

Working Title Natasha Hopkins

82H Project for Master of Biology (MBiol)University of York, UK

Project DirectorDr. Richard Maguire

Examination Date 18 April, 2022

Contents

Introduction	1
Materials and Methods	2
Cell Culture and Application of Shear Stress	2
RNA Extraction and Real-Time Quantitative PCR	2
Statistical Analysis	3
Results	3
Discussion	4
Acknowledgements	5
References	5

Shear stress alters endothelial responses via developmental pathways. This is thought to play a role in atherosclerosis formation in regions of low shear stress. One feature of atherosclerosis is an increase in angiogenesis.

Key Words: Atherosclerosis • Wnt/βeta-catenin Signalling Pathway • Shear Stress • Orbital Shaker • Angiopoietin-2 • Thrombospondin-1

(250 Words)

Introduction

Atherosclerosis is a chronic inflammatory disease characterised by the formation of arterial plaques. Haemodynamic shear stress has been identified as a modulator of site-specificity in atherosclerosis, which occurs preferentially in regions exposed to low, oscillatory shear stress (Stone et al., 2007). Whereas areas of high, laminar shear stress are atheroprotective (Timmins et al., 2017). Shear stress is an important factor in regulating gene expression in vascular endothelial cells (Ni et al., 2010), which is thought to contribute to the susceptibility of plaque formation in atheroprone sites. Multiple omics studies have implicated variations in flow with the regulation of developmental signalling pathways in atherosclerosis, including the Wnt Pathway (Souilhol et al., 2019; Gelfand et al., 2011).

Wnt is an evolutionarily conserved pathway with a critical role in axis patterning during embryonic development. In the absence of Wnt, axin forms a destruction complex with glycogen synthase kinase 3β (GSK-3) and adenomatous polyposis coli (APC), which phosphorylates β -catenin and targets it for degradation. However, in the active canonical Wnt pathway, Wnt ligands interact with Frizzled and LRP receptors. This leads to the translocation of axin, inhibiting the formation of the destruction complex, allowing β -catenin to accumulate and translocate to the nucleus, where it will activate the transcription of Wnt target genes (Gordon and Nusse, 2006). Of these includes axin, which acts as a negative regulator of Wnt signalling (Jho et al., 2002; Lustig et al., 2002).

Shear stress-mediated Wnt orchestrates a range of endothelial responses, including angiogenesis, which is increased in regions of low shear stress compared to high shear stress (Du and Li, 2018). One target of Wnt, angiopoietin-2 (ANGPT2), is an established growth factor involved in angiogenesis. Studies in both zebrafish and mice have shown that the increase in ANGPT2 contributes to the development of atherosclerosis (Li et al., 2014; Farhat et al., 2013a).

Thrombospondin-1 (THBS1) is a glycoprotein involved in endothelial cell interactions and is a potential target of Wnt. High levels of THBS1 has been correlated with the inhibition of tumour angiogenesis (Naumov et al., 2006), possibly by induction of apoptosis via the TGF- β pathway (Miao et al., 2001; Yee et al., 2004), or by inhibition of the VEGF pathway (Gupta et al., 1999; Kaur et al., 2010). Jo et al. (2005) demonstrated that activation of the Wnt pathway downregulates THBS1 in colon cancer. Thus, Wnt may mediate the expression of THBS1 in

- response to flow. Since atherosclerosis occurs in regions of increased angiogenesis, we would
- expect THSB1 to be downregulated in low shear stress. This has been confirmed in a study by Moura et al. (2008).
- This study was conducted to investigate the RNA expression of ANGPT2 and THBS1 in HU-
- ⁴⁶ VECs exposed to atheroprone low shear stress (LSS), compared to atheroprotective high shear
- 47 stress (HSS), and whether these genes are mediated by canonical Wnt signalling. We ad-
- dressed this using an orbital shaker model, along with canonical Wnt inhibitor, XAV939.

Materials and Methods

50 Cell Culture and Application of Shear Stress

49

HUVECs were cultured at 37°C in 8% M199, 0.15% sodium bicarbonate, 1 U/mL pen-strep, 0.1 ug/ml amphotericin B, 20% Hi-FBS, 30 ug/ml endothelial cell growth supplement (ECGS), and 10 U/ml heparin. After reaching ~80% confluence, passage 2 cells were incubated with 1ml of trypsin until cells thoroughly detached, and neutralised with 9ml of M199. They were then re-suspended in M199 media before transferring to 10mm radius 6 well plates coated in 1% gelatin. The canonical Wnt pathway was inhibited using XAV939. Once confluent, cells were treated with either 3ml of 0.1% DMSO in M199 or 0.1% XAV939 in M199 (Zhu et al., 2017). They were placed on an orbital shaker at 210 rpm for 72 hours, and exposed to low (~5 dynes/cm²) and high shear stress (~15 dynes/cm²) (Warboys, Ghim and Weinberg, 2019), with the exception of the static control.

61 RNA Extraction and Real-Time Quantitative PCR

62 Cells were isolated from the periphery and centre of the plates with cold PBS and centrifuged 63 for 5 minutes at 400g. Total mRNA was extracted using the RNEasy Mini Kit (Qiagen) and 64 the concentration was determined spectrophotometrically. cDNA synthesis was performed 65 using the Verso cDNA Synthesis Kit (Thermo Scientific) as per the manufacturers instruc-66 tions. ANGPT2, AXIN2, THSB1, and HPRT1 (reference gene) mRNA was quantified using 67 StepOne qPCR (Thermo Scientific) with SYBR Green, using oligonucleotide qPCR primers 68 from Ensembl (Howe et al., 2020) (Table 1). The amplification included 30 cycles at 95°C for 69 30s, 5°C for 30s, and 72°C for 45s, followed by 72 °C 10 min.

Table 1. Oligonucleotide qPCR primers from Ensembl.

Gene	Direction	Sequence
ANGPT2	L	CGGCTGTGATGATAGAAATAGGGA
ANGF12	R	GTTCCAAGAGCTGAAGTTCAAGTC
AXIN1	L	TGTCACTTACTTTTCTGTGGGGA
AAINI	R	TGTCACTTACTTTTCTGTGGGGA
HPRT1	L	TTGGTCAGGCAGTATAATCC
пгки	R	GGGCATATCCTACAACAAC
THSB1	L	AAAGATGGAGAATGCTGAGTTGGA
	R	GGTTCCAAAGACAAACCTCACATT

Statistical Analysis

Relative expression is expressed as $2^{\Delta\Delta Ct}$ fold change \pm SEM, normalised to the HPRT control. Normality was determined with Kolmogorov-Smirnov Tests. Comparison analysis was performed using the Student's t-test. All plots and analyses were performed in R (R Core Team, 2018).

Results

To asses the effect of low and high shear stress on gene express, HUVECs were exposed to flow using an orbital shaker system. Gene expression of ANGPT2, THSB1, and Wnt reporter AXIN2 was quantified by qPCR. Gene expression in HUVECs exposed LSS was compared to those exposed to HSS (Figure 1A). Low shear stress downregulated the expression of AXIN2, ANGPT2, and THBS1. AXIN2, a known Wnt target, decreased by 0.28-fold in low shear stress. Similarly, ANGPT2 was decreased by 0.15-fold, and THBS1 was decreased 0.12-fold. Using the same method, HUVECs were also treated with either DMSO or canonical Wnt inhibitor XAV929, to assess for regulation by Wnt signalling. Expression in the presence of XAV939 was then compared to DMSO (Figure 1B). Exposure to LSS with the addition XAV939 intensified the expression of AXIN2 by 18.82-fold, ANGPT by 33.35-fold, and THBS1 by 35.59-fold. Whereas XAV929 decreased expression in HSS. AXIN2 decreased 0.68-fold, ANGPT2 by 0.42-fold, and THBS1 by 0.87-fold. Results were analysed using the Students *t* test, which regarded them insignificant due to the sample size.

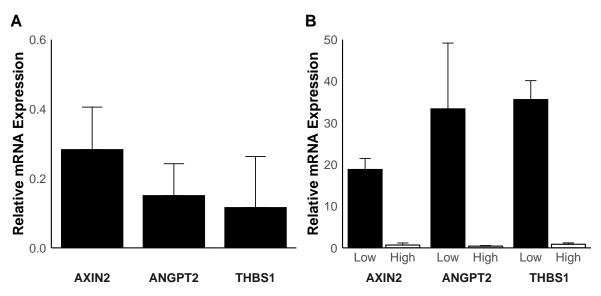


Figure 1. Low shear stress downregulates gene expression in the presence Wnt, and upregulates gene expression in the absence of Wnt.. Cells were treated with DMSO(-) or XAV939(+) and exposed to low or high shear stress. Levels of angiopoietin-2, axin-2, and thrombospondin-1 mRNA were quantified by qPCR. (A) Low shear stress upregulated the expression of all genes. AXIN: 0.28 ± 0.12 , ANGPT2: 0.15 ± 0.09 , THBS1: 0.12 ± 0.15 . Data is shown as fold change \pm SEM of low shear stress relative to high shear stress. (B) XAV939 upregulated expression of all genes in low shear stress and downregulated expression of all genes in high shear stress. AXIN: 18.82 ± 2.66 & 0.68 ± 0.50 , ANGPT2: 33.36 ± 15.80 & 0.42 ± 0.14 , THBS1: 35.59 ± 4.55 & 0.87 ± 0.33 . Data is shown as fold change \pm SEM of XAV939 relative to DMSO.

Discussion

89

104

105

106

107

108

109

110

116

117

118

119

120

121

126

In our study, we compare the effects of low shear stress and high shear stress on the expression of two regulators of angiogenesis, ANGPT2 and THBS1, and whether their expression is controlled by Wnt. This was achieved using an orbital shaker model, first described by Warboys et al. (2014). Direct Wnt target, AXIN2, was also measured to quantify Wnt expression (Jho et al., 2002). Finally, we compared gene expression in LSS to HSS, and XAV939 to DMSO.

Our findings indicate that AXIN2, ANGPT, and THBS1 is downregulated in HUVECs exposed to LSS (Figure 1A). The downregulation of ANGPT and AXIN2 in LSS differs from our expectations. Previous studies demonstrate that canonical Wnt is activated by low shear stress (Gelfand et al., 2011), as is its direct target, AXIN2 (Jho et al., 2002). ANGPT2, a positive regulator of angiogenesis, has also been identified as a positive target of Wnt in zebrafish (Li et al., 2014). Thus, ANGPT2 was also expected to be upregulated in LSS. However, the results imply that Wnt is downregulated by LSS compared to HSS, and in doing so, downregulates both ANGPT2 and AXIN2.

On the other hand, the lower expression of THSB1 in low shear stress was anticipated (Moura et al., 2008). Wnt has been shown to inhibit THBS1, a negative regulator of angiogenesis, in colonic tumours (Jo et al., 2005). Therefore, its expression should increase with the down-regulation of AXIN2. This was not the case, suggesting that it is not repressed by the Wnt pathway as previously reported.

• This is a very different context - you should make this clear, and if it's the basis of choosing it as a target mention it in the introduction. If it's not regulated by Wnt in this context - then that is fine but you need to be quite explicit in how you discuss it.

In the presence of XAV939, all genes were upregulated in both low and high shear stress (Figure 1B). Initially, this would imply that Wnt is an inhibitor of ANGPT2 and THSB1.

However, since AXIN2 is a direct inhibitor of Wnt, this suggests that the inhibitor in fact failed to inhibit Wnt. ?

- Maybe how could you find out? Also is this canonical or non-canonical Wnt? There is cross talk between the two kinds of pathway, but they are quite separate. XAV939 won't inhibit non-canonical Wnt signalling.
- If Inhibitor failed, why not consistent with the DMSO samples?
 - XAV939 works but issue with primers specificity? Bind to another protein upregulated in LSS. Wnt downregulated in HSS hence why fold changes smaller? Small sample size so downregulation seems more prominent then it actually is

There are many discrepancies between our results and prior studies, likely due to limitations and errors in our method...

- Possible Errors and Refine Methods
 - Small sample = not enough to confirm spectrophotometer was consistent

	 No prior PCR experience 	127
	• Melt curve suggests primer dimers & contamination (include images of curves or include in supplementary?)	128 129
	• Vascular tubule formation (Tressel et al., 2007)	130
	• en face staining for gene markers in inner curvature (Warboys et al., 2014)	131
	• Direct interaction with Wnt or via VEGF?	132
	- THSB1 inhibits VEGF by competing for heparin (which is used in our medium) (Gupta et al., 1999; Dias et al., 2012; Tolsma et al., 1993)	133 134
	Methods used in other studies	135
	• animal studies, zebrafish, mice, pigs (Serbanovic-Canic et al., 2017; Li et al., 2014; Farhat et al., 2013b; Moura et al., 2008; Moonen et al., 2015)	136 137
	• non-canonical wnt (Franco et al., 2016)	138
	• relative vs quantitative expression	139
	flow chamber instead of orbital shaker	140
	Acknowledgements	141
	Word Count: 1472 Words	142
	Word Count: 1472 Words References	142
1.	References Dias, J. V. et al. (2012). A motif within the N-terminal domain of TSP-1 specifically promotes the proangiogenic activity of endothelial colony-forming cells. <i>Biochemical Pharmacology</i> , 84	143 144
1.	References Dias, J. V. et al. (2012). A motif within the N-terminal domain of TSP-1 specifically promotes the proangiogenic activity of endothelial colony-forming cells. <i>Biochemical Pharmacology</i> , 84 (8), pp.1014–1023. [Online]. Available at: doi:10.1016/j.bcp.2012.07.006. Du, J. and Li, J. (2018). The role of wnt signaling pathway in atherosclerosis and its relationship with angiogenesis. <i>Experimental and Therapeutic Medicine</i> . [Online]. Available at:	143 144 145 146
	References Dias, J. V. et al. (2012). A motif within the N-terminal domain of TSP-1 specifically promotes the proangiogenic activity of endothelial colony-forming cells. <i>Biochemical Pharmacology</i> , 84 (8), pp.1014–1023. [Online]. Available at: doi:10.1016/j.bcp.2012.07.006. Du, J. and Li, J. (2018). The role of wnt signaling pathway in atherosclerosis and its relationship with angiogenesis. <i>Experimental and Therapeutic Medicine</i> . [Online]. Available at: doi:10.3892/etm.2018.6397. Farhat, N. et al. (2013a). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the</i>	143 144 145 146 147 148
2.	References Dias, J. V. et al. (2012). A motif within the N-terminal domain of TSP-1 specifically promotes the proangiogenic activity of endothelial colony-forming cells. <i>Biochemical Pharmacology</i> , 84 (8), pp.1014–1023. [Online]. Available at: doi:10.1016/j.bcp.2012.07.006. Du, J. and Li, J. (2018). The role of wnt signaling pathway in atherosclerosis and its relationship with angiogenesis. <i>Experimental and Therapeutic Medicine</i> . [Online]. Available at: doi:10.3892/etm.2018.6397. Farhat, N. et al. (2013a). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the American Heart Association</i> , 2 (3). [Online]. Available at: doi:10.1161/jaha.113.000201. Farhat, N. et al. (2013b). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the</i>	143 144 145 146 147 148 149 150
2.	References Dias, J. V. et al. (2012). A motif within the N-terminal domain of TSP-1 specifically promotes the proangiogenic activity of endothelial colony-forming cells. <i>Biochemical Pharmacology</i> , 84 (8), pp.1014–1023. [Online]. Available at: doi:10.1016/j.bcp.2012.07.006. Du, J. and Li, J. (2018). The role of wnt signaling pathway in atherosclerosis and its relationship with angiogenesis. <i>Experimental and Therapeutic Medicine</i> . [Online]. Available at: doi:10.3892/etm.2018.6397. Farhat, N. et al. (2013a). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the American Heart Association</i> , 2 (3). [Online]. Available at: doi:10.1161/jaha.113.000201.	143 144 145 146 147 148 149 150
 3. 4. 	References Dias, J. V. et al. (2012). A motif within the N-terminal domain of TSP-1 specifically promotes the proangiogenic activity of endothelial colony-forming cells. <i>Biochemical Pharmacology</i> , 84 (8), pp.1014–1023. [Online]. Available at: doi:10.1016/j.bcp.2012.07.006. Du, J. and Li, J. (2018). The role of wnt signaling pathway in atherosclerosis and its relationship with angiogenesis. <i>Experimental and Therapeutic Medicine</i> . [Online]. Available at: doi:10.3892/etm.2018.6397. Farhat, N. et al. (2013a). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the American Heart Association</i> , 2 (3). [Online]. Available at: doi:10.1161/jaha.113.000201. Farhat, N. et al. (2013b). Angiopoietin-Like 2 Promotes Atherogenesis in Mice. <i>Journal of the American Heart Association</i> , 2 (3). [Online]. Available at: doi:10.1161/jaha.113.000201. Franco, C. A. et al. (2016). Non-canonical Wnt signalling modulates the endothelial shear stress flow sensor in vascular remodelling. <i>eLife</i> , 5. [Online]. Available at:	143 144 145 146 147 148 149 150 151 152

- ¹⁵⁸ 8. Gupta, K. et al. (1999). *Angiogenesis*, 3 (2), pp.147–158. [Online]. Available at: doi:10.1023/a:1009018702832.
- Howe, K. L. et al. (2020). Ensembl 2021. Nucleic Acids Research, 49 (D1), pp.D884–D891. [Online]. Available at: doi:10.1093/nar/gkaa942.
- In the second of the Signaling Pathway. Molecular and Cellular Biology, 22 (4), pp.1172–1183. [Online]. Available at: doi:10.1128/mcb.22.4.1172-1183.2002.
- Jo, W.-S. et al. (2005). Wnt signaling can repress thrombospondin-1 expression in colonic tumorigenesis. *Cancer Biology & Therapy*, 4 (12), pp.1361–1366. [Online]. Available at: doi:10.4161/cbt.4.12.2201.
- Kaur, S. et al. (2010). Thrombospondin-1 Inhibits VEGF Receptor-2 Signaling by Disrupting Its Association with CD47. *Journal of Biological Chemistry*, 285 (50), pp.38923–38932. [Online].
 Available at: doi:10.1074/jbc.m110.172304.
- Li, R. et al. (2014). Shear stress-activated wnt-angiopoietin-2 signaling recapitulates vascular repair in zebrafish embryos. *Arteriosclerosis, thrombosis, and vascular biology,* 34 (10), pp.2268–2275.
- Lustig, B. et al. (2002). Negative Feedback Loop of Wnt Signaling through Upregulation of Conductin/Axin2 in Colorectal and Liver Tumors. *Molecular and Cellular Biology*, 22 (4), pp.1184–1193. [Online]. Available at: doi:10.1128/mcb.22.4.1184-1193.2002.
- 15. Miao, W. M. et al. (2001). Thrombospondin-1 type 1 repeat recombinant proteins inhibit tumor growth through transforming growth factor-beta-dependent and -independent mechanisms. *Cancer research*, 61 21, pp.7830–7839.
- Moonen, J.-R. A. J. et al. (2015). Endothelial-to-mesenchymal transition contributes to fibroproliferative vascular disease and is modulated by fluid shear stress. *Cardiovascular Research*, 108 (3), pp.377–386. [Online]. Available at: doi:10.1093/cvr/cvv175.
- 17. Moura, R. et al. (2008). Thrombospondin-1 Deficiency Accelerates Atherosclerotic Plaque Maturation in *ApoE* -/- Mice. *Circulation Research*, 103 (10), pp.1181–1189. [Online]. Available at: doi:10.1161/circresaha.108.185645.
- 18. Naumov, G. N. et al. (2006). A Model of Human Tumor Dormancy: An Angiogenic Switch From the Nonangiogenic Phenotype. *JNCI: Journal of the National Cancer Institute*, 98 (5), pp.316–325. [Online]. Available at: doi:10.1093/jnci/djj068.
- Ni, C.-W. et al. (2010). Discovery of novel mechanosensitive genes in vivo using mouse carotid artery endothelium exposed to disturbed flow. *Blood*, 116 (15), pp.e66–e73. [Online]. Available at: doi:10.1182/blood-2010-04-278192.
- R Core Team. (2018). *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. [Online]. Available at: https://www.R-project.org/.
- Serbanovic-Canic, J. et al. (2017). Zebrafish Model for Functional Screening of Flow-Responsive Genes. *Arteriosclerosis, Thrombosis, and Vascular Biology,* 37 (1), pp.130–143. [Online]. Available at: doi:10.1161/atvbaha.116.308502.
- Souilhol, C. et al. (2019). Endothelial responses to shear stress in atherosclerosis: a novel role for developmental genes. *Nature Reviews Cardiology*, 17 (1), pp.52–63. [Online]. Available at: doi:10.1038/s41569-019-0239-5.
- Stone, P. H. et al. (2007). Regions of low endothelial shear stress are the sites where coronary plaque progresses and vascular remodelling occurs in humans: an in vivo serial study. *European Heart Journal*, 28 (6), pp.705–710. [Online]. Available at: doi:10.1093/eurheartj/ehl575.
- Timmins, L. H. et al. (2017). Oscillatory wall shear stress is a dominant flow characteristic affecting lesion progression patterns and plaque vulnerability in patients with coronary artery disease. *Journal of The Royal Society Interface*, 14 (127), p.20160972. [Online]. Available at: doi:10.1098/rsif.2016.0972.
- Tolsma, S. et al. (1993). Peptides derived from two separate domains of the matrix protein thrombospondin-1 have anti-angiogenic activity. *Journal of Cell Biology*, 122 (2), pp.497–511. [Online]. Available at: doi:10.1083/jcb.122.2.497.

26.	Tressel, S. L. et al. (2007). Laminar Shear Inhibits Tubule Formation and Migration of Endothe-	194
	lial Cells by an Angiopoietin-2-Dependent Mechanism. Arteriosclerosis, Thrombosis, and Vascu-	
	lar Biology, 27 (10), pp.2150–2156. [Online]. Available at: doi:10.1161/atvbaha.107.150920.	195

27. Warboys, C. M. et al. (2014). Disturbed Flow Promotes Endothelial Senescence via a p53-Dependent Pathway. Arteriosclerosis, Thrombosis, and Vascular Biology, 34 (5), pp.985–995. [Online]. Available at: doi:10.1161/atvbaha.114.303415.

197

199

202

203

- 28. Warboys, C. M., Ghim, M. and Weinberg, P. D. (2019). Understanding mechanobiology in 198 cultured endothelium: A review of the orbital shaker method. Atherosclerosis, 285, pp.170–177. [Online]. Available at: doi:10.1016/j.atherosclerosis.2019.04.210.
- 29. Yee, K. O. et al. (2004). Expression of the Type-1 Repeats of Thrombospondin-1 Inhibits Tu- 200 mor Growth Through Activation of Transforming Growth Factor-β. The American Journal of Pathology, 165 (2), pp.541–552. [Online]. Available at: doi:10.1016/s0002-9440(10)63319-6.
- Zhu, J. et al. (2017). Regulation of angiogenic behaviors by oxytocin receptor through Gli1-30. indcued transcription of HIF-1? in human umbilical vein endothelial cells. Biomedicine & Pharmacotherapy, 90, pp.928–934. [Online]. Available at: doi:10.1016/j.biopha.2017.04.021.