

find_MMsat4

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Setup

```
setwd("D:/Krueger_Lab/Publications/miR181_paper/Figure2/MMsat4")
```

packages

```
library(AnnotationHub)
```

```
## Loading required package: BiocGenerics
```

```
##
```

```
## Attaching package: 'BiocGenerics'
```

```
## The following objects are masked from 'package:stats':
```

```
##
```

```
##      IQR, mad, sd, var, xtabs
```

```
## The following objects are masked from 'package:base':
```

```
##
```

```
##      anyDuplicated, aperm, append, as.data.frame, basename, cbind,  
##      colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,  
##      get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,  
##      match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,  
##      Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort,  
##      table, tapply, union, unique, unsplit, which.max, which.min
```

```
## Loading required package: BiocFileCache
```

```
## Loading required package: dbplyr
```

```
library(dplyr)
```

```
##
```

```
## Attaching package: 'dplyr'
```

```
## The following objects are masked from 'package:dbplyr':
```

```
##
```

```
##      ident, sql
```

```
## The following objects are masked from 'package:BiocGenerics':
```

```
##
```

```
##      combine, intersect, setdiff, union
```

```
## The following objects are masked from 'package:stats':
```

```
##
```

```
##      filter, lag
```

```
## The following objects are masked from 'package:base':
##
## intersect, setdiff, setequal, union
library(ggplot2)
library(ggpubr)
library(rtracklayer)

## Loading required package: GenomicRanges
## Loading required package: stats4
## Loading required package: S4Vectors
##
## Attaching package: 'S4Vectors'
## The following objects are masked from 'package:dplyr':
##
## first, rename
## The following objects are masked from 'package:base':
##
## expand.grid, I, unname
## Loading required package: IRanges
##
## Attaching package: 'IRanges'
## The following objects are masked from 'package:dplyr':
##
## collapse, desc, slice
## The following object is masked from 'package:grDevices':
##
## windows
## Loading required package: GenomeInfoDb
##
## Attaching package: 'rtracklayer'
## The following object is masked from 'package:AnnotationHub':
##
## hubUrl
```

data

```
RNA <- read.csv("D:/Krueger_Lab/Publications/miR181_paper/Figure3/RNA_masterframe.csv")
RPF <- read.csv("D:/Krueger_Lab/Publications/miR181_paper/Figure3/RPF_masterframe.csv")

#load the gtf file to compare genes
gff23 <- import.gff3("D:/Krueger_Lab/Ribo_Profiling/run15112022M23/ref_genome/gencode.vM23.annotation.g

#colours
farbeneg <- "#b4b4b4"
farbe1 <- "#0073C2FF"
farbe2 <- "#EFC000FF"
farbe3 <- "#CD534CFF"
```

```

farbe4 <- "#7AA6DCFF"
farbe5 <- "#868686FF"
farbe6 <- "#003C67FF"
farbe7 <- "#8F7700FF"
farbe8 <- "#3B3B3BFF"
farbe9 <- "#A73030FF"
farbe10 <- "#4A6990FF"
farbe11 <- "#FF6F00FF"
farbe12 <- "#C71000FF"
farbe13 <- "#008EAOFF"
farbe14 <- "#8A4198FF"
farbe15 <- "#5A9599FF"
farbe16 <- "#FF6348FF"

RNApcol <- "#b56504"
RNAncol <- "#027d73"
RPFpcol <- "#c4c404"
RPFncol <- "#8d0391"

```

Get annotation

```

# ah = AnnotationHub()
# query(ah, c("RepeatMasker", "Mus musculus"))
# repeat_masker <- ah[["AH99012"]]

# load the downloaded data
repeat_masker <- readRDS("D:/Krueger_Lab/Publications/miR181_paper/Figure2/MMSat4/repeat_masker.rds")
repeat_masker[repeat_masker$repName == "MMSAT4"]

```

```

## GRanges object with 1693 ranges and 11 metadata columns:
##           seqnames           ranges strand |   swScore   milliDiv
##           <Rle>             <IRanges> <Rle> | <integer> <numeric>
##      [1]      chr1    75624925-75625061   + |      347      293
##      [2]      chr1    75625261-75625377   + |      228      292
##      [3]      chr1    75625502-75625561   + |      257      233
##      [4]      chr1 116745797-116745936   + |      357      314
##      [5]      chr1 116745987-116746488   + |      600      228
##      ...      ...      ...      ...      ...      ...
## [1689] chr9_KB469738_fix      24254-24454   - |      304      289
## [1690] chr9_KB469738_fix      24588-24803   - |      440      281
## [1691] chr9_KB469738_fix      25005-25142   - |      399      270
## [1692] chr9_KB469738_fix      25257-25488   - |      389      275
## [1693] chrY_JH792832_fix      349767-349817   + |      230      180
##           milliDel milliIns   genoLeft   repName   repClass   repFamily
##           <numeric> <numeric> <integer> <character> <character> <character>
##      [1]          29          29 -119846910   MMSAT4   Satellite   Satellite
##      [2]          34          34 -119846594   MMSAT4   Satellite   Satellite
##      [3]           0           0 -119846410   MMSAT4   Satellite   Satellite
##      [4]           7           0 -78726035   MMSAT4   Satellite   Satellite
##      [5]          48          22 -78725483   MMSAT4   Satellite   Satellite
##      ...      ...      ...      ...      ...      ...
## [1689]          24          16   -186187   MMSAT4   Satellite   Satellite
## [1690]           0           0   -185838   MMSAT4   Satellite   Satellite

```

```
##      [1691]          7          7    -185499      MMSAT4  Satellite  Satellite
##      [1692]         21         21    -185153      MMSAT4  Satellite  Satellite
##      [1693]          0         20    -194372      MMSAT4  Satellite  Satellite
##              repStart    repEnd    repLeft
##              <integer> <integer> <integer>
##      [1]         32        168         0
##      [2]         32        148        -20
##      [3]         31         90        -78
##      [4]         27        167         -1
##      [5]          1        514          0
##      ...         ...         ...         ...
##      [1689]          0        202          1
##      [1690]          0        216          1
##      [1691]          0        168         31
##      [1692]          0        232          1
##      [1693]        115        164         -4
##      -----
##      seqinfo: 239 sequences (1 circular) from mm10 genome
```

find overlaps

```
# same strand
MMSAT4 <- repeat_masker[repeat_masker$repName == "MMSAT4"]

OLgenes <- as.data.frame(subsetByOverlaps(gff23, MMSAT4))

# opposite strand
antiMMSAT4 <- MMSAT4
strand(antiMMSAT4) <- ifelse(strand(MMSAT4) == '+', '-', '+')

antiOLgenes <- as.data.frame(subsetByOverlaps(gff23, antiMMSAT4))

dim(OLgenes)
```

```
## [1] 3806  28
```

```
dim(antiOLgenes)
```

```
## [1] 140  28
```

```
RNA$MMSat4 <- "No_MMsat4"
RNA$MMSat4[RNA$gene_symbol %in% OLgenes$gene_name] <- "MMsat4"
RNA$MMSat4[RNA$gene_symbol %in% antiOLgenes$gene_name] <- "anti-MMsat4"
table(RNA$MMSat4)
```

```
##
## anti-MMsat4      MMSat4    No_MMsat4
##           12         238       13051
```

```
RPF$MMSat4 <- "No_MMsat4"
RPF$MMSat4[RPF$gene_symbol %in% OLgenes$gene_name] <- "MMsat4"
RPF$MMSat4[RPF$gene_symbol %in% antiOLgenes$gene_name] <- "anti-MMsat4"
table(RPF$MMSat4)
```

```
##
```

## anti-MMsat4	MMsat4	No_MMsat4
##	12	225
		11132

plot into MA plots

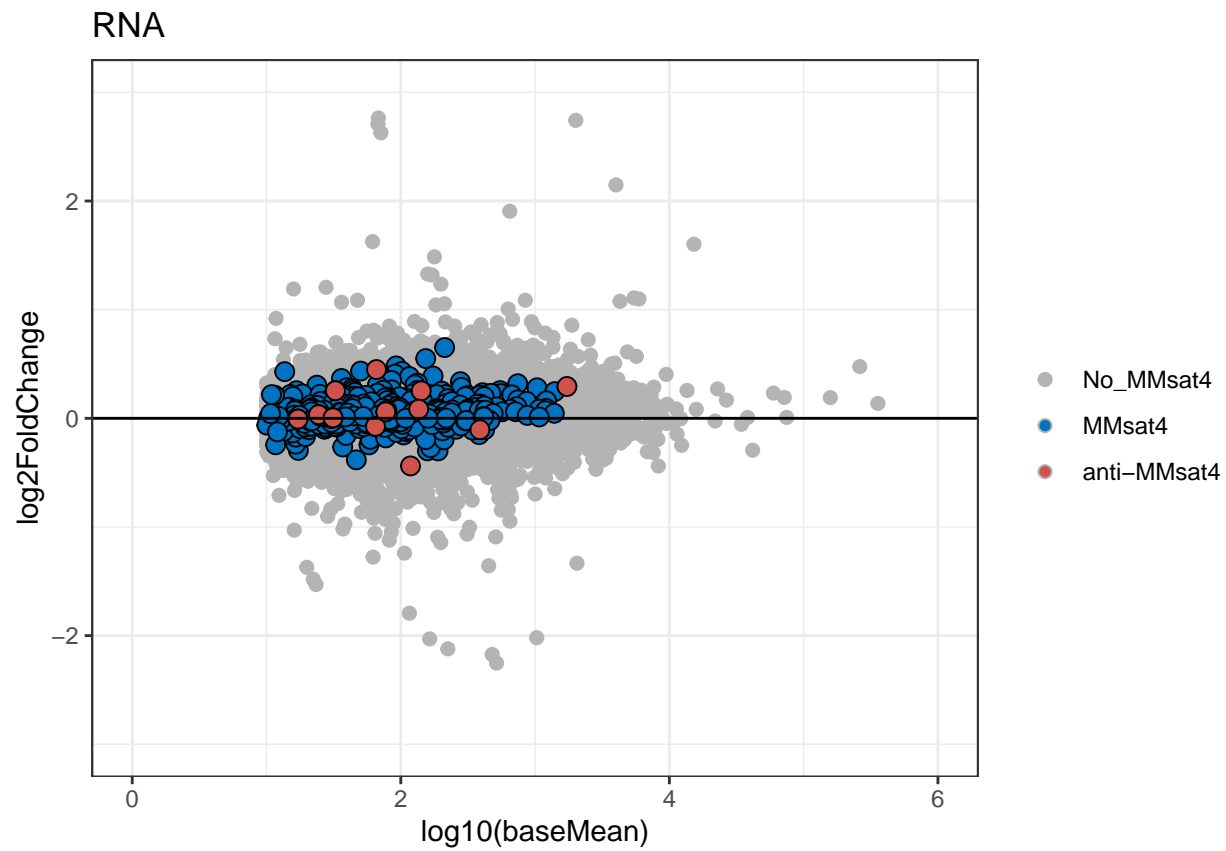
#RNA

```
RNAMA <- ggplot(RNA, aes(x=log10(baseMean), y=log2FoldChange, fill=factor(MMsat4, levels = c("No_MMsat4", "MMsat4", "anti-MMsat4")))) +
  geom_point(shape=21, size=2, colour=farbeneg) +
  scale_fill_manual(values = c(farbeneg, farbe1, farbe3)) +
  coord_cartesian(ylim = c(-3,3), xlim = c(0,6)) +
  geom_point(data=RNA[RNA$gene_symbol %in% OLgenes$gene_name,], aes(x=log10(baseMean), y=log2FoldChange)) +
  geom_point(data=RNA[RNA$gene_symbol %in% antiOLgenes$gene_name,], aes(x=log10(baseMean), y=log2FoldChange)) +
  geom_hline(yintercept = 0, colour= "black") +
  theme_bw() +
  theme(legend.title = element_blank()) +
  ggtitle("RNA")
```

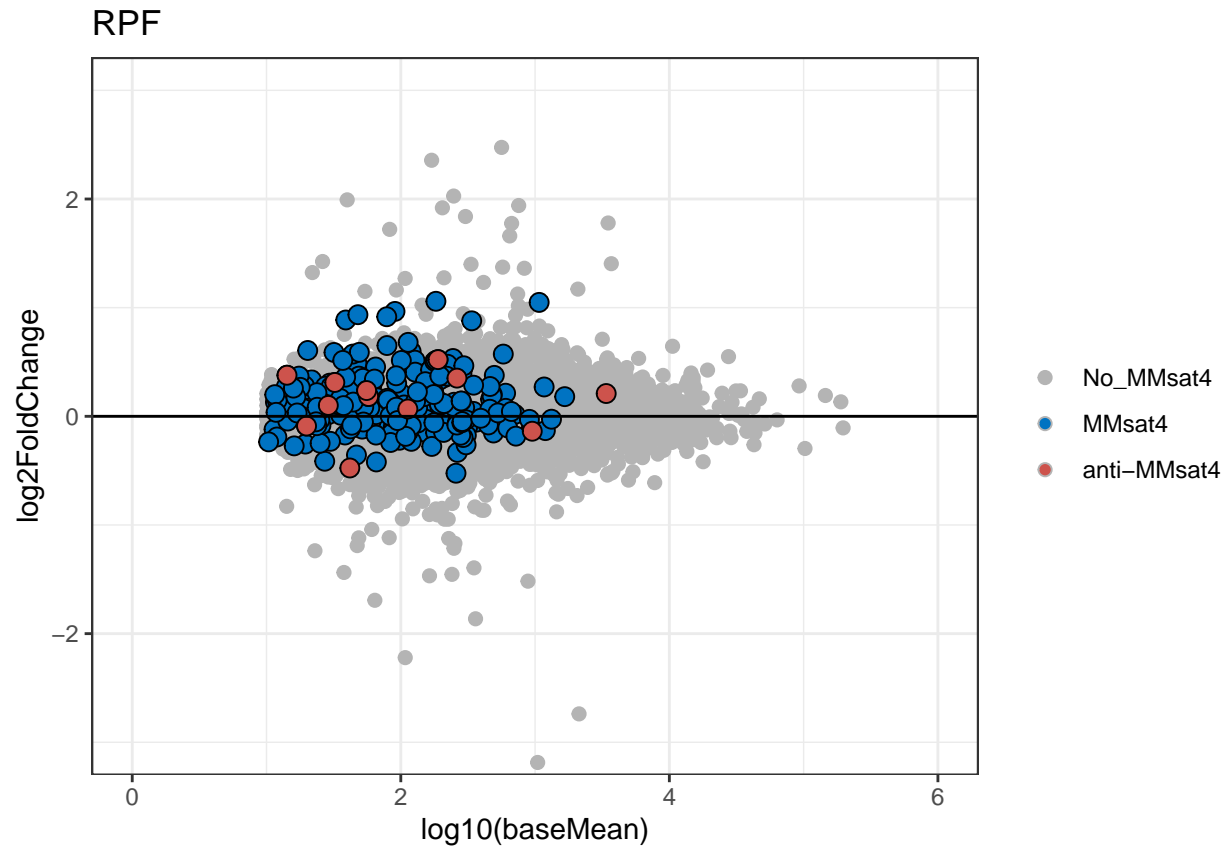
#RPF

```
RPFMA <- ggplot(RPF, aes(x=log10(baseMean), y=log2FoldChange, fill=factor(MMsat4, levels = c("No_MMsat4", "MMsat4", "anti-MMsat4")))) +
  geom_point(shape=21, size=2, colour=farbeneg) +
  scale_fill_manual(values = c(farbeneg, farbe1, farbe3)) +
  coord_cartesian(ylim = c(-3,3), xlim = c(0,6)) +
  geom_point(data=RPF[RPF$gene_symbol %in% OLgenes$gene_name,], aes(x=log10(baseMean), y=log2FoldChange)) +
  geom_point(data=RPF[RPF$gene_symbol %in% antiOLgenes$gene_name,], aes(x=log10(baseMean), y=log2FoldChange)) +
  geom_hline(yintercept = 0, colour= "black") +
  theme_bw() +
  theme(legend.title = element_blank()) +
  ggtitle("RPF")
```

RNAMA



RPFMA

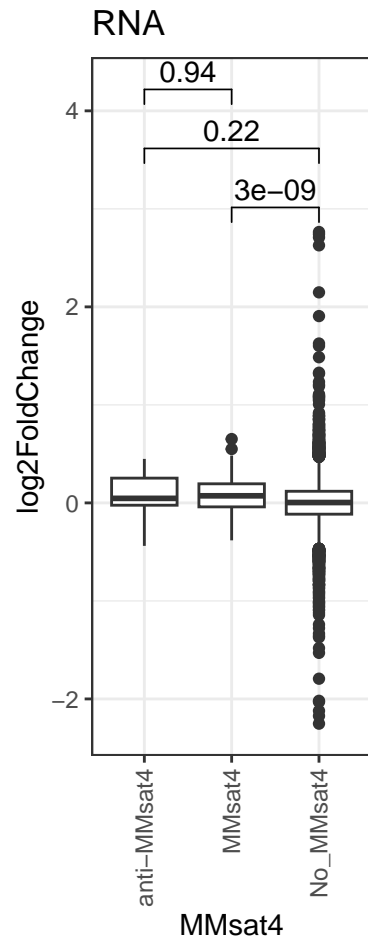


```
my_comparisons <- list( c("No_MMsat4", "MMsat4"), c("No_MMsat4", "anti-MMsat4"), c("MMsat4", "anti-MMsat4") )
```

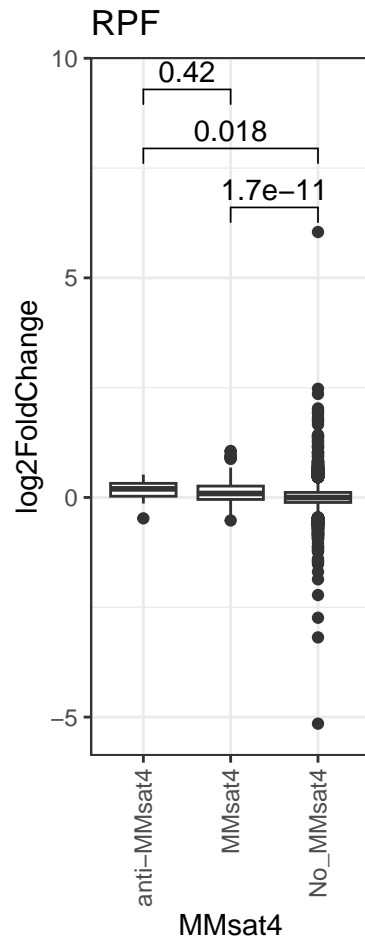
```
RNAbox <- ggplot(RNA, aes(x=MMsat4, y=log2FoldChange)) +
  geom_boxplot() +
  stat_compare_means(comparisons = my_comparisons) +
  theme_bw() +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  ggtitle("RNA")
```

```
RPFbox <- ggplot(RPF, aes(x=MMsat4, y=log2FoldChange)) +
  geom_boxplot() +
  stat_compare_means(comparisons = my_comparisons) +
  theme_bw() +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  ggtitle("RPF")
```

RNAbox



RPFbox



session info

```
sessionInfo()
```

```
## R version 4.2.3 (2023-03-15 ucrt)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 19045)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=German_Germany.utf8 LC_CTYPE=German_Germany.utf8
## [3] LC_MONETARY=German_Germany.utf8 LC_NUMERIC=C
## [5] LC_TIME=German_Germany.utf8
##
## attached base packages:
## [1] stats4      stats      graphics  grDevices  utils      datasets  methods
## [8] base
##
## other attached packages:
## [1] rtracklayer_1.58.0 GenomicRanges_1.50.2 GenomeInfoDb_1.34.9
## [4] IRanges_2.32.0      S4Vectors_0.36.2      ggpubr_0.6.0
```

```

## [7] ggplot2_3.4.2      dplyr_1.1.2      AnnotationHub_3.6.0
## [10] BiocFileCache_2.6.1 dbplyr_2.3.2      BiocGenerics_0.44.0
##
## loaded via a namespace (and not attached):
## [1] bitops_1.0-7      matrixStats_0.63.0
## [3] bit64_4.0.5      filelock_1.0.2
## [5] httr_1.4.6       tools_4.2.3
## [7] backports_1.4.1  utf8_1.2.3
## [9] R6_2.5.1         DBI_1.1.3
## [11] colorspace_2.1-0 withr_2.5.0
## [13] tidyselect_1.2.0 bit_4.0.5
## [15] curl_5.0.0       compiler_4.2.3
## [17] cli_3.6.0        Biobase_2.58.0
## [19] DelayedArray_0.23.2 labeling_0.4.2
## [21] scales_1.2.1     rappdirs_0.3.3
## [23] digest_0.6.31    Rsamtools_2.14.0
## [25] rmarkdown_2.21   XVector_0.38.0
## [27] pkgconfig_2.0.3  htmltools_0.5.4
## [29] MatrixGenerics_1.10.0 highr_0.10
## [31] fastmap_1.1.1    rlang_1.1.0
## [33] rstudioapi_0.14  RSQLite_2.3.1
## [35] shiny_1.7.4      farver_2.1.1
## [37] BiocIO_1.8.0     generics_0.1.3
## [39] BiocParallel_1.32.6 car_3.1-2
## [41] RCurl_1.98-1.12  magrittr_2.0.3
## [43] GenomeInfoDbData_1.2.9 Matrix_1.5-3
## [45] Rcpp_1.0.10      munsell_0.5.0
## [47] fansi_1.0.4      abind_1.4-5
## [49] lifecycle_1.0.3  yaml_2.3.7
## [51] carData_3.0-5    SummarizedExperiment_1.28.0
## [53] zlibbioc_1.44.0  grid_4.2.3
## [55] blob_1.2.4       parallel_4.2.3
## [57] promises_1.2.0.1 crayon_1.5.2
## [59] lattice_0.20-45  Biostings_2.66.0
## [61] KEGGREST_1.38.0  knitr_1.42
## [63] pillar_1.9.0     rjson_0.2.21
## [65] ggsignif_0.6.4   codetools_0.2-19
## [67] XML_3.99-0.14    glue_1.6.2
## [69] BiocVersion_3.16.0 evaluate_0.21
## [71] BiocManager_1.30.20 png_0.1-8
## [73] vctrs_0.6.2      httpuv_1.6.11
## [75] gtable_0.3.3     purrr_1.0.1
## [77] tidyr_1.3.0      cachem_1.0.8
## [79] xfun_0.37        mime_0.12
## [81] xtable_1.8-4     broom_1.0.4
## [83] restfulr_0.0.15  rstatix_0.7.2
## [85] later_1.3.1      tibble_3.2.1
## [87] GenomicAlignments_1.34.1 AnnotationDbi_1.60.2
## [89] memoise_2.0.1    ellipsis_0.3.2
## [91] interactiveDisplayBase_1.36.0

```