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# **End-to-End Deep Learning Model for Base Calling of MinION Nanopore Reads**

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*Umjesto ove stranice umetnite izvornik Vašeg rada.*  
*Kako biste uklonili ovu stranicu, obrišite naredbu \izvornik.*

*I would like to thank my mentor, Mile Šikić, for his patient guidance, encouragement and advice provided over the years.*

*I would also like to thank my family and friends for their continuous support.*

*In the end, honorable mentions go to Marko Ratković for his help with this thesis.*

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# 1. Introduction

In recent years, deep learning methods significantly improved the state-of-the-art in multiple domains such as computer vision, speech recognition and natural language processing (Lecun and Bengio, 1998; Krizhevsky et al., 2012) In this paper, we present application of deep learning for DNA basecalling problem.

Oxford Nanopore Technology's MinION nanopore sequencing platform Mikheyev and Tin (2014) is the first portable DNA sequencing device. It produces longer reads than competing technologies. In addition, it enables real-time data analysis which makes it suitable for various applications. Although MinION is able to produce long reads, even up to 882 kb Loman (a,b), they have an error rate of 10% or higher. This master thesis uses R9.4 pore model and compares previous techniques with novel auto-encoder multi-task training.

## 1.1. Organization

[TODO]: Write some fancy stuff once completed.

## 2. Background

Due to technical constraints, it's infeasible to sequence whole DNA in single strand. Every sequencing technology to date have an upper limit how big strand can it precisely sequence. This limit is considerably smaller than size of genome. For example E.Coli has 4.5 million base pairs in its DNA, while Sanger's sequencing maximum output is around 1000 base pairs max. To make DNA basecalling feasible technique called shotgun sequencing was invented. The strand is cloned number of times, then via chemical agent broken down into smaller fragments of appropriate length. Sequenced fragments are called reads.

Genome assembly is the process of reconstructing the original genome from reads and usually starts with finding overlaps between reads. The quality of reconstruction heavily depends on the length and the quality (accuracy) of the reads produced by the sequencer.

If we have reference sequence we usually align the reads on the reference to aid us into genome assembly. Otherwise we have to use many de novo assembly techniques.

The right analogy would be building a puzzle. Since we cannot scan the whole puzzle because our camera is too small or imprecise, we are scanning pieces of the whole picture. Puzzle pieces would represent fragments in this analogy. If we have a map, even a rough one, it shall aid us into assembling those puzzle pieces into complete pictures. Otherwise we're fiddling in the dark and using de novo assembly techniques.

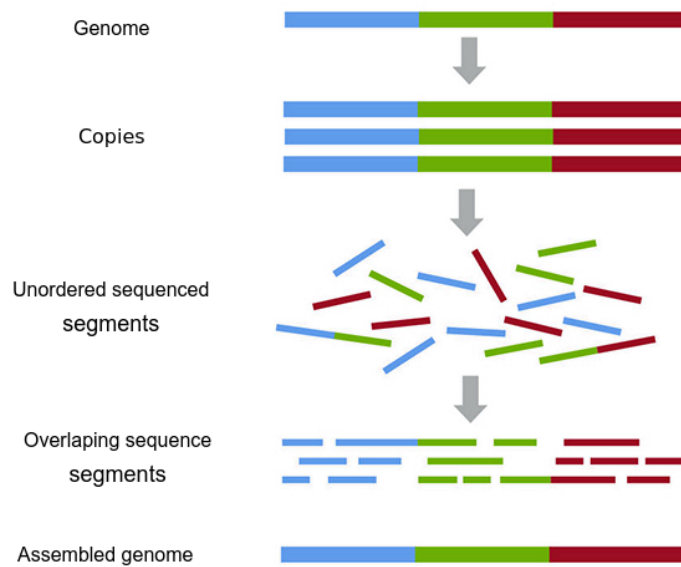
Figure 2.1 depicts process of sequencing visually.

In 1977, Frederick Sanger (Mile Šikić, 2013)(Pettersson et al., 2009) started development of sequencing technologies. It allowed read lengths up to 1000 bases with very high accuracy(99.9%) at the cost of 1\$ per 1000 bases. Later, second generation sequencing, like IAN Torrent and Illumina devices, reduced the price while keeping the accuracy high. However, they had a cost of shorter read lengths, about a few hundred base pairs, which makes resolving repetitive regions practically impossible.

Third generation sequencing technologies have longer read lengths at the accuracy's expense. PacBio, for example, developed technology with a few thousand bases with error rates of ~10-15%.

MinION sequences, which this master thesis use, made sequencing less expensive and even portable.





**Figure 2.1:** Depiction of the sequencing process. Taken from (Ratković, 2017) with authors permission

## 2.1. Oxford Nanopore MinION

The MinION device by Oxford Nanopore Technologies is the first portable DNA sequencing device. Its small weight, low cost, and long read length combined with decent accuracy yield promising results in various applications including full human genome assembly Jain et al. (2017) what could potentially lead to personalized genomic medicine. It weights only 87 grams, and its portability lead to uses on international space station and the antarctic among other places.

Under the hood, it has numerous nano-meter sized pores, thus a names Nanopore. Each pore has width of around 6 nucleotied. Under electric current DNA strand passed through the pore and changes its electric resistance. The sensor measures current through the pore multiple times a second <sup>1</sup>. This signal varies depending which k-mer is occupying the pore, and on its basis we're performing the basecalling. On figure 2.2 this process is visually depicted.

MinION devices can produce long reads, usually tens of thousand base pairs (with reported reads lengths of 100 thousand Loman (a) and even recently above 800 thousand base pairs Loman (b)), but with high sequencing error than older generations of sequencing technologies.

The sequecing resulting file is in FAST5 format, which is adapted HDF5 file format, popular in bioinformatics community. It stores raw signal, alongside various metadata. Un-

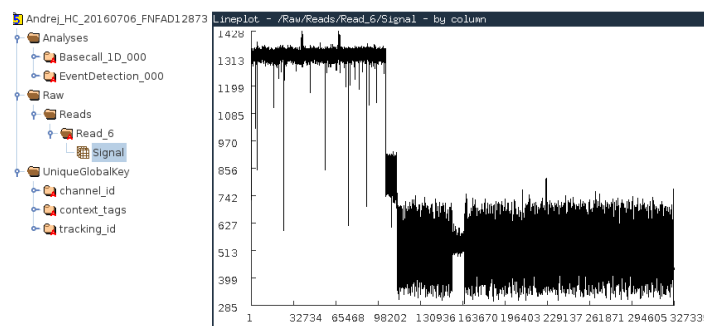
<sup>1</sup>The model we worked with had 4000 samples per seconds

<sup>2</sup>Figure adapted from <https://nanoporetech.com/how-it-works>



**Figure 2.2:** DNA strand being pulled through a nanopore <sup>2</sup>

fortunately, many basecallers, including the official ones, upon executing store their results in the FAST5 files. It leads to data and processing coupling in single file, and bloated file sizes.



**Figure 2.3:** Structure of FAST5 file and raw signal line plot show in *HDFView* <sup>3</sup>

## 2.2. Related work

### 2.2.1. Oxford Nanopore Technologies

This subsection covers various basecallers published by ONT, the MinION device maker.

#### Metrichor

Metrichor is now defunct cloud based basecaller. The older versions used *hidden Markov models* (HMM) as underlying algorithm. Preprocessing started with segmenting signal into smaller chunks called events with their start, end, length, mean signal strength and standard

<sup>3</sup><https://support.hdfgroup.org/products/java/hdfview/>

deviation. These events are observations in the HMM model, and the underlying generating hidden sequence is sequence of 6-mers. They build their HMM transition matrix with stay, skip 1 and skip 2 probabilities, that is underlying 6-mer cannot move for more than 2 nucleotides per event. Basecalling is performed using Viterbi algorithm. This approach showed poor results when calling long homopolymer stretches as the context in the pore remains the same Goodwin et al. (2016) Ip et al. (2015).

## **Nanonet**

Nanonet is ONT first generation neural network basecaller. It used to be available on github, but it's now defunct and unavailable.

## **Albacore**

Albacore is a production basecaller provided by Oxford Nanopore, and uses a command-line interface. It utilizes the latest in Recurrent Neural Network algorithms in order to interpret the signal data from the nanopore, and basecall the DNA or RNA passing through the pore. It implements stable features into Oxford Nanopore Technologies' software products, and is fully supported. It receives .fast5 files as an input, and is capable of producing:

- .fast5 files appended with basecalled information
- .fast5 files that have been processed, but basecall information present in a separate .fastq file

## **Guppy**

Guppy is ONT's new basecaller that can use GPUs to basecall much faster than Albacore. Both the GridION X5 and PromethION contain GPUs and use Guppy to basecall while sequencing. Guppy can also use CPUs and scales well to many-CPU systems, so it may run faster than Albacore even without GPUs. In the future it's intended to replace Albacore as production basecaller.

## **Scrappie**

Scrappie<sup>4</sup> is ONT's research basecaller. Scrappie is reported to be the first basecaller that specifically address homopolymer base calling. It became publicly available just recently in June, 2017 and supports R9.4 and future R9.5 data.

Unlike Albacore, Scrappie does not have fastq output, either directly or by writing it into the fast5 files – it only produces fasta reads.

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<sup>4</sup><https://github.com/nanoporetech/scrappie>

## 2.2.2. Third-party basecallers

### Nanocall

Nanocall (David et al., 2016) was the first third-party open source basecaller for nanopore data. It uses HMM approach like the original R7 Metrichor. Nanocall does not support newer chemistries after R7.3.

### DeepNano

DeepNano (Boža et al., 2017) was the first open-source basecaller based on neural networks. It uses bidirectional recurrent neural networks implemented in Python, using the Theano library. When released, originally only supported R7 chemistry, but support for R9 and R9.4 was added recently.

### basecRAWller

basecRAWller<sup>5</sup> is developed by Marcus Stoiber and James Brown at the Lawrence Berkeley National Laboratory.

### Chiron

Chiron (Teng et al., 2017) is developed by Haotian Teng and others in Lachlan Coin's group at the University of Queensland. They are basecalling from the raw signal, using first residual convolutional neural network, then LSTM and finally beam search or greedy decoder depending on chosen configuration.

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<sup>5</sup><https://basecrawller.lbl.gov/>

## **3. Methods**

In this chapter all key deep learning concepts shall be described.

## 4. System architecture

### 4.1. Data Preparation

Data has been downloaded from [https://data.genomicsresearch.org/Projects/online\\_dataset/train\\_set\\_all/](https://data.genomicsresearch.org/Projects/online_dataset/train_set_all/). The following species were provided there by the Chiron team:

- Human
- E. Coli
- Lambda Phage

The raw dataset is transformed using my repo, `minion-data`<sup>1</sup>. It defines common dataset training structure in the protobuf (Google) interface description language (IDL). The whole definition can be seen in figure 4.1.

For the concrete Chiron dataset, the re-squiggled preparation method was used. (The data was re-squiggled, that is after aligning the read on the reference, the read data is improved and each base pairs place on the raw signal is calculated.)

The re-squiggled basecalled data is located at `/Analyses/RawGenomeCorrected_000/BaseCa`. The interesting code fragments are in function `processDataPoint` of file `minion_data/preperation/` from `minion-data` python package.

After the gzipped dataset is prepared, it goes into the training pipeline. The whole training & testing pipeline is available open source on <https://github.com/nmiculenic/minion-basecaller>

### 4.2. Training pipeline

### 4.3. Hyperparameter optimization

---

<sup>1</sup><https://github.com/nmiculenic/minion-data>

```

1 syntax = "proto3";
2
3 package dataset;
4
5 enum BasePair {
6     A = 0;
7     C = 1;
8     G = 2;
9     T = 3;
10    BLANK = 4;
11 }
12
13 enum Cigar {
14     MATCH = 0;
15     MISMATCH = 1;
16     INSERTION = 2; // Insertion, soft clip, hard clip
17     DELETION = 3;  // Deletion, N, P
18 }
19
20 message DataPoint {
21     message BPConfidenceInterval {
22         uint64 lower = 1;
23         uint64 upper = 2;
24         BasePair pair = 3;
25     }
26     repeated float signal = 1;
27     repeated BasePair basecalled = 2; // What we basecalled
28     repeated BPConfidenceInterval labels = 3; // labels describe ←
        corrected basecalled signal for training
29 }
30

```

**Figure 4.1:** dataset protobuf description

## **5. Results**



## **6. Conclusion**

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## **End-to-End Deep Learning Model for Base Calling of MinION Nanopore Reads**

### **Abstract**

In the MinION device, single-stranded DNA fragments move through nanopores, which causes drops in the electric current. The electric current is measured at each pore several thousand times per second. Each event is described by the mean and variance of the current and by event duration. This sequence of events is then translated into a DNA sequence by a base caller. Develop a base-caller for MinION nanopore sequencing platform using a deep learning architecture such as convolutional neural networks and recurrent neural networks. Instead of events, use current waveform at the input. Compare the accuracy with the state-of-the-art basecallers. For testing purposes use publicly, available datasets and Graphmap or Minimap 2 tools for aligning called reads on reference genomes. Implement method using TensorFlow or similar library. The code should be documented and hosted on a publicly available Github repository.

**Keywords:** base calling, Oxford Nanopore Technologies, MinION, deep learning, seq2seq, convolutional neural network, residual network, CTC loss

### **S kraja na kraj model dubokog učenja za određivanje očitanih baza dobivenih uređajem za sekvenciranje MinION**

#### **Sažetak**

Unutar uređaja MinION, fragmenti jednostruke DNA prolaze kroz nanopore, što uzrokuje promjene u električnoj struji. Struja proizvedena na svakoj nanopori mjeri se nekoliko tisuća puta u sekundi. Svaki događaj opisan je srednjom vrijednosti i varijancom struje te svojim trajanjem. Postupak kojim se takav slijed događaja prevodi u niz nukleotida naziva se određivanje očitanih baza. Razviti alat za prozivanje baza za uređaj za sekvenciranje MinION koristeći modele dubokog učenje kao što su konvolucijske i povratne neuronske mreže. Umjesto događaja na ulazu koristi valni oblik struje. Usporediti dobivenu točnost s postojećim rješenjima. U svrhu testiranja koristiti javno dostupne skupove podataka i alate GraphMap ili Minimap 2 za poravnanje očitavanja na referentni genom. Alat implementirati koristeći programsku biblioteku TensorFlow (ili neku sličnu). Programski kod treba biti dokumentiran i javno dostupan preko repozitorija GitHub. **Ključne riječi:** određivanje

baza, Oxford Nanopore Technologies, MinION, duboko učenje, prevođenje, konvolucijske neuronske mreže, rezidualne mreže, CTC gubitak