

Table S1A. Bacterial strains used in this study.

Strain	Genotype	Reference
<i>E. coli</i>		
BL21 Star™ (DE3)	<i>F ompT hsdS_B(r_B-, m_B-) gal dcm rne131 (DE3)</i>	Invitrogen
S17-1	<i>Pro</i> , <i>res</i> ⁻ <i>hsdR17</i> (<i>rK</i> ⁻ <i>mK</i> ⁺) <i>recA</i> ⁻ , <i>RP4-2-Tc::Mu-Km::Tn7</i> , <i>Tp</i> ^r	(1)
<i>F. johnsoniae</i>		
UW101		(2)
Fl_082	UW101 <i>ΔgldL</i>	(3)
Rhj_074	UW101 <i>gldL</i> _{64-74GSS}	This study
Rhj_076	UW101 <i>gldL</i> _{Δ64-74}	This study

Table S1B. Plasmids used in this study

Plasmid	Description	Reference
pT12_SpaPQR3xFLAG	Encodes <i>Salmonella enterica</i> serovar Typhimurium SpaPQR operon with a C-terminal 3xFLAG tag on SpaR under the control of the <i>E. coli</i> rhaB promoter; ori cloDF13, Kan ^r	(4)
pMA-RQ_SthPKLMN	Synthetic plasmid with <i>S. thermophila</i> strain DSM 21410 genes RCX05217-05221 and 100 bp upstream and downstream in the pMA-RQ vector. Ori Col E1, Amp ^r	This study. Supplied by GeneArt (Invitrogen).
pRHJ012	pGEM-T <i>gldL</i>	(3)
pRHJ113	pMA-RQ_SthPKLMN with NcoI, NdeI and BamHI sites in GldL and GldM, respectively, removed by Quikchange mutagenesis	This study
pRHJ117	pT12 <i>S. thermophila gldL gldM</i> (1-229)- <i>twinstrep</i>	This study
pRHJ118	pT12 <i>P. gingivalis porL porM</i> (1-227)- <i>twinstrep</i>	This study
pRHJ170	pT12 <i>S. wenxiniae gldL gldM</i> (1-224)- <i>twinstrep</i>	This study
pRHJ174	pT12 <i>C. canimorsus gldL gldM</i> (1-330)- <i>twinstrep</i>	This study
pRHJ237	pGEM-T <i>gldL</i> _{64-74GSS}	This study
pRHJ238	pGEM-T <i>gldL</i> _{Δ64-74}	This study
pRHJ240	Suicide plasmid used to introduce the E64-L74 to GSSGSSGSSGS codon change into <i>gldL</i> ; <i>gldL</i> _{64-74GSS} in pYT354	This study

pRHJ241	Suicide plasmid used to delete E64-L74 from <i>gldL</i> ; <i>gldL</i> _{Δ64-74} in pYT354	This study
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1. Simon R, Priefer U, Pühler A. 1983. A broad host range mobilization system for in vivo genetic engineering: Transposon mutagenesis in Gram-negative bacteria. *Bio/Technology* 1:784–791.
2. McBride MJ, Braun TF. 2004. GldI is a lipoprotein that is required for *Flavobacterium johnsoniae* gliding motility and chitin utilization. *J Bacteriol* 186:2295–302.
3. Hennell James R, Deme JC, Kjær A, Alcock F, Silale A, Lauber F, Johnson S, Berks BC, Lea SM. 2021. Structure and mechanism of the proton-driven motor that powers type 9 secretion and gliding motility. *Nat Microbiol* 6:221–223.
4. Dietsche T, Tesfazgi Mebrhatu M, Brunner MJ, Abrusci P, Yan J, Franz-Wachtel M, Schärfe C, Zilkenat S, Grin I, Galán JE, Kohlbacher O, Lea S, Macek B, Marlovits TC, Robinson CV, Wagner S. 2016. Structural and Functional Characterization of the Bacterial Type III Secretion Export Apparatus. *PLOS Pathog* 12:e1006071.