

Cell Growth Model - v0.8

Table of Contents

Model Overview	2
Features	2
Version Modifications	2
Processes Models.....	3
Transcription	3
mRNA degradation	5
Translation Initiation Complex formation	6
Translation Elongation Complex formation	7
Translation Protein Synthesis	9
Ribosome Assembly	10
Energy production.....	11
Energy consumption	12
Phenotypes.....	14
Cell size	14
Growth rate	15

Model Overview

Features

Phenotypes

- Cell partition in 3 biomolecular species classes:
 - Metabolic
 - Ribosome-associated
 - Housekeeping
- Cell mass
 - Phenomenological model/Logistic function shaped
 - Variable mass dependent on intracellular energy levels
- Growth rate
 - Function of protein mass growth rate & cell mass size

Cell processes

- Translation models:
 - Initiation Complex formation
 - Elongation Complex maturation
 - Protein Synthesis
- Elongation complex maturation model:
 - Allows polysomes formation implicitly
 - Maturation dependent on cell energy levels
- Energy production & utilisation models
 - Simple one-step energy production model
 - Energy Utilised by Protein Synthesis & Elongation Complex Maturation
- Ribosome Assembly model
 - Ribosome formation from ribosome-associated proteins
 - No rRNA species involved
 - Allows for realistic numbers of ribosomes in cell

Version Modifications

- Derived from model v0.7
- Ribosome Assembly model
- Removed rRNA species

Processes Models

Transcription

Assumptions			
<ul style="list-style-type: none">Transcription is an energy dependent process, but it is not made to consume any energy as its consumption rate is small compared to the translation processes (Weiße et al., 2015)The threshold value for half-maximal activation of transcription occurs at higher intracellular energy levels for ribosomal protein gene expression, as it has been demonstrated that intracellular effector molecules inhibit transcription of ribosome-associated genes at poor intracellular metabolic environment (Barker et al., 2001; Lemke et al., 2011).			
Reaction			
Forward rate	$wx \cdot \text{mod_fcn_a} \rightarrow m_x$		
Reverse rate			
Energy modulating function	$\text{mod_fnc_a} = \left(\frac{a}{\text{theta} + a} \right)$ theta for hsk & met = thetax theta for rib = thetar		
Species			
m_x	RBS of class x = {rib, met, hsk}		
a	Intracellular energy molecule		
Parameters	Description	Value	Units
wx	transcription rate		molecules/minutes
Modifications to basic Model			
Assumptions			
<ul style="list-style-type: none">For hsk class:<ul style="list-style-type: none">For rbs/mRNA of housekeeping class, the transcription rate is inhibited by a modulation function based on the number of hsk proteins in the cell, as previous research suggests the existence of an autoregulation mechanism so that their relative proteome abundance is maintained across different growth conditions (Hwa et al., 2010)			
Modulating function (hsk class)	$\text{mod_fnc_txhsk} = \left(\frac{1}{1 + \left(\frac{p_{rib}}{K_{repr}} \right)} \right)$		
Parameters/Species	Description	Value	Units
p_rib	ribosomal proteins		molecules
Krepr	repression threshold		molecules
Fluxes			
$d(m_{rib})/dt = +wr \cdot \text{mod_fcn_a}$			

$$d(m_met)/dt = +wr* mod_fcn_a* mod_fnc_txhsk$$

$$d(m_hsk)/dt = +wr*mod_fcn_a$$

mRNA degradation

Assumptions			
<ul style="list-style-type: none">Only the free form of the mRNA is degraded (RBS)Degradation rate is the same for all mRNA classes			
Reaction			
Forward rate	$m_x \xrightarrow{dm} \text{null}$		
Reverse rate			
Species			
m_x	RBS of class x = {rib, met, hsk}		
Parameters	Description	Value	Units
dm	mRNA degradation rate		1/minute
Fluxes			
$d(m_{\text{rib}})/dt = -dm * m_{\text{rib}}$			
$d(m_{\text{met}})/dt = -dm * m_{\text{met}}$			
$d(m_{\text{hsk}})/dt = -dm * m_{\text{hsk}}$			

Translation Initiation Complex formation

Assumptions			
<ul style="list-style-type: none">The association rate constant between ribosomes and rbs_mRNAs is variable, while all classes exhibit the same dissociation kinetics. This simplification does not represent reality as experimental observation show both constant values can vary within an interval of 3 orders of magnitude (Gualerzi & Pon, 2015) but its implementation was necessary to reduce the search space and the number of dimensions in parameter fitting.			
Reaction			
Forward rate	$m_x + \text{ribo} \xrightleftharpoons[ku]{kb_x} ic_x$		
Reverse rate			
Species			
m_x	RBS of class x = {rib, met, hsk}		
ribo	Ribosomes		
ic_x	Translation Initiation complex of class x = {rib, met, hsk}		
Parameters	Description	Value	Units
kb_x	RBS/ribosome association rate		1/(molecules*minutes)
ku	Initiation Complex dissociation rate		1/minute
Modifications to basic Model			
Assumptions			
<ul style="list-style-type: none">For rib class:<ul style="list-style-type: none">The association rate of rbs_mRNA with ribosome for the formation of ic_rib is inhibited by a modulating function based on the amount of free ribosomal protein molecules in the cell, as negative feedback regulation at translation initiation stage is well established for the control of ribosomal protein expression (Nomura et al., 1980)			
Modulating function (rib class)	$\text{mod_fnc} = \left(\frac{1}{1 + \left(\frac{p_rib}{Krepr} \right)} \right)$		
Parameters/Species	Description	Value	Units
p_rib	ribosomal proteins		molecules
Krepr	repression threshold		molecules
Fluxes			
$\begin{aligned} d(ic_rib)/dt &= +kb_ribo * ribo * r_rib * (1 / (1 + (p_rib / Krepr))) \\ &\quad -ku*ic_rib \\ \\ d(ic_met)/dt &= +kb_met * ribo * r_cat \\ &\quad -ku*ic_cat \\ \\ d(ic_hsk)/dt &= +kb_others*ribo * r_hsk \\ &\quad -ku*ic_hsk \end{aligned}$			

Translation Elongation Complex formation

Assumptions			
<ul style="list-style-type: none">The maturation rate of IC to EC is energy dependent with an activating modulating function that depends on the cell energy levels. The modulating function half-activation threshold is set at the same value as that of translation elongation process. This parameter value can be independently modified but only towards a higher value to prevent ribosome overcrowding on the mRNA molecule.1 energy molecule is consumed per maturation event a GTP molecule is utilised by IF2 in the pathway for 70SIC formation (Gualerzi & Pon, 2015).			
Reaction			
Forward rate	$ic_x + a \xrightarrow{kc * mod_fcn_gamma} ec_x + m_x$		
Reverse rate			
Energy modulating function	$mod_fnc_gamma = \left(\frac{a}{Kgamma + a} \right)$		
Species			
ic_x	initiation complex of class x = {rib, met, hsk}		
ec_x	elongation complex of class x = {rib, met, hsk}		
m_x	RBS of class x = {rib, met, hsk}		
a	Intracellular energy molecule		
Parameters	Description	Value	Units
kc	rate constant of IC maturation		1/minutes
Kgamma	Threshold of half-maximal transpeptidation rate (based on energy levels)		molecules
Fluxes			
$d(ic_rib)/dt = -ic_rib * kc * mod_fcn_gamma$			
$d(ic_met)/dt = -ic_met * kc * mod_fcn_gamma$			
$d(ic_hsk)/dt = -ic_hsk * kc * mod_fcn_gamma$			
$d(a)/dt = -ic_rib * kc * mod_fcn_gamma - ic_met * kc * mod_fcn_gamma - ic_hsk * kc * mod_fcn_gamma$			
$d(ec_rib)/dt = +ic_rib * kc * mod_fcn_gamma$			
$d(ec_met)/dt = +ic_met * kc * mod_fcn_gamma$			

$$d(ec_hsk)/dt = +ic_hsk * kc * mod_fcn_gamma$$

$$d(m_rib)/dt = +ic_rib * kc * mod_fcn_gamma$$

$$d(m_met)/dt = +ic_met * kc * mod_fcn_gamma$$

$$d(m_hsk)/dt = +ic_hsk * kc * mod_fcn_gamma$$

Translation Protein Synthesis

Assumptions			
<ul style="list-style-type: none">Simplified translation elongation model developed in (Weiße et al., 2015). In short, the net rate is dependent on the amino acid length of the protein molecule and is controlled by a modulating function that is activated by increased cellular energy levels.			
Reaction			
Forward rate	$ec_x + a \cdot lenX \xrightarrow{gmax \cdot mod_fcn_gamma} p_x$		
Reverse rate			
Energy modulating function	$mod_fnc_gamma = \left(\frac{a}{Kgamma + a} \right)$		
Species			
ec_x	elongation complex of class x = {rib, met, hsk}		
a	Intracellular energy mollecule		
p_x	protein of class x = {rib, met, hsk}		
Parameters	Description	Value	Units
lenX {lenO,lenR,lenC}	Protein length {p_hsk, p_rib, p_met}		amino acids
Kgamma	Threshold of half-maximal transpeptidation rate (based on energy levels)		molecules
Fluxes			
$d(ec_rib)/dt = -ec_rib \cdot gmax / lenR \cdot mod_fcn_gamma$			
$d(ec_met)/dt = -ec_met \cdot gmax / lenC \cdot mod_fcn_gamma$			
$d(ec_hsk)/dt = -ec_hsk \cdot gmax / lenO \cdot mod_fcn_gamma$			
$d(a)/dt = -ec_rib \cdot gmax \cdot mod_fcn_gamma - ec_met \cdot gmax \cdot mod_fcn_gamma - ec_hsk \cdot gmax \cdot mod_fcn_gamma$			
$d(p_rib)/dt = + ec_rib \cdot gmax / lenR \cdot mod_fcn_gamma$			
$d(p_met)/dt = + ec_met \cdot gmax / lenC \cdot mod_fcn_gamma$			
$d(p_hsk)/dt = +ec_hsk \cdot gmax / lenO \cdot mod_fcn_gamma$			

Ribosome Assembly

Assumptions			
<ul style="list-style-type: none">• Ribosomal proteins instantaneously form sets of ribosomal proteins of size equal to the total amino acid mass of an assembled ribosome• Ribosome Assembly is a single association event where a ribosomal proteins set forms an assembled ribosome			
Reaction			
Forward rate	$p_rib_set \xrightarrow{k_form} ribo$		
Reverse rate			
Species			
p_rib p_rib_set ribo	Free ribosomal proteins $p_rib_set = \text{ribosomal proteins} / (\text{lenRibo}/\text{lenR})$ Ribosomes		
Parameters	Description	Value	Units
lenRibo	Size of ribosome with all its ribosomal proteins		aa
lenR	Size of ribosomal protein		aa
k_form	Rate constant for ribosome assembly		molecules/minute
Fluxes			
$d(rib)/dt = -k_form * [p_rib_set] * (\text{lenRibo}/\text{lenR})$			
$d(Ribo)/dt = +k_form * [p_rib_set]$			

Energy production

Assumptions			
<ul style="list-style-type: none">Energy production is limited by a single bottleneck enzyme in the metabolic pathway and is modelled based on Michaelis-Menten reaction rate modelEfficiency of energy generation is dictate by the quality of the media (ns). The ns variable linearly scales the effectiveness by which energy molecules are produced by the metabolic sector. This simplification has also been used by others (Weiße et al., 2015) and is necessary as the true relationship between media composition/quality and energy molecules production would be very hard to calculateThe production rate flux is determine by protein molecules that are able to carry out a series of chemical reactions that facilitate the conversion of an extracellular nutrient molecule to an intracellular energy molecule. Thus, the Vmax, Km, and ns values are not meant to represent any “real” values but adjustable quantities that facilitates the devotion of amino acid mass to the metabolic proteome sector for the needs of resources allocations			
Reaction			
Forward rate	$ns* ((Vmax *s0)/(Km + s0))$		
Reverse rate	$p_met \rightarrow a + met$		
Species			
a	Intracellular energy		
p_met	Proteins metabolic class		
Parameters	Description	Value	Units
ns	Nutrient quality		No units
Vmax	Catalytic rate of nutrient utilisation		1/minute
s0	Extracellular concentration of nutrients		molecules
Km	Half-maximal threshold of nutrient levels utilisation		molecules
Fluxes			
$d(a)/dt \quad = +p_met * ns* ((Vmax *s0)/(Km + s0))$			

Energy consumption

Assumptions			
<ul style="list-style-type: none">One energy molecule is consumed by each Elongation Complex maturation eventOne energy molecule is consumed by each transpeptidation event in protein synthesis, where the consumption of each protein synthesis event is determined by the aa length of the synthesized protein			
Process 1			
Forward rate	$ic_x + a \xrightarrow{kc * mod_fcn_gamma} ec_x + m_x$		
Reverse rate			
Energy modulating function	$mod_fnc_gamma = \left(\frac{a}{Kgamma + a} \right)$		
Species			
ic_x	initiation complex of class x = {rib, met, hsk}		
ec_x	elongation complex of class x = {rib, met, hsk}		
m_x	RBS of class x = {rib, met, hsk}		
a	Intracellular energy molecule		
Parameters	Description	Value	Units
kc	rate constant of EC formation from IC		1/minute
Kgamma	transpeptidation elongation rate threshold for translation based on energy levels		molecules
Process 2			
Forward rate	$ec_x + a * lenX \xrightarrow{gmax * mod_fcn_gamma} p_x$		
Reverse rate			
Energy modulating function	$mod_fnc_gamma = \left(\frac{a}{Kgamma + a} \right)$		
Species			
ec_x	elongation complex of class x = {rib, met, hsk}		
a	Intracellular energy molecule		
p_x	protein of class x = {rib, met, hsk}		

Parameters	Description	Value	Units
lenX {lenO,lenR,lenC}	Protein length {p_hsk, p_rib, p_met}		1/minute
Kgamma	Threshold of half-maximal transpeptidation rate (based on energy levels)		molecules
Fluxes			
tinitrate = - (ic_rib + ic_met + ic_hsk) * kc * mod_fcn_gamma ttrate = - (ec_rib + ec_met + ec_hsk) * gmax * mod_fcn_gamma d(a)/dt = - tinitrate - ttrate			

Phenotypes

Cell size

Assumptions			
<ul style="list-style-type: none">• Phenomenological model based on a logistic function• Cell size is increases with increasing availability of intracellular energy (metabolic precursors)• Cell size is equal to an intracellular amino acid count, while other biomolecules are not considered (ex. Lipids, carbohydrates, DNA, RNA etc). This is because the model here aims to describe the competition between various cellular processes for proteome space and protein based biomolecules.• facilitates the dilution of intracellular species and redistribution of cellular content according to the reaction fluxes.			
Phenotype Equation			
cell size	$\text{minimal_mass} + \left(\frac{\text{max_inf}}{1 + \exp(- \text{inflation_gradient} * (a - \text{mid_inf}))} \right)$		
Species			
a	Intracellular energy molecule		
Parameters	Description	Value	Units
min_mass	Minimal cell mass that is allowed for the cell to assume even in the absence of intracellular energy		amino acids
max_inf	Maximum inflation of cell size upon high levels of cell energy		amino acids
mid_inf	Energy levels inflection point of half maximal cell size inflation		molecules

Growth rate

Assumptions			
<ul style="list-style-type: none">• Growth of the system is defined as the number of transpeptidation (amino acid incorporation in protein molecules) events per unit of time.• Normalised to 1 cell unit set by cell mass variable.• facilitates the dilution of intracellular species and redistribution of cellular content according to the reaction fluxes.			
Phenotype Equation			
Cell growth rate	$\lambda_{\text{am}} = \text{ttrate} / \text{cell size}$ <p>where:</p> $\text{ttrate} = (\text{ec_rib} + \text{ec_met} + \text{ec_hsk}) * \text{gmax} * \text{mod_fcn_gamma}$		
Species			
ec_x	Elongation complexes of class x = {rib, met, hsk}		
Parameters	Description	Value	Units
Kgamma	Threshold of half-maximal transpeptidation rate (based on energy levels)		molecules