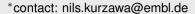
Bioconductor packages for analysis of Thermal Proteome Profiling data

Nils Kurzawa^{1,*}, Dorothee Childs^{1,2}, Holger Franken², Mikhail M. Savitski¹ & Wolfgang Huber¹

¹Genome Biology Unit, European Molecular Biology Laboratory

²Cellzome GmbH, GlaxoSmithKline





Thermal proteome profiling (TPP) is a mass spectrometry-based technology which was originally developed to identify drug (off) -targets on a proteome-wide scale. The assay is built on the principle of the cellular thermal shift assay which is that proteins inside living cells may be modulated in their thermal stability by ligand binding. This can be read out by shifts of melting profiles of respective proteins. Soon after TPP was established, a first Bioconductor package TPP was released, which implemented function to infer ligand-protein interactions using shifts of melting points. The realization that this procedure lacks sensitivity to detect proteins with e.g. very high thermal stability lead to the development of the NPARC package. Further, it was realized that TPP data can also inform on protein-protein interaction (PPI) dynamics. Another Bioconductor package Rtpca was thus implemented to accommodate respective analyses. Lastly, the experimental setup of TPP experiments was expanded to profile ligand dose and temperature ranges together to improve sensitivity. To accomodate the different data structure and different required statistical analysis the package TPP2D was created. Here, the different functionalities of these packages are presented.

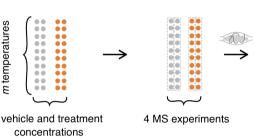
Thermal Proteome Profiling (TPP)

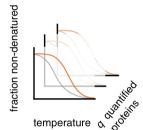
TPP over a temperature range (TPP-TR) [1]

Grow cells in presence and absence of ligand in duplicates

Heat treatment and extraction of remaining soluble proteins TMT labelling of tryptic peptides and LC-MS/MS analysis

Protein identification and quantification and melting curve comparison



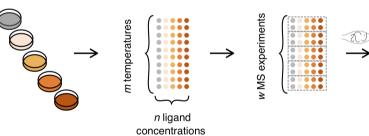


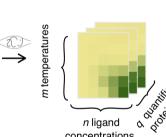
Ligand dose and temperature range TPP (2D-TPP) [2]

Grow cells with n different ligand concentrations

Heat treatment and extraction of remaining soluble proteins TMT labelling of tryptic peptides and LC-MS/MS analysis

Protein Identification and Quantification

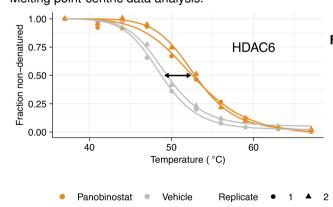




concentrations

TPP-TR analysis with TPP [3]

Melting point-centric data analysis.

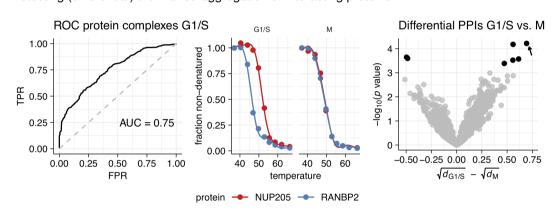


Features of the TPP package:

- TPP-TR data import into ExpressionSet structure
- data normalization and melting curve fitting, detection of melting point shifts
- · analysis of isothermal concentartion range TPP experiments

Detecting differential PPIs with Rtpca [5]

Detecting (differential) thermal co-aggregation of interacting proteins.

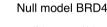


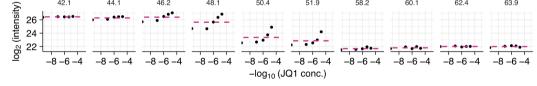
Features of the Rtpca package:

- test for co-aggregation of annotated protein-protein interactors (PPI), compute receiver operating characteristics (ROC)
- perform differential testing to find changes in PPIs between different conditions

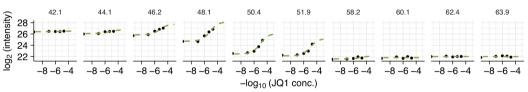
2D-TPP data analysis with TPP2D [6]

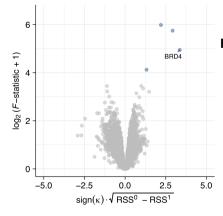
FDR-controlled detection of ligand-protein interactions from 2D thermal profiles.





Alternative model BRD4





Features of the TPP2D package:

- hypothesis test on curves: intercept model vs. dose-response model contrained across tempera tures
- bootstrapping approach to calibrate F-statistic in terms of false discovery-rate (FDR)
- · several functions for data visualization

TPP-TR analysis with NPARC [4]

Nonparametric analysis of response curves comparing exploiting all measurements.

Features of the NPARC package:

- light-weight package for performing sensitive differential melting curve analysis
- nested model comparison: one sigmoidal across conditions vs. sigmoidals per condition
- *F*-statistic based on explained variance of each model
- method for calibration of retrieved F-statistics

References

- [1] M. M. Savitski, F. B. M. Reinhard, H. Franken, T. Werner, M. F. Savitski, D. Eberhard, D. Martinez Molina, R. Jafari, R. B. Dovega, S. Klaeger, B. Kuster, P. Nordlund, M. Bantscheff, and G. Drewes, "Tracking cancer drugs in living cells by thermal profiling of the proteome.," *Science*, vol. 346, p. 1255784, oct 2014.
- . Becher, T. Werner, C. Doce, E. A. Zaal, I. Tögel, C. A. Khan, A. Rueger, M. Muelbaier, E. Salzer, C. R. Berkers, P. F. Fitzpatrick, M. Bantscheff, and M. M. Savitski, "Thermal profiling reveals phenylalanine hydroxylase as an off-target of panobinostat.," Nat Chem
- Biol, vol. 12, pp. 908-910, nov 2016. H. Franken, T. Mathieson, D. Childs, G. M. A. Sweetman, T. Werner, I. Tögel, C. Doce, S. Gade, M. Bantscheff, G. Drewes, F. B. M. Reinhard, W. Huber, and M. M. Savitski, "Thermal proteome profiling for unbiased identification of direct and indirect drug targets
- using multiplexed quantitative mass spectrometry.," Nat Protoc, vol. 10, pp. 1567-1593, oct 2015. [4] D. Childs, K. Bach, H. Franken, S. Anders, N. Kurzawa, M. Bantscheff, M. Savitski, and W. Huber, "Non-parametric analysis of thermal
- proteome profiles reveals novel drug-binding proteins.," *Mol Cell Proteomics*, oct 2019.

 [5] N. Kurzawa, A. Mateus, and M. M. Savitski, "Rtpca: an r package for differential thermal proximity coaggregation analysis.," *Bioinfor-*
- [6] N. Kurzawa, I. Becher, S. Sridharan, H. Franken, A. Mateus, S. Anders, M. Bantscheff, W. Huber, and M. M. Savitski, "A computational method for detection of ligand-binding proteins from dose range thermal proteome profiles.," Nat Commun, vol. 11, p. 5783, nov 2020.

This work was funded by EMBL and GSK. The poster is available at https://github.com/nkurzaw/EuroBioc2020Poster

