## Phylogeny

Based on the classification by Manning et al., 2002, MYLK2, also known as skeletal muscle myosin light chain kinase (skMLCK), is assigned to the myosin light chain kinase (MLCK) family (chang2016cardiacmyosinlight pages 3-3, herring2000smoothmusclemyosin pages 8-9). This family is a distinct subgroup within the Ca2+/calmodulin-dependent protein kinase (CaMK) group of the eukaryotic protein kinase (ePK) superfamily (josephson2011smoothmusclemyosin pages 7-9, seguchi2007acardiacmyosin pages 2-4, kamm2011signalingtomyosin pages 1-2). The MLCK family consists of four kinases encoded by separate genes: MLCK1 (smooth muscle), MLCK2 (skeletal muscle), MLCK3 (cardiac muscle), and MLCK4 (chang2016cardiacmyosinlight pages 1-1, chang2016cardiacmyosinlight pages 1-3). The use of transgenic mouse models to study related gene mutations and the study of MLCK in species such as rat and guinea pig indicate the existence of orthologs across mammals (unknownauthors2019exploringmyosinrlc pages 118-121, wang2010mutationsinmyosin pages 2-3, yu2016phosphorylationofthe pages 13-15).

## Reaction Catalyzed

MYLK2 catalyzes the phosphorylation of the regulatory light chain (RLC) of myosin II by transferring the γ-phosphate group from ATP to the hydroxyl group of a specific serine residue on the RLC (chang2016cardiacmyosinlight pages 3-3, hong2011biochemistryofsmooth pages 1-2, thiriet2013cytoplasmicproteinserinethreonine pages 69-73). In skeletal muscle, the specific site of phosphorylation on the RLC is Serine 15 (stull2011myosinlightchain pages 1-2, tsukamoto2013biochemicalandphysiological pages 2-5).

## Cofactor Requirements

The catalytic activity of MYLK2 requires the binding of calcium ions (Ca2+) and the calcium-binding protein calmodulin (CaM) (josephson2011smoothmusclemyosin pages 7-9, kamm2011signalingtomyosin pages 1-2). Activation is achieved when four Ca2+ ions bind to CaM, forming a (Ca2+)4-calmodulin complex that then binds to MYLK2 (stull2011myosinlightchain pages 1-2, unknownauthors2023theinfluenceof pages 26-30). In addition, Mg2+ is an essential cofactor required for the kinase to coordinate with the ATP substrate and facilitate the phosphoryl transfer reaction (stull2011myosinlightchain pages 2-3, temmerman2013structuralandfunctional pages 10-12, chang2016cardiacmyosinlight pages 1-3).

## Substrate Specificity

The specific consensus substrate motif and detailed amino acid preferences from positions P-5 to P+5 for MYLK2 were determined by Johnson et al. (2023) using positional scanning peptide arrays (johnson2023anatlasof pages 1-2). This comprehensive dataset, including position-specific scoring matrices (PSSMs) for MYLK2, is located in the supplementary materials of the publication, specifically noted as being in Supplementary Table 3 and associated Extended Data figures (johnson2023anatlasof pages 2-3, johnson2023anatlasof pages 3-4, johnson2023anatlasof pages 1-2). The provided excerpts from the main text of the article do not contain the explicit motif for MYLK2 (johnson2023anatlasof pages 18-20, johnson2023anatlasof pages 2-3).

## Structure

MYLK2 is a monomeric protein composed of an N-terminal sequence of unknown function, a prototypical bi-lobed catalytic kinase core, and a C-terminal regulatory segment that contains both an autoinhibitory sequence and a calmodulin-binding sequence (stull2011myosinlightchain pages 1-2). The 3D structure, as predicted by the AlphaFold model (AF-Q9H1R3-F1), shows a conserved kinase fold featuring key catalytic and regulatory elements (fang2023molecularinsightsinto pages 1-3, temmerman2013structuralandfunctional pages 9-10, stull2011myosinlightchain pages 2-3). These include a structured activation loop, C-helix, and hydrophobic regulatory (R) spine that stabilizes the active conformation (temmerman2013structuralandfunctional pages 10-12, fang2023molecularinsightsinto pages 16-19). As a member of the DMT kinase family, MYLK2’s activation segment contains a hydrophobic HF/LD motif cluster instead of the canonical HRD motif, and the phenylalanine of the invariant DFG motif adopts an ‘in’ conformation characteristic of an active state (temmerman2013structuralandfunctional pages 10-12). The AlphaFold model also shows the autoinhibitory domain occupying the active site in the absence of calmodulin (fang2023molecularinsightsinto pages 6-8).

## Regulation

MYLK2 activity is principally regulated by allosteric and intrasteric mechanisms (stull2011myosinlightchain pages 1-2). In its inactive state, the enzyme is autoinhibited by its C-terminal regulatory segment, which binds to the catalytic core and physically occludes the substrate-binding site (stull2011myosinlightchain pages 1-2, kamm2011signalingtomyosin pages 1-2). Activation occurs upon the stoichiometric, high-affinity binding of the (Ca2+)4-calmodulin complex to the calmodulin-binding motif (stull2011myosinlightchain pages 2-3). This binding event induces a conformational change that displaces the autoinhibitory segment, exposing the catalytic cleft for substrate phosphorylation (chang2016cardiacmyosinlight pages 1-1, stull2011myosinlightchain pages 2-3). Inactivation is coupled to Ca2+ removal, which leads to a slow dissociation of calmodulin and a return to the autoinhibited state (stull2011myosinlightchain pages 1-2). The overall level of RLC phosphorylation is determined by the balance between MYLK2 activity and the counteracting activity of myosin light chain phosphatase (MLCP) (stull2011myosinlightchain pages 1-2, thiriet2013cytoplasmicproteinserinethreonine pages 69-73).

## Function

MYLK2 is a dedicated Ca2+/calmodulin-dependent serine-threonine protein kinase predominantly expressed in skeletal muscle, with the highest levels found in fast-contracting fibers (stull2011myosinlightchain pages 1-2, kamm2011signalingtomyosin pages 1-2). Its primary role is to phosphorylate the RLC of sarcomeric myosin (stull2011myosinlightchain pages 1-2). This phosphorylation modulates muscle contraction by altering myosin cross-bridge kinetics and increasing the Ca2+ sensitivity of the contractile apparatus (stull2011myosinlightchain pages 1-2). MYLK2 activity also contributes to the proper assembly and organization of sarcomeres in muscle cells (seguchi2007acardiacmyosin pages 2-4).

## Inhibitors

Several experimental small-molecule inhibitors of MLCK have been described, although their specificity for the MYLK2 isoform varies. ML-7 and ML-9 are widely used ATP-competitive inhibitors (kumar2024identificationandbenchmarking pages 1-5, xiong2017myosinlightchain pages 2-3). In a kinase screen, ML-7 inhibited human MYLK2 by 7%, while a more selective probe for MLCK1, Myokinasib-II, inhibited MYLK2 by 17% (kumar2024identificationandbenchmarking pages 17-20). KT5926 is another potent ATP-competitive inhibitor (Ki = 18 nM), but it is not commercially available (kumar2024identificationandbenchmarking pages 1-5). A nonapeptide, rkkykyrrk-NH2 (D-PIK), has also been used experimentally to reduce RLC phosphorylation (yu2016phosphorylationofthe pages 13-15).

## Other Comments

Mutations in the MYLK2 gene are associated with hypertrophic cardiomyopathy (HCM) (li2019advancedevolutionof pages 6-8, qin2021flncandmylk2 pages 10-13). MYLK2 is not typically associated with left ventricular non-compaction (LVNC) (li2019advancedevolutionof pages 6-8). A truncating mutation (p.E380X) has been identified that is predicted to undergo nonsense-mediated decay and has been associated with dilated cardiomyopathy (DCM) and heart failure, suggesting loss of function as a pathogenic mechanism (qin2021flncandmylk2 pages 10-13).

References

1. (chang2016cardiacmyosinlight pages 3-3): Audrey N. Chang, Pravin Mahajan, Stefan Knapp, Hannah Barton, H. Lee Sweeney, Kristine E. Kamm, and James T. Stull. Cardiac myosin light chain is phosphorylated by ca 2+ /calmodulin-dependent and -independent kinase activities. Proceedings of the National Academy of Sciences, 113:E3824-E3833, Jun 2016. URL: https://doi.org/10.1073/pnas.1600633113, doi:10.1073/pnas.1600633113. This article has 59 citations.
2. (hong2011biochemistryofsmooth pages 1-2): Feng Hong, Brian D. Haldeman, Del Jackson, Mike Carter, Jonathan E. Baker, and Christine R. Cremo. Biochemistry of smooth muscle myosin light chain kinase. Archives of biochemistry and biophysics, 510 2:135-46, Jun 2011. URL: https://doi.org/10.1016/j.abb.2011.04.018, doi:10.1016/j.abb.2011.04.018. This article has 149 citations and is from a peer-reviewed journal.
3. (johnson2023anatlasof pages 1-2): Jared L. Johnson, Tomer M. Yaron, Emily M. Huntsman, Alexander Kerelsky, Junho Song, Amit Regev, Ting-Yu Lin, Katarina Liberatore, Daniel M. Cizin, Benjamin M. Cohen, Neil Vasan, Yilun Ma, Konstantin Krismer, Jaylissa Torres Robles, Bert van de Kooij, Anne E. van Vlimmeren, Nicole Andrée-Busch, Norbert F. Käufer, Maxim V. Dorovkov, Alexey G. Ryazanov, Yuichiro Takagi, Edward R. Kastenhuber, Marcus D. Goncalves, Benjamin D. Hopkins, Olivier Elemento, Dylan J. Taatjes, Alexandre Maucuer, Akio Yamashita, Alexei Degterev, Mohamed Uduman, Jingyi Lu, Sean D. Landry, Bin Zhang, Ian Cossentino, Rune Linding, John Blenis, Peter V. Hornbeck, Benjamin E. Turk, Michael B. Yaffe, and Lewis C. Cantley. An atlas of substrate specificities for the human serine/threonine kinome. Nature, 613:759-766, Jan 2023. URL: https://doi.org/10.1038/s41586-022-05575-3, doi:10.1038/s41586-022-05575-3. This article has 446 citations and is from a highest quality peer-reviewed journal.
4. (johnson2023anatlasof pages 2-3): Jared L. Johnson, Tomer M. Yaron, Emily M. Huntsman, Alexander Kerelsky, Junho Song, Amit Regev, Ting-Yu Lin, Katarina Liberatore, Daniel M. Cizin, Benjamin M. Cohen, Neil Vasan, Yilun Ma, Konstantin Krismer, Jaylissa Torres Robles, Bert van de Kooij, Anne E. van Vlimmeren, Nicole Andrée-Busch, Norbert F. Käufer, Maxim V. Dorovkov, Alexey G. Ryazanov, Yuichiro Takagi, Edward R. Kastenhuber, Marcus D. Goncalves, Benjamin D. Hopkins, Olivier Elemento, Dylan J. Taatjes, Alexandre Maucuer, Akio Yamashita, Alexei Degterev, Mohamed Uduman, Jingyi Lu, Sean D. Landry, Bin Zhang, Ian Cossentino, Rune Linding, John Blenis, Peter V. Hornbeck, Benjamin E. Turk, Michael B. Yaffe, and Lewis C. Cantley. An atlas of substrate specificities for the human serine/threonine kinome. Nature, 613:759-766, Jan 2023. URL: https://doi.org/10.1038/s41586-022-05575-3, doi:10.1038/s41586-022-05575-3. This article has 446 citations and is from a highest quality peer-reviewed journal.
5. (josephson2011smoothmusclemyosin pages 7-9): Matthew P. Josephson, Laura A. Sikkink, A. Penheiter, T. Burghardt, and K. Ajtai. Smooth muscle myosin light chain kinase efficiently phosphorylates serine 15 of cardiac myosin regulatory light chain. Biochemical and biophysical research communications, 416 3-4:367-71, Dec 2011. URL: https://doi.org/10.1016/j.bbrc.2011.11.044, doi:10.1016/j.bbrc.2011.11.044. This article has 19 citations and is from a peer-reviewed journal.
6. (kamm2011signalingtomyosin pages 1-2): K. Kamm and J. Stull. Signaling to myosin regulatory light chain in sarcomeres\*. The Journal of Biological Chemistry, 286:9941-9947, Jan 2011. URL: https://doi.org/10.1074/jbc.r110.198697, doi:10.1074/jbc.r110.198697. This article has 142 citations.
7. (seguchi2007acardiacmyosin pages 2-4): O. Seguchi, S. Takashima, S. Yamazaki, M. Asakura, Y. Asano, Y. Shintani, M. Wakeno, T. Minamino, Hiroya Kondo, H. Furukawa, K. Nakamaru, A. Naito, Tomoko Takahashi, Toshiaki Ohtsuka, K. Kawakami, T. Isomura, S. Kitamura, H. Tomoike, N. Mochizuki, and M. Kitakaze. A cardiac myosin light chain kinase regulates sarcomere assembly in the vertebrate heart. The Journal of clinical investigation, 117 10:2812-24, Oct 2007. URL: https://doi.org/10.1172/jci30804, doi:10.1172/jci30804. This article has 188 citations.
8. (stull2011myosinlightchain pages 1-2): J. Stull, K. Kamm, and R. Vandenboom. Myosin light chain kinase and the role of myosin light chain phosphorylation in skeletal muscle. Archives of biochemistry and biophysics, 510 2:120-8, Jun 2011. URL: https://doi.org/10.1016/j.abb.2011.01.017, doi:10.1016/j.abb.2011.01.017. This article has 218 citations and is from a peer-reviewed journal.
9. (unknownauthors2023theinfluenceof pages 26-30): The Influence of a High Fat Diet on Mice with and without Myosin Light Chain Kinase: Implications for Muscle Thermogenesis and Obesity
10. (chang2016cardiacmyosinlight pages 1-1): Audrey N. Chang, Pravin Mahajan, Stefan Knapp, Hannah Barton, H. Lee Sweeney, Kristine E. Kamm, and James T. Stull. Cardiac myosin light chain is phosphorylated by ca 2+ /calmodulin-dependent and -independent kinase activities. Proceedings of the National Academy of Sciences, 113:E3824-E3833, Jun 2016. URL: https://doi.org/10.1073/pnas.1600633113, doi:10.1073/pnas.1600633113. This article has 59 citations.
11. (chang2016cardiacmyosinlight pages 1-3): Audrey N. Chang, Pravin Mahajan, Stefan Knapp, Hannah Barton, H. Lee Sweeney, Kristine E. Kamm, and James T. Stull. Cardiac myosin light chain is phosphorylated by ca 2+ /calmodulin-dependent and -independent kinase activities. Proceedings of the National Academy of Sciences, 113:E3824-E3833, Jun 2016. URL: https://doi.org/10.1073/pnas.1600633113, doi:10.1073/pnas.1600633113. This article has 59 citations.
12. (fang2023molecularinsightsinto pages 1-3): Xuan Fang, Vladimir Bogdanov, Jonathan P. Davis, and Peter M. Kekenes-Huskey. Molecular insights into the mlck activation by cam. Journal of Chemical Information and Modeling, 63:7487-7498, Nov 2023. URL: https://doi.org/10.1021/acs.jcim.3c00954, doi:10.1021/acs.jcim.3c00954. This article has 8 citations and is from a peer-reviewed journal.
13. (fang2023molecularinsightsinto pages 16-19): Xuan Fang, Vladimir Bogdanov, Jonathan P. Davis, and Peter M. Kekenes-Huskey. Molecular insights into the mlck activation by cam. Journal of Chemical Information and Modeling, 63:7487-7498, Nov 2023. URL: https://doi.org/10.1021/acs.jcim.3c00954, doi:10.1021/acs.jcim.3c00954. This article has 8 citations and is from a peer-reviewed journal.
14. (fang2023molecularinsightsinto pages 6-8): Xuan Fang, Vladimir Bogdanov, Jonathan P. Davis, and Peter M. Kekenes-Huskey. Molecular insights into the mlck activation by cam. Journal of Chemical Information and Modeling, 63:7487-7498, Nov 2023. URL: https://doi.org/10.1021/acs.jcim.3c00954, doi:10.1021/acs.jcim.3c00954. This article has 8 citations and is from a peer-reviewed journal.
15. (herring2000smoothmusclemyosin pages 8-9): B. Paul Herring, Shelley Dixon, and Patricia J. Gallagher. Smooth muscle myosin light chain kinase expression in cardiac and skeletal muscle. American Journal of Physiology-Cell Physiology, 279:C1656-C1664, Nov 2000. URL: https://doi.org/10.1152/ajpcell.2000.279.5.c1656, doi:10.1152/ajpcell.2000.279.5.c1656. This article has 78 citations.
16. (johnson2023anatlasof pages 18-20): Jared L. Johnson, Tomer M. Yaron, Emily M. Huntsman, Alexander Kerelsky, Junho Song, Amit Regev, Ting-Yu Lin, Katarina Liberatore, Daniel M. Cizin, Benjamin M. Cohen, Neil Vasan, Yilun Ma, Konstantin Krismer, Jaylissa Torres Robles, Bert van de Kooij, Anne E. van Vlimmeren, Nicole Andrée-Busch, Norbert F. Käufer, Maxim V. Dorovkov, Alexey G. Ryazanov, Yuichiro Takagi, Edward R. Kastenhuber, Marcus D. Goncalves, Benjamin D. Hopkins, Olivier Elemento, Dylan J. Taatjes, Alexandre Maucuer, Akio Yamashita, Alexei Degterev, Mohamed Uduman, Jingyi Lu, Sean D. Landry, Bin Zhang, Ian Cossentino, Rune Linding, John Blenis, Peter V. Hornbeck, Benjamin E. Turk, Michael B. Yaffe, and Lewis C. Cantley. An atlas of substrate specificities for the human serine/threonine kinome. Nature, 613:759-766, Jan 2023. URL: https://doi.org/10.1038/s41586-022-05575-3, doi:10.1038/s41586-022-05575-3. This article has 446 citations and is from a highest quality peer-reviewed journal.
17. (johnson2023anatlasof pages 3-4): Jared L. Johnson, Tomer M. Yaron, Emily M. Huntsman, Alexander Kerelsky, Junho Song, Amit Regev, Ting-Yu Lin, Katarina Liberatore, Daniel M. Cizin, Benjamin M. Cohen, Neil Vasan, Yilun Ma, Konstantin Krismer, Jaylissa Torres Robles, Bert van de Kooij, Anne E. van Vlimmeren, Nicole Andrée-Busch, Norbert F. Käufer, Maxim V. Dorovkov, Alexey G. Ryazanov, Yuichiro Takagi, Edward R. Kastenhuber, Marcus D. Goncalves, Benjamin D. Hopkins, Olivier Elemento, Dylan J. Taatjes, Alexandre Maucuer, Akio Yamashita, Alexei Degterev, Mohamed Uduman, Jingyi Lu, Sean D. Landry, Bin Zhang, Ian Cossentino, Rune Linding, John Blenis, Peter V. Hornbeck, Benjamin E. Turk, Michael B. Yaffe, and Lewis C. Cantley. An atlas of substrate specificities for the human serine/threonine kinome. Nature, 613:759-766, Jan 2023. URL: https://doi.org/10.1038/s41586-022-05575-3, doi:10.1038/s41586-022-05575-3. This article has 446 citations and is from a highest quality peer-reviewed journal.
18. (kumar2024identificationandbenchmarking pages 1-5): Gautam Kumar, Prema Kumari Agarwala, Aswin T. Srivatsav, Ashok Ravula, G. Ashmitha, Sreenath Balakrishnan, Shobhna Kapoor, and Rishikesh Narayan. Identification and benchmarking of myokinasib-ii as a selective and potent chemical probe for exploring mlck1 inhibition. ACS Chemical Biology, 19:2165-2175, Sep 2024. URL: https://doi.org/10.1021/acschembio.4c00336, doi:10.1021/acschembio.4c00336. This article has 2 citations and is from a domain leading peer-reviewed journal.
19. (kumar2024identificationandbenchmarking pages 17-20): Gautam Kumar, Prema Kumari Agarwala, Aswin T. Srivatsav, Ashok Ravula, G. Ashmitha, Sreenath Balakrishnan, Shobhna Kapoor, and Rishikesh Narayan. Identification and benchmarking of myokinasib-ii as a selective and potent chemical probe for exploring mlck1 inhibition. ACS Chemical Biology, 19:2165-2175, Sep 2024. URL: https://doi.org/10.1021/acschembio.4c00336, doi:10.1021/acschembio.4c00336. This article has 2 citations and is from a domain leading peer-reviewed journal.
20. (stull2011myosinlightchain pages 2-3): J. Stull, K. Kamm, and R. Vandenboom. Myosin light chain kinase and the role of myosin light chain phosphorylation in skeletal muscle. Archives of biochemistry and biophysics, 510 2:120-8, Jun 2011. URL: https://doi.org/10.1016/j.abb.2011.01.017, doi:10.1016/j.abb.2011.01.017. This article has 218 citations and is from a peer-reviewed journal.
21. (temmerman2013structuralandfunctional pages 10-12): Koen Temmerman, Bertrand Simon, and Matthias Wilmanns. Structural and functional diversity in the activity and regulation of dapk‐related protein kinases. The FEBS Journal, Nov 2013. URL: https://doi.org/10.1111/febs.12384, doi:10.1111/febs.12384. This article has 42 citations.
22. (temmerman2013structuralandfunctional pages 9-10): Koen Temmerman, Bertrand Simon, and Matthias Wilmanns. Structural and functional diversity in the activity and regulation of dapk‐related protein kinases. The FEBS Journal, Nov 2013. URL: https://doi.org/10.1111/febs.12384, doi:10.1111/febs.12384. This article has 42 citations.
23. (thiriet2013cytoplasmicproteinserinethreonine pages 69-73): Marc Thiriet. Cytoplasmic protein serine/threonine kinases. Biomathematical and Biomechanical Modeling of the Circulatory and Ventilatory Systems, pages 175-310, Jul 2013. URL: https://doi.org/10.1007/978-1-4614-4370-4\_5, doi:10.1007/978-1-4614-4370-4\_5. This article has 12 citations.
24. (tsukamoto2013biochemicalandphysiological pages 2-5): Osamu Tsukamoto and Masafumi Kitakaze. Biochemical and physiological regulation of cardiac myocyte contraction by cardiac-specific myosin light chain kinase. Circulation journal : official journal of the Japanese Circulation Society, 77 9:2218-25, Aug 2013. URL: https://doi.org/10.1253/circj.cj-13-0627, doi:10.1253/circj.cj-13-0627. This article has 34 citations.
25. (unknownauthors2019exploringmyosinrlc pages 118-121): Exploring Myosin RLC Phosphorylation as a Potential Novel Therapeutic Target for Hypertrophic Cardiomyopathy
26. (li2019advancedevolutionof pages 6-8): Chia-Jung Li, Chien-Sheng Chen, Giou-Teng Yiang, A. Tsai, Wan-Ting Liao, and Meng-Yu Wu. Advanced evolution of pathogenesis concepts in cardiomyopathies. Journal of Clinical Medicine, Apr 2019. URL: https://doi.org/10.3390/jcm8040520, doi:10.3390/jcm8040520. This article has 22 citations and is from a peer-reviewed journal.
27. (qin2021flncandmylk2 pages 10-13): Xianyu Qin, Ping Li, H. Qu, Yichuan Liu, Yu Xia, Shaoxian Chen, Yongchao Yang, Shufang Huang, Pengju Wen, Xianwu Zhou, Xiao-Fei Li, Yonghua Wang, Lifeng Tian, H. Hakonarson, Yueheng Wu, and Zhuang Jian. Flnc and mylk2 gene mutations in a chinese family with different phenotypes of cardiomyopathy. International heart journal, May 2021. URL: https://doi.org/10.1101/2020.05.10.20097519, doi:10.1101/2020.05.10.20097519. This article has 11 citations and is from a peer-reviewed journal.
28. (wang2010mutationsinmyosin pages 2-3): Li Wang, Dong-chuan Guo, Jiumei Cao, Limin Gong, Kristine E. Kamm, Ellen Regalado, Li Li, Sanjay Shete, Wei-Qi He, Min-Sheng Zhu, Stephan Offermanns, Dawna Gilchrist, John Elefteriades, James T. Stull, and Dianna M. Milewicz. Mutations in myosin light chain kinase cause familial aortic dissections. American journal of human genetics, 87 5:701-7, Nov 2010. URL: https://doi.org/10.1016/j.ajhg.2010.10.006, doi:10.1016/j.ajhg.2010.10.006. This article has 372 citations and is from a highest quality peer-reviewed journal.
29. (xiong2017myosinlightchain pages 2-3): Yongjian Xiong, Chenou Wang, Liqiang Shi, Liang Wang, Zijuan Zhou, Dapeng Chen, Jingyu Wang, and Hui-shu Guo. Myosin light chain kinase: a potential target for treatment of inflammatory diseases. Frontiers in Pharmacology, May 2017. URL: https://doi.org/10.3389/fphar.2017.00292, doi:10.3389/fphar.2017.00292. This article has 72 citations and is from a peer-reviewed journal.
30. (yu2016phosphorylationofthe pages 13-15): Haiyang Yu, Samya Chakravorty, Weihua Song, and Michael A. Ferenczi. Phosphorylation of the regulatory light chain of myosin in striated muscle: methodological perspectives. European Biophysics Journal, 45:779-805, Apr 2016. URL: https://doi.org/10.1007/s00249-016-1128-z, doi:10.1007/s00249-016-1128-z. This article has 45 citations.