22 21 20 19 18 17 16 15 14 13 12 11 10 9 8 7 6 5 4 3 2 1
## 1004  0  0  0  0  1  1  1  1  1  1  1  1  0  0  0  1  1  1  1  1  1  0
## 1005  0  0  0  0  1  1  1  1  1  1  1  1  0  0  0  1  1  1  1  1  1  0
## 1035  0  0  0  0  0  0  0  0  0  0  0  0  0  0  0  0  1  1  1  1  1  0
## 1043  0  0  0  0  1  1  0  1  0  0  1  1  1  0  1  1  1  1  1  1  1  0
## 1050  0  0  0  0  0  0  1  1  1  1  0  1  1  0  1  1  1  0  1  1  1  1
## 1054  0  0  0  1  1  1  1  0  1  1  1  1  1  0  1  1  1  1  0  1  1  0
```

```
#reverse column order
heatframeA <-heatframeA[,23:1]
```

Heatmap

Colours

```
hmcols1 <- c("white", "black")
hmcols2 <- colorRamp2(c(-2, 2), c("white", "red"))
```

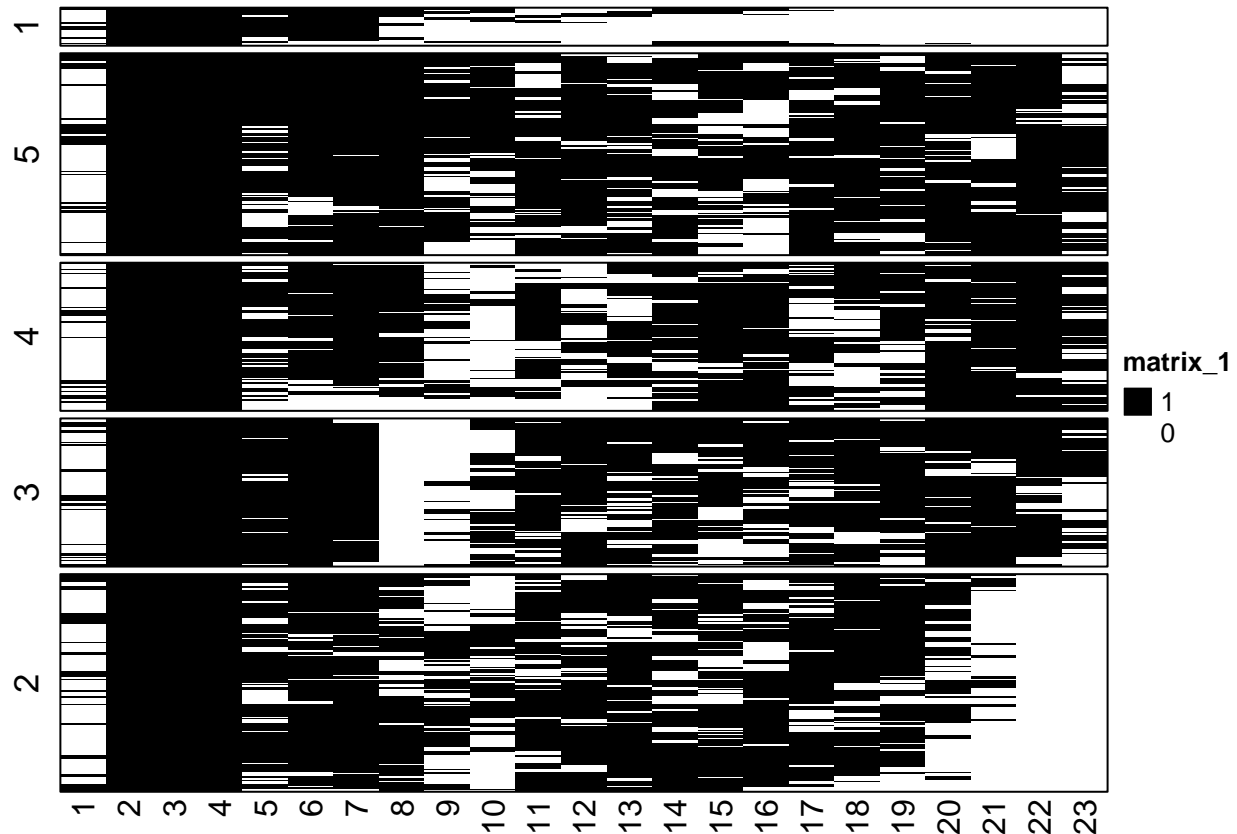
Heatmap of all the single reads

make heatmap without column clustering but with row clustering

```
set.seed(123)
```

```
HMA <- Heatmap(heatframeA, cluster_columns = F, col = hmcols1, row_km = 5, show_row_names = F, show_row
```

```
HMA
```



session info

```
sessionInfo()
```

```
## R version 4.2.3 (2023-03-15 ucrt)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 19045)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=German_Germany.utf8  LC_CTYPE=German_Germany.utf8
## [3] LC_MONETARY=German_Germany.utf8 LC_NUMERIC=C
## [5] LC_TIME=German_Germany.utf8
##
## attached base packages:
## [1] grid      stats4    stats      graphics  grDevices  utils      datasets
## [8] methods  base
##
```



```

## other attached packages:
## [1] stringr_1.5.0                      seqinr_4.2-30
## [3] ComplexHeatmap_2.15.2              circlize_0.4.15
## [5] ggplot2_3.4.2                      dplyr_1.1.1
## [7] BSgenome.Mmusculus.UCSC.mm10_1.4.3 BSgenome_1.66.3
## [9] rtracklayer_1.58.0                 Biostrings_2.66.0
## [11] XVector_0.38.0                     GenomicRanges_1.50.2
## [13] GenomeInfoDb_1.34.9                IRanges_2.32.0
## [15] S4Vectors_0.36.2                   BiocGenerics_0.44.0
##
## loaded via a namespace (and not attached):
## [1] MatrixGenerics_1.10.0              Biobase_2.58.0
## [3] foreach_1.5.2                      highr_0.10
## [5] GenomeInfoDbData_1.2.9             Rsamtools_2.14.0
## [7] yaml_2.3.7                         pillar_1.9.0
## [9] lattice_0.20-45                    glue_1.6.2
## [11] digest_0.6.31                      RColorBrewer_1.1-3
## [13] colorspace_2.1-0                   htmltools_0.5.4
## [15] Matrix_1.5-3                       XML_3.99-0.14
## [17] pkgconfig_2.0.3                    GetoptLong_1.0.5
## [19] magick_2.7.4                       zlibbioc_1.44.0
## [21] scales_1.2.1                       BiocParallel_1.32.6
## [23] tibble_3.2.1                       generics_0.1.3
## [25] withr_2.5.0                        SummarizedExperiment_1.28.0
## [27] cli_3.6.0                          magrittr_2.0.3
## [29] crayon_1.5.2                       evaluate_0.20
## [31] fansi_1.0.4                        doParallel_1.0.17
## [33] MASS_7.3-58.2                      Cairo_1.6-0
## [35] tools_4.2.3                        GlobalOptions_0.1.2
## [37] BiocIO_1.8.0                       lifecycle_1.0.3
## [39] matrixStats_0.63.0                 mgsub_1.7.3
## [41] munsell_0.5.0                      cluster_2.1.4
## [43] DelayedArray_0.23.2                ade4_1.7-22
## [45] compiler_4.2.3                     rlang_1.1.0
## [47] RCurl_1.98-1.12                    iterators_1.0.14
## [49] rstudioapi_0.14                    rjson_0.2.21
## [51] bitops_1.0-7                       rmarkdown_2.21
## [53] restfulr_0.0.15                    gtable_0.3.3
## [55] codetools_0.2-19                   R6_2.5.1
## [57] GenomicAlignments_1.34.1           knitr_1.42
## [59] fastmap_1.1.1                      utf8_1.2.3
## [61] clue_0.3-64                        shape_1.4.6
## [63] stringi_1.7.12                     parallel_4.2.3
## [65] Rcpp_1.0.10                        vctrs_0.6.1
## [67] png_0.1-8                          tidyrselect_1.2.0
## [69] xfun_0.37

```