

Working Title Natasha Hopkins

Stage 3 Project for Master of Biology (MBiol)University of York, UK

Project DirectorDr. Richard Maguire

Examination Date 18 April, 2022

Contents

Introduction	1
Hypothesis	2
Methods	2
Orbital Shaker	2
mRNA Isolation and qPCR	2
Statistical Analysis	3
Results	3
Discussion	3
Summarise Method & Results	3
Inhibitors & Axin	3
Unreliable Data	3
Refine Methods	3
Future Plans	5
Acknowledgements	5
References	5

🛮 🐧 / ith diam quis enim lobortis scelerisque fermentum dui faucibus in ornare quam viverra orci sagittis eu volutpat odio facilisis mauris sit amet massa vitae tortor condimentum lacinia quis vel eros donec ac odio tempor orci dapibus ultrices in iaculis nunc sed augue lacus viverra vitae congue eu consequat ac felis donec et odio pellentesque diam volutpat commodo sed egestas egestas fringilla phasellus faucibus scelerisque eleifend donec pretium vulputate sapien nec sagittis aliquam malesuada bibendum arcu vitae elementum curabitur vitae nunc sed velit dignissim sodales ut eu sem integer vitae justo eget magna fermentum iaculis eu non diam phasellus vestibulum lorem sed risus ultricies tristique nulla aliquet enim tortor at auctor urna nunc id cursus metus aliquam eleifend mi in nulla posuere sollicitudin aliquam ultrices sagittis orci a scelerisque purus semper eget duis at tellus at urna condimentum mattis pellentesque id nibh tortor id aliquet lectus proin nibh nisl condimentum id venenatis a condimentum vitae sapien pellentesque habitant morbi tristique senectus et netus et malesuada fames ac turpis egestas sed tempus urna et pharetra pharetra massa massa ultricies mi quis hendrerit dolor magna eget est lorem ipsum dolor sit amet consectetur adipiscing elit pellentesque habitant morbi tristique senectus et netus et malesuada fames ac turpis egestas integer eget aliquet nibh praesent tristique magna sit amet purus gravida quis blandit turpis cursus in hac habitasse platea dictumst quisque sagittis purus sit amet volutpat consequat mauris nunc congue nisi vitae suscipit tellus mauris a diam maecenas sed enim ut sem viverra aliquet eget sit.

Key Words: Atherosclerosis • Wnt/βeta-catenin Signalling Pathway • Shear Stress • Orbital Shaker • Angiopoietin-2 • Thrombospondin-1

(250 Words)

6

14

15

16

17

20

21 22

24

27

31

33

34

35

36

37

39

40

41

Introduction

Atherosclerosis is an chronic inflammatory disease characterised by the formation of arterial plaques. Haemodynamic shear stress has been identified as a modulator of site specificity in atherosclerosis, which occurs preferentially in regions exposed to low, oscillatory shear stress (Stone et al., 2007). Whereas areas of high, laminar shear stress is atheroprotective (Timmins et al., 2017). Shear stress is an important factor in regulating gene expression in vascular endothelial cells (Ni et al., 2010), which is though to contribute to the susceptibility of plaque formation in atheroprone sites. Multiple omics studies have implicated variations in flow with the regulation of developmental signalling pathways in atherosclerosis, including the Wnt Pathway (Souilhol et al., 2019; Gelfand et al., 2011).

Wnt is an evolutionarily conserved pathway with a critical role in axis patterning during embryonic development. In the absence of Wnt, axin forms a destruction complex with glycogen synthase kinase 3β (GSK-3) and adenomatous polyposis coli (APC), which phosphorylates β -catenin and targets it for degradation. However, in the active canonical Wnt pathway, Wnt ligands interact with Frizzled and LRP receptors. This leads to the translocation of axin, inhibiting the formation of the destruction complex, allowing β -catenin to accumulate and

- translocate to the nucleus, where it will activate the transcription of Wnt target genes (Gordon and Nusse, 2006). Of these includes axin, which acts as a negative regulator of Wnt signalling (Jho et al., 2002; Lustig et al., 2002).
- Targets of Wnt alter endothelial responses to shear stress, for instance, by activating angiogenesis. One target, angiopoietin-2 (ANGPT2), is an established growth factor involved in angiogenesis. Studies in both zebrafish and mice have shown that the ANGPT2 contributes to the development of atherosclerosis (Li et al., 2014; Farhat et al., 2013).
- Thrombospondin-1 (THBS1) is a glycoprotein involved in endothelial cell interactions, and a possible target of Wnt. High levels of THBS1 has been correlated with the inhibition of tumour angiogenesis (Naumov et al., 2006), possibly by selective induction of apoptosis in cells undergoing angiogenesis (Guo et al., 1997). Jo et al. (2005) demonstrated that activation of the Wnt pathway downregulates THBS1 in colon cancer.

55 Hypothesis

This study was conducted to investigate the expression ANGPT2 and THBS1 in low and high shear stress in human umbilical vein endothelial cells (HUVECs), and whether these genes are mediated by canonical Wnt signalling. We addressed this using an orbital shaker model, along with canonical Wnt inhibitor, XAV939.

60 Methods

61 Orbital Shaker

HUVECs were cultured in complete growth medium containing M199, sodium bicarbonate, pen-strep, amphotericin B, Hi-FBS, endothelial cell growth supplement (ECGS), and heparin. When ~80% confluent, cells were incubated with 1ml of trypsin until cells thoroughly detached, and neutralised with 9ml of M199. Cells were spun for 5 minutes at 400g to discard the supernatant, then re-suspended in M199 media before transferring to 10mm radius 6 well plates. Once confluent, 3ml (Warboys, Ghim and Weinberg, 2019) of 0.1% DMSO in M199 or 0.1% XAV939 in M199 were each added to half of the plates (Zhu et al., 2017). Cells were then subjected to flow using a orbital shaker at 210 rpm for 72 hours, with the exception of a static controls.

mRNA Isolation and qPCR

72 Cells were isolated from the periphery and centre of the plates with cold PBS and centrifuged 73 for 5 minutes at 400g to remove the supernatant. Total mRNA was extracted using the 74 RNEasy Mini Kit (Qiagen) and the concentration was determined spectrophotometrically. 75 cDNA synthesis was performed using the Verso cDNA Synthesis Kit (Thermo Scientific) with 76 5.5μl of 0.01067% mRNA. *ANGPT2*, *AXIN2*, *THSB1*, and *HPRT1* mRNA was quantified using 77 StepOne qPCR (Thermo Scientific) with SYBR Green, using oligonucleotide qPCR primers 78 from Ensembl (Howe et al., 2020) (Table 1).

Table 1. Oligonucleotide qPCR primers from Ensembl.

Gene	Direction	Sequence
ANGPT2	L	CGGCTGTGATGATAGAAATAGGGA
ANGF12	R	GTTCCAAGAGCTGAAGTTCAAGTC
AXIN1	L	TGTCACTTACTTTTCTGTGGGGA
AAINI	R	TGTCACTTACTTTTCTGTGGGGA
HPRT1	L	TTGGTCAGGCAGTATAATCC
ПРКП	R	GGGCATATCCTACAACAAC
THSB1	L	AAAGATGGAGAATGCTGAGTTGGA
111301	R	GGTTCCAAAGACAAACCTCACATT

Statistical Analysis

Relative expression is expressed as $2^{\Delta\Delta Ct}$ fold change \pm SEM normalised to the HPRT control. Normality was determined with Kolmogorov-Smirnov Tests. Comparison analysis was performed using the Student's *t*-test. All analyses were performed in R (R Core Team, 2018).

Results

79

90

92

93

In the orbital shaker system, low shear stress downregulated the expression of AXIN2, ANGPT2, and THBS1 in human umbilical vein endothelial cells. Expression of *AXIN*2, a known Wnt target, decreased by 0.28-fold in low shear stress. Similarly, ANGPT2 was decreased by ~0.15-fold, and THBS1 was decreased 0.12-fold.

Exposure to LSS with the addition of a Wnt inhibitor (XAV939) intensified the expression of *AXIN2* by 18.8-fold, ANGPT by 33.4-fold, and THBS1 by 35.6-fold. Conversely, XAV929 decreased expression in high shear stress. AXIN2 decreased 0.68-fold, ANGPT2 by 0.42-fold, and THBS1 by 0.87-fold.

Discussion

Summarise Method & Results

Inhibitors & Axin

Unreliable Data

Refine Methods

Methods used in other studies

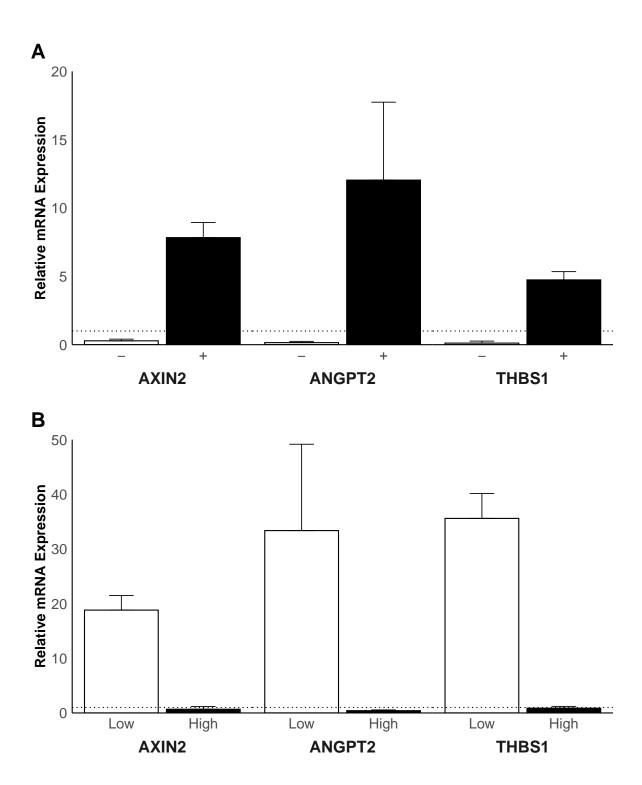


Figure 1. Cells were treated with DMSO(-) or XAV939(+) and exposed to low or high shear stress. Levels of angiopoeitin-2, axin-2, and thrombospondin-1 mRNA quantified by qPCR. (A) Data is shown as fold change \pm SEM of low shear stress relative to high shear stress. (B) Data is shown as fold change \pm SEM of XAV939 relative to DMSO.

Future Plans 98

Acl	kno	wl	ed	σ e1	ner	ıts
ΔC	KILO	AAT	Cu	χCI	1161	ILO

(732 Words) remove headers !!! 100

125

-	•	
ĸ	eferenc	OC
1/	CICICIIC	C 3

	Keferences	101
1.	Farhat, N. et al. (2013). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the American Heart Association</i> , 2 (3). [Online]. Available at: doi:10.1161/jaha.113.000201.	102
2.	Gelfand, B. D. et al. (2011). Hemodynamic Activation of ?-Catenin and T-Cell-Specific Transcription Factor Signaling in Vascular Endothelium Regulates Fibronectin Expression. <i>Arteriosclerosis, Thrombosis, and Vascular Biology,</i> 31 (7), pp.1625–1633. [Online]. Available at: doi:10.1161/atvbaha.111.227827.	
3.	Gordon, M. D. and Nusse, R. (2006). Wnt Signaling: Multiple Pathways, Multiple Receptors, and Multiple Transcription Factors. <i>Journal of Biological Chemistry</i> , 281 (32), pp.22429–22433. [Online]. Available at: doi:10.1074/jbc.r600015200.	106
4.	Guo, N. et al. (1997). Thrombospondin 1 and type i repeat peptides of thrombospondin 1 specifically induce apoptosis of endothelial cells. <i>Cancer research</i> , 57 (9), pp.1735–1742.	108
5.	Howe, K. L. et al. (2020). Ensembl 2021. <i>Nucleic Acids Research</i> , 49 (D1), pp.D884–D891. [Online]. Available at: doi:10.1093/nar/gkaa942.	
6.	Jho, E. et al. (2002). Wnt/?-Catenin/Tcf Signaling Induces the Transcription of Axin2, a Negative Regulator of the Signaling Pathway. <i>Molecular and Cellular Biology</i> , 22 (4), pp.1172–1183. [Online]. Available at: doi:10.1128/mcb.22.4.1172-1183.2002.	112 113
7.	Jo, WS. et al. (2005). Wnt signaling can repress thrombospondin-1 expression in colonic tumorigenesis. <i>Cancer Biology & Therapy</i> , 4 (12), pp.1361–1366. [Online]. Available at: doi:10.4161/cbt.4.12.2201.	
8.	Li, R. et al. (2014). Shear stress-activated wnt-angiopoietin-2 signaling recapitulates vascular repair in zebrafish embryos. <i>Arteriosclerosis, thrombosis, and vascular biology,</i> 34 (10), pp.2268–2275.	116
9.	Lustig, B. et al. (2002). Negative Feedback Loop of Wnt Signaling through Upregulation of Conductin/Axin2 in Colorectal and Liver Tumors. <i>Molecular and Cellular Biology</i> , 22 (4), pp.1184–1193. [Online]. Available at: doi:10.1128/mcb.22.4.1184-1193.2002.	118
10.	Naumov, G. N. et al. (2006). A Model of Human Tumor Dormancy: An Angiogenic Switch From the Nonangiogenic Phenotype. <i>JNCI: Journal of the National Cancer Institute</i> , 98 (5), pp.316–325. [Online]. Available at: doi:10.1093/jnci/djj068.	120
11.	Ni, CW. et al. (2010). Discovery of novel mechanosensitive genes in vivo using mouse carotid artery endothelium exposed to disturbed flow. <i>Blood</i> , 116 (15), pp.e66–e73.	122

R Core Team. (2018). R: A language and environment for statistical computing. Vi- 124

enna, Austria: R Foundation for Statistical Computing. [Online]. Available at: https://or.

[Online]. Available at: doi:10.1182/blood-2010-04-278192.

//www.R-project.org/.

12.

- Souilhol, C. et al. (2019). Endothelial responses to shear stress in atherosclerosis: a novel role for developmental genes. *Nature Reviews Cardiology*, 17 (1), pp.52–63. [Online]. Available at: doi:10.1038/s41569-019-0239-5.
- Stone, P. H. et al. (2007). Regions of low endothelial shear stress are the sites where coronary plaque progresses and vascular remodelling occurs in humans: an in vivo serial study. *European Heart Journal*, 28 (6), pp.705–710. [Online]. Available at: doi:10.1093/eurheartj/ehl575.
- Timmins, L. H. et al. (2017). Oscillatory wall shear stress is a dominant flow characteristic affecting lesion progression patterns and plaque vulnerability in patients with coronary artery disease. *Journal of The Royal Society Interface*, 14 (127), p.20160972. [Online]. Available at: doi:10.1098/rsif.2016.0972.
- Warboys, C. M., Ghim, M. and Weinberg, P. D. (2019). Understanding mechanobiology in cultured endothelium: A review of the orbital shaker method. *Atherosclerosis*, 285, pp.170–177. [Online]. Available at: doi:10.1016/j.atherosclerosis.2019.04.210.
- ¹³⁴ 17. Zhu, J. et al. (2017). Regulation of angiogenic behaviors by oxytocin receptor through Gli1-indcued transcription of HIF-1? in human umbilical vein endothelial cells. *Biomedicine & Pharmacotherapy*, 90, pp.928–934. [Online]. Available at: doi:10.1016/j.biopha.2017.04.021.