

Gviz_fig1

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setup

directory

```
setwd("D:/Krueger_Lab/Publications/miR181_paper/Figure1/Genome_tracks")
```

packages

```
library(Gviz)
```

```
## Loading required package: S4Vectors
```

```
## Loading required package: stats4
```

```
## Loading required package: BiocGenerics
```

```
##
```

```
## Attaching package: 'BiocGenerics'
```

```
## The following objects are masked from 'package:stats':
```

```
##
```

```
##      IQR, mad, sd, var, xtabs
```

```
## The following objects are masked from 'package:base':
```

```
##
```

```
##      anyDuplicated, aperm, append, as.data.frame, basename, cbind,  
##      colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,  
##      get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,  
##      match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,  
##      Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort,  
##      table, tapply, union, unique, unsplit, which.max, which.min
```

```
##
```

```
## Attaching package: 'S4Vectors'
```

```
## The following objects are masked from 'package:base':
```

```
##
```

```
##      expand.grid, I, unname
```

```
## Loading required package: IRanges
```

```
##
```

```
## Attaching package: 'IRanges'
```

```
## The following object is masked from 'package:grDevices':
```

```
##
```

```
##      windows
```

```

## Loading required package: GenomicRanges
## Loading required package: GenomeInfoDb
## Loading required package: grid
library(GenomicRanges)
library(rtracklayer)
library(dplyr)

##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:GenomicRanges':
##
##   intersect, setdiff, union
## The following object is masked from 'package:GenomeInfoDb':
##
##   intersect
## The following objects are masked from 'package:IRanges':
##
##   collapse, desc, intersect, setdiff, slice, union
## The following objects are masked from 'package:S4Vectors':
##
##   first, intersect, rename, setdiff, setequal, union
## The following objects are masked from 'package:BiocGenerics':
##
##   combine, intersect, setdiff, union
## The following objects are masked from 'package:stats':
##
##   filter, lag
## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union
library(plyranges)

##
## Attaching package: 'plyranges'
## The following objects are masked from 'package:dplyr':
##
##   between, n, n_distinct
## The following object is masked from 'package:IRanges':
##
##   slice
## The following object is masked from 'package:stats':
##
##   filter

```

Import data

```
# diferencial results
diffres <- rtracklayer::import(con = "D:/Krueger_Lab/miReCLIP/Mirco/DifferentialBinding/BsDifferentialR

# non enriched non chimeric
IPK0minus <- import.bw("D:/Krueger_Lab/miReCLIP/Melina/pipe_output_22_02_14/pipe_output_22_02_14/non-ch
IPK0plus <- import.bw("D:/Krueger_Lab/miReCLIP/Melina/pipe_output_22_02_14/pipe_output_22_02_14/non-chi
IPWTminus <- import.bw("D:/Krueger_Lab/miReCLIP/Melina/pipe_output_22_02_14/pipe_output_22_02_14/non-ch
IPWTplus <- import.bw("D:/Krueger_Lab/miReCLIP/Melina/pipe_output_22_02_14/pipe_output_22_02_14/non-chi

# enriched chimeric
IP181chimWTminus <- import.bw("D:/Krueger_Lab/miReCLIP/Melina/pipe_output_22_02_14/pipe_output_22_02_14/
IP181chimWTplus <- import.bw("D:/Krueger_Lab/miReCLIP/Melina/pipe_output_22_02_14/pipe_output_22_02_14/
```

setting regions

Use this panel to change the coordinates in all following plots

```
scoord = 20405350
ecoord = 20406237
tchr = "chr13"
clim = 10
```

Zfp36l1: chr12 80107716-80108070 Zfp36l2: chr17 84184290-84184345 Cblb: chr16:52171980-52172133 elmo1:
chr13:20405350-20406237

Tracks

Annotation track

```
gatrack <- GenomeAxisTrack()
```

pureclip (merged)

Always just activate or inactivate the strand that actually contains data

```
# -

# #KO minus
# KOM <- as.data.frame(IPK0minus)
# KOM <- KOM[KOM$seqnames == tchr & KOM$start >= scoord & KOM$end <= ecoord,]
# KOM$seqnames <- as.character(KOM$seqnames)
# KOM <- makeGRangesFromDataFrame(KOM, keep.extra.columns = T)
#
# KOMT <- DataTrack(KOM, name="KO-", ylim = c(0,clim))
#
#
# #WT minus
# WTM <- as.data.frame(IPWTminus)
# WTM <- WTM[WTM$seqnames == tchr & WTM$start >= scoord & WTM$end <= ecoord,]
# WTM$seqnames <- as.character(WTM$seqnames)
# WTM <- makeGRangesFromDataFrame(WTM, keep.extra.columns = T)
#
```

```

# WTmT <- DataTrack(WTm, name="WT-", ylim = c(0,clim))

# +

#KO plus
KOp <- as.data.frame(IPKOpplus)
KOp <- KOp[KOp$seqnames == tchr & KOp$start >= scoord & KOp$end <= ecoord,]
KOp$seqnames <- as.character(KOp$seqnames)
KOp <- makeGRangesFromDataFrame(KOp, keep.extra.columns = T)

KOpT <- DataTrack(KOp, name="KO+", ylim = c(0,clim))

#WT plus
WTP <- as.data.frame(IPWTplus)
WTP <- WTP[WTP$seqnames == tchr & WTP$start >= scoord & WTP$end <= ecoord,]
WTP$seqnames <- as.character(WTP$seqnames)
WTP <- makeGRangesFromDataFrame(WTP, keep.extra.columns = T)

WTPt <- DataTrack(WTP, name="WT+", ylim = c(0,clim))

```

chimeric reads

```

#minus
# chim181m <- as.data.frame(IP181chimWTminus)
# chim181m <- chim181m[chim181m$seqnames == tchr & chim181m$start >= scoord & chim181m$end <= ecoord,]
# chim181m$seqnames <- as.character(chim181m$seqnames)
# chim181m <- makeGRangesFromDataFrame(chim181m, keep.extra.columns = T)
#
# chim181mT <- DataTrack(chim181m, name="WT181rich-", ylim = c(0,clim))

#plus
chim181p <- as.data.frame(IP181chimWTplus)
chim181p <- chim181p[chim181p$seqnames == tchr & chim181p$start >= scoord & chim181p$end <= ecoord,]
chim181p$seqnames <- as.character(chim181p$seqnames)
chim181p <- makeGRangesFromDataFrame(chim181p, keep.extra.columns = T)

chim181pT <- DataTrack(chim181p, name="WT181rich-", ylim = c(0,clim))

```

differential binding

```

diffresf <- diffres %>% filter(seqnames == tchr)

diffresT <- AnnotationTrack(diffresf, name = "dBS", id=diffresf$name)

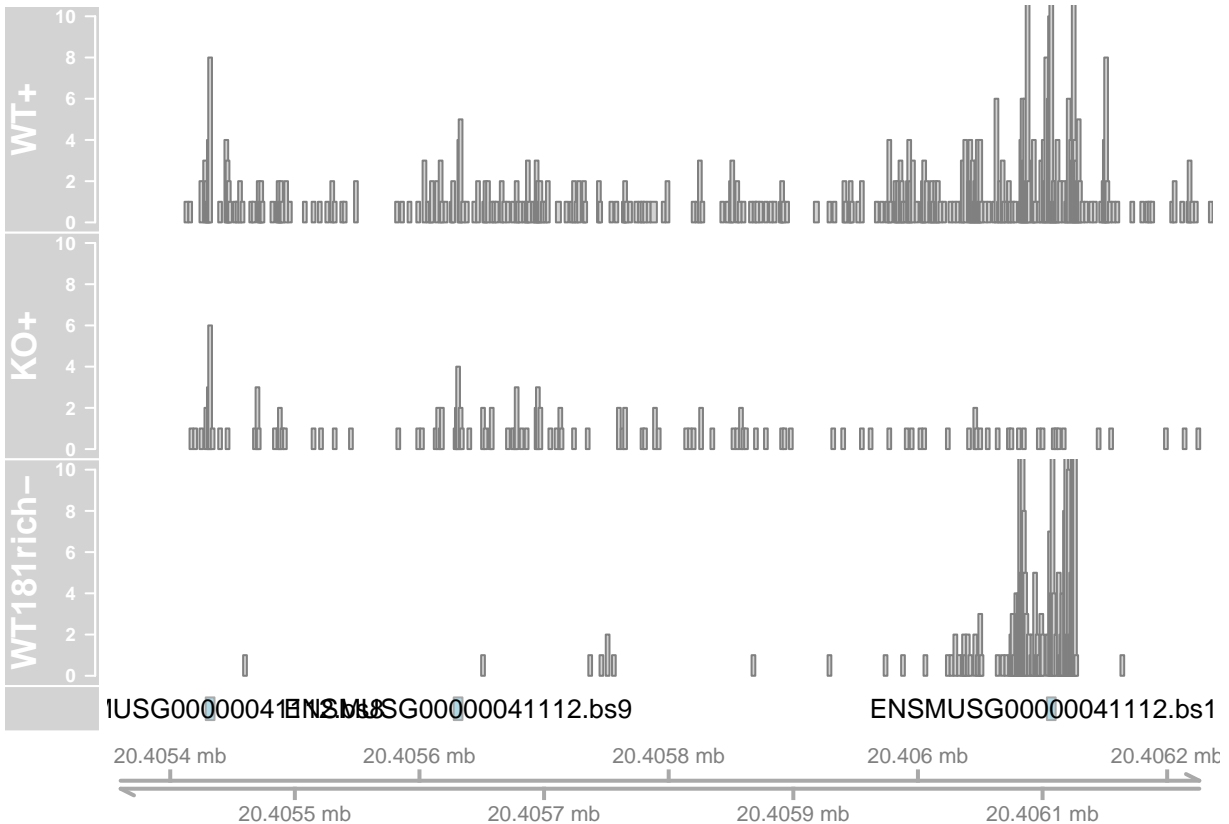
```

plot

```

plotTracks(list(WTPt, KOpT, chim181pT, diffresT, gatrack), from = scoord, to= ecoord, type="histogram"
            fontsize.feature=10, fontcolor.feature = "black")

```



session info

```
sessionInfo()
```

```
## R version 4.2.3 (2023-03-15 ucrt)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 19045)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=German_Germany.utf8  LC_CTYPE=German_Germany.utf8
## [3] LC_MONETARY=German_Germany.utf8 LC_NUMERIC=C
## [5] LC_TIME=German_Germany.utf8
##
## attached base packages:
## [1] grid      stats4    stats      graphics  grDevices  utils      datasets
## [8] methods  base
##
## other attached packages:
## [1] plyranges_1.18.0      dplyr_1.1.1          rtracklayer_1.58.0
## [4] Gviz_1.42.1           GenomicRanges_1.50.2 GenomeInfoDb_1.34.9
## [7] IRanges_2.32.0        S4Vectors_0.36.2     BiocGenerics_0.44.0
##
## loaded via a namespace (and not attached):
```

```

## [1] ProtGenerics_1.30.0      bitops_1.0-7
## [3] matrixStats_0.63.0        bit64_4.0.5
## [5] filelock_1.0.2            RColorBrewer_1.1-3
## [7] progress_1.2.2            httr_1.4.5
## [9] tools_4.2.3               backports_1.4.1
## [11] utf8_1.2.3                R6_2.5.1
## [13] rpart_4.1.19              lazyeval_0.2.2
## [15] Hmisc_5.0-1               DBI_1.1.3
## [17] colorspace_2.1-0          nnet_7.3-18
## [19] tidyselect_1.2.0          gridExtra_2.3
## [21] prettyunits_1.1.1         bit_4.0.5
## [23] curl_5.0.0                compiler_4.2.3
## [25] cli_3.6.0                 Biobase_2.58.0
## [27] htmlTable_2.4.1           xml2_1.3.3
## [29] DelayedArray_0.23.2       scales_1.2.1
## [31] checkmate_2.1.0           rappdirs_0.3.3
## [33] stringr_1.5.0             digest_0.6.31
## [35] Rsamtools_2.14.0          foreign_0.8-84
## [37] rmarkdown_2.21            XVector_0.38.0
## [39] jpeg_0.1-10               dichromat_2.0-0.1
## [41] base64enc_0.1-3           pkgconfig_2.0.3
## [43] htmltools_0.5.4           MatrixGenerics_1.10.0
## [45] highr_0.10                ensemblDb_2.22.0
## [47] dbplyr_2.3.2              fastmap_1.1.1
## [49] BSgenome_1.66.3           htmlwidgets_1.6.2
## [51] rlang_1.1.0               rstudioapi_0.14
## [53] RSQLite_2.3.1             BiocIO_1.8.0
## [55] generics_0.1.3            BiocParallel_1.32.6
## [57] VariantAnnotation_1.44.1   RCurl_1.98-1.12
## [59] magrittr_2.0.3            GenomeInfoDbData_1.2.9
## [61] Formula_1.2-5             interp_1.1-4
## [63] Matrix_1.5-3              Rcpp_1.0.10
## [65] munsell_0.5.0             fansi_1.0.4
## [67] lifecycle_1.0.3           stringi_1.7.12
## [69] yaml_2.3.7                SummarizedExperiment_1.28.0
## [71] zlibbioc_1.44.0           BiocFileCache_2.6.1
## [73] blob_1.2.4                parallel_4.2.3
## [75] crayon_1.5.2              deldir_1.0-6
## [77] lattice_0.20-45           Biostrings_2.66.0
## [79] GenomicFeatures_1.50.4    hms_1.1.3
## [81] KEGGREST_1.38.0           knitr_1.42
## [83] pillar_1.9.0              rjson_0.2.21
## [85] codetools_0.2-19          biomaRt_2.54.1
## [87] XML_3.99-0.14             glue_1.6.2
## [89] evaluate_0.20             latticeExtra_0.6-30
## [91] biovizBase_1.46.0         data.table_1.14.8
## [93] png_0.1-8                 vctrs_0.6.1
## [95] gtable_0.3.3              cachem_1.0.7
## [97] ggplot2_3.4.2             xfun_0.37
## [99] AnnotationFilter_1.22.0    restfulr_0.0.15
## [101] tibble_3.2.1              GenomicAlignments_1.34.1
## [103] AnnotationDbi_1.60.2      memoise_2.0.1
## [105] cluster_2.1.4

```