Supporting Information

Biological processes in Sansevieria trifasciata reduce its particulate matter removal efficiency

Ning Yuan Lee Samuel Lai Swee Yang Low

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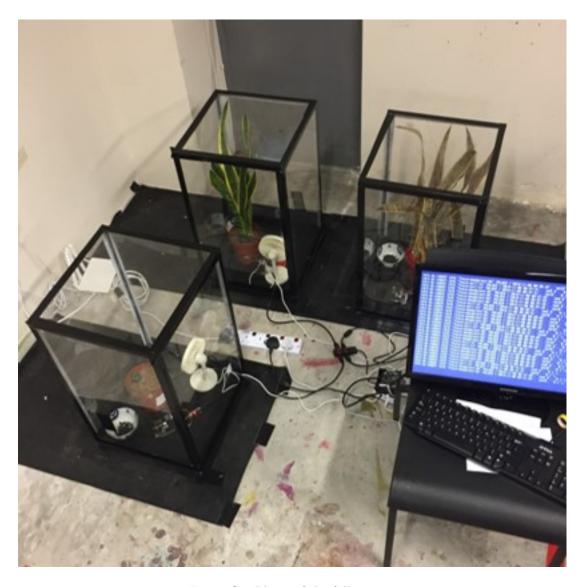


Figure S1: Photo of the full set-up.

1 Import and Tidy Data

We have 60 sets of data in total. Each set represents a single trial of $PM_{2.5}$, PM_{10} readings for one group from the four control and treatment groups. Each trial was conducted using one of three sensor-box combinations (1a, 2b, or 3d).

1.1 Define an import Function

Here, we define a function import which will

- 1. Import a trial from csv file,
- 2. Add a column to documents its sensor-box and another for its group,
- 3. Place the $PM_{2.5}$ and PM_{10} readings into their own tibbles, then for each of the two reading types:
- 4. Trim away all the readings before the first time the sensor desaturates, and
- 5. Set the time column as being the seconds since the first desaturated reading.

By desaturation, we mean the first reading which falls within the sensor's specified range. For our model of

PM sensor, this is defined as $[0,999.9] \, \mu \text{g m}^{-3}$.

```
library(tidyverse)
import <- function(filename) {</pre>
  # Import a trial from csv file
  data <- read_csv(filename, col_names=c('time', 'type', 'value'))</pre>
  # Place the PM2.5 and PM10 readings into their own tibbles
  data <- list(</pre>
    pm2.5=(data %>%
      subset(type=='pm2.5') %>%
      mutate(value=as.numeric(value)) %>%
      mutate(time=as.numeric(as.POSIXct(time)))),
    pm10=(data %>%
      subset(type=='pm10') %>%
      mutate(value=as.numeric(value)) %>%
      mutate(time=as.numeric(as.POSIXct(time))))
  )
  # Add a column to documents its group
  if (grepl('nothing', filename, fixed=T)) {
    data$group <- 'nothing'</pre>
  else if (
    grepl('plant', filename, fixed=T) &
    !grepl('deadplant', filename, fixed=T)
  ) {
    data$group <- 'plant'</pre>
  else if (grepl('deadplant', filename, fixed=T)) {
    data$group <- 'deadplant'</pre>
  } else if (grepl('soil', filename, fixed=T)) {
    data$group <- 'soil'</pre>
  } else {
    stop(paste('import: Invaid group in', filename))
  }
  # Add a column to documents its sensor-box
  if(grepl('1[aA]', filename)) {
    data$sensorbox <- '1a'
  } else if (grepl('2[bB]', filename)) {
    data$sensorbox <- '2b'</pre>
  } else if (grepl('3[dD]', filename)) {
    data$sensorbox <- '3d'
  } else {
    stop(paste('import: Invalid sensor-box in', filename))
  # Trim away all the readings before the first time the sensor desaturates
  pm2.5_last_saturated <- last(which(data$pm2.5$value>=999.9))
  data$pm2.5 <- data$pm2.5[-(1:pm2.5_last_saturated),]</pre>
  pm10_last_saturated <- last(which(data$pm10$value>=999.9))
  data$pm10 <- data$pm10[-(1:pm10_last_saturated),]</pre>
```

```
# Set the time column as being the seconds since the first desaturated reading
data$pm2.5$time <- data$pm2.5$time - data$pm2.5$time[1]
data$pm10$time <- data$pm10$time - data$pm10$time[1]
return(data)
}</pre>
```

1.2 Apply the import Function

Then, we import all our data by first building a vector of their filenames.

```
filenames <- c(
  '19-octa-nothing-1a.csv', '19-octa-nothing-2b.csv', '19-octa-nothing-3d.csv',
  '20octa-nothing1a.csv', '20octa-nothing2b.csv', '20octa-nothing3d.csv',
  '21octa-nothing-1a.csv', '21octa-nothing-2b.csv',
                                                      '21octa-nothing-3d.csv',
  '21octb-nothing-1a.csv', '21octb-nothing-2b.csv',
                                                      '21octb-nothing-3d.csv',
  '22octa-nothing-1a.csv', '22octa-nothing-2b.csv',
                                                      '22octa-nothing-3d.csv',
  '25octa-deadplant-1a.csv', '25octa-plant-3D.csv',
                                                      '25octa-soil-2b.csv',
  '25octb-deadplant-1a.csv', '25octb-plant-3D.csv',
                                                      '25octb-soil-2b.csv',
  '26octa-deadplant-1a.csv', '26octa-plant-3d.csv',
                                                      '26octa-soil-2b.csv',
  '27octa-deadplant-1a.csv', '27octa-plant-3d.csv',
                                                      '27octa-soil-2b.csv',
  '28octa-deadplant-1a.csv', '28octa-plant-3d.csv',
                                                      '28octa-soil-2b.csv',
  '2nova-deadplant-2b.csv', '2nova-plant-1a.csv',
                                                      '2nova-soil-3d.csv',
  '2novb-deadplant-2b.csv', '2novb-plant-1a.csv',
                                                      '2novb-soil-3d.csv',
  '30octa-deadplant-2b.csv', '30octa-plant-1a.csv',
                                                      '30octa-soil-3d.csv',
  '30octb-deadplant-2b.csv', '30octb-plant-1a.csv',
                                                      '30octb-soil-3d.csv',
  '31octb-deadplant-2b.csv', '31octb-plant-1a.csv',
                                                      '31octb-soil-3d.csv',
  '3nova-deadplant-3d.csv', '3nova-plant-2b.csv',
                                                      '3nova-soil-1a.csv',
  '4nova-deadplant-3d.csv', '4nova-plant-2b.csv',
                                                      '4nova-soil-1a.csv',
  '5nova-deadplant-3d.csv', '5nova-plant-2b.csv',
                                                      '5nova-soil-1a.csv',
  '6nova-deadplant-3d.csv', '6nova-plant-2b.csv',
                                                      '6nova-soil-1a.csv',
  '7nova-plant-2b.csv', '7nova-deadplant-3d.csv', '7nova-soil-1a.csv'
```

Followed by an lapply of the import function over the filenames vector to obtain a list of lists. Each nested list is a trial.

```
data <- lapply(filenames, function(fn) {
  import(paste('05-analysis-proper/', fn, sep=''))})</pre>
```

2 Calibrate Sensors

We observed that each sensor is not calibrated with the others, and varies sometimes in a non-linear fashion (and so cannot be easily controlled for in the linear model). In order to calibrate these sensors, we conducted another trial where

- Each sensor is placed in the same box and reads during the same session, and
- Each sensor reads within 5s of each other.

An arbitrary sensor labelled 3 was selected as the baseline for calibration. We then ran a quadratic regression in a pairwise manner between the sensors, in order to obtain estimates for a calibration function.

2.1 Import and Tidy the Calibration Data

First, load the readings for the individual sensors. Then, combine them into one tibble. Note that each sensor's time stamp is synchronised because they are given by the same raspberry pi zero computer.

```
import_cdata <- function(filename) {</pre>
  data <- filename %>%
    read_csv(col_names=c('time', 'type', 'value')) %>%
    subset(type != 'comment') %>%
    mutate(value=as.numeric(value)) %>%
    mutate(time=as.numeric(as.POSIXct(time)))
}
# The three sensors were set-up such that they all began at the same time, +- 1s.
sensor1 <- import_cdata('03-sensor-calibration/16octc-nothing-1bigbox.csv')</pre>
sensor2 <- import cdata('03-sensor-calibration/16octc-nothing-2bigbox.csv')</pre>
sensor3 <- import_cdata('03-sensor-calibration/16octc-nothing-3bigbox.csv')</pre>
# Sometimes, the an interval (supposedly per-minute) fails to read, and a row is missing.
# So, we want to make sure that each row index of sensor 1/2 aligns with sensor 3.
grouped pm2.5 <- tibble(s1=double(), s2=double(), s3=double())
sensor1_pm2.5 <- subset(sensor1, type=='pm2.5')</pre>
sensor2_pm2.5 <- subset(sensor2, type=='pm2.5')</pre>
sensor3_pm2.5 <- subset(sensor3, type=='pm2.5')</pre>
# Also, trim the pre-desaturation data. The 999.9 are outliers with high leverage, and
# the data leading up to saturation is high variance.
last_saturated <- max(</pre>
  last(which(sensor1_pm2.5$value >= 999.9)),
  last(which(sensor2_pm2.5$value >= 999.9)),
  last(which(sensor3_pm2.5$value >= 999.9))
sensor1_pm2.5 <- sensor1_pm2.5[-(1:last_saturated),]</pre>
sensor2_pm2.5 <- sensor2_pm2.5[-(1:last_saturated),]</pre>
sensor3_pm2.5 <- sensor3_pm2.5[-(1:last_saturated),]</pre>
for (i in seq(1, nrow(sensor3_pm2.5))) {
  s3 <- sensor3 pm2.5[i,]
  s1 <- sensor1_pm2.5 %>% mutate(time=time-s3[1,]$time) %>% subset(abs(time) <= 5)
  s2 <- sensor2_pm2.5 %>% mutate(time=time-s3[1,]$time) %>% subset(abs(time) <= 5)
  if (nrow(s1) == 0 \& nrow(s2) == 0) {
    next
  } else if (nrow(s1) == 0) {
    grouped_pm2.5 <- bind_rows(</pre>
      grouped_pm2.5, list(s1=NA, s2=s2[1,]$value, s3=s3[1,]$value))
```

```
} else if (nrow(s2) == 0) {
    grouped_pm2.5 <- bind_rows(</pre>
      grouped_pm2.5, list(s1=s1[1,]$value, s2=NA, s3=s3[1,]$value))
    grouped_pm2.5 <- bind_rows(</pre>
      grouped_pm2.5, list(s1=s1[1,]$value, s2=s2[1,]$value, s3=s3[1,]$value))
}
# Repeat for PM10...
grouped_pm10 <- tibble(s1=double(), s2=double(), s3=double())</pre>
sensor1_pm10 <- subset(sensor1, type=='pm10')</pre>
sensor2_pm10 <- subset(sensor2, type=='pm10')</pre>
sensor3_pm10 <- subset(sensor3, type=='pm10')</pre>
last_saturated <- max(</pre>
  last(which(sensor1_pm10$value >= 999.9)),
  last(which(sensor2_pm10$value >= 999.9)),
  last(which(sensor3_pm10$value >= 999.9))
)
sensor1_pm10 <- sensor1_pm10[-(1:last_saturated),]</pre>
sensor2_pm10 <- sensor2_pm10[-(1:last_saturated),]</pre>
sensor3_pm10 <- sensor3_pm10[-(1:last_saturated),]</pre>
for (i in seq(1, nrow(sensor3_pm10))) {
  s3 <- sensor3_pm10[i,]</pre>
  s1 <- sensor1 pm10 %>% mutate(time=time-s3[1,]$time) %>% subset(abs(time) <= 5)
  s2 <- sensor2_pm10 %>% mutate(time=time-s3[1,]$time) %>% subset(abs(time) <= 5)
  if (nrow(s1) == 0 & nrow(s2) == 0) {
    next
  } else if (nrow(s1) == 0) {
    grouped_pm10 <- bind_rows(</pre>
      grouped_pm10, list(s1=NA, s2=s2[1,]$value, s3=s3[1,]$value))
  } else if (nrow(s2) == 0) {
    grouped_pm10 <- bind_rows(</pre>
      grouped_pm10, list(s1=s1[1,]$value, s2=NA, s3=s3[1,]$value))
  } else {
    grouped_pm10 <- bind_rows(</pre>
      grouped_pm10, list(s1=s1[1,]$value, s2=s2[1,]$value, s3=s3[1,]$value))
}
data_calibration <- list(pm2.5=grouped_pm2.5, pm10=grouped_pm10)
```

Similar to the previous section, readings before the saturation points were trimmed away.

A quick graphical check shows that the sensors are indeed different:

```
ggplot(data_calibration$pm2.5) +
  geom_point(mapping=aes(x=s3, y=s1), size=.2, colour='blue') +
  geom_point(mapping=aes(x=s3, y=s2), size=.2, colour='red') +
  geom_abline(linetype='longdash') +
  ylab('s1/s2')
```

```
## Warning: Removed 10 rows containing missing values (geom_point).
```

^{##} Warning: Removed 7 rows containing missing values (geom_point).

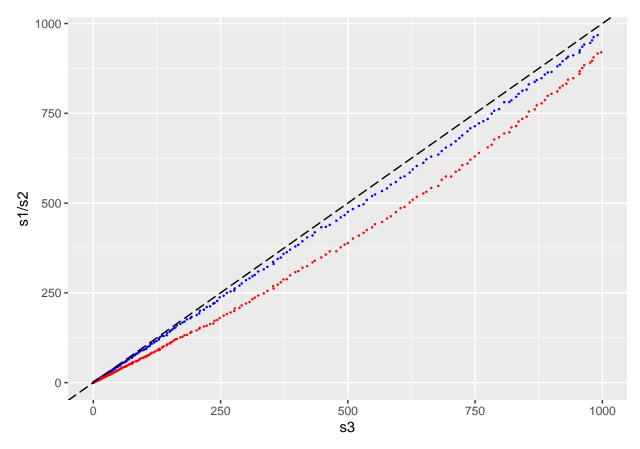


Figure S2: PM2.5 readings of sensor s1 (blue) and s2 (red) for readings of sensor s3 on the x-axis, in the same enclosure, taken at roughly the same time. The solid black line is the x=y line.

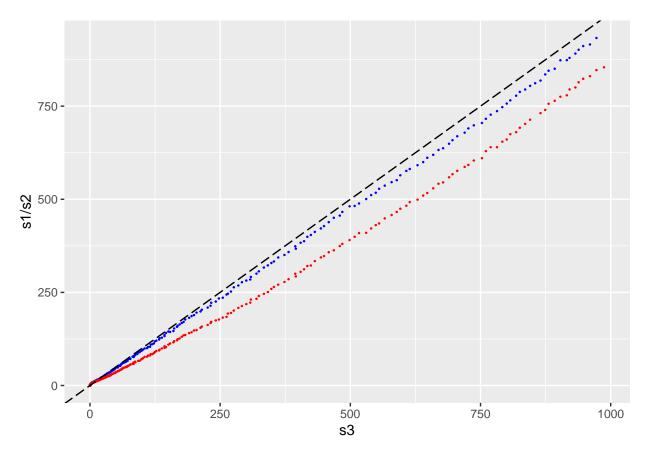


Figure S3: PM10 readings of sensor s1 (blue) and s2 (red) for readings of sensor s3 on the x-axis, in the same enclosure, taken at roughly the same time. The solid black line is the x = y line.

```
ggplot(data_calibration$pm10) +
  geom_point(mapping=aes(x=s3, y=s1), size=.2, colour='blue') +
  geom_point(mapping=aes(x=s3, y=s2), size=.2, colour='red') +
  geom_abline(linetype='longdash') +
  ylab('s1/s2')
```

Warning: Removed 10 rows containing missing values (geom_point).

Warning: Removed 7 rows containing missing values (geom_point).

The missing values are a result of a glitch in obtaining readings from the PM sensors (sometimes, readings fail to be obtained from the sensors).

2.2 Calibration via Regression

Now, the calibration via regression.

```
calibration <- list(
  pm2.5=list(
    s1=lm(s1~s3+I(s3^2), data=data_calibration$pm2.5),
    s2=lm(s2~s3+I(s3^2), data=data_calibration$pm2.5)
),
  pm10=list(
    s1=lm(s1~s3+I(s3^2), data=data_calibration$pm10),</pre>
```

```
s2=lm(s2~s3+I(s3^2), data=data_calibration$pm10)
 )
)
summary(calibration$pm2.5$s1)
##
## Call:
## lm(formula = s1 ~ s3 + I(s3^2), data = data_calibration$pm2.5)
## Residuals:
                 1Q
                     Median
                                   3Q
## -11.9730 -0.3237 -0.2314 0.2253 10.5947
##
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 3.696e-01 7.396e-02
                                   4.997 7.19e-07 ***
              9.236e-01 1.121e-03 823.930 < 2e-16 ***
## I(s3^2)
              4.531e-05 1.392e-06 32.559 < 2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 1.851 on 783 degrees of freedom
     (10 observations deleted due to missingness)
## Multiple R-squared: 0.9999, Adjusted R-squared: 0.9999
## F-statistic: 5.05e+06 on 2 and 783 DF, p-value: < 2.2e-16
summary(calibration$pm2.5$s2)
##
## Call:
## lm(formula = s2 ~ s3 + I(s3^2), data = data_calibration$pm2.5)
## Residuals:
       Min
                 1Q
                     Median
                                   3Q
## -14.6745 -0.4492 -0.3537 -0.0691 13.3057
##
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 5.581e-01 9.175e-02
                                    6.083 1.84e-09 ***
## s3
              6.517e-01 1.384e-03 471.028 < 2e-16 ***
## I(s3^2)
              2.618e-04 1.705e-06 153.538 < 2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 2.303 on 786 degrees of freedom
     (7 observations deleted due to missingness)
## Multiple R-squared: 0.9999, Adjusted R-squared: 0.9999
## F-statistic: 2.642e+06 on 2 and 786 DF, p-value: < 2.2e-16
summary(calibration$pm10$s1)
##
## Call:
## lm(formula = s1 ~ s3 + I(s3^2), data = data_calibration$pm10)
```

```
##
## Residuals:
##
       Min
                 1Q
                     Median
## -10.1701 -0.8193 -0.3620 0.7817 11.9315
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 1.140e+00 6.993e-02
                                    16.31
                                             <2e-16 ***
## s3
              9.297e-01 1.130e-03 822.51
                                             <2e-16 ***
## I(s3^2)
              2.480e-05 1.478e-06
                                   16.78
                                             <2e-16 ***
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 1.74 on 763 degrees of freedom
     (10 observations deleted due to missingness)
## Multiple R-squared: 0.9999, Adjusted R-squared: 0.9999
## F-statistic: 4.267e+06 on 2 and 763 DF, p-value: < 2.2e-16
summary(calibration$pm10$s2)
##
## Call:
## lm(formula = s2 ~ s3 + I(s3^2), data = data_calibration$pm10)
## Residuals:
##
       Min
                 1Q
                      Median
                                   3Q
                                           Max
## -10.4328 -0.9058 -0.6122
                               0.8488
                                        8.6659
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
##
                                    15.08
## (Intercept) 1.070e+00 7.097e-02
                                             <2e-16 ***
              6.773e-01 1.137e-03 595.55
                                             <2e-16 ***
              1.937e-04 1.476e-06 131.26
## I(s3^2)
                                             <2e-16 ***
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 1.772 on 766 degrees of freedom
     (7 observations deleted due to missingness)
## Multiple R-squared: 0.9999, Adjusted R-squared: 0.9999
## F-statistic: 3.118e+06 on 2 and 766 DF, p-value: < 2.2e-16
```

2.3 Define a calibrate Function

Finally, package it all into a function.

```
calibrate_one <- function(pm, sensorbox, x) {
  if (is.na(x)) return(NA)
  if (pm == 'pm2.5' & sensorbox == '1a') {
    lm_obj <- calibration$pm2.5$s1
} else if (pm == 'pm2.5' & sensorbox == '2b') {
    lm_obj <- calibration$pm2.5$s2
} else if (pm == 'pm10' & sensorbox == '1a') {
    lm_obj <- calibration$pm10$s1
} else if (pm == 'pm10' & sensorbox == '2b') {
    lm_obj <- calibration$pm10$s2</pre>
```

```
} else {
    stop(paste('Invalid pm/sensorbox: ', pm, sensorbox))
  coeff <- lm_obj$coefficients - c(x, 0, 0)</pre>
  roots <- Re(polyroot(coeff))</pre>
  root <- roots[roots>=0]
 if (length(root) == 1) return(root)
 return(NA) # would use NULL, but can't have NA in the middle of a vector
}
calibrate <- function(pm, sensorbox, xs) {</pre>
  return(unlist(Map(
    function(x) calibrate_one(pm, sensorbox, x), xs)))
}
Some closing graphical sanity checks:
ggplot(data_calibration$pm2.5 %>% mutate(s1.fix=calibrate('pm2.5', '1a', s1))) +
  geom point(mapping=aes(x=s3, y=s1), colour='red', size=.2) +
  geom_point(mapping=aes(x=s3, y=s1.fix), colour='darkgreen', size=.2) +
  geom_abline()
## Warning: Removed 10 rows containing missing values (geom_point).
## Warning: Removed 127 rows containing missing values (geom_point).
ggplot(data_calibration$pm2.5 %>% mutate(s2.fix=calibrate('pm2.5', '2b', s2))) +
  geom_point(mapping=aes(x=s3, y=s2), colour='red', size=.2) +
  geom_point(mapping=aes(x=s3, y=s2.fix), colour='darkgreen', size=.2) +
 geom_abline()
## Warning: Removed 7 rows containing missing values (geom point).
## Warning: Removed 377 rows containing missing values (geom_point).
ggplot(data_calibration$pm10 %>% mutate(s1.fix=calibrate('pm10', '1a', s1))) +
  geom_point(mapping=aes(x=s3, y=s1), colour='red', size=.2) +
  geom point(mapping=aes(x=s3, y=s1.fix), colour='darkgreen', size=.2) +
 geom abline()
## Warning: Removed 10 rows containing missing values (geom_point).
## Warning: Removed 274 rows containing missing values (geom_point).
ggplot(data calibration$pm10 %>% mutate(s2.fix=calibrate('pm10', '2b', s2))) +
  geom_point(mapping=aes(x=s3, y=s2), colour='red', size=.2) +
  geom_point(mapping=aes(x=s3, y=s2.fix), colour='darkgreen', size=.2) +
  geom_abline()
## Warning: Removed 7 rows containing missing values (geom_point).
## Warning: Removed 399 rows containing missing values (geom_point).
```

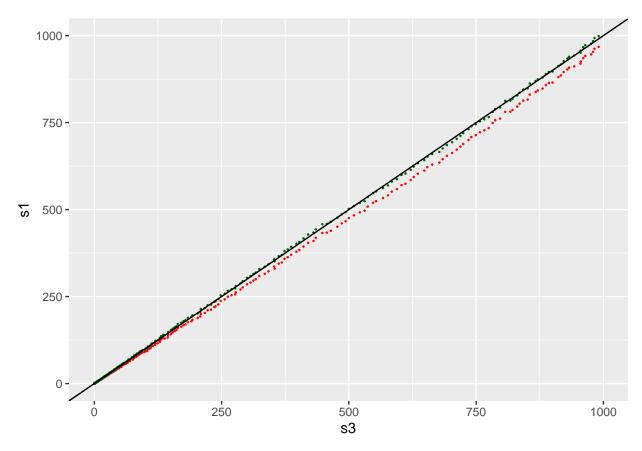


Figure S4: PM2.5 readings of s1 versus s3 after calibration (dark green), and before (red). The solid black line is the x=y line.

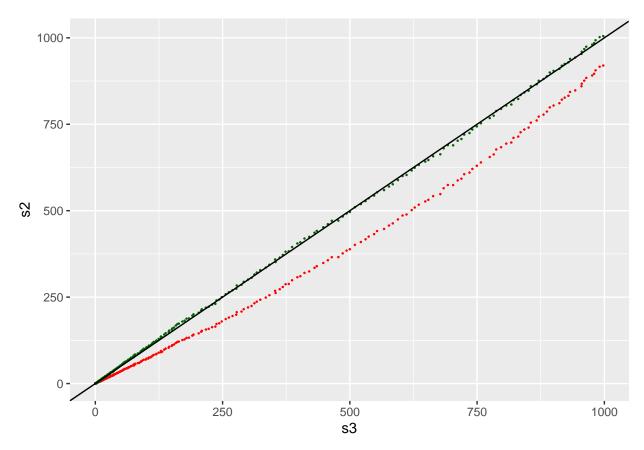


Figure S5: PM2.5 readings of s2 versus s3 after calibration (dark green), and before (red). The solid black line is the x=y line.

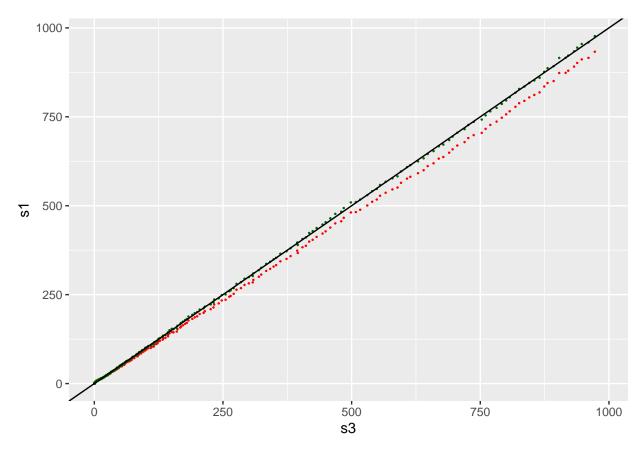


Figure S6: PM10 readings of s1 versus s3 after calibration (dark green), and before (red). The solid black line is the x=y line.

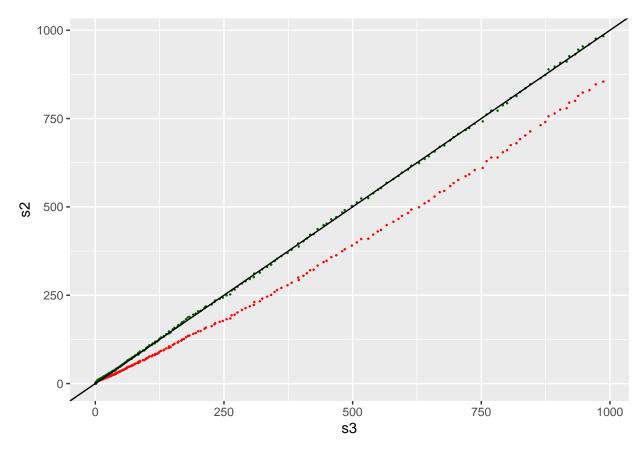


Figure S7: PM10 readings of s2 versus s3 after calibration (dark green), and before (red). The solid black line is the x=y line.

3 Box Plot & Regression

First, we consider only $PM_{2.5}$ in §3.1–3.2. Then, we repeat the analysis for PM_{10} in §3.3.

3.1 Transform Data

First, calibrate the data using the previously-defined calibrate function.

```
data_calibrated <- data %>%
  lapply(function(trial) {
    if (trial$sensorbox == '1a') {
        trial$pm2.5$value <- calibrate('pm2.5', '1a', trial$pm2.5$value)
        trial$pm10$value <- calibrate('pm10', '1a', trial$pm10$value)
    } else if (trial$sensorbox == '2b') {
        trial$pm2.5$value <- calibrate('pm2.5', '2b', trial$pm2.5$value)
        trial$pm10$value <- calibrate('pm10', '2b', trial$pm10$value)
    } else if (trial$sensorbox == '3d') {
        # noop
    } else {
        stop(paste('Invalid sensorbox', trial$sensorbox))
    }
    return(trial)
})</pre>
```

Now, create a new tibble which records the time taken for $PM_{2.5}$ and PM_{10} to decrease from $900 \,\mu g \, m^{-3}$ to $450 \,\mu g \, m^{-3}$. Since our readings are limited in resolution (per-minute), we pick the readings closest to $900 \,\mu g \, m^{-3}$ and $450 \,\mu g \, m^{-3}$.

```
pm2.5_aggregated <- lapply(data_calibrated, function(trial) {</pre>
  # This function is applied for each trial.
  # First, find the index of the 900 reading.
  index 900 <- which.min(abs(trial$pm2.5$value - 900))
  index_450 <- which.min(abs(trial$pm2.5$value - 450))</pre>
  time <- (trial pm2.5[index_450,] time - trial pm2.5[index_900,] time) / 60
  list(sensorbox=trial$sensorbox, group=trial$group, time=time)
})
pm2.5_decrease <- tibble(</pre>
  sensorbox=sapply(pm2.5_aggregated, function(xs) xs$sensorbox),
  group=sapply(pm2.5_aggregated, function(xs) xs$group),
  time=sapply(pm2.5_aggregated, function(xs) xs$time)
pm2.5_decrease$has_soil <- ifelse(</pre>
  pm2.5_decrease$group %in% c('soil', 'deadplant', 'plant'), 1, 0)
pm2.5_decrease$has_sa <- ifelse(</pre>
  pm2.5_decrease$group %in% c('deadplant', 'plant'), 1, 0)
pm2.5_decrease$has_life <- ifelse(</pre>
 pm2.5_decrease$group == 'plant', 1, 0)
```

3.2 Regression

```
pm2.5_decrease.lm <- lm(
   time~has_soil + has_sa + has_life + sensorbox, data=pm2.5_decrease)
summary(pm2.5_decrease.lm)</pre>
```

```
##
## Call:
## lm(formula = time ~ has_soil + has_sa + has_life + sensorbox,
       data = pm2.5_decrease)
##
## Residuals:
                      Median
        Min
                  10
                                    30
                                4.7758 15.1956
## -14.7500 -5.1403 -0.0528
##
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
                             2.345 37.827 < 2e-16 ***
                 88.707
## (Intercept)
                             2.708 -6.106 1.15e-07 ***
                -16.536
## has_soil
                             2.708 -3.473 0.00102 **
## has_sa
                 -9.406
## has_life
                             2.708
                                     2.955 0.00463 **
                  8.001
## sensorbox2b -20.957
                             2.345
                                    -8.936 3.13e-12 ***
               20.650
                                     8.806 5.05e-12 ***
## sensorbox3d
                             2.345
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 7.416 on 54 degrees of freedom
## Multiple R-squared: 0.8841, Adjusted R-squared: 0.8733
## F-statistic: 82.36 on 5 and 54 DF, p-value: < 2.2e-16
confint(pm2.5_decrease.lm)
##
                    2.5 %
                              97.5 %
## (Intercept) 84.005042 93.408292
## has_soil
               -21.964524 -11.106587
## has sa
               -14.834524 -3.976587
## has_life
                 2.572142 13.430080
## sensorbox2b -25.658292 -16.255042
## sensorbox3d 15.948375 25.351625
(confint(pm2.5_decrease.lm)[,2] - confint(pm2.5_decrease.lm)[,1]) / 2
## (Intercept)
                  has_soil
                                          has_life sensorbox2b sensorbox3d
                                has_sa
##
      4.701625
                  5.428969
                              5.428969
                                          5.428969
                                                       4.701625
                                                                   4.701625
      Repeat for PM_{10}
3.3
pm10_aggregated <- lapply(data_calibrated, function(trial) {</pre>
  index_900 <- which.min(abs(trial$pm10$value - 900))</pre>
  index_450 <- which.min(abs(trial$pm10$value - 450))</pre>
  time <- (trial pm10[index_450,] time - trial pm10[index_900,] time) / 60
  list(sensorbox=trial$sensorbox, group=trial$group, time=time)
})
pm10_decrease <- tibble(</pre>
  sensorbox=sapply(pm10_aggregated, function(xs) xs$sensorbox),
  group=sapply(pm10_aggregated, function(xs) xs$group),
  time=sapply(pm10_aggregated, function(xs) xs$time)
pm10_decrease$has_soil <- ifelse(</pre>
```

```
pm10_decrease$group %in% c('soil', 'deadplant', 'plant'), 1, 0)
pm10_decrease$has_sa <- ifelse(</pre>
  pm10_decrease$group %in% c('deadplant', 'plant'), 1, 0)
pm10_decrease$has_life <- ifelse(</pre>
  pm10_decrease$group == 'plant', 1, 0)
pm10_decrease.lm <- lm(time~has_soil + has_sa + has_life + sensorbox, data=pm10_decrease)
summary(pm10 decrease.lm)
##
## Call:
## lm(formula = time ~ has_soil + has_sa + has_life + sensorbox,
       data = pm10_decrease)
##
##
## Residuals:
       Min
                  1Q
                     Median
                                    3Q
                                            Max
## -11.3003 -4.3371 -0.0531
                                4.3937 10.4322
## Coefficients:
               Estimate Std. Error t value Pr(>|t|)
## (Intercept)
                77.568
                             1.898 40.877 < 2e-16 ***
## has soil
                -15.464
                             2.191 -7.058 3.32e-09 ***
                -7.873
                             2.191 -3.593 0.000707 ***
## has_sa
## has_life
                 6.672
                             2.191
                                    3.045 0.003592 **
                             1.898 -9.618 2.67e-13 ***
## sensorbox2b -18.251
## sensorbox3d 16.151
                             1.898
                                    8.511 1.49e-11 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
## Residual standard error: 6.001 on 54 degrees of freedom
## Multiple R-squared: 0.893, Adjusted R-squared: 0.8831
## F-statistic: 90.17 on 5 and 54 DF, p-value: < 2.2e-16
confint(pm10 decrease.lm)
##
                    2.5 %
                             97.5 %
## (Intercept) 73.763334 81.37222
## has_soil
              -19.857438 -11.07145
## has_sa
               -12.266327
                          -3.48034
## has_life
                 2.279229 11.06522
## sensorbox2b -22.055277 -14.44639
## sensorbox3d 12.346389 19.95528
(confint(pm10_decrease.lm)[,2] - confint(pm10_decrease.lm)[,1]) / 2
## (Intercept)
                 has_soil
                                has_sa
                                          has_life sensorbox2b sensorbox3d
##
      3.804444
                  4.392994
                              4.392994
                                          4.392994
                                                      3.804444
                                                                  3.804444
     Box Plot
3.4
library(ggpubr)
pm10_decrease$pm <- 'PM[10]'</pre>
pm2.5_decrease$pm <- 'PM[2.5]'
# monotone print friendly
```

```
ggboxplot(bind_rows(pm10_decrease, pm2.5_decrease),
  x='group', y='time', fill='sensorbox') +
  facet_grid(. ~ pm, labeller=label_parsed) +
 ylab(expression(atop("Time for PM = 900 "*mu*"g m"^-3*" to half (mins)", ""))) +
 xlab("\nControl/Treatment Group") +
  guides(fill=guide_legend(title='Box')) +
  scale_x_discrete(labels=c(
      'nothing'=expression(
       atop(NA, textstyle('Nothing'))),
      'soil'=expression(atop(
       atop(NA, textstyle('Substrate')), atop(textstyle('Only'), NA))),
      'deadplant'=expression(
        atop(atop(NA, textstyle('Dried')), atop(textstyle('Specimen'), NA))),
      'plant'=expression(
        atop(atop(NA, textstyle('Live')), atop(textstyle('Specimen'), NA))))) +
      scale_fill_discrete(
       palette=function(x) c('#f1a340', '#f7f7f7', '#998ec3'),
       labels=c('Box 1', 'Box 2', 'Box 3')) +
  theme_bw() +
  theme(legend.position="top")
```

ggsave('boxplot.eps', width=9, height=5)