

Lab8 PCA

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PCA of UK food data

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url, row.names = 1)

dim(x)
```

```
## [1] 17 4
```

Check data, preview the first six rows

```
head(x)
```

```
##           England Wales Scotland N.Ireland
## Cheese           105    103      103        66
## Carcass_meat      245    227      242       267
## Other_meat        685    803      750       586
## Fish              147    160      122        93
## Fats_and_oils      193    235      184       209
## Sugars             156    175      147       139
```

Fix the rownames:

```
#rownames(x) <- x[,1]
#x <- x[,-1]
#head(x)

#fixed the above by specifying the row names while reading the csv

#recheck the dimensions
dim(x)
```

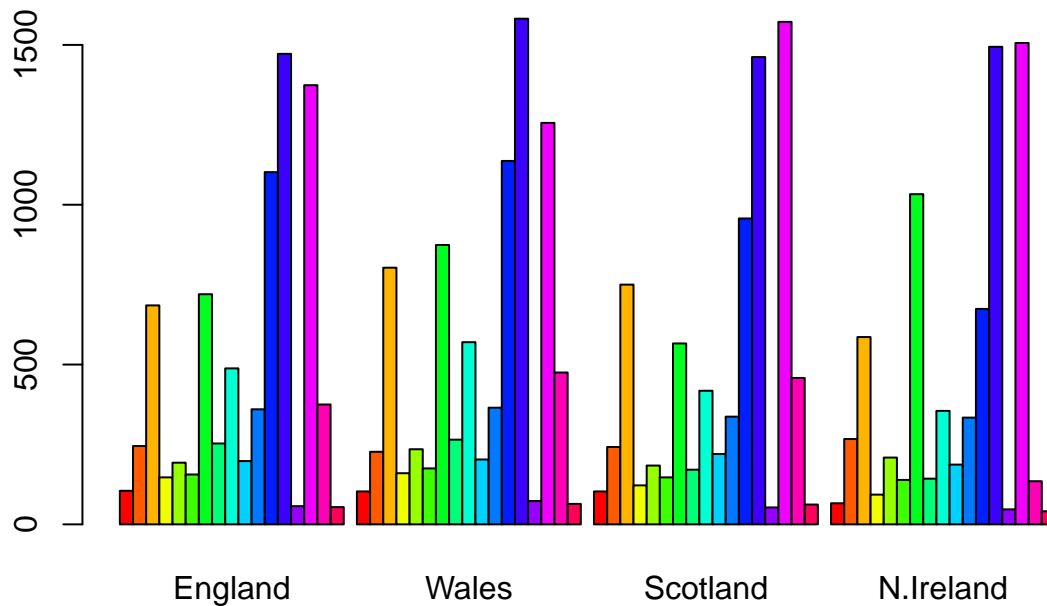
```
## [1] 17 4
```

Q2. Which approach to solving the ‘row-names problem’ mentioned above do you prefer and why? Is one approach more robust than another under certain circumstances?

I would prefer the second approach, which is to assign the row names when first reading the file, because if you run `x <- x[,-1]` multiple times it will keep removing columns which will mess with our data.

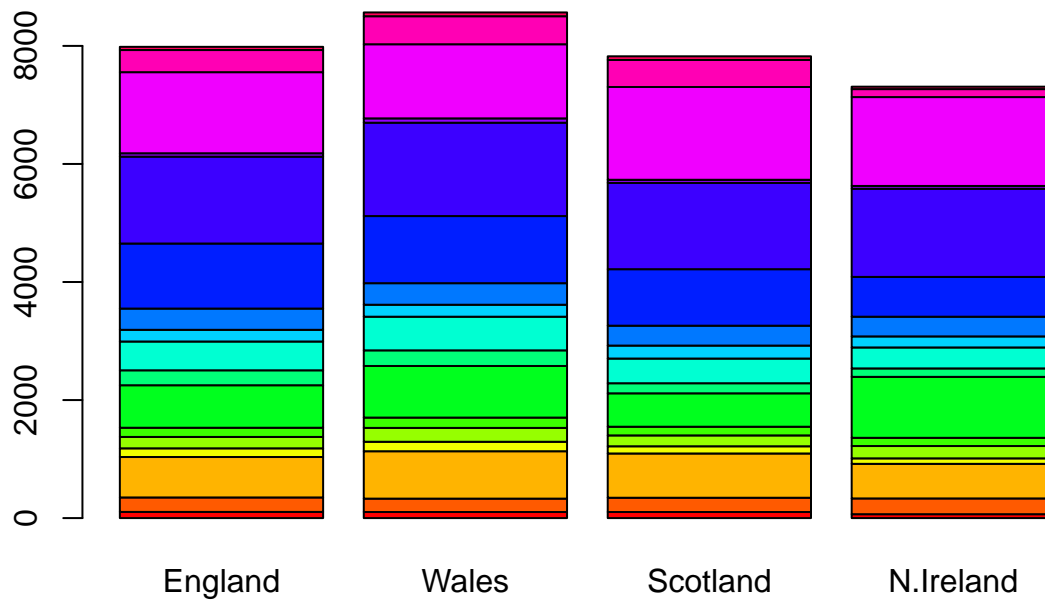
Plot the results:

```
barplot(as.matrix(x), beside=T, col=rainbow(nrow(x)))
```



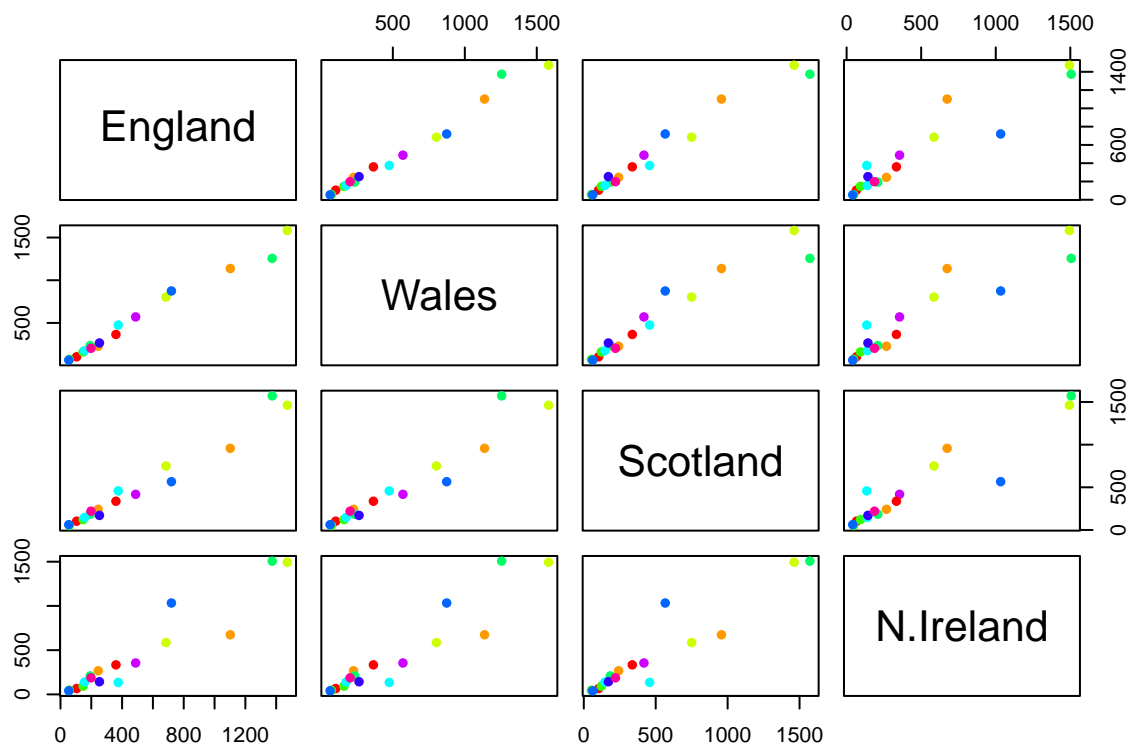
One can set the `beside` parameter to false in order to not display the results side by side.

```
barplot(as.matrix(x), beside=F, col=rainbow(nrow(x)))
```



Q5: Generating all pairwise plots may help somewhat. Can you make sense of the following code and resulting figure? What does it mean if a given point lies on the diagonal for a given plot?

```
pairs(x, col=rainbow(10), pch=16)
```



If a point aligns along the diagonal, it means that that value is similar for the two countries.

We use PCA to make sense of the data

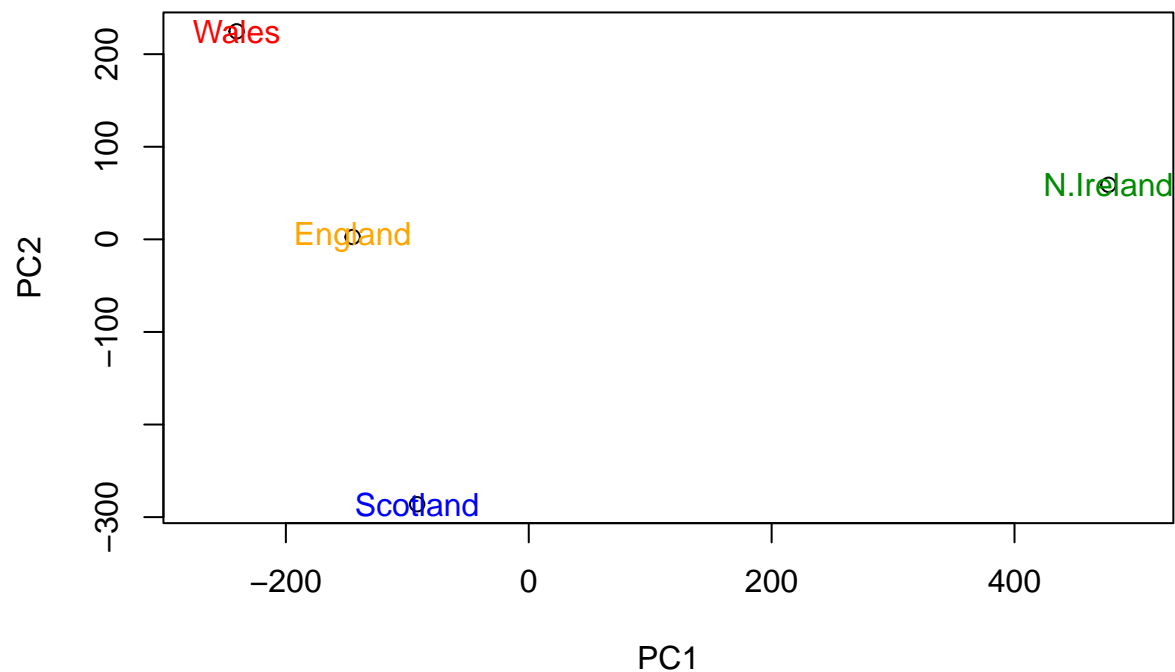
```
# Use the prcomp() PCA function
pca <- prcomp( t(x) )
summary(pca)
```

Importance of components:

##	PC1	PC2	PC3	PC4
## Standard deviation	324.1502	212.7478	73.87622	4.189e-14
## Proportion of Variance	0.6744	0.2905	0.03503	0.000e+00
## Cumulative Proportion	0.6744	0.9650	1.00000	1.000e+00

Generate a plot of PC1 vs PC2

```
# Plot PC1 vs PC2
plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500))
text(pca$x[,1], pca$x[,2], colnames(x), col=c("ORANGE", "RED", "BLUE", "GREEN4"))
```



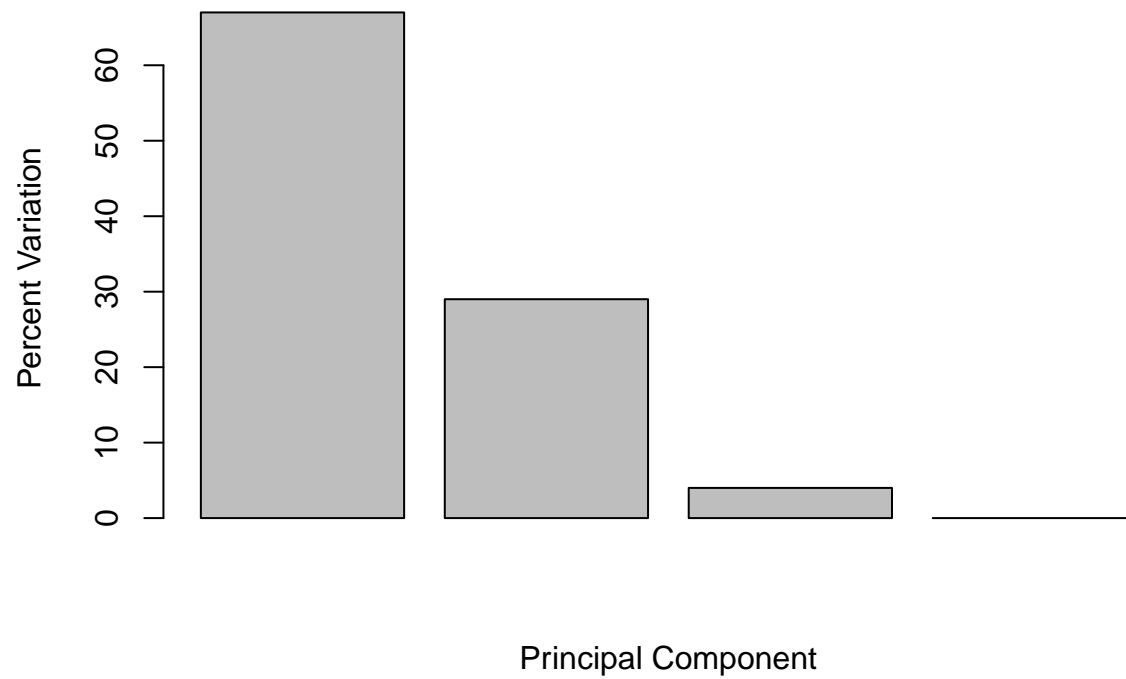
Calculate the variation within the original data that each PC accounts for (rounded up)

```
v <- round( pca$sdev^2/sum(pca$sdev^2) * 100 )
v
```

```
## [1] 67 29 4 0
```

Summarizing plot of the variances (visually representation of above info)

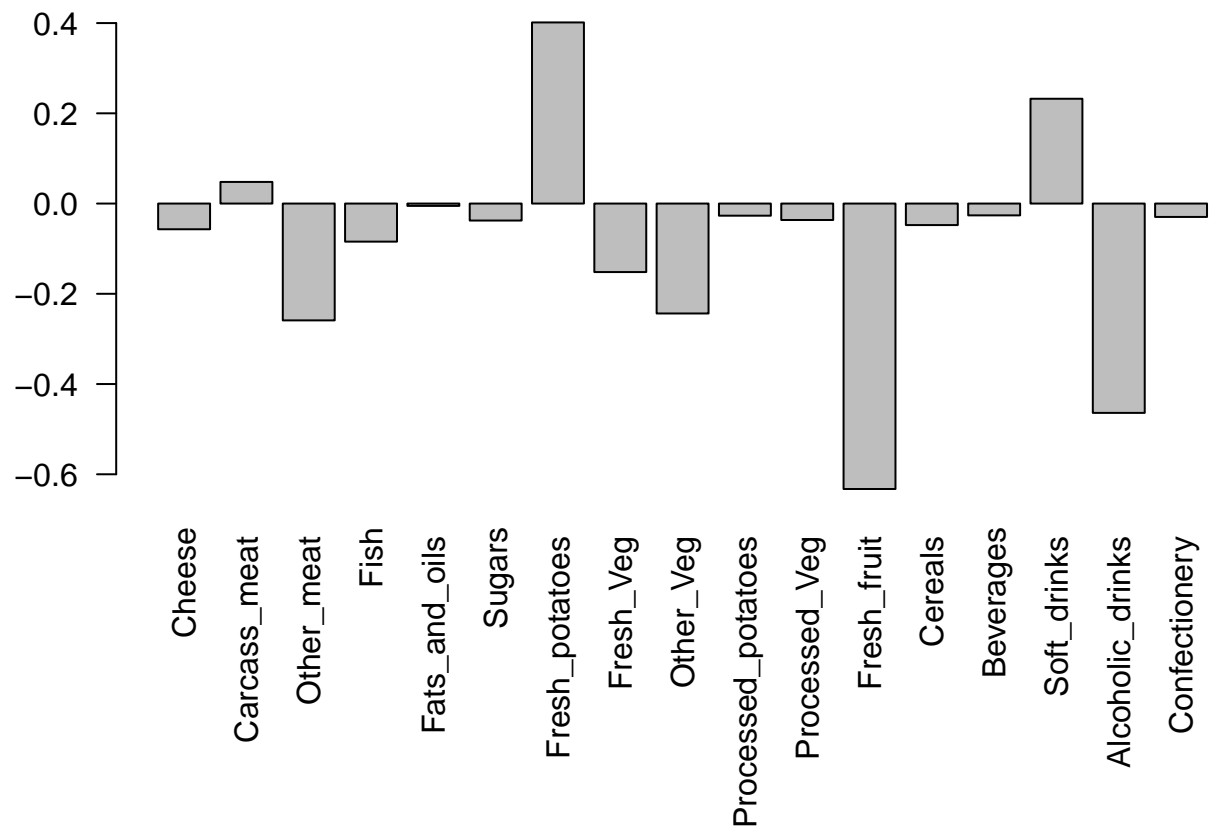
```
barplot(v, xlab="Principal Component", ylab="Percent Variation")
```



Finding and representing loading scores:

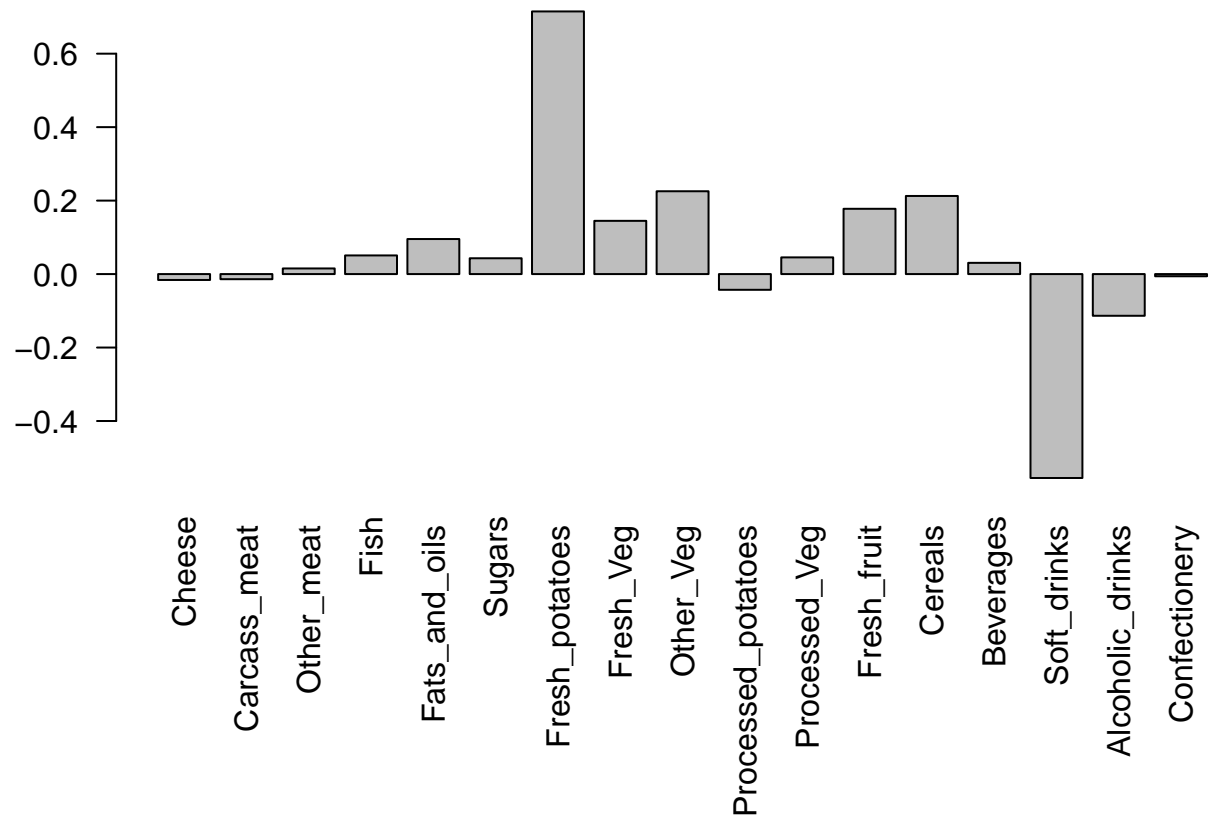
```
## Lets focus on PC1 as it accounts for > 90% of variance
par(mar=c(10, 3, 0.35, 0))

#Barplot for PC1
barplot( pca$rotation[,1], las=2 )
```



Q9: Generate a similar 'loadings plot' for PC2. What two food groups feature prominently and what does PC2 mainly tell us about?

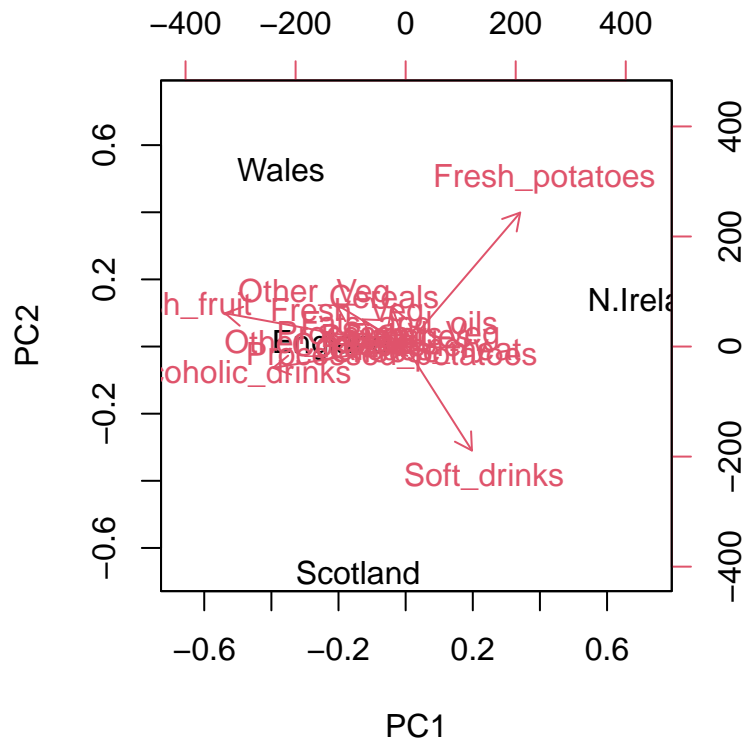
```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,2], las=2 )
```



The two prominent food groups are potatoes and soft drinks.

Representing data using a Biplot:

```
biplot(pca)
```

```
//Part 2// RNA-seq data
```

```
url2 <- "https://tinyurl.com/expression-CSV"
rna.data <- read.csv(url2, row.names=1)
head(rna.data)
```

##		wt1	wt2	wt3	wt4	wt5	ko1	ko2	ko3	ko4	ko5
##	gene1	439	458	408	429	420	90	88	86	90	93
##	gene2	219	200	204	210	187	427	423	434	433	426
##	gene3	1006	989	1030	1017	973	252	237	238	226	210
##	gene4	783	792	829	856	760	849	856	835	885	894
##	gene5	181	249	204	244	225	277	305	272	270	279
##	gene6	460	502	491	491	493	612	594	577	618	638

Q10: How many genes and samples are in this data set?

```
dim(rna.data)
```

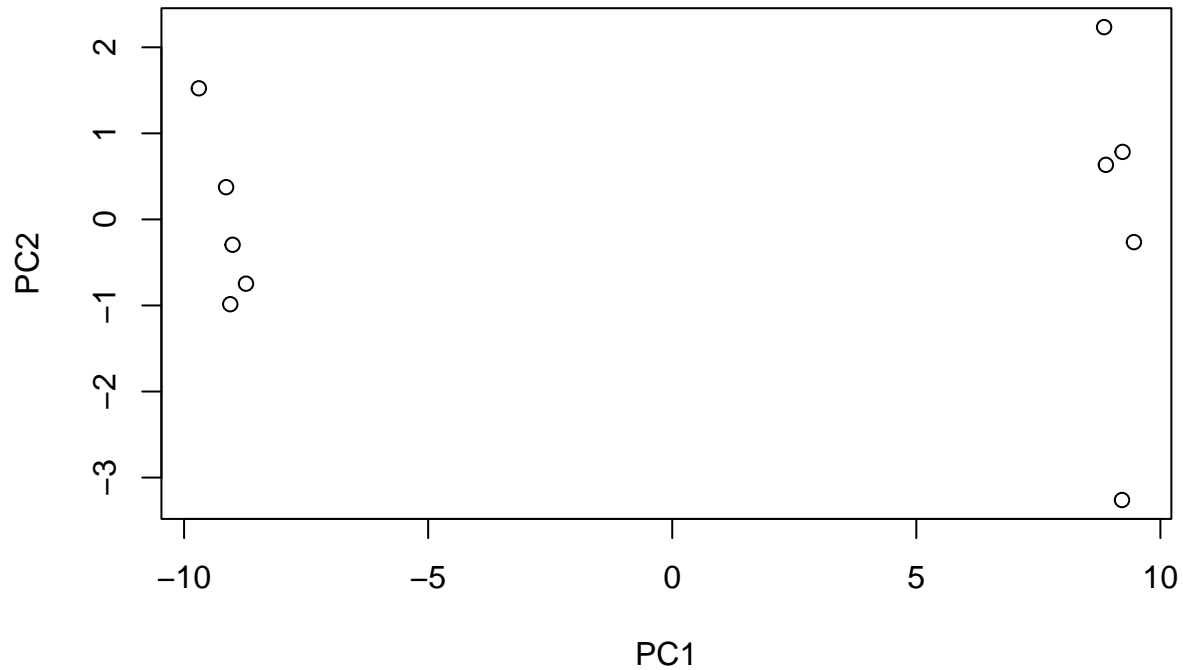
```
## [1] 100 10
```

We have 100 genes, and 10 samples.

Use PCA to analyze data

```
## Again we have to take the transpose of our data
pca <- prcomp(t(rna.data), scale=TRUE)
```

```
## Simple un polished plot of pc1 and pc2
plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2")
```



```
summary(pca)
```

```
## Importance of components:
##              PC1      PC2      PC3      PC4      PC5      PC6      PC7
## Standard deviation  9.6237 1.5198 1.05787 1.05203 0.88062 0.82545 0.80111
## Proportion of Variance 0.9262 0.0231 0.01119 0.01107 0.00775 0.00681 0.00642
## Cumulative Proportion 0.9262 0.9493 0.96045 0.97152 0.97928 0.98609 0.99251
##              PC8      PC9      PC10
## Standard deviation  0.62065 0.60342 3.348e-15
## Proportion of Variance 0.00385 0.00364 0.000e+00
## Cumulative Proportion 0.99636 1.00000 1.000e+00
```

Scree barplot to represent variance

```
plot(pca, main="Quick scree plot")
```

Quick scree plot



More in-depth Scree plot with our calculated values

```
## Variance captured per PC
```

```
pca.var <- pca$sdev^2
```

```
## Percent variance is often more informative to look at
```

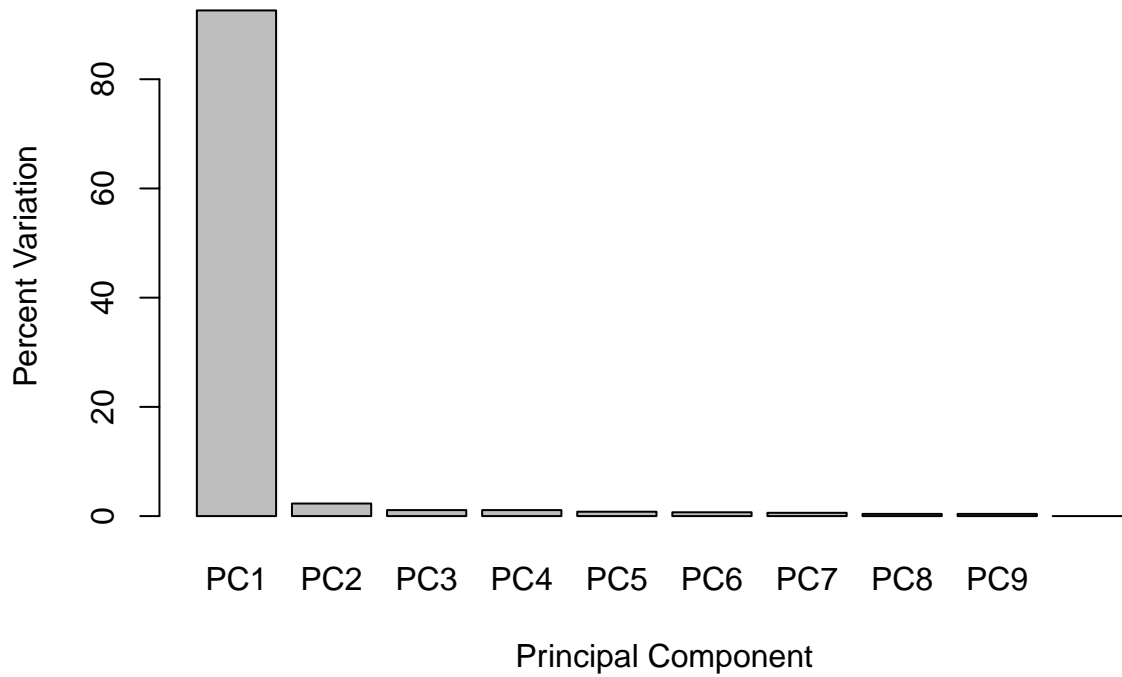
```
pca.var.per <- round(pca.var/sum(pca.var)*100, 1)
```

```
pca.var.per
```

```
## [1] 92.6 2.3 1.1 1.1 0.8 0.7 0.6 0.4 0.4 0.0
```

```
barplot(pca.var.per, main="Scree Plot",  
        names.arg = paste0("PC", 1:10),  
        xlab="Principal Component", ylab="Percent Variation")
```

Scree Plot

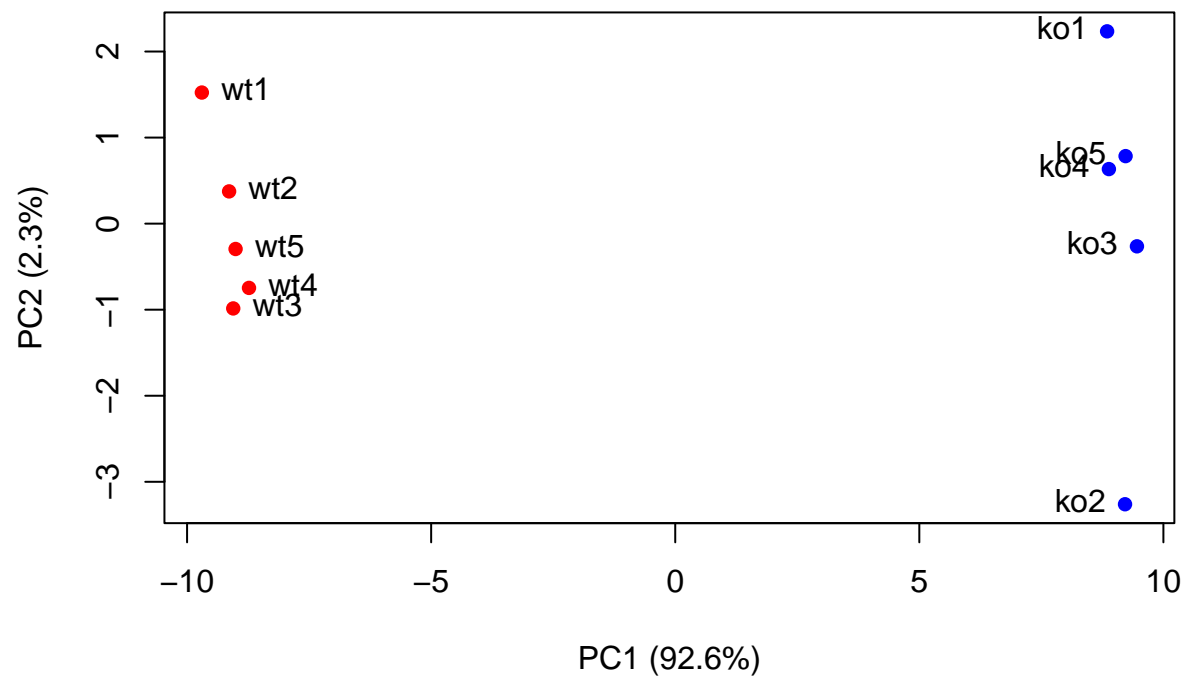


Add details to original PCA plot:

```
## A vector of colors for wt and ko samples
colvec <- colnames(rna.data)
colvec[grep("wt", colvec)] <- "red"
colvec[grep("ko", colvec)] <- "blue"

plot(pca$x[,1], pca$x[,2], col=colvec, pch=16,
     xlab=paste0("PC1 (", pca.var.per[1], "%)"),
     ylab=paste0("PC2 (", pca.var.per[2], "%)"))

text(pca$x[,1], pca$x[,2], labels = colnames(rna.data), pos=c(rep(4,5), rep(2,5)))
```

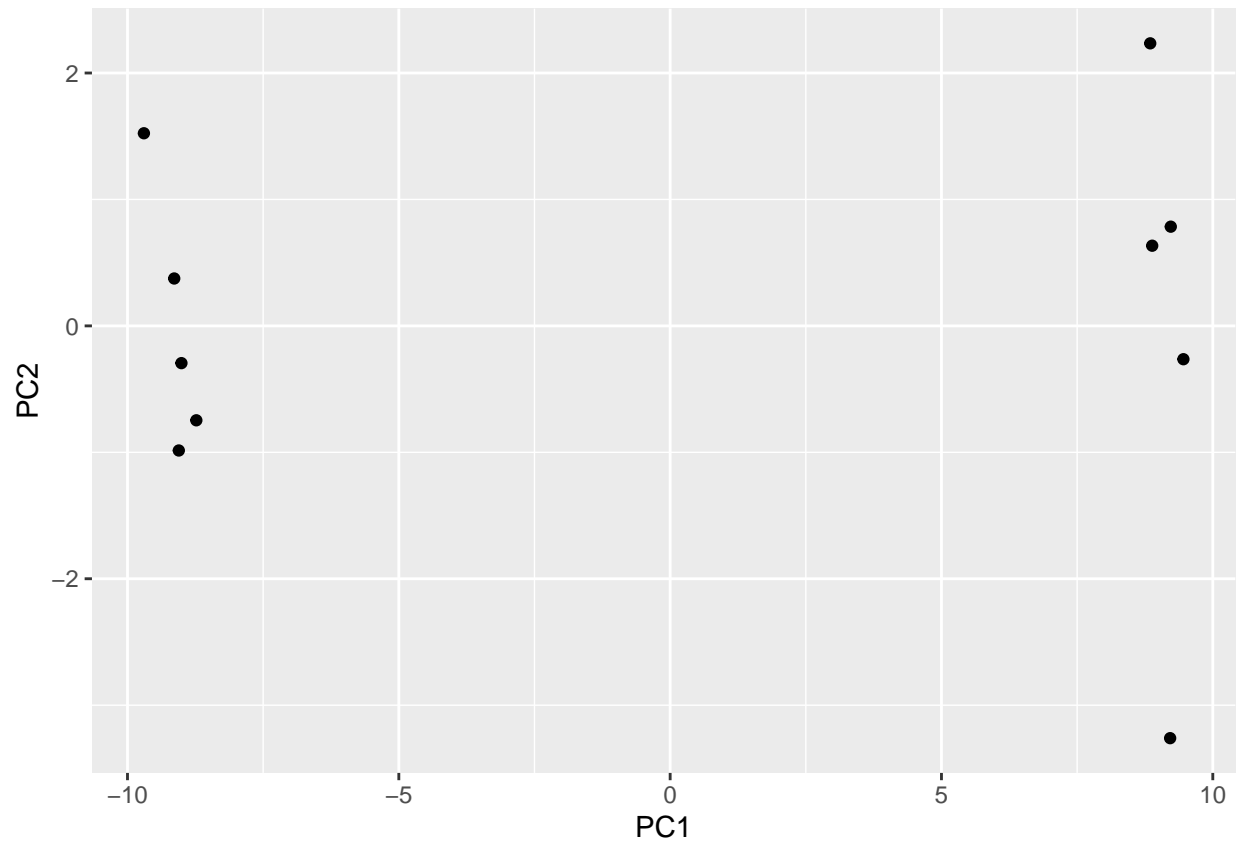


Building a similar plot using ggplot2

```
library(ggplot2)

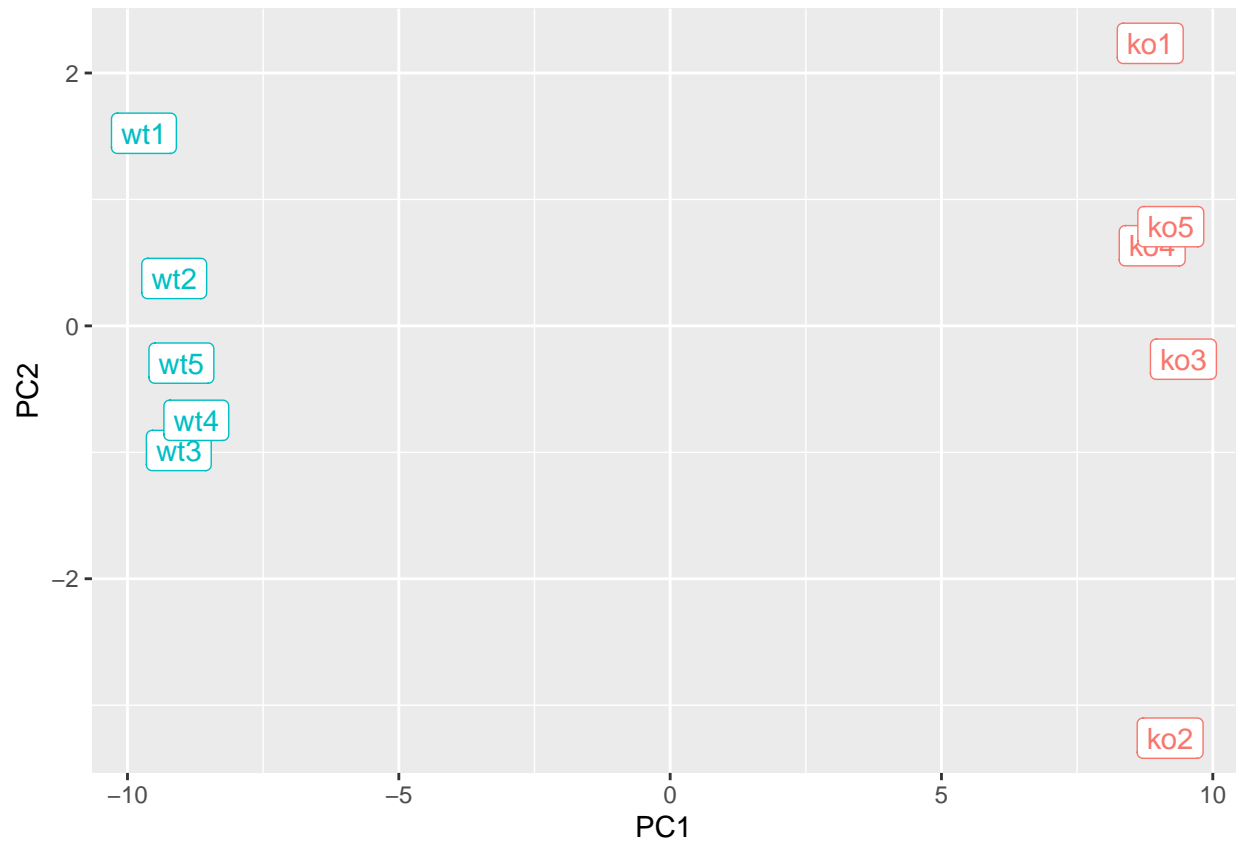
df <- as.data.frame(pca$x)

# Our first basic plot
ggplot(df) +
  aes(PC1, PC2) +
  geom_point()
```



```
# Add a 'wt' and 'ko' "condition" column
df$samples <- colnames(rna.data)
df$condition <- substr(colnames(rna.data),1,2)

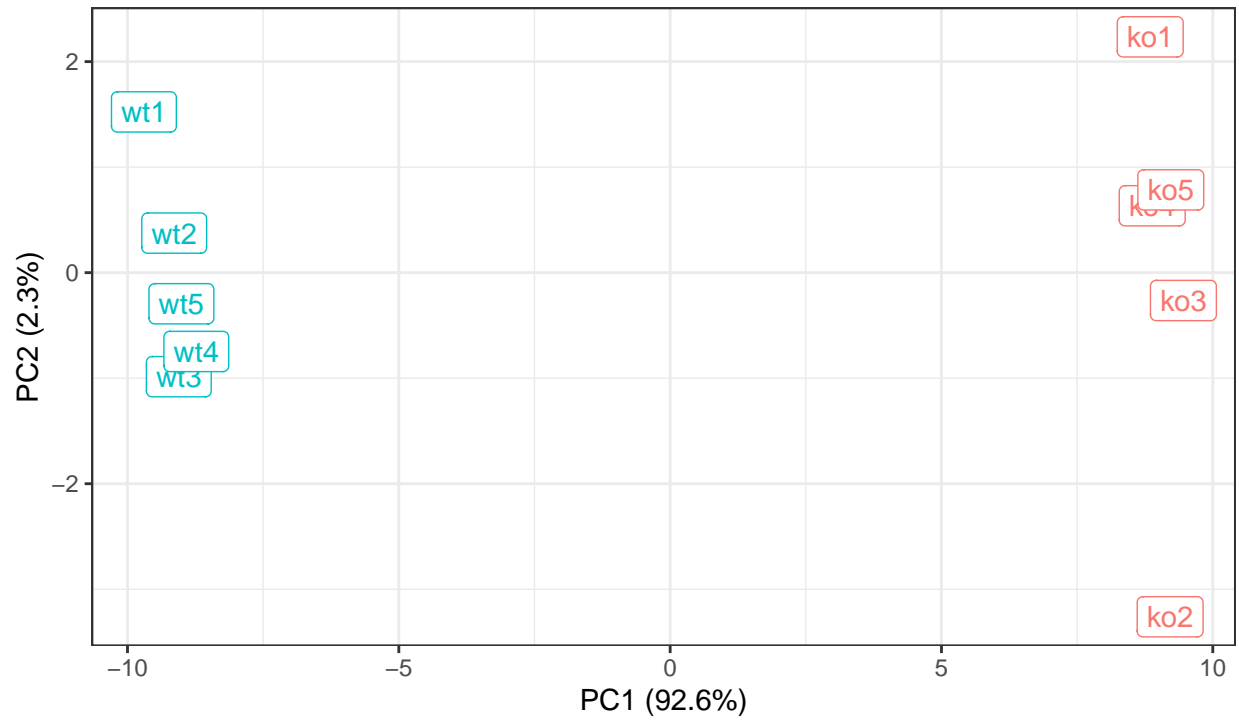
p <- ggplot(df) +
  aes(PC1, PC2, label=samples, col=condition) +
  geom_label(show.legend = FALSE)
p
```



```
p + labs(title="PCA of RNASeq Data",
  subtitle = "PC1 clealy seperates wild-type from knock-out samples",
  x=paste0("PC1 (", pca.var.per[1], "%)"),
  y=paste0("PC2 (", pca.var.per[2], "%)"),
  caption="BIMM143 example data") +
theme_bw()
```

PCA of RNASeq Data

PC1 clearly separates wild-type from knock-out samples



BIMM143 example data