HSC1_MUT vs. HSC1_WT

NP

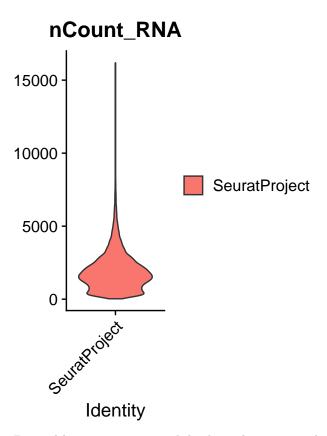
1/20/2022

```
# https://satijalab.org/signac/articles/pbmc_multiomic.html
counts <- Read10X_h5("filtered_feature_bc_matrix.h5")</pre>
fragpath <- "atac_fragments.tsv.gz"</pre>
# Get gene annotations for HG38
annotation <- GetGRangesFromEnsDb(ensdb = EnsDb.Hsapiens.v86)
genome(annotation) <- "hg38"</pre>
seqlevelsStyle(annotation) <- "UCSC"</pre>
# Create a Seurat object containing the RNA data
npm1 <- CreateSeuratObject(</pre>
        counts = counts$`Gene Expression`,
        assay = "RNA"
)
# npm1[["percent.mt"]] <- PercentageFeatureSet(npm1, pattern = "^MT-")</pre>
# # create ATAC assay and add it to the object
#
# npm1[["ATAC"]] <- CreateChromatinAssay(</pre>
         counts = counts$Peaks,
#
          sep = c(":", "-"),
#
          fragments = fragpath,
#
          annotation = annotation
#
# )
```

Violin plot of RNA counts

```
DefaultAssay(npm1) <- "RNA"

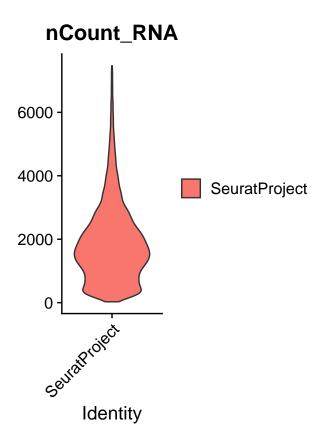
VlnPlot(
  object = npm1,
  features = c("nCount_RNA"),
  ncol = 4,
  pt.size = 0
)</pre>
```



Discard low count genes and display subsequent violin plot.

```
npm1 <- subset(
    x = npm1,
    subset = nCount_RNA < 7500
)

VlnPlot(
    object = npm1,
    features = c("nCount_RNA"),
    ncol = 4,
    pt.size = 0
)</pre>
```



Dimensionality reduction: SC transform and PCA

```
#npm1 <- SCTransform(npm1) %>% RunPCA() %>% RunUMAP(dims = 1:50, reduction.name = 'umap.rna', reduction
npm1 <- SCTransform(npm1)</pre>
##
##
npm1 <- RunPCA(npm1)</pre>
Add cell identity status from Seurat object to NPM1 Seurat metadata.
reference <- readRDS("~/Downloads/namlab/NPM1_seurat/NPM1_seurat.rds")</pre>
npm1 <- AddMetaData(</pre>
        object = npm1,
        metadata = reference@meta.data %% select(Cell.Ident_Mutation.Status) %>% filter(rownames(.) %i
Idents(npm1) <- "Cell.Ident_Mutation.Status"</pre>
Find DEGs using FindMarkers(). Convert gene symbols to ENTREZ IDs and add to dataframe.
```

```
DefaultAssay(npm1) <- "SCT"</pre>
stem_cell_markers_1 <- FindMarkers(npm1, ident.1 = "HSC1_MUT", ident.2 = "HSC1_WT", only.pos = FALSE, 1
stem_cell_markers_1$entrez = mapIds(org.Hs.eg.db, rownames(stem_cell_markers_1), 'ENTREZID', 'SYMBOL')
stem_cell_markers_1 = na.omit(stem_cell_markers_1)
```

Split DEG list into upregulated and downregulated genes.

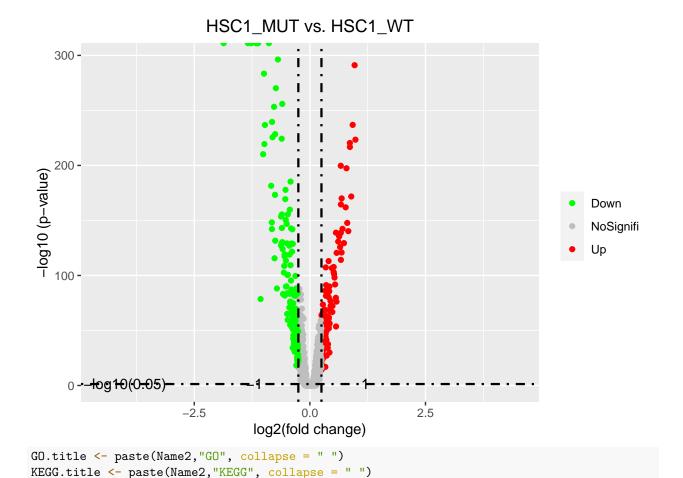
```
upreg = subset(stem_cell_markers_1, subset = avg_log2FC > 0.25 & p_val_adj < 0.01)</pre>
downreg = subset(stem_cell_markers_1, subset = avg_log2FC < -0.25 & p_val_adj < 0.01)</pre>
Create sorted list of upregulated and downregulated genes.
upreg_list <- sign(upreg$avg_log2FC)*(-log10(upreg$p_val_adj))</pre>
names(upreg_list) <- rownames(upreg)</pre>
upreg_list <- upreg_list[na.exclude(names(upreg_list))]</pre>
upreg_list <- sort(upreg_list, decreasing = T)</pre>
downreg_list <- sign(downreg$avg_log2FC)*(-log10(downreg$p_val_adj))</pre>
names(downreg list) <- rownames(downreg)</pre>
downreg_list <- downreg_list[na.exclude(names(downreg_list))]</pre>
downreg_list <- sort(downreg_list, decreasing = T)</pre>
downreg_list = replace(downreg_list, c(which(downreg_list %in% -Inf)),-(.Machine$double.xmax/100))
full list = sign(stem cell markers 1$avg log2FC)*(-log10(stem cell markers 1$p val adj))
names(full list) <- rownames(stem cell markers 1)</pre>
full_list <- full_list[na.exclude(names(full_list))]</pre>
full_list <- sort(full_list, decreasing = T)</pre>
full_list = replace(full_list, c(which(full_list %in% -Inf)),-(.Machine$double.xmax/100))
```

Volcano plot

```
Name2 = "HSC1_MUT vs. HSC1_WT"

stem_cell_markers_1$threshold <- as.factor(ifelse(stem_cell_markers_1$p_val_adj < 0.05 & abs(stem_cell_second))

ggplot(data=stem_cell_markers_1, aes(x=avg_log2FC, y=-log10(p_val_adj), colour=threshold)) +
geom_point(alpha=1, size=1.5) +
scale_color_manual(values=c("green", "grey", "red")) +
xlim(c(-4.5, 4.5)) +
geom_vline(xintercept=c(-.25, .25), lty=4,col="black",lwd=0.8) +
geom_hline(yintercept=-log10(0.05), lty=4,col="black",lwd=0.8) +
annotate("text", x=c(-1.2, 1.2), y=1.8, label=c("-1", "1")) +
annotate("text", x=-4, y=1.8, label="-log10(0.05)") +
labs(x="log2(fold change)", y="-log10 (p-value)", title=Name2) +
theme(plot.title=element_text(hjust=0.5), legend.position="right", legend.title=element_blank())</pre>
```



GSEA

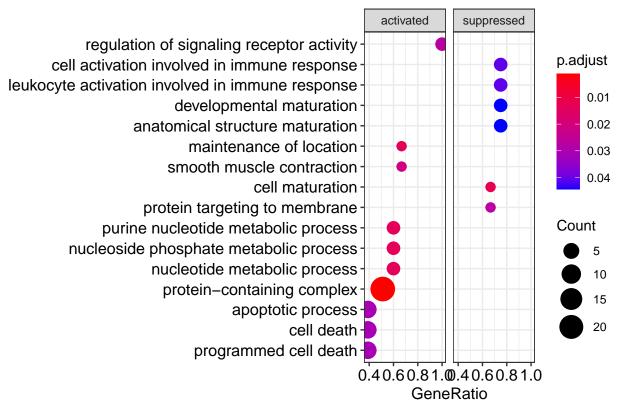
Perform GSEA on up- and downregulated genes.

```
gse_up <- gseGO(geneList=upreg_list,</pre>
             ont ="ALL",
             keyType = "SYMBOL",
             minGSSize = 3,
             maxGSSize = 800,
             pvalueCutoff = 0.05,
             verbose = TRUE,
             OrgDb = org.Hs.eg.db,
             pAdjustMethod = "none")
gse_down <- gseGO(geneList=downreg_list,</pre>
             ont ="ALL",
             keyType = "SYMBOL",
             minGSSize = 3,
             maxGSSize = 800,
             pvalueCutoff = 0.05,
             verbose = TRUE,
             OrgDb = org.Hs.eg.db,
             pAdjustMethod = "none")
```

Dot plots

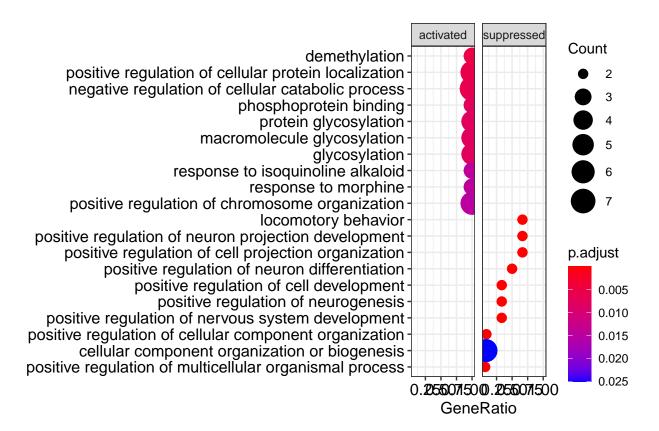
Upregulated

```
require(DOSE)
dotplot(gse_up, showCategory=10, split=".sign") + facet_grid(.~.sign)
```



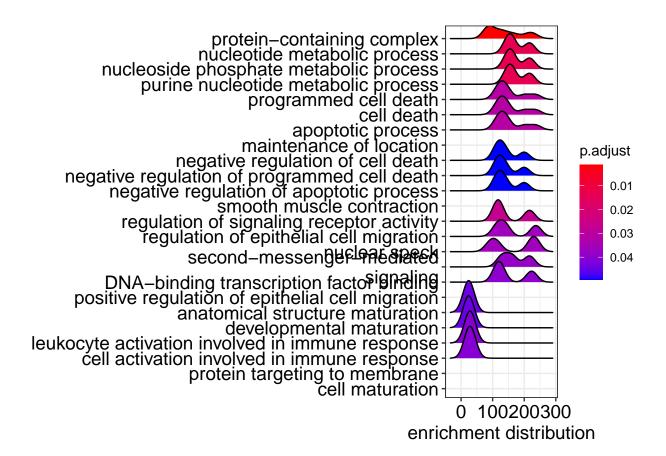
Downregulated

```
require(DOSE)
dotplot(gse_down, showCategory=10, split=".sign") + facet_grid(.~.sign)
```



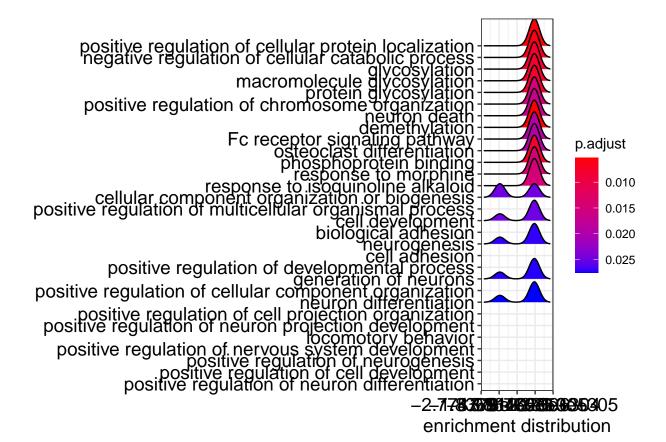
Ridge plot (frequency of fold values per gene within each set) Upregulated

```
ridgeplot(gse_up) + labs(x = "enrichment distribution")
```



Downregulated

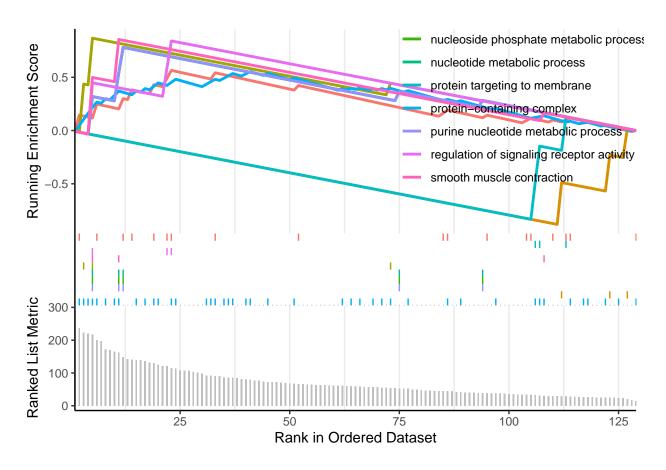
ridgeplot(gse_down) + labs(x = "enrichment distribution")



GSEA plot

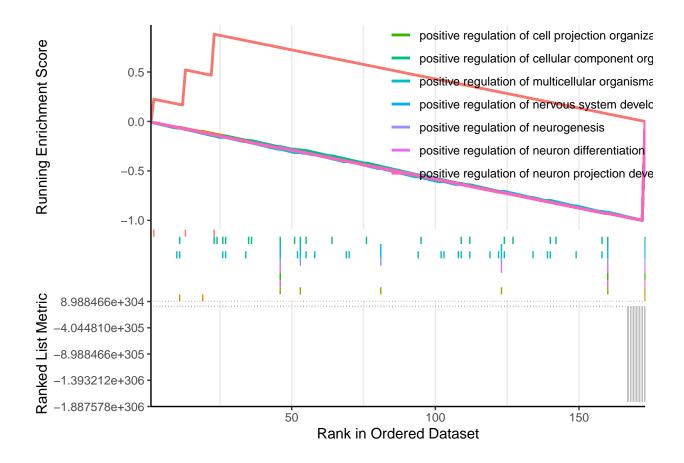
Upregulated

```
#gseaplot(gse, by = "all", title = gse$Description[3], geneSetID = 3)
gseaplot2(gse_up, geneSetID=1:10)
```



Downregulated

```
#gseaplot(gse, by = "all", title = gse$Description[3], geneSetID = 3)
gseaplot2(gse_down, geneSetID=1:10)
```



KEGG GSEA

Create gseKEGG objects

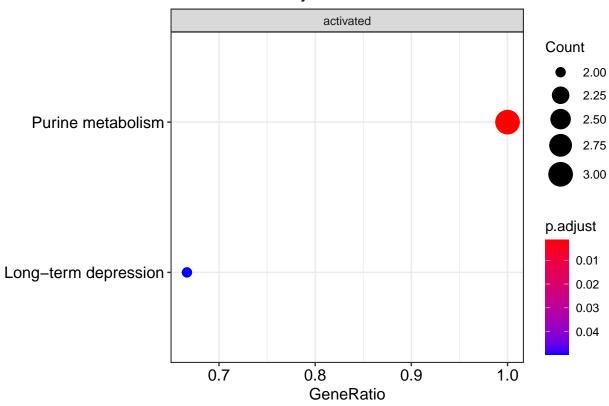
```
kegg organism = "hsa"
names(upreg_list) = mapIds(org.Hs.eg.db, names(upreg_list), 'ENTREZID', 'SYMBOL')
names(downreg_list) = mapIds(org.Hs.eg.db, names(downreg_list), 'ENTREZID', 'SYMBOL')
kk2_up <- gseKEGG(geneList = upreg_list,
               organism
                            = kegg_organism,
               minGSSize
                            = 3,
               maxGSSize
                            = 800.
               pvalueCutoff = 0.05,
               pAdjustMethod = "none",
                             = "ncbi-geneid")
               keyType
kk2_down <- gseKEGG(geneList = downreg_list,
                            = kegg_organism,
               organism
               minGSSize
               maxGSSize
                            = 800,
               pvalueCutoff = 0.05,
               pAdjustMethod = "none",
               keyType
                           = "ncbi-geneid")
```

Dot plot

Upregulated

dotplot(kk2_up, showCategory = 10, title = "Enriched Pathways" , split=".sign") + facet_grid(.~.sign)

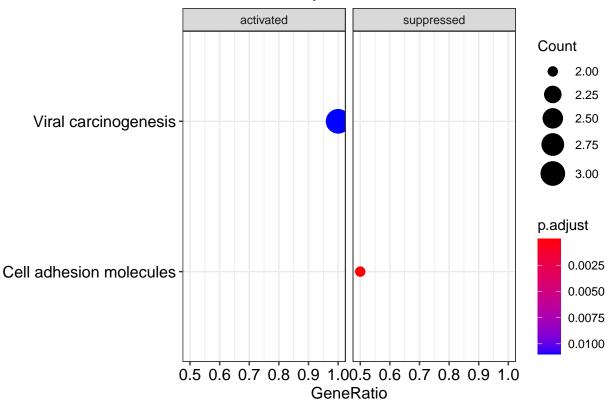
Enriched Pathways



Downregulated

dotplot(kk2_down, showCategory = 10, title = "Enriched Pathways" , split=".sign") + facet_grid(.~.sign)

Enriched Pathways



MSigDB

Hallmark

```
#all_gene_sets = msigdbr(species = "Homo sapiens")
h_gene_sets = msigdbr(species = "human", category = "H")
pathwaysH = split(x = h_gene_sets$entrez_gene, f = h_gene_sets$gs_name)
```

Upregulated - NONE with padj < 0.05

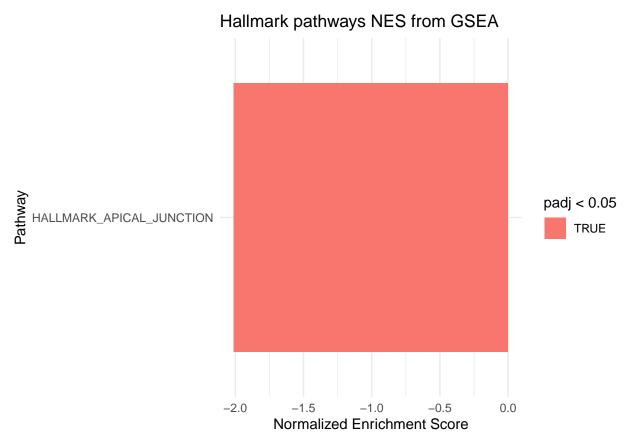
```
theme_minimal()
```

Hallmark pathways NES from GSEA

Pathway

Normalized Enrichment Score

Downregulated



| Pathway | | Gen | e ranks | 3 | NES | pval | padj |
|-----------------------------|---|-----|---------|-----|-------|---------|---------|
| "IVE_OXYGEN_SPECIES_PATHWAY | | | | | -1.37 | 3.4e-01 | 9.8e-01 |
| HALLMARK_NOTCH_SIGNALING | | | | | -1.40 | 3.2e-01 | 9.8e-01 |
| MARK_XENOBIOTIC_METABOLISM | | | ** | | -1.33 | 2.5e-01 | 9.8e-01 |
| HALLMARK_ADIPOGENESIS | | | | | -1.33 | 2.5e-01 | 9.8e-01 |
| HALLMARK_COAGULATION | | | | | -1.46 | 2.4e-01 | 9.8e-01 |
| HALLMARK_COMPLEMENT | | | | | -1.30 | 2.2e-01 | 9.8e-01 |
| LLMARK_PANCREAS_BETA_CELLS | | | | • | -1.76 | 1.5e-01 | 9.8e-01 |
| ARK_PI3K_AKT_MTOR_SIGNALING | | | | | -1.87 | 9.4e-02 | 9.8e-01 |
| _LMARK_ALLOGRAFT_REJECTION | • | | | | -1.94 | 7.1e-02 | 9.8e-01 |
| HALLMARK_APICAL_JUNCTION | | | | | -2.01 | 1.0e-10 | 3.9e-09 |
| | 0 | 50 | 100 | 150 | | | |

All

Biocarta

```
#all_gene_sets = msigdbr(species = "Homo sapiens")
b_gene_sets = msigdbr(species = "human", category = "C2", subcategory = "CP:BIOCARTA")
pathwaysB = split(x = b_gene_sets$entrez_gene, f = b_gene_sets$gs_name)
```

Upregulated - NONE with padj < 0.05

Downregulated - NONE with padj < 0.05

All - NONE with padj < 0.05

Pathway

Normalized Enrichment Score

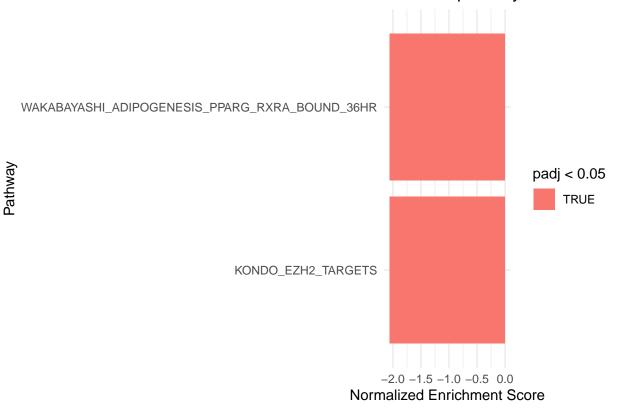
CGP

```
#all_gene_sets = msigdbr(species = "Homo sapiens")
c_gene_sets = msigdbr(species = "human", category = "C2", subcategory = "CGP")
pathwaysC = split(x = c_gene_sets$entrez_gene, f = c_gene_sets$gs_name)
```

Upregulated - NONE with padj < 0.05

Downregulated

Hallmark pathways NES from (



All