

An image-computable spatio-chromatic receptive
field model of the midget retinal ganglion mosaic
across the retina

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Abstract

Image-computable models of retinal ganglion cell (RGC) mosaics that are synthesized and constrained jointly by optical, anatomical and physiological properties, and which operate on images defined by their spatial-spectral radiance, do not currently exist. Here, we deploy a novel computational framework which synthesizes mosaics of linear spatio-chromatic receptive fields (RFs) of ON midget RGCs (mRGCs) by integrating published anatomical, physiological, and optical quality measurements, all varying with eccentricity. We use the synthesized mRGC mosaics to simulate both *in vivo* and *in vitro* physiological experiments and demonstrate the model's consistency with published data. The model enables computation of how visual performance is shaped by the representation of visual information provided by the linear spatiochromatic processing stage of midget RGCs. The developed computational framework carefully accounts for the effect of physiological optics on mRGC responses, enables comparison of *in vivo* and *in vitro* data, and allows exploration of how different assumptions about RF organization, such as selectivity for the type of cones pooled by the RF center mechanism, affect physiological responses and psychophysical performance. The open-source and freely available implementation provides a platform for understanding how the linear spatiochromatic receptive field representation of the mRGCs shapes visual performance, as well as a foundation for future work that incorporates response nonlinearities, temporal filtering, and extends to additional RGC mosaics.

Keywords: retinal ganglion cells, receptive field, model

047 **1 Introduction**

048

049 An important aim in computational visual neuroscience is to create accurate computer
050 simulations of how neurons in the visual pathways encode and respond to visual scenes.
051 These simulations, often called digital twins, are a quantitative description of the
052 visual system. They enable links between the neural representation and perception
053 and provide a tool for evaluating the effects of blinding disease and its treatment.

054 Over the last ten years we have built an open-source software platform, ISETBio
055 (Image Systems Engineering Tools for Biology) [1], which serves as a digital twin for
056 the initial stages of the human visual system. Previously, we described how ISETBio
057 models (a) the formation of the retinal image, (b) the excitation of the cone pho-
058 toreceptors, (c) phototransduction, and (d) fixational eye movements [2–4]. We and
059 others have employed ISETBio to model human vision, including sensitivity to spa-
060 tial contrast [2, 3], the impact of chromatic aberration on acuity [5], the encoding of
061 information from natural images captured by cones [6], the effects of optics and cone
062 density across the visual field on performance [7], and the influence of initial visual sig-
063 nals on tasks like judging surface properties and lighting [8, 9]. We also used ISETBio
064 to help interpret experimental measurements of retinal ganglion cells [10].

065 Here, we describe an extension of ISETBio to model the mosaic of a class of retinal
066 ganglion cells (RGCs), the midget RGC (mRGC) mosaic. RGCs are the only pathway
067 for information transmission from the retina to the brain, and their properties surely
068 impact visual performance on many tasks. The spike trains transmitted via the axons
069 of one million RGCs that form the human optic nerve, represent the signals from
070 roughly 6.5 million cones and 110 million rods [11, 12]. Of these RGCs, mRGCs are
071 a particularly important subtype, comprising 80% of the perifoveal RGCs and 45%
072 of the peripheral RGCs. In the very central fovea, it has been estimated that the
073 mRGCs are 95% of the RGC population [13].

074 The role of the mRGCs in limiting spatial and color vision is still debated [14].
075 Simulation of performance using image computable models of the mRGC mosaic offers
076 a powerful tool for understanding the visual information encoded by these cells, espe-
077 cially because they are very hard to measure and isolate experimentally. We have four
078 primary goals for this human retina model.

079 First, the model must distinguish the contributions of the eye’s optics and pho-
080 toreceptors from the subsequent post-receptoral retinal circuitry. This separation is
081 crucial for incorporating key physiological measurements, some of which are made *in*
082 *vitro* without the eye’s optics. Failing to isolate the optical effects would prevent us
083 from using this vital collection of data.

084 Second, the model must capture responses across a large portion of central
085 retina. This is important because we and others are interested in how the retinal
086 representation shapes performance not just in the fovea but also for peripheral viewing.

087 Third, the model must integrate diverse data types, including optical, anatomical,
088 and physiological measurements. A comprehensive formulation is necessary because
089 retinal ganglion cell (RGC) responses are shaped by all three of these factors.

090 Fourth, we aim for an extensible framework. The current implementation uses a
091 linear spatiochromatic receptive field, which serves as a good initial approximation.
092

The framework is designed to incorporate future extensions—such as response non-linearities, temporal dynamics, and additional RGC classes—to improve the model’s accuracy over time. The following points describe how our implementation achieves these goals.	093
1. <i>Separating representations.</i> Our mRGC model operates on the cone mosaic signals. This design isolates the post-receptoral circuitry (cone-to-mRGC), which is the pathway measured in <i>in vitro</i> experiments where the eye’s optics are removed [15, 16]. This separation is also valuable for interpreting experiments that use adaptive optics to eliminate optical blur [10]. While the components are separable, our implementation integrates the optics, cone sampling, and mRGC circuitry into a complete, image-computable pipeline. This full pathway allows us to simulate the transformation of a visual stimulus into an mRGC response, matching the conditions of <i>in vivo</i> measurements [17–19] and enabling predictions of human performance under natural viewing conditions.	094
2. <i>Representation across the visual field.</i> Visual performance varies across the visual field, and a key contribution of our model is that it allows computation of the mRGC representation continuously across the retina from the fovea out to 30°, along any meridian. Achieving this goal required implementation of novel algorithms for synthesizing mRGC RF mosaics.	095
3. <i>Multiple data types.</i> By explicitly representing different biological stages, our model enables algorithms that combine anatomical, physiological, and optical data. Incorporation of multiple types of measurements from the literature is critical because at present no one type of data sufficiently constrains mRGC properties across the visual field.	096
4. <i>Extensible.</i> The current implementation is a linear spatial pooling model, a useful approximation for stimuli with modest contrast. The software’s modular design provides a foundation for future extensions. We can incorporate known nonlinear properties that shape mRGC responses, including phototransduction effects [20]; spatial and static nonlinearities, which often differ between ON and OFF pathways [21–24]; temporal dynamics [25]; and response noise [26]. Furthermore, the mRGC model is a suitable base for developing models of other types of RGCs, such as parasol and bistratified cells [27].	097
1.1 Model overview	098
Fig. 1 provides a model overview. Computation begins with the image spectral radiance, such as produced by a calibrated monitor. A model of the human optics (including chromatic aberrations) and spectral filtering by the lens is used to compute the retinal irradiance. Retinal irradiance is spectrally filtered by the macular pigment and then spatially and spectrally sampled by the cone photoreceptor mosaic. The parameters of the optics, macular pigment and cone mosaic all vary across the visual field, according to measurements in the literature [2].	099
The mRGC mosaic extension is composed of spatial receptive fields (RFs) whose center and surround responses are weighted sums of signals from the cone mosaic.	100
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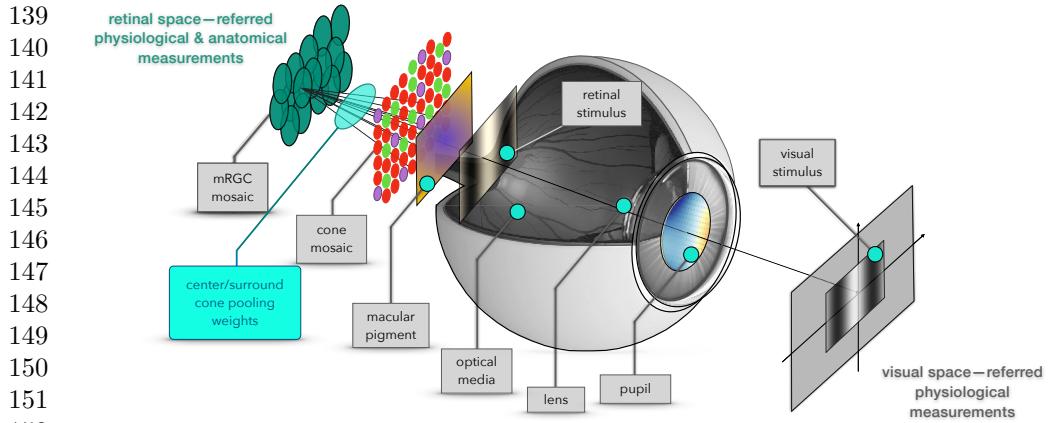


Fig. 1 Model overview. The extant ISETBio model computes the mosaic of cone excitations. The model mRGCs are obtained by connecting their RFs to the cone mosaic. The connectivity matrix is constrained by anatomy and optimized through forward simulation of physiological measurements, so that the synthetic mRGCs are consistent with optical, anatomical and physiological data across the visual field.

The wiring between the input cone mosaic and the mRGC mosaic is initially determined based on anatomical constraints, such as cone and mRGC densities, and is subsequently refined using optimization algorithms that align the model's spatial RF properties with physiological measurements.

A key challenge is the scarcity of *in vitro* physiological data across the visual field which could be used to directly determine the wiring between the two mosaics. To address this, our algorithm primarily leverages more widely available *in vivo* data for the optimization, while validating the model against *in vitro* data where it exists. The resulting model is simultaneously consistent with cone light encoding, anatomical properties (including those of mRGCs and H1 horizontal cells), and both *in vitro* and *in vivo* physiological data. This makes the model versatile for simulating visual stimulation under *in vivo*, *in vitro*, and adaptive optics paradigms.

1.1.1 Relationship to previous computational models of RGCs

To our knowledge, no previous model of RGCs has attempted to realistically capture the effects of the front end encoding in the visual system, specifically the eccentricity and wavelength-varying nature of physiological optics, and the eccentricity-varying spatio-chromatic properties of the cone mosaic. Instead previous models of RGCs have either completely ignored the impact of physiological optics [28, 29], or employed very simplistic models of the eye's optics [30]. Moreover, previous RGC models were designed to either operate directly on visual space, completely ignoring the spatio-spectral filtering by the tri-chromatic cone mosaic [28], or have employed simplistic implementations of the cone mosaic [30]. Finally, none of the previous models are constructed to operate on stimuli defined in terms of their physical spatial-spectral radiance, as they are designed to operate on light intensity defined stimuli [28, 30]. As

such, previous models can not capture the rich spatio-chromatic interactions between stimuli and physiological optics, and how their combined effects shape RGC responses. Indeed, we have recently shown that spatio-chromatic interactions between stimuli and physiological optics can have profound effects of the response properties of midget ganglion cells [31].	185 186 187 188 189
On the other hand, previous computational models of RGCs have focused on other, also important, components of the RGC circuit, that our linear spatio-chromatic model does not currently address, such as processing by retinal interneurons [28–30, 32], temporal dynamics [28–30, 32], contrast grain control [28, 33], and spike generation [28, 29, 33]. We plan to extend our linear spatiochromatic model of mRGCs to include several of these components, as described in section 4.2.2.	190 191 192 193 194 195 196
1.1.2 Paper organization	197
The remainder of this paper is organized as follows.	198 199
• In section 2 we describe the model’s construction stages, including, how the mRGC receptive field lattice is generated from anatomical data (section 2.1), how cones get connected to the mRGC RF centers using anatomical and physiological constraints (section 2.2), and how cone connections to mRGC RF surrounds are derived by optimizing against <i>in vivo</i> data (section 2.3).	200 201 202 203 204
• In section 3 we present, validate, and discuss a first application of the model. Specifically, we illustrate examples of synthesized mRGC mosaics (section 3.1), confirm that the model mRGC spatial RFs are consistent with <i>in vivo</i> (section 3.2), and <i>in vitro</i> data (section 3.3), demonstrate the significant impact of physiological optics (section 3.4), and how simpler Difference-of-Gaussians models can fail to capture the true surround pooling (section 3.5), and finally we present first applications of the developed model, which illustrate how the model can be used to estimate the contribution of the mRGC mosaic to spatiochromatic contrast sensitivity across the visual field (section 3.6).	205 206 207 208 209 210 211 212 213
• In section 4, we summarize our work, discuss ongoing applications of the model in its current stage, and discuss the model’s present limitations and planned expansions.	214 215 216
2 Methods	217 218
The synthesis of mRGC RF mosaics occurs in three stages. In the first stage, we generate spatial lattices representing the RF centers of cells in the mRGC mosaic and the position of cones of the cone mosaic that provides input to the mRGC mosaic. In the second stage, we connect the input cone mosaic to the RF centers of the mRGC mosaic. In the third stage, we connect the input cone mosaic to the RF surrounds of the mRGC mosaic.	219 220 221 222 223 224 225 226 227 228 229 230

231 **2.1 Generating the spatial position lattice of mRGC RF
232 centers (Stage 1)**

233 We begin by generating a lattice that represents the (x, y) positions of mRGC RF
234 centers. This process comprises three sub-stages, components of which are illustrated
235 in Fig. 2.

- 236 • **Stage 1A:** We estimate the mRGC RF center densities along the four principal
237 meridians (0° , 90° , 180° , and 270°). These estimates are based on human data
238 [34, 35]. We take the ON-mRGC density to be half of the total mRGC density,
239 ignoring the possible density differences between ON- and OFF-center mRGCs.
240 The meridian functions are depicted in Fig. 2A.
- 241 • **Stage 1B:** We generate a continuous, two-dimensional map representing the mRGC
242 RF density map, depicted in Fig. 2B. This map is created by linearly interpolating
243 the meridian estimates, and it serves as a target for the algorithm in the next stage.
- 244 • **Stage 1C:** We synthesize a sampling lattice that represents the (x, y) positions of
245 the mRGC RF centers. The lattice is created using the iterative algorithm that we
246 introduced earlier [2] for generating cone mosaics, replacing the two-dimensional
247 cone density map with the target mRGC RF density map. A typical lattice of mRGC
248 RF positions is obtained after about 1,300 iterations and has a density that varies
249 smoothly over space, matching the target density, as illustrated in Fig. 2C. Example
250 patches of mRGC RF center mosaics synthesized at eccentricities of 0° and 20°
251 along the temporal horizontal meridian, are depicted in Figs. 2D & 2E, respectively.
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253 The same procedure is used to generate the lattice that represents the (x, y) posi-
254 tions of cones, but, in this case, using the meridian densities of cone photoreceptors in
255 human retina [36] as targets. The density of cones in the synthesized cone lattice also
256 varies smoothly over space and matches closely the target cone density, as illustrated
257 in Fig. 2F.

258 **2.2 Connecting cones to mRGC RF centers (Stage 2)**

259 The connections between cones and mRGC centers are constrained by (1) anatomical
260 data across the retina, specifically, the ratio of densities of mRGC RF centers to cones
261 [34], and (2) *in-vitro* physiological data from peripheral retina, that (a) indicate that,
262 unlike OFF-center mRGCs, which draw indiscriminately from all three cone types
263 [15, 37, 38], ON-center mRGCs draw only from L- and M-cones, and (b) quantify the
264 degree of RF center overlap in neighboring mRGCs [39]. In the present version of the
265 model, we only model ON-center mRGCs. The cone to mRGC RF center connectivity
266 is established in 3 sub-stages, summarized here.

- 267 • **Stage 2A:** In the first substage, each L- and M-cone gets connected to a single
268 mRGC RF center; single RF centers can receive input from more than one cone.
269 At this substage, each connected cone has unit connection weight. S-cones are not
270 connected because they do not contribute to ON-center mRGCs. This initial cone-
271 to-RF center connectivity often results in inhomogeneities in the composition of
272 neighboring mRGCs RF centers, which are dealt with in the next stage. Algorithmic
273 details regarding this substage are provided in Supplemental Section A.1.

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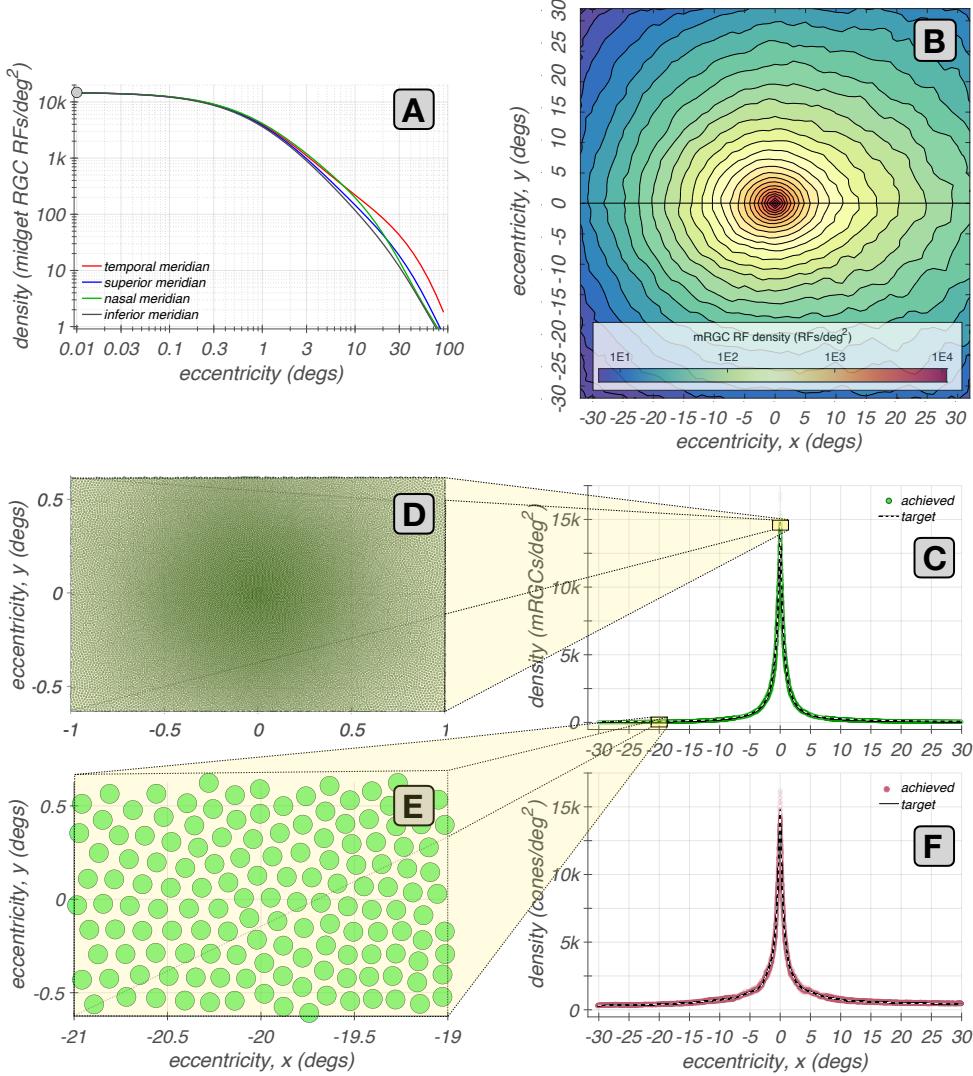


Fig. 2 Eccentricity-varying mRGC RF position lattices. **A:** Meridian density functions of mRGC RFs [34]. **B:** Two-dimensional mRGC RF density map obtained by interpolating the four meridian density functions. **C:** Achieved and target densities of mRGC RF centers along the horizontal meridian (green disks and white dashed line, respectively). **D & E:** Examples of $2^\circ \times 1^\circ$ mosaics of mRGC RF centers at eccentricities of 0° and 20° , respectively, along the temporal meridian. **F:** Achieved and target densities of cones along the horizontal meridian (maroon disks and white dashed line, respectively).

- **Stage 2B:** This substage refines the center connections to establish a balance between the spectral purity and spatial compactness of the mRGC RF centers, which is quantified by a single parameter, ϕ . For the body of this work, all mRGC

mosaics are generated by maximizing spatial compactness, but the option to maximize spectral purity allows testing of different scenarios where mRGC RF centers may be biased to some extent towards cone type selective pooling [15, 16]. At this substage, connected cones retain their unit connection weights. Algorithmic details regarding this substage are provided in Supplemental Section A.2.

- **Stage 2C:** Finally, the mutual exclusivity constraint enforced in substages 2A and 2B is lifted, and single cones are permitted to connect to multiple nearby mRGC RF centers. The extent of divergence varies with retinal eccentricity, being minimal in the fovea and increasing towards the periphery to match experimental observations [39]. This is done by varying the exponent of a supra-Gaussian distribution that describes the spatial weighting profile of cone connections to the RF centers which at this substage become non-binary. Algorithmic details regarding this substage are provided in Supplemental Section A.3.

We illustrate Stage 2 by examining key properties of synthesized mRGC RF center mosaics at each of the three substages.

2.2.1 Mosaics with convergent-only cone connections (stage 2A)

Example mosaics of RF centers synthesized at four eccentricities along the temporal horizontal meridian at the end of this substage are depicted in Fig. 3, where each green ellipse represents the spatial extent of the RF center of a single mRGC. At this stage, the pooling weight of each cone is set to unit.

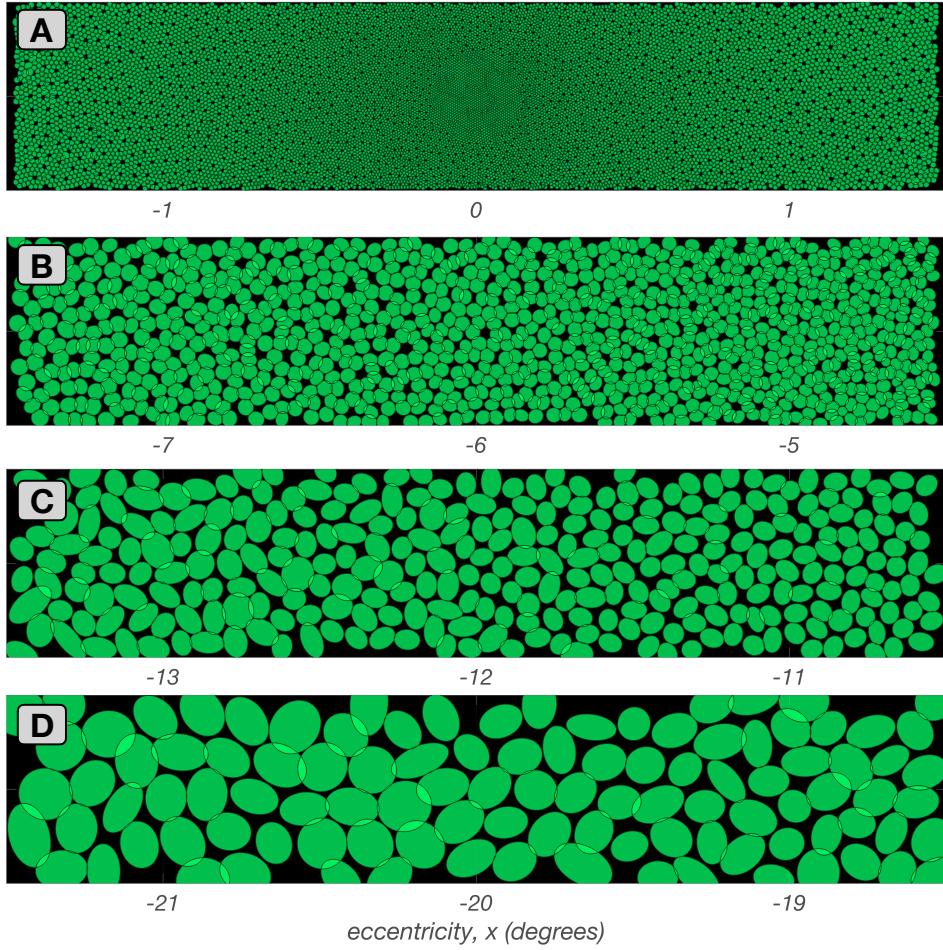
For the foveal mosaic depicted in Fig. 3A, RF centers connect to just a single cone. Note how RF center sizes increase as we move towards parafoveal regions to the left and right sides of Fig. 3A. This is due to the continuously increasing, with eccentricity, cone aperture in the input cone mosaic. The empty regions in this foveal mRGC RF center mosaic correspond to the location of S-cones which are not pooled by the model.

In the parafoveal mosaic depicted in Fig. 3B, RF centers mostly receive inputs from two cones, whereas in the more peripheral mosaics depicted in Figs 3C & 3D, RF centers connect to multiple cones. Note that the number of cones connecting to RF centers does not correspond precisely to RF center size, because cone aperture and inter-cone spacing both increase with eccentricity. At all eccentricities, however, mRGC RF center mosaics tile the retinal space with no spatial overlap or voids, except at the sparse positions where S-cones are located.

2.2.2 Mosaics synthesized under different spatial compactness/spectral purity tradeoffs (stage 2B)

This substage allows for different optimizations of cone pooling within the mRGC RF centers, which is controlled by the spatial compactness/spectral purity tradeoff parameter, ϕ . At this stage, the pooling weight of each cone is still set to unit, independent of the value of ϕ .

Fig. 4 depicts examples of mRGC RF center mosaics all synthesized at a single eccentricity (12° along the temporal meridian), but under different values of ϕ . The mosaic synthesized under $\phi = 1$, where spatial compactness is maximal and spectral purity constraint is not enforced, is depicted in Fig. 4A. Note that the RF centers



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Fig. 3 Stage 2A mRGC RF mosaics. Each panel shows a $3.0^\circ \times 0.5^\circ$ mosaic of synthesized mRGC RF centers at a different visual field location from fovea to periphery. The green ellipses depict a spatial region that encompasses all cones pooled by single RF centers. **A:** Foveal mosaic, in which RF centers receive signals from a single L- or M-cone. **B:** Mosaic centered at 6.0° along the temporal horizontal meridian, in which RF centers receive signals from 2–3 L/M-cones. **C:** Mosaic centered at 12.0° along the temporal horizontal meridian, in which RF centers receive signals from 3–4 L/M-cones. **D:** Mosaic centered at 20.0° along the temporal horizontal meridian, in which RF centers receive signals from 6–9 cones.

tile the visual field relatively uniformly with no overlap. Figures 4B and 4C depict mosaics synthesized as ϕ decreases to 0.5 and 0.0, respectively, which increasingly enforces center connections to cones of the same type. Note that this occurs at the cost of reduced spatial compactness, as is evident by the increased spatial disorder and overlap in the RF centers.

By varying ϕ we can examine the effect that cone-selective pooling may have on mRGC RF spatial structure, as well as on the spatio-chromatic processing in the

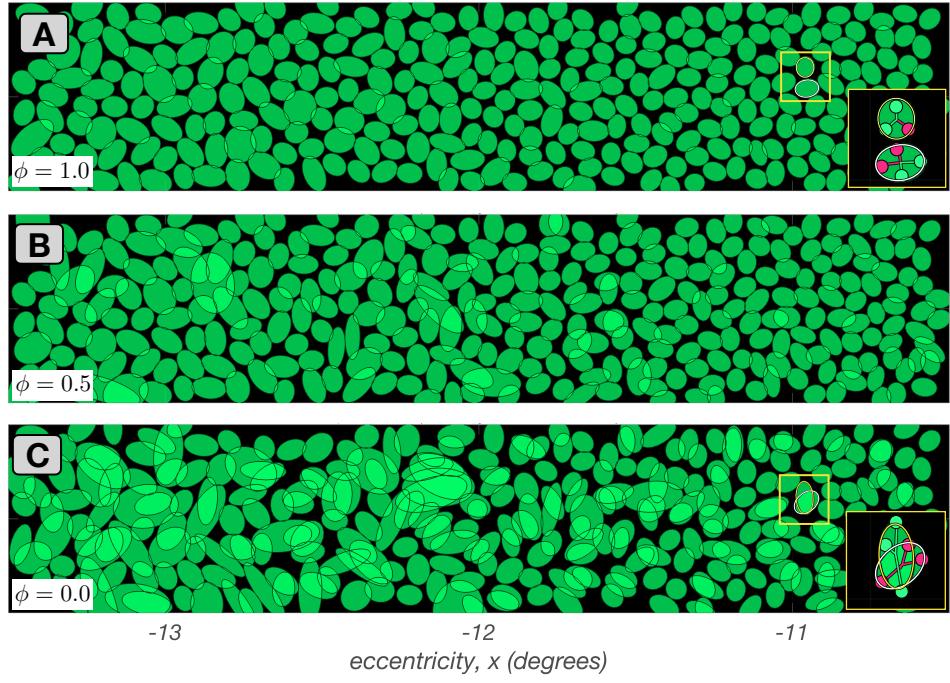
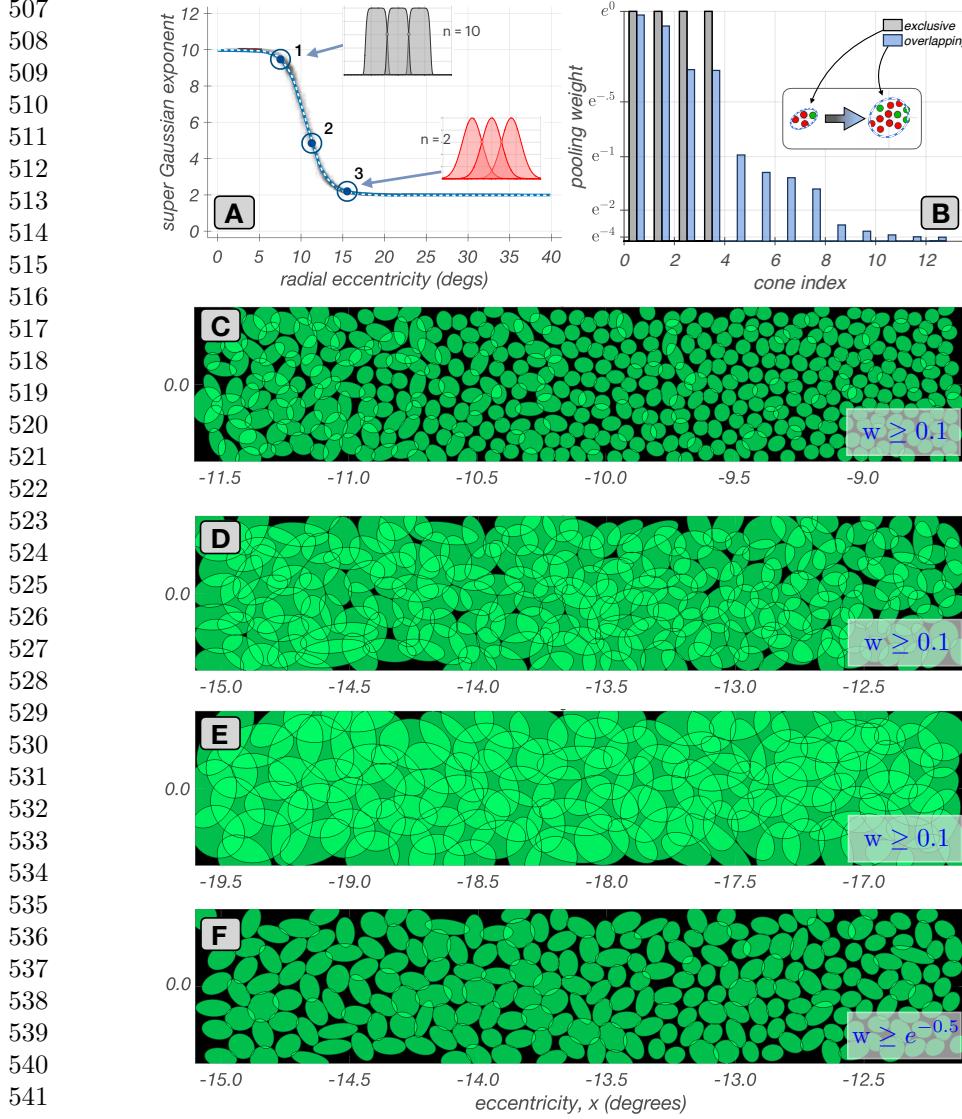


Fig. 4 Mosaics of mRGC RF centers at the end of stage 2B. Depicted here are $3.0^\circ \times 0.5^\circ$ mRGC mosaics, each centered at 12° along the temporal horizontal meridian, but synthesized under different values of tradeoff between spatial compactness and spectral purity, ϕ . **A:** $\phi = 1.0$ (maximal spatial compactness). **B:** $\phi = 0.5$. **C:** $\phi = 0$ (maximal spectral purity). Insets in A and C depict pooling of cones within the RF centers of the two mRGC RF centers contained within the yellow square. The inset in C illustrates how RF center overlap and spatial disorder is introduced as the algorithm avoids cones of different types that are close to the RF center in order to maximize the spectral purity of RF centers.

mRGC pathway. Current electrophysiological evidence favors little selective cone pooling, i.e., a ϕ value of ≈ 1 , in RF centers of peripheral mRGCs [15, 16, 40]. However, the degree of cone type selectivity in more central locations is not known with as much certainty. For example, there is anatomical evidence that ON-center mRGCs in the fovea contact multiple ON-cone bipolars, as opposed to OFF-center mRGCs, which contact single OFF-cone bipolars [41], and also electrophysiological evidence that the RF centers of parafoveal mRGCs appear to be pooling from more than one cones [42]. In general, the question of whether foveal mRGCs that pool from more than one cone in the RF centers are doing so selectively remains unanswered. Our modeling approach allows exploration of the benefits and tradeoffs of cone-selective pooling at any retinal eccentricity, although we do not pursue such exploration in this paper.

2.2.3 Mosaics with divergent cone connections (stage 2C)	461
In the final substage of establishing the wiring of mRGC RF centers to the cone mosaic, the mutual exclusivity constraint is lifted and single cones are permitted to connect to multiple nearby mRGC RF centers. This divergence of cone connections is enabled by replacing the binary distribution of cone pooling weights in the mRGC RF centers with a supra Gaussian distribution, as illustrated in Fig. 5.	462
Fig. 5A depicts how a progressively increasing overlap in neighboring mRGC RF centers with eccentricity is accomplished by varying the exponent of the supra-Gaussian distribution. In central retina, the exponent is kept at 10, which results in a flat top distribution of weights with minimal overlap between neighboring RF centers (gray histograms in the inset of Fig. 5A). As eccentricity increases beyond 7°, the exponent decreases, reaching a value of 2 at around 15°, which results in Gaussian distributions of weights and a significant overlap between neighboring RF centers (red histograms in the inset of Fig. 5A).	463
To our knowledge, there is no physiological data on the variation with eccentricity of the divergence of cone connections to nearby mRGC RF centers. Therefore the varying, with eccentricity, supra-Gaussian distribution of cone weights is an arbitrary mechanism. It's intent is to capture the fact that in the fovea, input to mRGC RF centers comes exclusively or mostly [41, 42] from a single cone, whereas in the periphery, <i>in vitro</i> measurements reveal that neighboring mRGC RF centers abut at approximately one standard deviation of their Gaussian RF profile [39].	464
The transformation of cone pooling weights from binary and mutually exclusive to graduated and shared is depicted in Fig. 5B for an mRGC located at an eccentricity of 12°, with gray and blue histograms depicting the spatial distributions of cone pooling weights before and after substage 2C.	465
Figs. 5C–5E depict mosaics with divergent connections synthesized at three eccentricities. In these mosaic depictions, each green ellipse represents the spatial extent that encompasses all cones that are pooled by the RF center of a single mRGC with weights ≥ 0.1 . For the mosaic centered at 10° (Fig. 5C), divergence of cone connections has just begun. The overlap in RF centers due to the divergence of connections increases as we move in eccentricity from 9° on the right side to 11°, on the left side. For the mosaic centered at around 13° (Fig. 5D), cone divergence and RF center overlap is higher and again increases with increasing eccentricity. For the mosaic centered at around 18° (Fig. 5E), divergence of cone connections has assymptoted, and we have a constant RF center overlap.	466
Finally, Fig. 5F provides a visualization comparable to the visualization commonly reported by <i>in vitro</i> RF mapping studies [39]. It depicts the same mosaic as Fig. 5D, but with ellipses encompassing cones that are pooled with weights $\geq e^{-1/2} \approx 0.67$. This depiction choice makes the overlap less visually salient.	467
2.3 Connecting cones to mRGC RF surrounds (Stage 3)	501
Overview	502
In the last stage of mRGC mosaic synthesis, we derive the cone pooling weights for the mRGC RF surrounds. Since there are no clear anatomical data on surround	503
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543 **Fig. 5 Mosaics of mRGC RF centers with divergent cone connections (stage 2C).**
 544 **A:** Variation of supra-Gaussian exponent with eccentricity. The exponent is set to 10 in the central
 545 retina, resulting in flat top weight distributions with zero overlap (gray histograms). As eccen-
 546 tricity is increased, the exponent is gradually decreased, achieving a value of 2.0, at around 15° (red
 547 histograms). **B:** Transformation of cone pooling weights, from binary, in mutually exclusive con-
 548 nections, (gray histogram) to non-binary in shared cone connections, (blue histogram) due to the
 549 supra-Gaussian distribution for an example mRGC. Insets depict the spatial arrangement of cones
 550 that are connected with binary and non-binary weights. **C, D & E:** Mosaics at 10°, 13°, and
 551 18°, along the temporal horizontal meridian with divergent cone connections. The RF center ellipses
 552 encompass the ensemble of cones with pooling weights ≥ 0.1 . **F:** Same mosaic as **C**, but with ellipses
 553 showing cones with pooling weights $\geq e^{-0.5}$.

sizes, these weights are determined using <i>in vivo</i> characterizations of macaque mRGC visual space-referred spatial transfer functions, $vSTF(\omega)$, i.e., the variation in response amplitude of mRGC cells as a function of spatial frequency, ω . We use the measurements of Croner & Kaplan [17], who characterized $vSTF(\omega)$ for populations of mRGCs across a wide range of eccentricities.	553
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We incorporate these data into the model using numerical optimization. More specifically, we determine the cone-to-mRGC RF surround connections such that a forward simulation of the <i>in vivo</i> physiological experiments of Croner & Kaplan through the model best reproduces the experimental data. This approach allows us to use data collected through physiological optics, which blur the stimulus in an eccentricity and wavelength dependent manner, to determine the wiring of cones to mRGC RF surrounds.	558
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Importantly, the optimization is achieved while adhering to the already established connectivity between the cone mosaic and mRGC RF centers. Simultaneously, the parametric form of the surrounds is constrained based on Packer & Dacey's characterizations of the spatial RF of macaque H1 horizontal cells [43], which are the main components of the linear spatial mRGC RF surrounds [44]. The use of optimization around forward simulation of an experiment to integrate data from multiple non-commensurate sources is an important innovation of our RGC modeling approach. Stage 3 proceeds in three sub-stages.	565
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• Stage 3A: We begin by computing the visual space-referred cone mosaic responses to stimuli used to measure vSTFs in macaque mRGCs. This is done by presenting achromatic gratings of different spatial frequencies which are delivered to the retina through human physiological optics [45]. We use human optics as a proxy of how macaque optics would have blurred the stimuli employed by the <i>in vivo</i> characterizations of Croner & Kaplan [17], which were collected with stimuli viewed through the animal's natural optics.	573
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• Stage 3B: We derive surround cone pooling functions for a subset of target synthetic mRGCs, which span the extent of the synthesized mRGC mosaic. This optimization is done so that the ensuing target cells (a) have vSTF characteristics that are well approximated by a Difference of Gaussians (DoG) model, with DoG model parameters reasonably matching the DoG model parameters reported by Croner & Kaplan at corresponding eccentricities, and (b) have surround cone pooling weights that maintain macaque H1-like spatial properties as characterized by Packer & Dacey.	581
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• Stage 3C: We compute surround cone pooling weights for all cells in the synthesized mRGC mosaic by evaluating the derived surround cone pooling functions at the vicinity of each mRGC's input cone mosaic and finally interpolating the computed weights. A small amount of jitter in the ratio of the surround to center weights is added to simulate the variance in integrated surround to center ratio at each eccentricity seen in the macaque data.	589
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599 **2.3.1 Computation of visual space-referred cone mosaic responses**
600 **to stimuli used to measure vSTFs in macaque mRGCs**
601 **(Stage 3A)**

602 We employ the ISETBio machinery to compute the excitation of the input cone mosaic
603 to achromatic gratings of different spatial frequencies delivered to the retina via phys-
604 iological optics. This process captures several crucial spatio-chromatic effects in the
605 transformation of scene radiance into cone responses: spatial and chromatic filtering
606 by physiological optics, spectral filtering by the eye's inert pigments, and sampling by
607 the interdigitated trichromatic cone mosaic. To mimic the phototransduction process,
608 cone excitation responses are converted to cone modulation responses.

609 In these computations, we employ human physiological optics matched to the
610 eccentricity of each synthesized mRGC, but we adjust the defocus term of the mod-
611 eled optics so as to maximize the Strehl ratio. The Strehl ratio is defined as the ratio
612 of peak sensitivity of the point spread function (PSF) at the wavelength of focus, here
613 550 nm, to the peak sensitivity of a diffraction-limited PSF. This is done as a proxy to
614 the experimental paradigm of Croner & Kaplan, in which corrective lenses were used
615 to maximize cell responses at high spatial frequencies (personal communication with
616 the late Ehud Kaplan).

617 **2.3.2 Deriving surround cone pooling functions for a subset of**
618 **target synthetic mRGCs (Stage 3B)**

619 Croner & Kaplan reported summaries of spatial RF characteristics across populations
620 of mRGCs at different eccentricities which were derived by fitting a DoG model to the
621 measured vSTF. The DoG model defined in spatial frequency, ω , domain is given by:

622
$$\text{DoG}(\omega) = K_c \cdot R_c^2 \cdot \exp[-\pi \cdot R_c \cdot \omega]^2 - K_s \cdot R_s^2 \cdot \exp[-\pi \cdot R_s \cdot \omega]^2 \quad (1)$$

623 where K_c and K_s are the peak sensitivities of the RF center and RF surround mecha-
624 nisms, and R_c and R_s are the corresponding characteristic radii. The vSTF of a typical
625 macaque mRGC is depicted in Fig. 6A with cyan disks. The solid heavy line depicts
626 the fitted DoG model, with the center and surround components depicted by the thin
627 solid and dashed lines, respectively.

628 The shape of the vSTF is determined by two key measures, the ratio of surround
629 to center characteristic radii, R_s/R_c , and the ratio of surround to center integrated
630 sensitivities, $K_s/K_c \times (R_s/R_c)^2$. The distributions of these two ratios as a function of
631 eccentricity in the population of mRGCs recorded by Croner & Kaplan are depicted
632 by the gray squares in Figs 6B1 & 6B2. The mean variation in these two ratios, shown
633 as dashed lines, are the target values used to derive the surround cone pooling weights
634 in the synthetic mRGCs.

635 The optimization process is illustrated schematically in Fig. 6C. The vSTF of
636 the target synthetic mRGC is computed by forward simulation of the experiment of
637 Croner & Kaplan. The time course of responses of L- and M-cones in the input cone
638 mosaic to a drifting grating stimulus of spatial frequency ω , $R_{\text{cones}}(\omega, t)$, (computed
639 in Stage 3A) are depicted by the red and green traces in the rectangular panel of

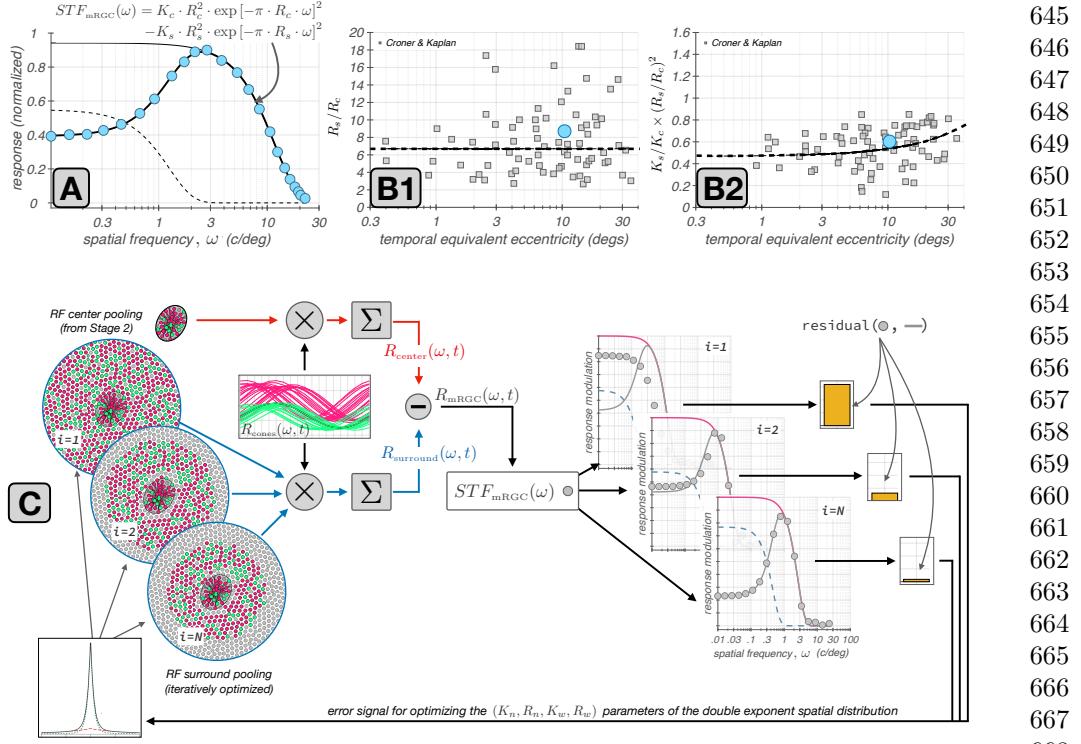


Fig. 6 Derivation of cone weights to mRGC surrounds. **A:** Typical macaque mRGC vSTF (cyan disks) fitted with a Difference of Gaussians model (thick black line), whose center and surround components are depicted by the thin black and the dashed line, respectively. **B1 & B2:** Ratios of surround to center characteristic radii, R_s/R_c , and ratios of surround to center integrated sensitivities, $K_s/K_c \times R_s^2/R_c^2$ as a function of eccentricity in the population of mRGCs recorded by Croner & Kaplan [17]. The dashed lines represent the trends in these two ratios as a function of eccentricity. The cyan disks depict the ratios for the example vSTF depicted in A. **C:** Depiction of the iterative estimation of surround cone pooling weights by forward simulation of the Croner & Kaplan vSTF measurements. See description in text for more details.

Fig. 6C. A spatially weighted sum of these cone responses using the RF center cone pooling weights (computed in Stage 2), is used to compute the response of the RF center, $R_{\text{center}}(\omega, t)$. This operation, which is depicted by the red computation arm in Fig. 6C, is fixed throughout the optimization of the surround.

In the computation of the spatial distribution of surround cone pooling weights, we impose a parametric form that is described by the sum of a narrow and a wide exponential spatial component, based on characterizations of the spatial RF properties of H1 horizontal cells by Packer & Dacey [43]. Specifically,

$$W_s(r) = K_{\text{wide}} \times \exp[-r/R_{\text{wide}}] + K_{\text{narrow}} \times \exp[-r/R_{\text{narrow}}] \quad (2)$$

where r is the radial distance from the RF center, K_{wide} and K_{narrow} are the peak sensitivities of the wide and the narrow components, respectively, and R_{wide} and R_{narrow} are the corresponding characteristic radii.

691 Beginning with a random initial value for the parameters of the double exponential
692 distribution, we compute an initial estimate of the surround cone weights by evalu-
693 ating $W_s(r)$ at the vicinity of the input cone mosaic that surrounds the RF center.
694 These weights are depicted in the top-left circular panel of Fig. 6C (labeled as $i = 1$,
695 with i denoting iteration). Using these initial weights we compute a weighted sum of
696 the surround cone responses to derive the initial estimate of the surround response,
697 $R_{\text{surround}}(\omega, t)$. This operation is depicted by the blue computation arm in Fig. 6C.

698 The composite response of the synthesized mRGC is obtained by instantaneously
699 subtracting the surround response from the center response, as follows:

700

$$R_{\text{mRGC}}(\omega, t) = R_{\text{center}}(\omega, t) - R_{\text{surround}}(\omega, t) \quad (3)$$

702

703 The amplitude modulation of $R_{\text{mRGC}}(\omega, t)$ is taken as the value of the cell's
704 vSTF_{mRGC}(ω). Repeating over a range of spatial frequencies, we obtain the initial
705 estimate of the full vSTF_{mRGC}, which is depicted by the gray disks in the top-right
706 rectangular panel of Fig. 6C, labeled as $i = 1$.

707 Following the experimental procedure of Croner & Kaplan, we fit the computed
708 vSTF_{mRGC}(ω) with a DoG model. This fit is depicted by the solid gray line in the top-
709 right rectangular panel of Fig. 6C. Note that in this procedure we constrain the DoG
710 model fit so that its shape parameters, R_s/R_c , and $K_s/K_c \times R_s^2/R_c^2$, both lie within
711 a narrow range of the mean values of R_s/R_c , and $K_s/K_c \times R_s^2/R_c^2$ ratios reported
712 for macaque mRGCs at corresponding eccentricities [17]. Due to this constrain, in the
713 first iteration the residual between the computed vSTF and the DoG model fit to it,
714 is large.

715 This residual $\|v\text{STF}_{\text{mRGC}} - \text{DoG}\|$, depicted by the yellow bar in the right-most
716 panel of Fig. 6C, serves as an error signal. The optimization algorithm minimizes
717 this error signal by adjusting the parameters of $W_s(r)$, which controls the surround
718 weights. This adjustment is also constrained, so that the parameters of $W_s(r)$ remain
719 within a range of the values reported in macaque H1 horizontal cells [43].

720 When the $\|v\text{STF}_{\text{mRGC}} - \text{DoG}\|$ reaches a minimum value, labeled as $i = N$,
721 in Fig. 6C, we obtain the optimized surround cone pooling function for the target
722 synthetic mRGC. Additional details about this surround optimization method are
723 provided in Supplemental Section B.1.

724

725 2.4 Deriving surround cone pooling weights for each cell in 726 the mosaic(stage 3C)

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728 The optimization of the surround cone pooling functions is a computationally expensive
729 process. It is therefore conducted on a sparse spatial grid (with N_{xy} nodes), which
730 encompasses the spatial extent of the synthesized mRGC mosaic. At each node of the
731 spatial grid, we determine the range of cone numerosities in the RF centers of nearby
732 synthetic mRGCs, and we derive optimized surround cone pooling functions for each
733 of the encountered RF center cone numerosities (N_c), and we do this twice, once for
734 L-cone dominated RF centers, and once for M-cone dominated RF centers.

735 Once these $N_{xy} \times N_c \times 2$ surround cone pooling functions are derived, we compute
736 surround cone pooling weights for all synthetic mRGCs. For each target mRGC we

determine the 3 nearest spatial grid nodes, and extract the optimized surround cone	737
functions that were derived at this node for the cone numerosity that matches that	738
of the target mRGC, for both L- and M-center cone dominance variants. Then we	739
evaluate the six optimized surround pooling functions at the input cone mosaic in the	740
vicinity of the target mRGC, deriving six sets of surround cone pooling weights. The	741
target cell's surround cone pooling weights are determined by interpolating the 6 sets	742
of weights spatially, weighted inversely proportionally by the distance between the	743
location of the target mRGC and the location of the optimized model, and spectrally,	744
weighted based on the relative L-/M- cone weight ratio in the RF center of the target	745
mRGC.	746
	747
2.4.1 Adjusting the surround pooling variance	748
The final step in the generation of the mRGC RF surrounds is to apply a noisy scalar	749
multiplier to all surround pooling weights of individual mRGCS. The value of this	750
scalar is chosen so that the variance in the ratio of surround to center integrated sensi-	751
tivities, $K_s/K_c \times (R_s/R_c)^2$, of the synthetic mRGCS matches the variance observed in	752
the population of macaque mRGCS recorded by Croner & Kaplan. The manipulation	753
in $K_s/K_c \times (R_s/R_c)^2$ variance does not require re-computing the surround pooling	754
functions. This is unlike manipulating the variance in the R_s/R_c ratio, which requires	755
re-computing the surround pooling functions.	756
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2.5 Computing mRGC responses from cone mosaic responses	759
A fully synthesized mRGC mosaic consists of two connectivity matrices: $P_{center}(i, k)$,	760
determined in synthesis stage 2, and $P_{surround}(i, k)$, determined in synthesis stage	761
3, which capture the weights by which the RF center and surround mechanisms,	762
respectively, of the k^{th} - cell in the mRGC mosaic pools signals from the i^{th} cone in	763
the input cone mosaic.	764
Since the current version of the mRGC model does not contain a temporal compo-	765
nent, the response of the k^{th} mRGC to some stimulus at time instant, t , $R_{stim}(k, t)$,	766
is computed instantaneously from the response of the input cones to that stimulus at	767
time t , $C_{stim}(i, t)$, as follows:	768
	769
$R_{stim}(k, t) = \frac{1}{\sum_{i=1}^n P_{center}(i, k)} \times \dots$	770
$\left(\sum_{i=1}^n P_{center}(i, k) \cdot C_{stim}(i, t) - \sum_{j=1}^m P_{surround}(j, k) \cdot C_{stim}(j, t) \right)$	771
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To mimic adaptation to the background stimulus, the mRGC mosaic typically oper-	778
ates on cone contrast responses, instead of cone excitation responses, so the $C_{stim}(i, t)$	779
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783 term in the above equation is computed as follows:

784

785
$$C_{\text{stim}}(i, t) = \frac{E_{\text{stim}}(i, t) - E_{\text{bkgnd}}(i)}{E_{\text{bkgnd}}(i)}$$
 (5)

786

787 where $E_{\text{stim}}(i, t)$ is the excitation response of the i^{th} cone to the examined stimulus at
788 time t , and $E_{\text{bkgnd}}(i)$ is that cone's excitation response to a uniform field, zero con-
789 trast stimulus, whose mean chromaticity and luminance match those of the examined
790 stimulus.

791

792 3 Results

793

794 A key feature of our model is its dual representation of mRGC receptive field (RF)
795 properties, which separates neural circuitry from optical effects. The first representa-
796 tion, in *retinal space*, models the direct pooling of cone signals by the RF center and
797 surround. This describes the cell's intrinsic spatio-chromatic filtering and is directly
798 comparable to anatomical data and physiological measurements that bypass the eye's
799 optics (e.g., *in vitro* or adaptive optics experiments [10, 46]). In contrast, the second
800 representation, in *visual space*, models the end-to-end processing of a stimulus as it
801 passes through the eye's optics to the mRGC mosaic. This representation is applicable
802 to conventional *in vivo* physiology and psychophysical assessments of visual function.

803 The ability to go back and forth between cone and visual space is critical to under-
804 standing how retinal cone pooling interacts with physiological optics to generate the
805 processing characteristics of cells in visual space, which is what ultimately determines
806 natural visual performance. This ability is also critical in interpreting results from *in*
807 *vivo* physiology in terms of the underlying retinal wiring [31], as well as to relating
808 results obtained under adaptive optics viewing conditions to results obtained under
809 natural viewing conditions [10].

810 In this section we illustrate and contrast spatial RF characteristics of synthetic
811 mRGCs in the two representations and validate the properties of synthetic mRGCs
812 against those of macaque mRGCs as characterized by *in vivo* and *in vitro* physiological
813 studies.

814

815 3.1 Spatial characteristics of synthesized mRGC receptive 816 fields

817

818 Spatial characteristics of cells in an mRGC mosaic synthesized at 4.5° along the tem-
819 poral horizontal meridian are depicted in Fig. 7. The employed mosaic is depicted in
820 Fig. 7A. The numbered positions in Fig. 7A identify the locations of three selected
821 cells whose spatial RF characteristics are explored in detail next.

822 The cone pooling maps of these exemplar mRGCs are depicted in Figs 7B1–B3.
823 Here, pink and cyan disks depict L- and M-cones, respectively, that are pooled by
824 the RF center with a weight ≥ 0.1 , or by the RF surround with a pooling weight \geq
825 0.005, and gray disks depict cones that are either not pooled at all or pooled with
826 a weight less than the threshold for labeling. The solid and dashed lines depict the
827 spatial pooling extents of the RF center and surround mechanisms, respectively.

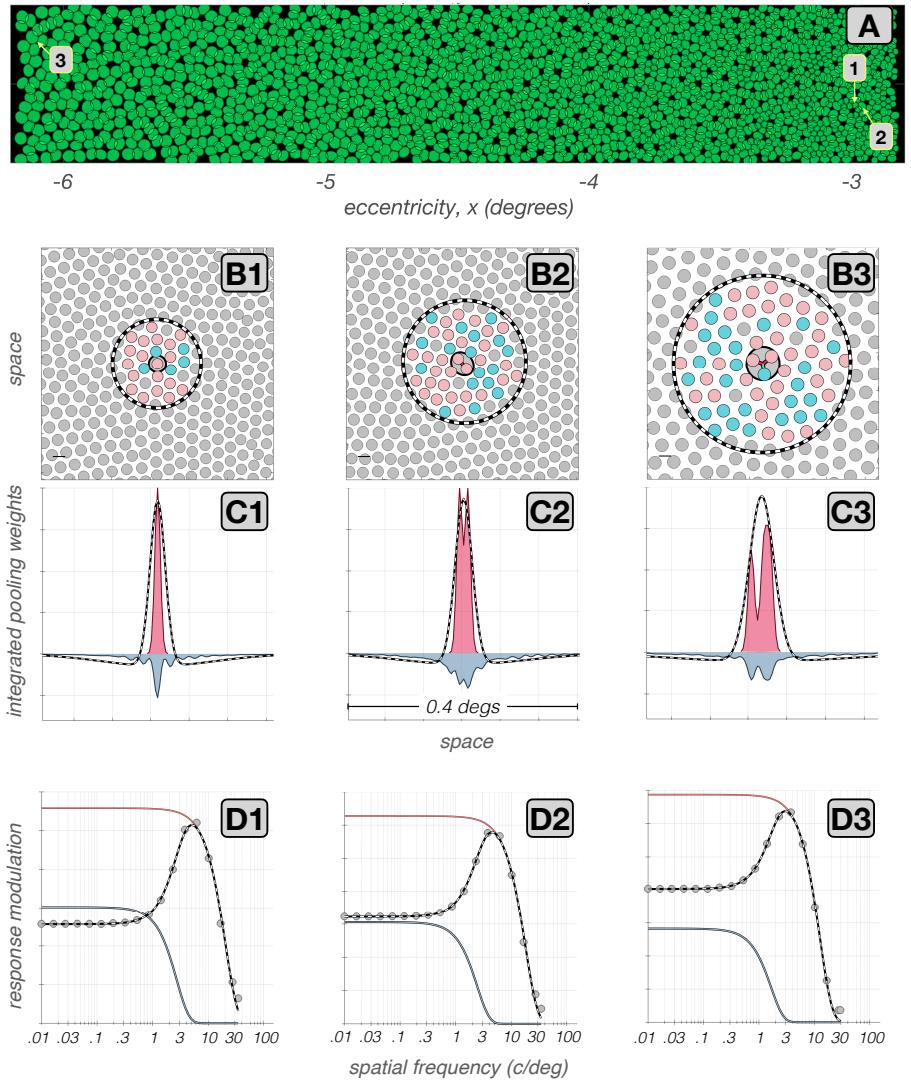


Fig. 7 Spatial RF characteristics of synthetic mRGCs **A:** Mosaic of RF centers of an mRGC mosaic synthesized at 4.5° along the temporal horizontal meridian. **B1–B3:** Cone pooling maps for 3 exemplar cells whose positions within the mRGC mosaic are labeled in A. Pink and cyan disks depict L- and M-cones, respectively, with RF center pooling weights ≥ 0.1 , or with RF surround weights ≥ 0.005 . Gray disks represent either S-cones, which are not pooled in our model, or L-/M-cones with pooling weights lower than the labeling thresholds. The solid and dashed black lines depict the extents of the RF center and surround pooling regions. **C1–C3:** Y-axis integrated cone pooling weight profiles within the RF center (maroon) and surround (slate). The dashed lines depict the visual space-referred line weighting functions as derived by fitting Difference of Gaussians (DoG) models to each cell's vSTF. **D1–D3:** The vSTFs of the exemplar mRGCs, computed under physiological optics, are depicted by the gray disks. The gray lines depict the DoG model fits to these vSTFs, and the maroon and slate lines depict the models' center and surround components, respectively.

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875 The cell depicted in Fig. 7B1 is located at an eccentricity of 3° . Its RF center
876 pools from a single L-cone and its RF surround pools from a total of 16 L- and M-
877 cones. The cell depicted in Fig. 7B2, also located at 3° , pools from two L-cones in its
878 RF center, and its RF surround is larger, pooling from 44 L- and M-cones. The cell
879 depicted in Fig. 7B3 is located at 6° . Its RF center, which pools from 2 L-cones and
880 1 M-cone, and its surround are both larger than those of the first 2 cells. The cone
881 pooling maps depicted in Figs 7B1–B3 illustrate the spatial connectivity between the
882 input cone mosaic and the center and surround subregions of mRGC RFs, but do not
883 depict the strength of these connections. In this sense, these maps depict the type of
884 information that is available from detailed anatomical studies.

885 Figs 7C1–C3 add to this view by providing information about the strength of the
886 cone inputs for the three exemplar cells. Here, the maroon and slate histograms depict
887 the cells' spatially integrated (along the y-axis) cone pooling weights for the RF center
888 and the surround mechanisms, respectively. Note that in the cell depicted in Fig. 7C1,
889 the double exponential spatial profile of the surround cone pooling mechanism, with
890 a sharp peak around the RF center and more shallow weights in peripheral regions, is
891 prominent. In the two other cells shown, this feature is less prominent.

892 This observation, where cells with larger RF centers have less peaked surround
893 weights than cells with smaller RF centers is seen commonly in our synthetic mRGCs.
894 The variation in surround pooling characteristics with RF center size results from
895 constraints in the model, which maintain vSTF shape parameters that are consistent *in*
896 *vivo* measurements [17] while at the same time remaining consistent with the surround
897 parametric form indicated by measurements of H1 receptive fields [43].

898 Visual space-referred spatial transfer functions are commonly measured in *in vivo*
899 physiological assessments to estimate spatial RF properties of mRGCs [17, 18]. The
900 vSTFs of the three examined synthetic mRGCs are depicted by the gray disks in
901 Figs 7D1–7D3. The corresponding DoG model fits are depicted by the solid gray lines,
902 and the spatial RF profiles corresponding to these DoG model fits are depicted by the
903 dashed lines in Figs 7C1–C3. Contrasting these inferred spatial RF profiles with the
904 actual cone pooling profiles, it becomes evident that one cannot use characterizations
905 obtained under physiological optics viewing to directly infer the characteristics of
906 spatial pooling of cone signals in the retina. We discuss this issue further in later
907 sections.

908

909 **3.2 Validation against *in vivo* physiology across the visual field**

910

911 To validate our model, we synthesized mRGC mosaics across a wide region of the
912 retina, and computed vSTFs of individual mRGCs by probing them with drifting
913 achromatic gratings of different spatial frequencies delivered to the retina under phys-
914 iological optics appropriate for the eccentricity of the examined cells, simulating the
915 experimental paradigm of Croner & Kaplan [17].

916

917 To compare synthetic and macaque mRGCs we fitted the synthetic cell vSTFs
918 with the DoG model employed by Croner & Kaplan and compared the ratios of sur-
919 round to center characteristic radii, R_s/R_c , and ratios of surround to center integrated
920 sensitivities, $K_s/K_c \times R_s^2/R_c^2$, to those reported by Croner & Kaplan.

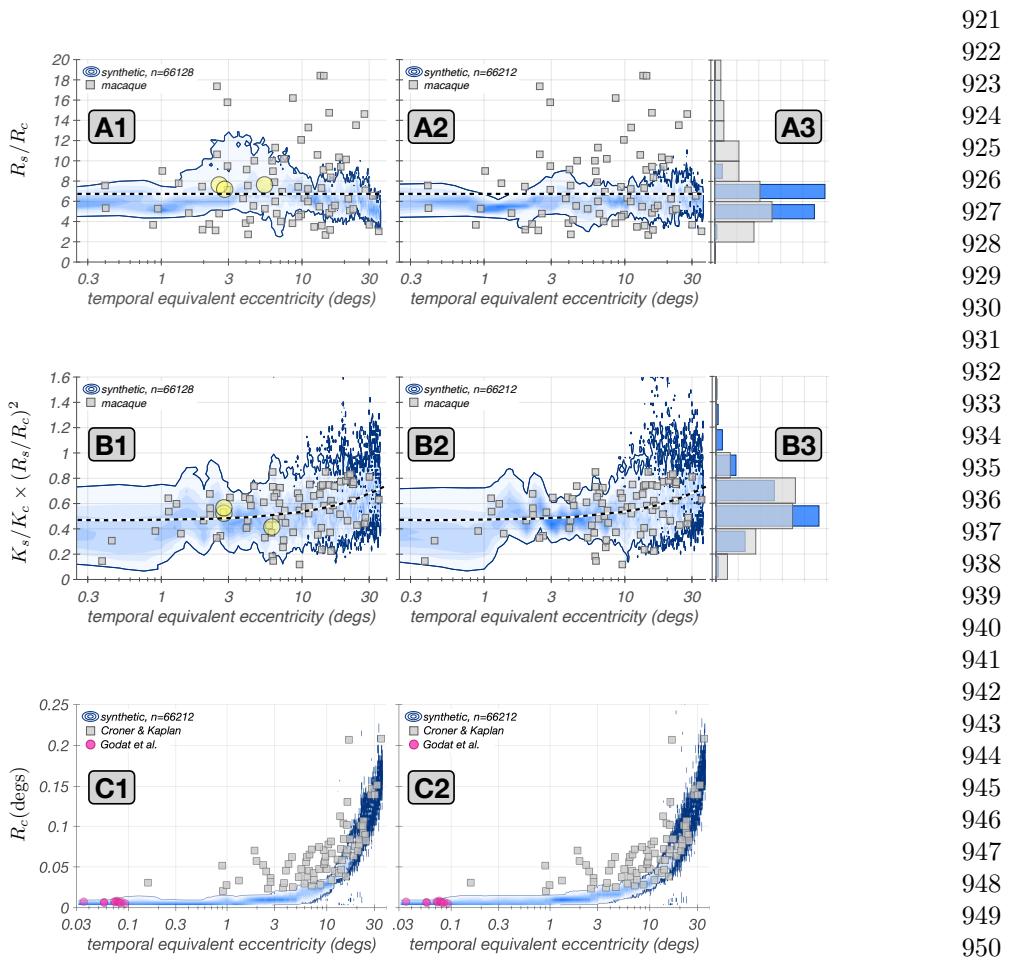


Fig. 8 Validation against *in vivo* measurements. In all panels, gray squares depict data from the population of macaque mRGCs recorded by Croner & Kaplan [17]. Blue contours depict the probability density function of the examined parameter in a population of 66,128 synthetic mRGCs with color saturation encoding probability level. Solid blue lines represent the 5% – 95% percentile range of examined parameter. Left and right panels are for mosaics synthesized under physiological optics of two different human subjects. **A1–A2:** Correspondence in ratio of surround-to-center characteristic radii, R_s/R_c , across eccentricity. The dashed line represents the target value that is in effect during the optimization of the synthetic mRGC surrounds, which is the mean value of R_s/R_c across the population of all mRGCs recorded by Croner & Kaplan. **A3:** Marginal histograms of R_s/R_c for macaque (gray) and synthetic mRGCs (blue). **B1–B2:** Correspondence in ratio of surround-to-center integrated sensitivities, $K_s/K_c \times (R_s/R_c)^2$, across eccentricity. The dashed line represents the target values in effect during the optimization of the synthetic mRGC surrounds, which is the trend observed with eccentricity in the population of the macaque mRGCs recorded by Croner & Kaplan. **B3:** Marginal histograms of $K_s/K_c \times (R_s/R_c)^2$ for macaque (gray) and synthetic mRGCs (blue). **C1–C2:** Correspondence in RF center characteristic radius, R_c , across eccentricity. The fuschia disks represent the R_c of foveolar mRGCs recorded by Godat *et al.* [10], back-projected in visual space using the monkey's own physiological optics.

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967 The results of this analysis are depicted in Fig. 8, in which the left and right
968 panels depict data from mRGC mosaics synthesized under the physiological optics
969 of two different human observers. Figs. 8A1 and 8A2 compare the distribution of
970 R_s/R_c ratios. Gray squares depict the macaque mRGC data and the blue density plots
971 depict the 5%–95% percentile range of the R_s/R_c ratios in a population of 66,128
972 synthetic mRGCs. The three yellow disks in Fig. 8A1 correspond to the three exemplar
973 cells illustrated in Fig. 7. Note that the R_s/R_c ratios in synthetic mRGCs follow the
974 macaque data across eccentricity for both human subjects. The synthetic data do
975 not, however, capture the full variance seen in the macaque data, as is evident by
976 the marginal histograms (Fig. 8A3). To capture the full variance seen in the macaque
977 data, we could consider synthesizing multiple surround pooling functions, each with
978 different target values of R_s/R_c , and then randomly selecting for each synthesized
979 mRGC from the multiple sets.

980 On the other hand, the integrated sensitivity ratios, $K_s/K_c \times R_s^2/R_c^2$, of the
981 synthetic mRGC population, depicted in Figs 8B1–B3, capture both the trend with
982 eccentricity and the variance of the macaque data. The variance match was achieved
983 by enforcing a target variance in the $K_s/K_c \times R_s^2/R_c^2$ ratio of the synthetic cells as
984 described earlier.

985 Note that, although we did use the mean variation with eccentricity of macaque
986 R_s/R_c and $K_s/K_c \times R_s^2/R_c^2$ ratios during construction of the model, the model was
987 derived using additional constraints: those imposed by the densities of cones and
988 mRGC RFs, by the spatial characteristics of H1 horizontal cells, and by the influence
989 of human optics. These validations, therefore, check both that we have not over con-
990 strained our model in a manner that makes it inconsistent with the macaque data,
991 and that our method of interpolating surround pooling weights from models derived
992 at a set of discrete retinal locations works well.

993 We next examined the correspondence between synthetic and macaque mRGCs in
994 terms of their visual space-referred RF center sizes, R_c . Recall that in synthesizing
995 mRGC mosaics, the RF centers are constructed independently of the Croner & Kaplan
996 physiological data, using only anatomical data and estimates of RF center overlap
997 obtained from *in vitro* physiology in the periphery [39]. Figs. 8C1–C2 compare the
998 distributions of R_c between the macaque and synthetic mRGCs. Note that R_c in the
999 synthetic mRGCs follows the trend seen in macaque mRGCs with eccentricity, with
1000 good agreement at eccentricities above 10° for both subjects. In more central locations,
1001 however, the synthetic mRGC RF center sizes are 2–3 times smaller than those in the
1002 macaque. We believe that the discrepancy at central locations is not a deficiency of
1003 our model, but rather results from several factors.

1004 First, the cone mosaic in our model has a peak density of 288,000 cones/mm²
1005 which is near the high end of densities reported in humans [36], whereas the average
1006 macaque peak cone density is around 200,000 cones/mm² [47, 48]. The higher cone
1007 mosaic density in humans implies smaller cone apertures, which in turn would bias
1008 our synthetic mRGCs towards somewhat smaller RF centers.

1009 Second, in acute macaque experiments, optical refraction is not necessarily perfect,
1010 so there could be residual blur due to errors in refraction, as well as due to corneal
1011 edema from the contact lens used in typical multi-day acute experiments. This would
1012

increase the size of the RF centers in the physiological data relative to those in our model in central retina. Moreover, residual eye movements can occur in acute experiments, despite the ocular muscle paralysis (personal observations by N.P. Cottaris). Such residual movements would artificially enlarge estimates of RF center size for central retina mRGCs.

Finally, in the macaque mRGC vSTF characterizations of Croner & Kaplan, stimulus orientation was not optimized to match any orientation bias in the RF of macaque mRGCs (Lisa Croner, personal communication), whereas in the simulated experiments, stimulus orientation was matched to the cell's visual-space referred orientation bias, which results in the smallest possible estimate of RF center size. Indeed, in additional analyses (not shown) in which we computed vSTFs under random grating orientations as well as a fixed orientation (as was done by Croner & Kaplan), we found enlarged estimates of R_c . However, these enlarged estimates still fall short of those reported by Croner & Kaplan, so the first two factors that we mentioned above must also be at play.

Support for our assertion is provided by *in vivo* data from foveal macaque mRGC vSTFs obtained under adaptive optics viewing conditions [10]. The center sizes of these cells, blurred by the optics measured for the monkey subjects studied, are depicted by the purple disks in Figs. 8C1 & 8C2. Note that these align well with the R_c values of our synthetic mRGCs.

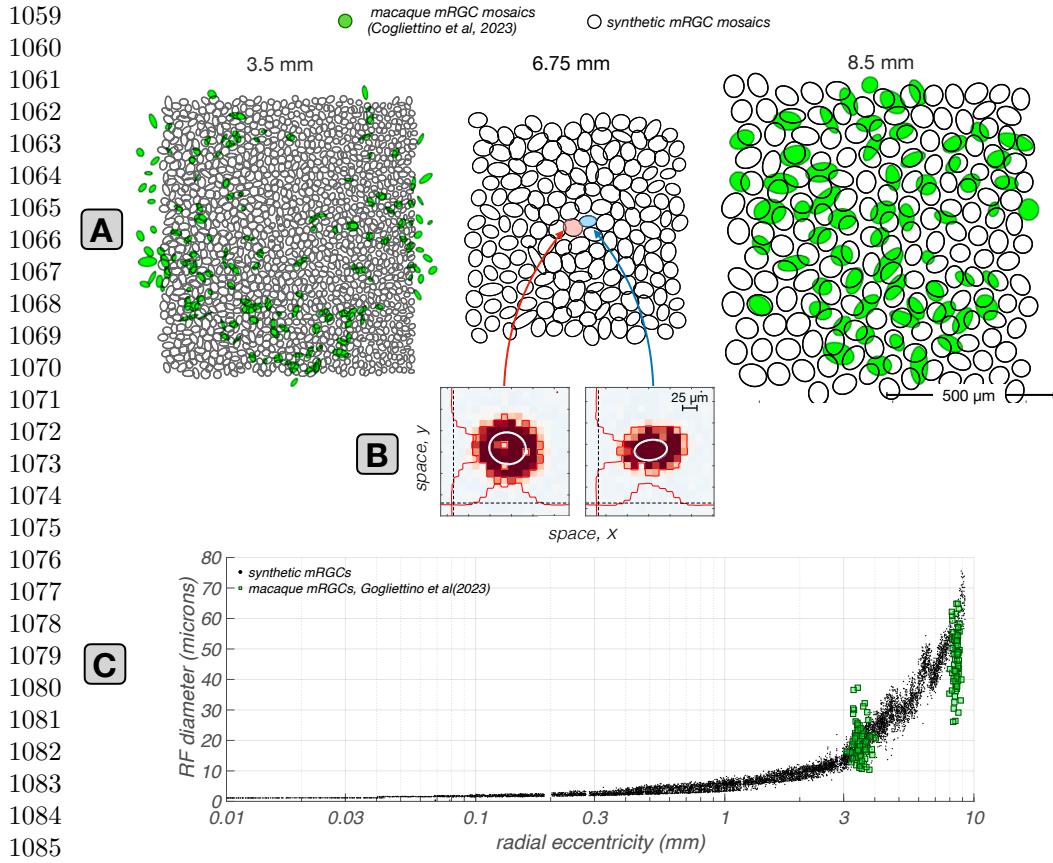
3.3 Validation against *in vitro* physiology in the periphery

We also compared spatial RF properties of synthetic mRGCs against macaque data from *in vitro* mRGC recordings. Since the *in vitro* data are not subject to optical blur, they may be compared directly to the retinal-space characteristics of our model. Data of this sort are currently only available in the peripheral retina.

The first study considered is that of Gogliettino *et al.* [49], in which the spatial RFs of mosaics of macaque mRGCs were mapped using white noise stimulation. To simulate their experiments, we probed synthetic mRGCs with white noise modulated achromatic checkerboard stimuli delivered to the retina under diffraction limited optics. To compute the spatial RFs of synthetic mRGCs, we cross-correlated the synthetic mRGC responses with the white noise stimulus sequence. Results of this analysis are depicted in Fig. 9.

The spatial RFs of cells in synthetic mRGC mosaics at three eccentricities, 3.5 mm, 6.75 mm and 8.5 mm, are illustrated by the black ellipses in the three top panels of Fig. 9A. The superimposed green filled ellipses depict spatial RFs from macaque mRGC mosaics located at 3.5 mm and 8.5 mm. Note that at both eccentricities, there is good correspondence in RF center size and coverage between the synthetic and the macaque mRGC mosaics.

To quantify the retinal space-referred RF center sizes in synthetic mRGCs, we computed the diameter of their RF centers as $2 \times \sqrt{\sigma_{\text{minor}} \times \sigma_{\text{major}}}$, where σ_{minor} and σ_{major} are the standard deviations of the fitted Gaussian ellipsoid along its minor and major axes. The results of this analysis across eccentricity are depicted by the black dots in Fig. 9C, along with the RF center sizes of mosaics of macaque mRGCs located at 3.5 mm and 8.5 mm, which are depicted by the green squares.



1087 **Fig. 9 Retinal space-referred RF center sizes: synthetic vs. macaque mRGCs recorded**
 1088 ***in vitro*. A:** Mosaics of synthetic mRGCs synthesized at three eccentricities, 3.5, 6.75, and 8.5 mm
 1089 along the temporal meridian. The black contours depict Gaussian ellipsoid fits to the increment-
 1090 excitatory regions of the computed RF maps, drawn at the e^{-1} normalized sensitivity level. Only
 1091 the increment-excitatory region of the RF map is fitted. Green contours depict RF maps from two
 1092 macaque mRGCs mosaics from the *in vitro* recordings of Gogliettino *et al.* [49]. **B:** Example spatial
 1093 RF maps of two synthetic mRGCs located at 6.75 mm, computed via white noise stimulation deliv-
 1094 ered to the retina under diffraction limited optics. Regions excitatory to light increments, i.e. the
 1095 RF centers, and to light decrements, i.e., the RF surrounds, are indicated by red and blue colors,
 1096 respectively. The scattered zero excitation spots within the light-increment RF centers correspond to
 1097 the location of S-cones. White lines depict iso-contour plots of Gaussian ellipsoids fitted to the light
 1098 increment-excitatory RF center region, drawn at the e^{-1} normalized sensitivity level. **C:** Compari-
 1099 son of synthetic against macaque mRGC RF center sizes across eccentricity. Black dots depict the RF
 1100 diameters of synthetic mRGCs, computed from the Gaussian ellipsoid fits as $2 \times \sqrt{\sigma_{\text{minor}} \times \sigma_{\text{major}}}$,
 1101 and green squares depict the RF diameters of macaque mRGCs at the two eccentricities where the
 1102 *in vitro* measurements are available.

1100
1101 Note that the correspondence between synthetic and macaque data is excellent at
1102 3.5 mm, whereas at 8.5mm, the RF diameters of the synthetic mRGCs are, on average,
1103 30–40% larger than the RF diameters of macaque mRGCs. The deviation in RF size
1104 at the far periphery may occur because human and macaque retinas differ somewhat

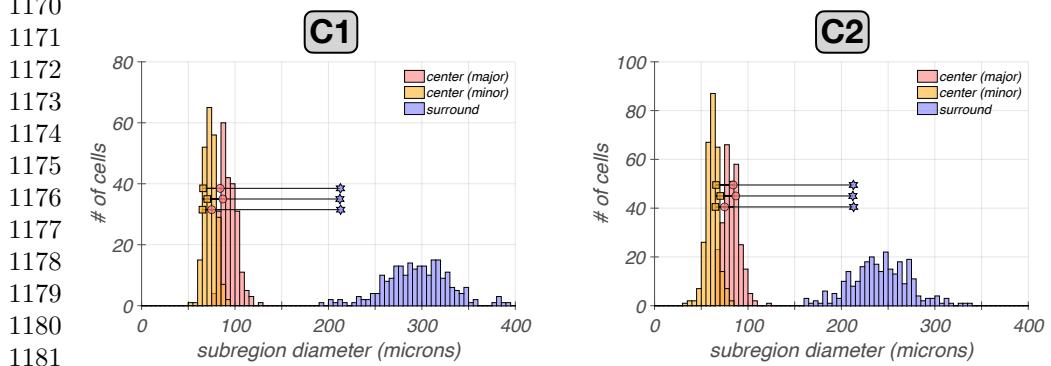
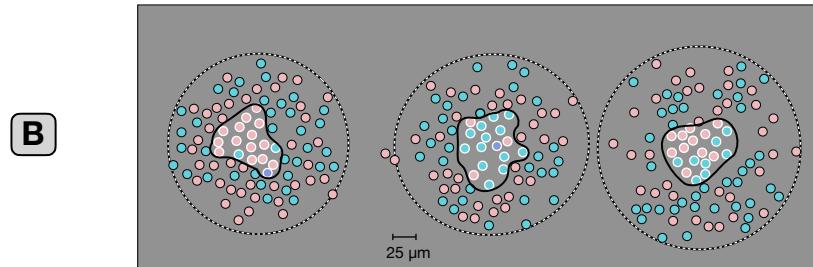
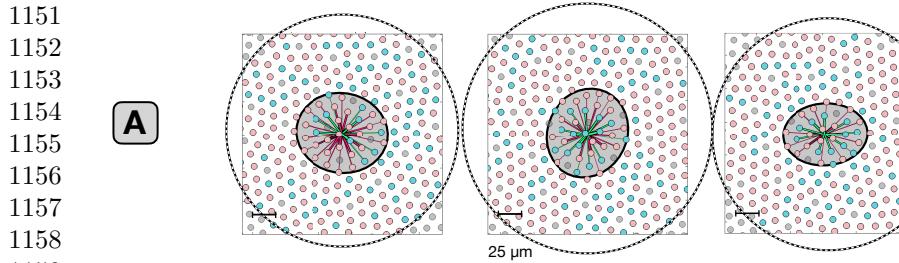
in the periphery. For example, in the human retina, cone density does not change much for eccentricities > 5mm, whereas in the macaque retina it continues to drop as eccentricity increases [50]. The RF size deviation we observe could be the result of a higher mRGC density in the peripheral macaque retina, relative to the human retina. 1105
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The second *in vitro* study we validated our synthetic mRGCs against, is that of Field *et al.* [15], which examined the spatial layout of single cone inputs to the RF centers and surrounds in peripheral macaque mRGCs. Results of this comparison are depicted in Fig. 10. The cone pooling maps of three synthetic mRGCs at a temporal eccentricity of 6.75 mm are depicted in Fig. 10A. The spatial distribution of cone pooling weights in three macaque mRGCs at the same eccentricity from the study of Field *et al.* [15], adapted from their Fig. 4, are shown in Fig. 10B. For both synthetic and macaque mRGCs, the visualized surround cones have pooling weights > 0.005 × the peak center cone weight (Greg Field, personal communication).

Note the general agreement between synthetic and macaque mRGCs in the extent of both their RF centers and surrounds, although again, synthetic mRGCs appear to have slightly larger RFs than their macaque counterparts. Also notable is that the density of cones in the synthetic mRGC cone pooling maps is higher than that seen in the macaque mRGCs. This occurs because our model is based on human cone mosaics, and human cone density is higher than macaque cone density at temporal eccentricities above 5 mm [50], which is where these comparisons are made.

To contrast the relationship in center and surround cone pooling regions between synthetic and macaque mRGCs more quantitatively we compared the diameters of cone pooling regions of the three depicted macaque mRGCs against those of populations of synthetic mRGCs at two eccentricities: 6.75 mm, and 6.0 mm. Results of this analysis are depicted in Fig. 10C1 and 10C2. The minor and major diameters of the center pooling mechanism and the diameter of the surround pooling mechanism for the 3 macaque mRGC cells are depicted by the yellow squares, pink circles and magenta stars, respectively. The corresponding distributions in populations of synthetic mRGCs are depicted by the yellow, pink and magenta histograms, respectively. Note that at 6.75 mm (Fig. 10C1), the cone pooling regions of the synthetic mRGCs are consistently larger than those of the three macaque mRGCs. However, at the slightly less peripheral eccentricity of 6.0 mm (Fig. 10C2) a better agreement exists between model and macaque mRGCs.

These observations highlight an inherent issue in building our mRGC model, namely that we had to employ a mixture of human and macaque data sources: human data regarding the density of cones and the density of mRGC RFs across visual space, human data regarding the characteristics of physiological optics across the retina, and macaque data regarding the spatial characteristics of mRGC RFs and of H1 horizontal cells, with our validations done against macaque data. This is not ideal, as there are some differences between human and macaque retinas [50]. But, it is unavoidable given the lack of complete data in either species. The modeling framework that we devised however, which incorporates data from different sources, can be easily modified as new data become available.



1183 **Fig. 10 Cone pooling maps in RF centers and surrounds: synthetic vs. macaque mRGCs**
1184 **recorded *in vitro*.** **A:** Center and surround cone pooling weight maps for three synthetic mRGCs
1185 at an eccentricity of 6.75 mm along the temporal raphe. Solid and dashed contours include cones
1186 pooled by the RF center and the RF surround, respectively, with pooling weights $> 0.005 \times$ the peak
1187 center weight. **B:** Center and surround cone pooling weights for three macaque mRGCs recorded in
1188 *vitro* at an eccentricity of 6.75 mm along the temporal raphe. White and black disks indicate cones
1189 pooled by the RF center and the RF surround respectively, with same threshold pooling weights as
1190 in A. The macaque mRGCs are from the *in vitro* recordings of Field *et al.* [15]. **C1 &C2:** Compari-
1191 son of minor and major diameters of the center pooling mechanism (yellow squares and pink circles)
1192 and of the surround pooling mechanism (purple stars) in the 3 macaque mRGC cells against corre-
1193 sponding distributions (yellow, pink and magenta histograms) in populations of synthetic mRGCs at
1194 eccentricities of 6.75 mm (C1) and 6.0 mm (C2).

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3.4 Visual *vs.* retinal space– referred RFs: the impact of physiological optics

In this section we characterize how physiological optics interacts with the retinal cone pooling within the RFs of mRGCs to shape their visual space–referred RF properties. Fig. 11 illustrates examples of this interaction at five horizontal eccentricities, $x = [-16^\circ, -8^\circ, 0^\circ, +8^\circ, +16^\circ]$, and 3 vertical eccentricities, $y = [-8^\circ, 0^\circ, +8^\circ]$. The yellow ellipses in each panel of the 3×5 grid of Fig. 11A depict Gaussian ellipsoids fitted to the retinal space–referred RF maps of synthetic mRGCs at the examined eccentricities. The small and non-systematic orientation biases in the retinal space–referred RF maps emerge due to the pooling of multiple cones by the RF center mechanism and are reminiscent of RGC mosaics mapped *in vitro* [39].

The blue ellipses in Fig. 11B depict Gaussian ellipses fitted to the visual space–referred RF maps of the same cells. Note that there are striking and systematic orientation biases in these visual space–referred RF maps, which emerge due to the characteristics of physiological optics, whose PSFs are depicted in Fig. 11C. Clearly, the shape of the PSFs, especially at peripheral locations is a major determinant of the visual space–referred RFs in mRGCs.

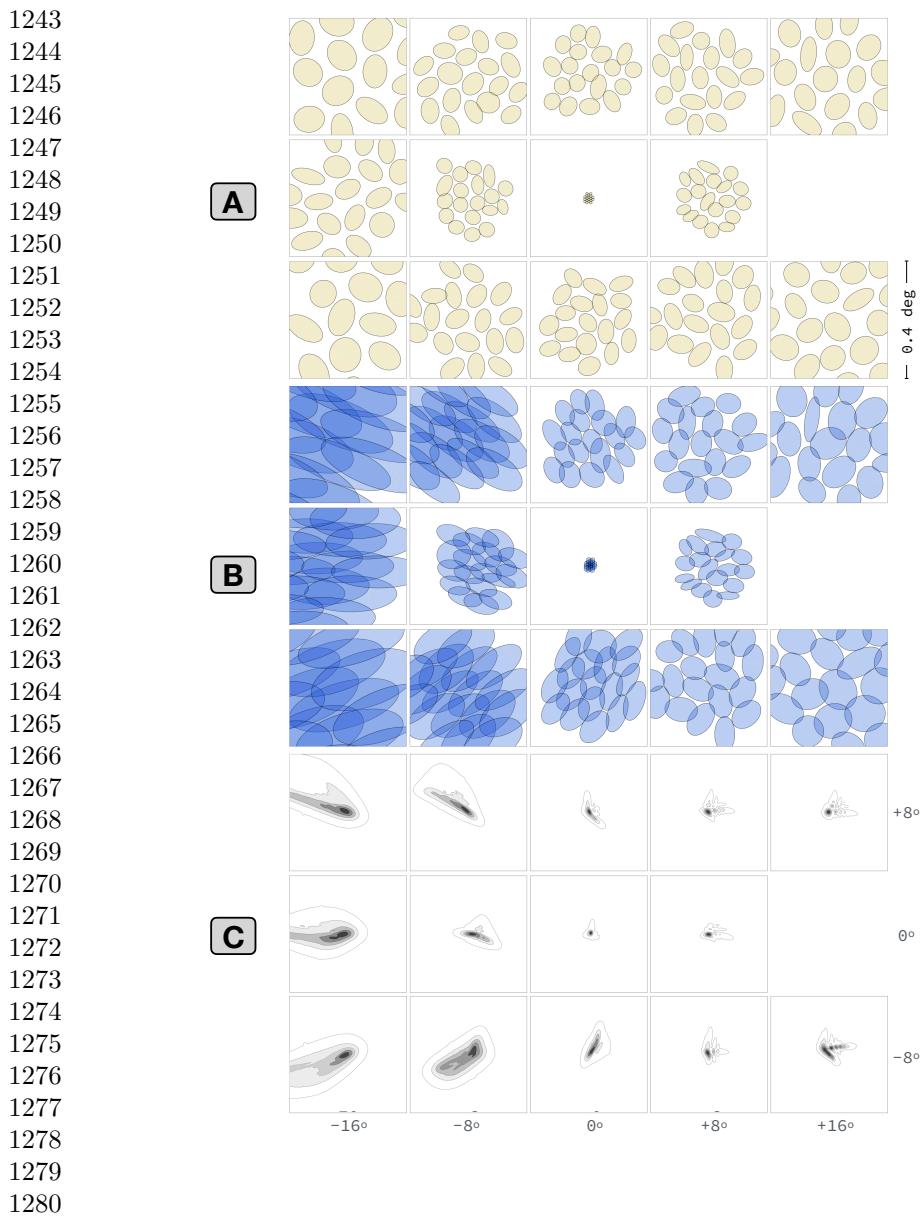
Overall, this analysis demonstrates that there can be substantial differences between *in vivo* and *in vitro* estimates of the spatial RFs of mRGCs, and, once again highlights the notion that inferences regarding retinal wiring from *in vivo* measurements must be evaluated in the context of the effect of the physiological optics. Indeed, in recent on-going work, [31], we have shown the importance of such analyses in assessing inferences regarding cone wiring to the surround subregions of mRGCs based on *in vivo* measurements of their spatio-chromatic RFs.

3.5 Validity of the Difference of Gaussians model applied to *in vitro* responses of mRGCs in retrieving their spatial pooling characteristics

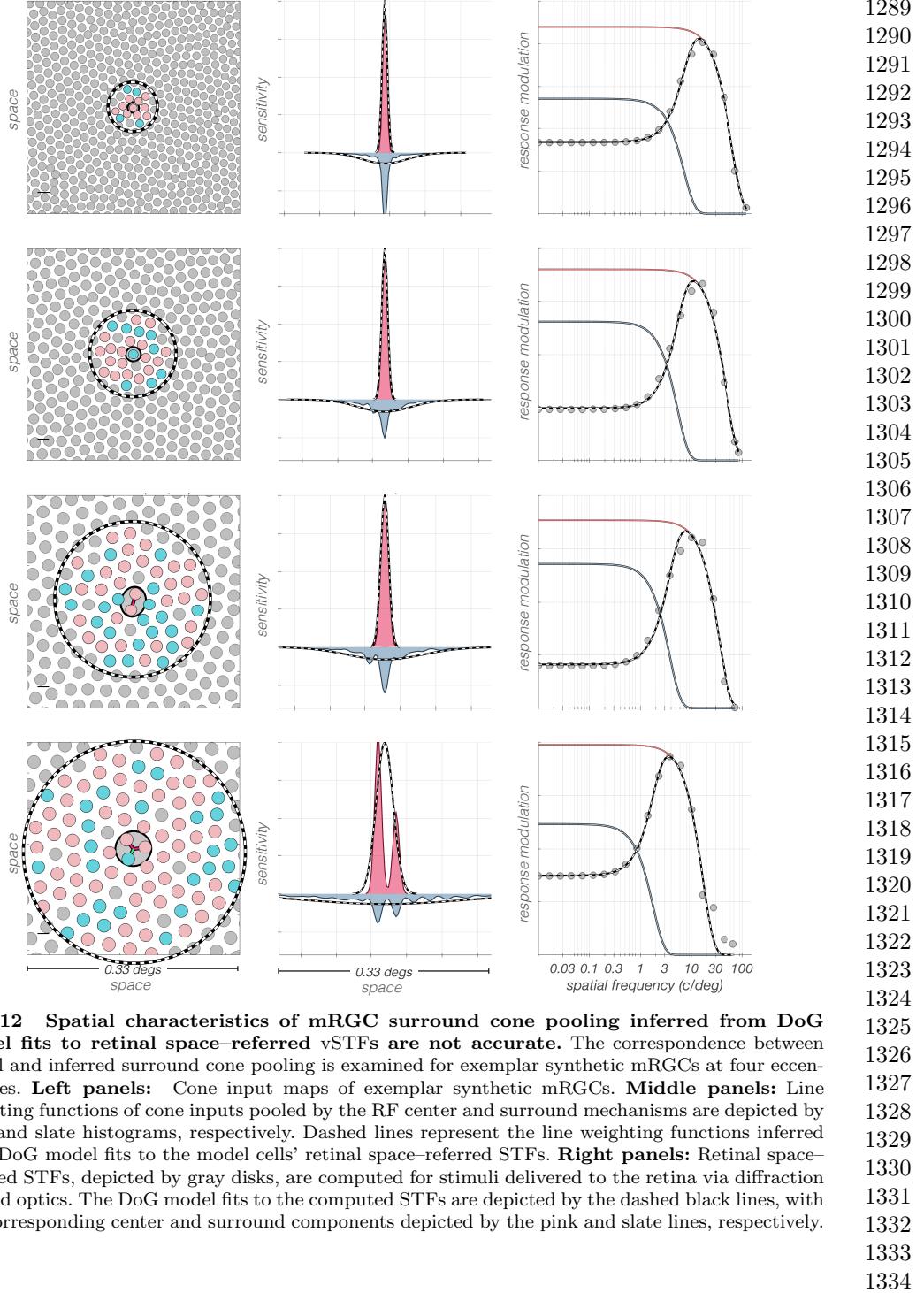
In our synthetic mRGCs, the spatial characteristics of cone pooling within the RF center and the RF surround *component* mechanisms are known by design. This allows us to test how well one can predict these characteristics from Difference of Gaussian model fits to *in vitro* measurements of mRGC STFs, where the RF center and surround mechanisms are driven simultaneously in the absence of optics [16].

Results of this analysis are illustrated in Fig. 12. The cone pooling maps of four exemplar mRGCs are depicted in the left column. The cells in the top two rows both have RF centers with a single cone input, whereas the cell in the third row has a 2-cone RF center, and the cell in the fourth row has a 3-cone RF center.

The pink and maroon histograms depicted in the middle column of Fig. 12, are the y-axis integrated cone pooling weights within these cells’ RF center and surround sub-regions, respectively. The superimposed dashed lines depict the center and surround line weighting profiles, as estimated by the DoG model fit to the cells’ retinal space–referred STFs, which are depicted by the gray disks in the right column of Fig. 12. Note that although the DoG model fits to the computed retinal space–referred STFs (solid lines in right column) are good for all cells, the inferred spatial RF profiles, (dashed



1281 **Fig. 11 Retinal vs. visual space-referred mRGC RF maps across the retina.** Illustration
 1282 of the effect of physiological optics on visual space-referred spatial RF maps of synthetic mRGCs
 1283 across eccentricity. **A:** Retinal space-referred spatial RF maps at different (x,y) eccentricities. Within
 1284 each panel, yellow contours depict Gaussian ellipsoid fits to RF maps of up to 19 cells from a single
 1285 **B:** Visual space-referred spatial RF maps of the same cells, computed under physiological optics
 1286 of one human subject at corresponding eccentricities. **C:** Point spread functions of the employed
 1287 physiological optics at corresponding eccentricities.
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1335 lines in the middle column), do not capture accurately the cone pooling regions of
1336 the RF surrounds (slate histograms in the middle column). The discrepancy between
1337 actual and inferred surround pooling is most obvious in the two top cells which have
1338 single-cone RF centers, and becomes less pronounced as RF center size increases. The
1339 discrepancy involves both the spatial extent and the peak sensitivity of the inferred
1340 surround pooling, which is estimated by the DoG model to be more diffuse with a
1341 weaker peak sensitivity than the cell's actual surround cone pooling.

1342 It is perhaps not surprising that the DoG model does not do a good job of fitting
1343 the model cell surrounds, given that they were constructed as double exponentials to
1344 match the spatial properties of H1 horizontal cells. The key point, however, is that the
1345 DoG model fits to the observable composite STFs are quite good. These observations
1346 suggest that caution should be exercised when inferring mRGC RF surround properties
1347 from DoG model fits to *in vitro* STF measurements.

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1349 3.6 Applications

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1351 We [2, 3, 6], and others [51–53] have reported on how the representation of visual
1352 information at the level of the cone mosaic shapes visual performance, in our case by
1353 exploiting the ISETBio image computable model of cone excitations. The transforma-
1354 tion from cone excitations to RGC responses further shapes the information available
1355 for perceptual decisions, and we can interrogate our linear spatio-chromatic RF model
1356 of the ON-center mRGC mosaic to understand how the information available from
1357 this neuron class differs from that at the cone mosaic.

1358 In this section, we present two example computations of this nature. Our goal is to
1359 illustrate how our model may be exploited in this way, and not to present a full analysis
1360 in either case. Even these initial calculations, however, provide interesting insight.

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1362 3.6.1 Achromatic and chromatic spatial contrast sensitivity

1363 We used a computational observer approach to compute spatial contrast sensitivity
1364 functions (CSFs) for achromatic and L-M cone opponent stimuli, based both on the
1365 representation at the cone mosaic and on the representation at the mRGC mosaic. To
1366 do so, we computed responses to drifting gratings of varying spatial frequency, ω .

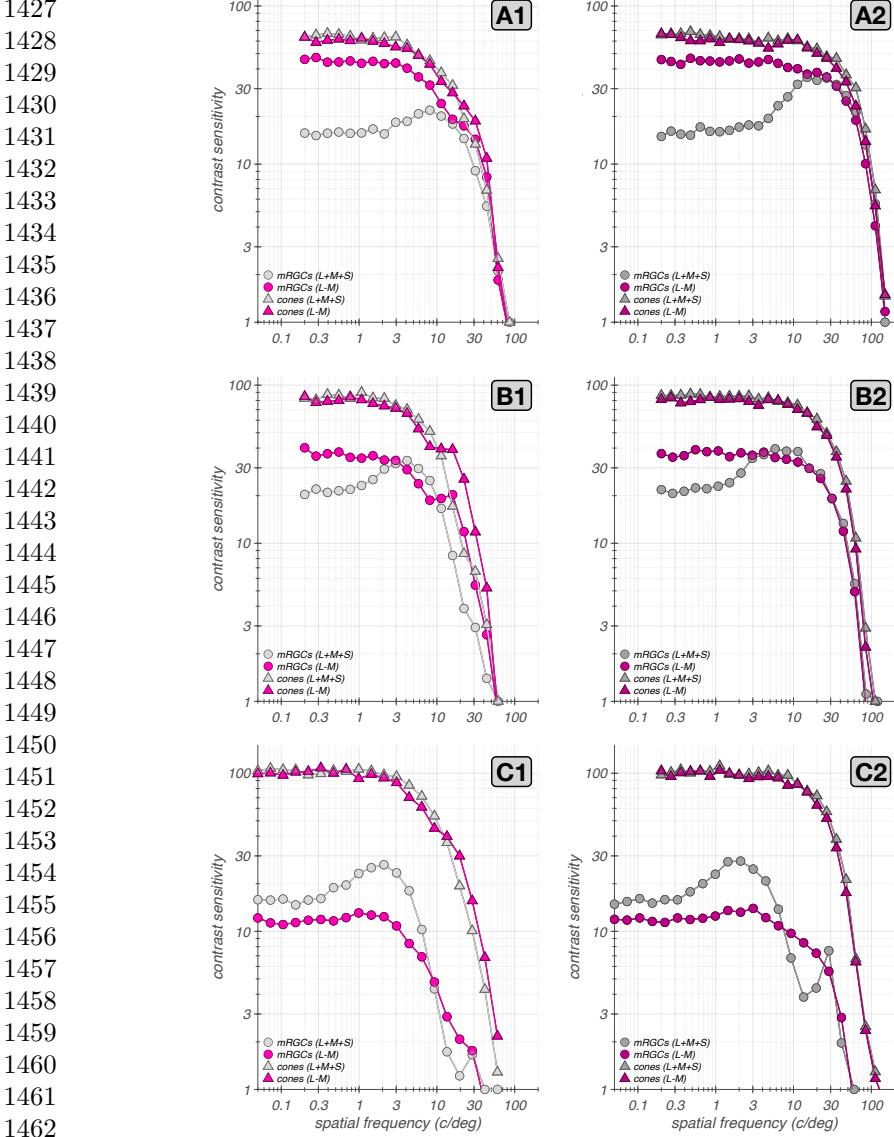
1367 For the achromatic gratings, the L-, M- and S-cone contrast component gratings
1368 were in phase, $C^L(\omega, x, y) = C^M(\omega, x, y) = C^S(\omega, x, y)$. For the L – M gratings, the
1369 L- and M-cone contrast components were in antiphase, $C^L(\omega, x, y) = -C^M(\omega, x, y)$,
1370 and $C^S(\omega, x, y) = 0$. For all stimuli, the mean (x, y) chromaticity was $(0.30, 0.32)$ and
1371 the mean luminance was 100 cd/m^2 . Stimuli were simulated as presented on a typical
1372 CRT monitor, but with 20-bit channel DACs, to avoid intrusion of quantization effects.

1373 For each eccentricity we studied, we oriented the gratings so that they were aligned
1374 with the axis of elongation of the optical point spread function at that eccentricity.
1375 Stimulus size was specified so that it extended over the area spanned by the input
1376 cone mosaic of the employed mRGC mosaic. The size of the mRGC mosaics was varied
1377 between eccentricities so as to achieve nearly equal numbers of mRGCs for mosaics
1378 between which we wished to compare performance.

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Cone fundamentals vary with eccentricity because of variation in macular pigment density and photopigment axial density, and this variation is captured by ISET-Bio. Therefore, in these computations, stimuli were designed using cone fundamentals specific to the eccentricity of the employed mRGC mosaic.	1381
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	1384
At present, our mRGC model does not include spike generation or response noise. Therefore, in these computations we modeled response variability by adding zero mean Gaussian noise to the noise-free model mRGC responses. This approximation allows us to examine relative sensitivity across stimuli and eccentricity, but the overall level of predicted sensitivity is arbitrary. Given the choice of Gaussian noise, we used a template matching computational observer decision rule, with templates provided by the noise-free responses to the stimuli being discriminated. For comparing computational observer performance at the mRGCs with that at the cones, we also adopted the Gaussian noise approximation for the cone excitations, and used the template matching decision rule.	1385
To estimate contrast sensitivity, we varied, for each tested spatial frequency, ω , the contrast of the test stimulus and identified threshold contrast, $C_{\text{threshold}}(\omega)$, as that for which the probability of correctly identifying the test versus a zero contrast stimulus was 80.6%. Contrast sensitivity was defined as $\text{CSF}(\omega) = 1/C_{\text{threshold}}(\omega)$.	1386
Estimates of so computed contrast sensitivities at three eccentricities are depicted in Fig. 13. The contrast sensitivities for stimuli viewed through typical human optics are shown in the left panels of Fig. 13, with disks and triangles depicting sensitivity at the mRGC mosaic and at its input cone mosaic, respectively. For comparison, the right panels of Fig. 13 depict corresponding calculations for stimulus viewed under diffraction-limited optics with no chromatic aberration, as might be measured using adaptive optics. The comparison between left and right panels helps understand which effects in the computed CSFs have their origin in the optics or sampling by the cone mosaic, and which should be attributed to retinal processing through to the mRGCs.	1387
At the fovea, the CSFs at the cone excitation level (triangles in Fig. 13A1), are low pass for both achromatic and L-M stimuli. This is expected because there is no spatial antagonism at the level of the photopigment excitations, and because we do not incorporate spatio-temporal coupling that arises because of interactions between fixational eye movements and post-receptor temporal filtering [54, 55].	1388
On the other hand, the achromatic CSF at the mRGC mosaic exhibits a mild low-spatial frequency attenuation, which is due to the spatial antagonism between mRGC RF centers and surrounds. Note that the low frequency attenuation appears weaker than what is observed under diffraction limited optics (Fig. 13A2). This occurs because physiological optical blur carves sensitivity at the high frequency regime, thereby reducing the apparent effect of the mRGC surrounds on the CSF. We observed a similar effect in foveal macaque mRGCs whose responses were measured under adaptive optics conditions [10].	1389
The L – M opponent CSF of the mRGC mosaic lacks the low-frequency attenuation seen for achromatic modulations because in foveal mRGCs, L – M cone opponent stimuli do not induce substantial spatial antagonism between their single cone RF centers and their surrounds. These observations, which are consistent with what is known regarding the L – M chromatic contrast sensitivity of the mRGC pathway	1390
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1464 **Fig. 13 Computational observer spatial CSFs.** Left column depicts CSFs computed with
 1465 stimuli that are delivered to the retina through typical human optics. Right column depict CSFs
 1466 computed with stimuli that are delivered to the retina under diffraction limited optics. **A1:** CSFs
 1467 for a $0.6^\circ \times 0.6^\circ$ foveal mRGC mosaic and of its input cone mosaic, depicted by disks and triangles,
 1468 respectively. Gray: achromatic; pink: L-M. This mosaic contains 4628 mRGCs. **A2:** Diffraction-limited
 1469 CSFs of the same foveal mRGC mosaic. **B1 & B2:** CSFs for a $2.1^\circ \times 2.1^\circ$ parafoveal mRGC mosaic
 1470 synthesized at an eccentricity of 4° along the temporal meridian. This mosaic contains 4633 mRGCs.
C1 & C2: CSFs, respectively for a $4.1^\circ \times 4.1^\circ$ peripheral mRGC mosaic synthesized at an eccentricity
 1471 of 14° along the temporal meridian. This mosaic contains 2195 mRGCs.

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[16, 56], demonstrate that L – M sensitivity exceeds achromatic sensitivity at low spatial frequencies, consistent with the literature [57].

At high spatial frequencies there is little difference between computational observer sensitivity to achromatic and L – M modulations. This is not true of human observers, where sensitivity drops more rapidly as a function of spatial frequency for red-green isoluminant gratings than for achromatic gratings either with [56] or without typical optical blur [58]. Although our L – M opponent CSFs are not precisely equivalent to the red-green isoluminant CSFs measured in many human experiments, this is not the primary source of the difference between computational and human observers. Rather, it is known that compared to ideal observers, humans lose foveal information available from the cones more rapidly as a function of spatial frequency for red-green than for achromatic gratings [53].

Our example calculation here suggests that this information loss should not be attributed to the linear receptive fields of the mRGCs. We believe this is because optical blur dominates computational observer performance at high spatial frequencies and the single cone RF centers of foveal mRGCs transmit information about each type of stimulus equally well; the surrounds have little effect at high spatial frequencies. Also, we do note that in the present calculations the specific resolution limit, i.e., the spatial frequency at which sensitivity drops to 1, depends on the variance of the added Gaussian noise and is thus somewhat arbitrary. We have chosen a noise level that is low relative to human observers so that our computations show the behavior in the high-spatial frequency regime more fully than would psychophysics conducted through natural optics.

As we move to more peripheral locations, additional features of the CSF emerge. Figs. 13B1 and 13C1 depict results of computations at 4°. Note that under physiological optics viewing (Fig. 13B1) there is a spatial frequency regime in which L – M sensitivity exceeds the corresponding achromatic sensitivity, with the L – M CSF having a notched shape. We have reported this observation in conference abstract form [59]. It occurs because of the wavelength dependent defocus that is introduced by longitudinal chromatic aberration (LCA), which can change the spatial phase of the L– and M–cone stimulus components in the retinal image. Consistent with this interpretation, the notch is present in the CSFs both at the cones and at the mRGCs on the left, but not under diffraction-limited optics (Fig. 13B2), where LCA is zero. Similar effects have been observed for S-cone CSFs [60]. We have presented in abstract form experimental results that suggest that these effects occur in measurements of the human L – M spatial CSF [61].

Comparison of the cone-based CSFs in Fig. 13A1 with those in Fig. 13B1 and Fig. 13C1 also reveals the effect of stronger optical blur with eccentricity, which increases the rolloff of the CSFs with spatial frequency. Similar comparison of the mRGC-based CSFs shows additional rolloff introduced by the increasing size of mRGC RF centers with eccentricity.

Additional observations are notable at 14° (Figs. 13C1 and 13C2). First, a notch arises in the achromatic CSF at high spatial frequencies for the mRGC CSF that is not apparent in the cone CSF. This seems unlikely to be an optical effect, because it is more salient in Fig. 13C2 where optical effects are not present. To explore the origin

1519 of this effect, we computed CSFs at different orientations (not shown), which show
1520 that this notch is orientation dependent and has to do with the precise alignment of
1521 individual cones with the receptive field of an mRGC. We do not explore it further
1522 here.

1523 We also note, once again, that our computational observer is with respect to a noise
1524 level that makes it more sensitive than the human observer, so that the notch shown
1525 in Fig. 13C1 would be unlikely to be revealed with psychophysics conducted with
1526 natural optics. Indeed, in further simulations (data not shown) conducted with twice
1527 the noise variance, we observed that, in addition to an overall reduction in sensitivity,
1528 the high frequency notch disappeared under both physiological and adaptive optics
1529 conditions. It is an interesting question as to whether such effects could be observed
1530 experimentally under adaptive optics conditions.

1531 Finally, note that the L – M advantage over the achromatic CSF is reversed at 14°
1532 of eccentricity. This is because at such high eccentricities, the L – M signal is reduced
1533 by the increased mixing of L– and M–cone signals within the larger mRGC RF centers
1534 and surrounds. Careful comparison of this effect with computational observer predic-
1535 tions for various choices of the model’s spatial homogeneity/spectral purity tradeoff
1536 parameter, ϕ , is an interesting future direction.

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1538 3.6.2 Chromatic contrast sensitivity of synthetic mRGC mosaics: 1539 dependence on eccentricity

1540 As a second example application, we examined chromatic sensitivity for uniform fields
1541 modulated in different directions in the LM-cone contrast plane. We used the same
1542 computational observer approach described above, and evaluated threshold for stimuli
1543 whose contrast was modulated in time. The cone contrasts of stimuli at different chro-
1544 matic directions, θ , on the LM plane were: $C^L(\theta) = \rho \cdot \cos(\theta)$; $C^M(\theta) = \rho \cdot \sin(\theta)$; $C^S(\theta) =$
1545 0. For each θ , we varied ρ to find its threshold value for discriminating that modula-
1546 tion direction from a zero contrast stimulus with a probability of 0.806. To summarize
1547 the computed thresholds across the different chromatic directions, we fit ellipses to
1548 the locus of threshold contrast points.

1549 Fig. 14 depicts computational observer thresholds for synthetic mRGC mosaics and
1550 for their input cone mosaics at different eccentricities. Note that how computational
1551 observer sensitivity changes with eccentricity depends on how stimulus size is covaried
1552 with eccentricity, as does human sensitivity (e.g. [62]). Comparison of the magnitude
1553 of sensitivity for cone- and mRGC-based computational observers depends on how the
1554 noise levels are chosen. For these example calculations, we focus on the shape rather
1555 than magnitude of the elliptical threshold contours. Therefore, each contour shown in
1556 Fig. 14 is normalized so that the threshold along the M cone direction is equal to one.

1557 A few observations are notable. First, the normalized contours for the cone-based
1558 observer are similar across eccentricities and align with the L– and M–cone contrast
1559 axes. They are more elongated in the M–cone direction because our mosaics have more
1560 L cones than M cones. The alignment with the axes is expected [63], and the similarity
1561 of the normalized shapes occurs because this shape depends primarily on the relative
1562 numbers of L and M cones.

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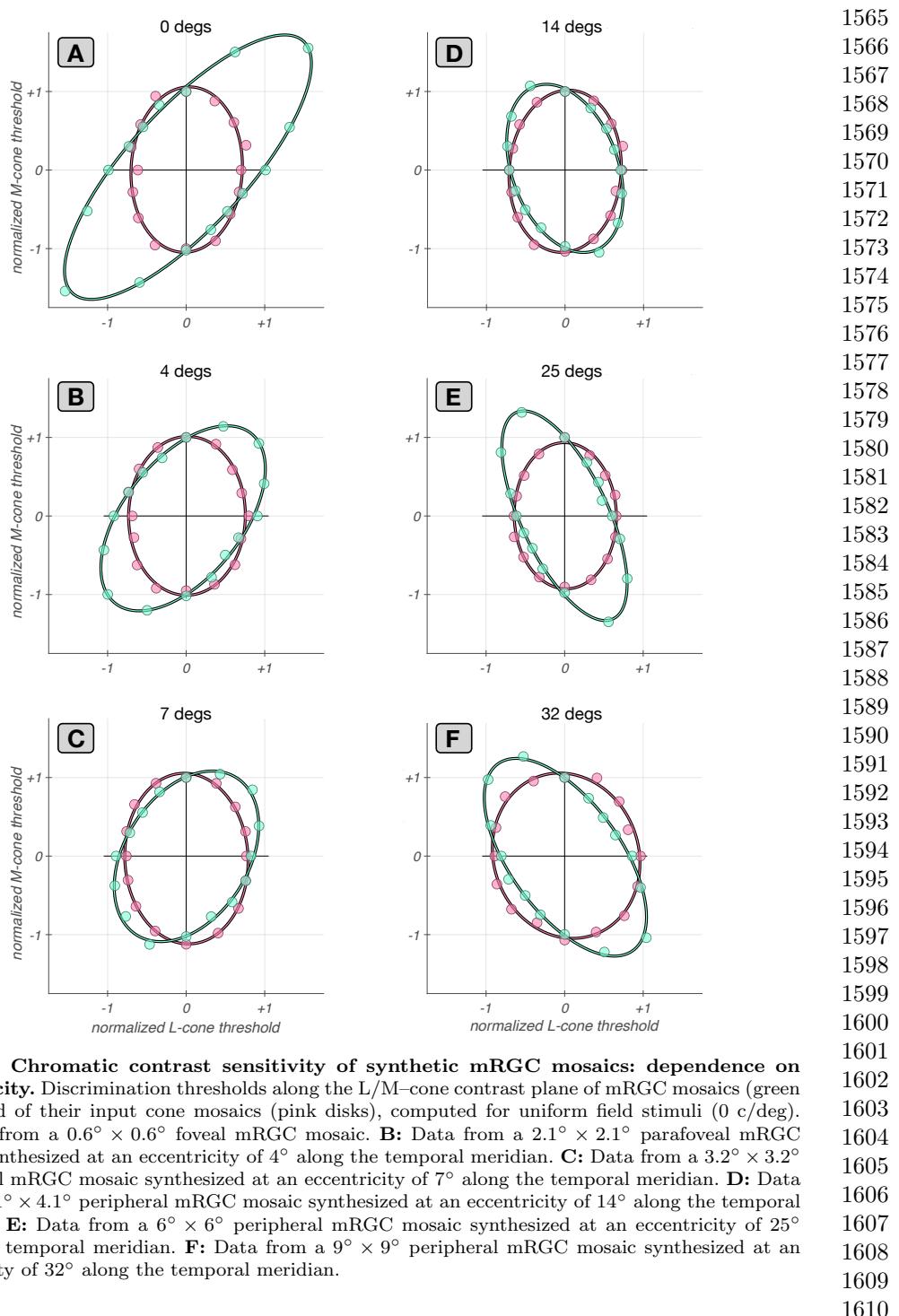


Fig. 14 Chromatic contrast sensitivity of synthetic mRGC mosaics: dependence on eccentricity. Discrimination thresholds along the L/M-cone contrast plane of mRGC mosaics (green disks) and of their input cone mosaics (pink disks), computed for uniform field stimuli (0 c/deg). **A:** Data from a $0.6^\circ \times 0.6^\circ$ foveal mRGC mosaic. **B:** Data from a $2.1^\circ \times 2.1^\circ$ parafoveal mRGC mosaic synthesized at an eccentricity of 4° along the temporal meridian. **C:** Data from a $3.2^\circ \times 3.2^\circ$ parafoveal mRGC mosaic synthesized at an eccentricity of 7° along the temporal meridian. **D:** Data from a $4.1^\circ \times 4.1^\circ$ peripheral mRGC mosaic synthesized at an eccentricity of 14° along the temporal meridian. **E:** Data from a $6^\circ \times 6^\circ$ peripheral mRGC mosaic synthesized at an eccentricity of 25° along the temporal meridian. **F:** Data from a $9^\circ \times 9^\circ$ peripheral mRGC mosaic synthesized at an eccentricity of 32° along the temporal meridian.

1611 Second, in contrast, the mRGC-based threshold contours change markedly with
1612 eccentricity. For the foveal mosaic, the threshold ellipse is highly elongated along 45°
1613 in the L/M-cone contrast plane, indicating that the highest discrimination thresholds
1614 occur when $c^L = c^M$ and lowest thresholds occur when $c^L = -c^M$. This difference
1615 in comparison to the cone-based computations is a consequence of the chromaticic-
1616 opponency of foveal mRGC RFs, which have single cone centers, and thus opponency
1617 between their centers and the surrounds as the surrounds draw on mixed cone-types
1618 [64, 65]. This opponency leads to cancellation of non-opponent L- and M-cone signals
1619 for low spatial frequency stimuli and thus the observed contour elongation along 45°
1620 [63, 66].

1621 Third, as eccentricity increases, the contours first become less elongated and then
1622 elongation starts increasing again but along the 135° rather than the 45° axis. This
1623 is because the cone non-selective wiring model we implemented leads to progressively
1624 less opponency with increasing RF center size [16, 64, 65].

1625 Although the qualitative features that emerge from this example calculation are
1626 understood in the literature, the example illustrates that our model enables this type
1627 of calculation to be made quantitatively in a way that takes chromatic aberration,
1628 stimulus size and spatial frequency and retinal position into account. Of particular
1629 interest to us will be exploring how this type of threshold contour varies with the the
1630 tradeoff between spatial homogeneity and spectral purity of mRGC RF centers (the
1631 center wiring parameter ϕ of our model).

1632

1633 4 Discussion

1634

1635 We developed an image computable model of the linear spatio-chromatic RF mosaic
1636 of mRGCs across the retina. The model extends our image-computable cone mosaic
1637 model [2, 3] by adding a layer of mRGCs which pool signals directly from the cone
1638 mosaic. The connectivity between cones and mRGCs is derived using a simulation
1639 framework that integrates anatomical, physiological and optical quality data, all of
1640 which vary across eccentricity.

1641 By explicitly modeling the optics and photoreceptors, rather than directly express-
1642 ing the RFs in terms of the stimulus, we are able to link our model with both *in-vitro*
1643 and *in-vivo* data, and to make predictions over a range of experimental conditions
1644 that are otherwise difficult to compare. These include psychophysical and physiolog-
1645 ical measurements made through physiological optics (natural viewing conditions),
1646 interferometric and adaptive optics techniques that bypass or correct for optical
1647 aberrations, and *in-vitro* physiology, where the natural optics are not present.

1648 To build the model we had to overcome the challenge that current data about
1649 mRGC properties are incomplete and, where they exist, may come from different
1650 species, different measurement modalities, and from different eccentricities. For exam-
1651 ple, there are *in-vivo* measurements of mRGC linear receptive fields across the retina
1652 [17], but physiological optics blur the stimuli so that they do not constrain mRGC
1653 input at the cone-by-cone resolution we seek. On the other hand, although there is
1654 single cone-resolution connectivity data from *in-vitro* physiology [15], these data are
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currently limited to large eccentricities ($\geq 25^\circ$). Thus, we developed a modeling framework that allows integration of data from multiple sources. This framework is an important contribution in its own right; we expect it will be useful to us and others, for incorporating new data that become available and for modeling other RGC classes.	1657
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We showed that the model captures visual space-referred spatial RF properties of macaque mRGCs recorded <i>in-vivo</i> across eccentricities, as well as retinal space-referred spatial RF properties of macaque mRGCs recorded <i>in-vitro</i> . We also showed that physiological optics plays a major role in shaping the visual space-referred spatial RF properties, so that inferences regarding retinal circuitry made from <i>in-vivo</i> measurements need to be evaluated in the context of the optics. Further, we showed that even under <i>in-vitro</i> conditions, where the optics are eliminated, the traditional approach of fitting a Difference of Gaussian model to spatial responses can lead to incorrect assessments of the properties of cone pooling in the mRGC surrounds.	1662
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1703 not been our goal, we would have had to bring in human data to characterize the
1704 physiological optics across the visual field, as such data are not currently available in
1705 macaque. At the same time, not all the required data are available for human: although
1706 measurements of cone and mRGC density and physiological optics across the retina
1707 are available, physiological characterizations come from the macaque.

1708 The need to mix data across the two closely related species produces tension in
1709 cases where the parameters for the two species differ. An example is the different
1710 cone densities in the far periphery [50], which intrudes on the interpretation of the
1711 comparison between our model and *in-vitro* physiology in that retinal region. As more
1712 data become available in both species, and as species differences come more fully into
1713 focus [67], our approach should allow more fully differentiated models to be developed
1714 targeted at each.

1715

1716 4.2.2 Noise, nonlinearities and temporal dynamics

1717 Although the current model captures fundamental aspects of the visual representation
1718 at the level of the mosaic of ON mRGCs, there are known characteristics of mRGCs
1719 that it does not account for. These include static and spatial nonlinearities, temporal
1720 filtering, spike generation, and physiologically constrained response noise. The modeling
1721 framework we developed is extensible however, so that these components may be
1722 included through future work.

1723 Response variability models are available for macaque mRGCs, as descriptions of
1724 spike generation mechanisms [26, 33, 68]. In addition, we can incorporate nonlinearities,
1725 such as (a) adaptation effects introduced through the phototransduction cascade
1726 [69], (b) compressive and expansive static nonlinearities in the output of mRGCs
1727 [23, 33], and (c) spatial nonlinearities introduced by rectifying sub-units within the
1728 RFs of mRGCs [21, 22]. Explicit inclusion of photocurrent-based responses in the
1729 input to the mRGCs introduces a temporal component to the response model [69]. In
1730 addition, a second temporal filter may be added, such that when combined with the
1731 photocurrent filter will yield the bandpass filter characteristics observed in macaque
1732 mRGCs [25].

1733 Our current model does not represent explicitly the properties of the retinal circuitry (horizontal, bipolar, and amacrine cells) that produces the mRGC response
1734 properties, as we have opted instead to work towards a functional model that describes
1735 those properties. A complementary mRGC modeling approach that does consider some
1736 of these cell types explicitly has recently been published [30], and there are other
1737 modeling efforts that have examined the influence of the various retinal interneurons
1738 on RGC response properties [28, 32]. We note however, that some of the processing
1739 performed by these other retinal cell types is incorporated implicitly in the current
1740 cone-to-mRGC model, such as the parametric form of the surrounds inherited from
1741 H1 cells.

1742 The framework we developed is designed so that it would be possible to interpose
1743 explicit models of intermediate retinal cell types. Representing the action of different
1744 cell types explicitly may in the longer run be an effective way to account for response
1745 nonlinearities in the mRGCs, or in other classes of retinal ganglion cells. Moreover,
1746 using our framework to model other cell classes may be of interest to those seeking to

interpret responses of those classes *per se*, or in the retinal mechanisms that produce RGC response properties. 1749
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1751

4.2.3 OFF mRGC mosaic

Because we model the linear RF, the distinction between ON and OFF mRGCs is subtle. However, our model should be thought of as a model of only the ON mRGCs because the synthetic cells only pool signals from L- and M-cones. This is believed true for ON mRGCs, but recent evidence suggests that OFF mRGCs draw upon all three types of cones in their RF centers [15, 37, 38]. Incorporating S-cone input into an OFF-center mRGC model is straightforward.

Another question that arises when considering a model of OFF mRGC mosaic is how to split the density of mRGCs in two populations at different eccentricities. In the current model, the ON mRGC density was assumed to be half of all mRGCs across all eccentricities. This seems reasonable for central retina where mRGC centers draw primarily on a single cone and where anatomical evidence suggests that each cone provides input to the center of one ON and one OFF midget bipolar cell. However, there is evidence that the RFs of peripheral ON midget (and parasol) RGCs are larger than their OFF counterparts in both human and macaque retinas [40]. This implies that the density of ON RGC cells might be lower in the periphery than that of OFF cells, given that ON and OFF mRGCs have similar RF overlap [39]. One idea is to treat the asymmetry between ON and OFF mRGC RF densities in an eccentricity-dependent manner, similar to the way we encoded a variable-with-eccentricity RF center overlap.

Finally, when adding an OFF mRGC mosaic one should allow for the possibility of coordination between the ON and the OFF submosaics, to account for recent observations regarding systematic shifts in the spatial layouts of ON and OFF mRGCs [70].

Using the software

The developed software for synthesizing ON mRGCm mosaics across the retina and for computing with them is part of ISETbio and is freely available at

<https://github.com/isetbio/isetbio>. An introduction to using the mRGCmosaic software is available at:

[https://github.com/isetbio/isetbio/wiki/Retinal-ganglion-cell-\(RGC\)-mosaics](https://github.com/isetbio/isetbio/wiki/Retinal-ganglion-cell-(RGC)-mosaics), and a number of MATLAB tutorials specific to the mRGCmosaic can be found at <https://github.com/isetbio/isetbio/tree/main/tutorials/mrgc>.

These tutorials demonstrate (a) how to use mosaics of ON mRGCs that have been synthesized at a number of eccentricities, and (b) how to build and validate mRGC mosaics at any desired eccentricity, using a number of design choices. A summary of current available tutorials is shown in Table 1.

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1795 **Table 1 List of tutorials for computing with mRGC mosaics and de novo**
 1796 **synthesis of mRGC mosaics.**

1797	Tutorial name	Scope
<i>Computing with mRGC mosaics</i>		
1799	<code>t_mRGCMosaicVisualizeWithOptics.m</code>	Visualizes a previously synthesized mRGC mosaic and the optics that were used for its synthesis
<i>Synthesizing mRGC mosaics</i>		
1800	<code>t_mRGCMosaicSynthesizeAtStage1.m</code>	Denovo synthesis of the spatial position lattices of cones and mRGC RF centers (stage 1)
1801	<code>t_mRGCMosaicSynthesizeAtStage2.m</code>	Denovo synthesis of an mRGC mosaic at different sub-stages of cone-to-mRGC RF center connectivity (stage 2)
1802	<code>t_mRGCMosaicSynthesizeAtStage3.m</code>	Denovo synthesis of an mRGC mosaic at different sub-stages of cone-to-mRGC RF surround connectivity (stage 3)
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1818 **Declarations**

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 1823

1824 **Conflict of interest/Competing interests**

1825 Not applicable

1827

1828 **Ethics approval and consent to participate**

1829 Not applicable

1830 **Consent for publication**

1833 Not applicable

1834

1835 **Data availability**

1836 Datasets (mRGCmosaics) generated during the current study are available at:
 1838 <https://github.com/isetbio/isetbio/tree/main>

1839

1840

Materials availability	1841
Not applicable	1842
	1843
Code availability	1844
The code used to generate the data, and various tutorials on how to use the software are available at: https://github.com/isetbio/isetbio/tree/main	1846
	1847
	1848
An introduction to using the software is available at:	1849
https://github.com/isetbio/isetbio/wiki/Retinal-ganglion-cell-(RGC)-mosaics	1850
	1851
	1852
Author contribution	1853
NPC: conceptualization, algorithm development, coding, data curation, validation, visualization, writing of original draft	1855
DHB: conceptualization, coding, reviewing and editing of manuscript	1856
BW: conceptualization, coding, reviewing and editing of manuscript	1857
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1887 **Appendix A Deriving cone weights to the mRGC
1888 RF centers**
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1890 **A.1 Local topology-based convergent connections (stage 2A)**
1891

1892 During the first sub-stage of cone to mRGC RF center connectivity, cones are con-
1893 nected to single mRGC RF centers based on the local topology of their respective
1894 lattices. Starting with the cell whose RF center is at most central location of the
1895 mRGC lattice, we connect $n_{pool}(\epsilon)$ number of L- and M-cones to it, where:

1896
1897
$$n_{pool}(\epsilon) = \lfloor \frac{D_{cones}(\epsilon)}{D_{mRGCRF}(\epsilon)} \rfloor \quad (A1)$$

1898

1899 with $D_{cones}(\epsilon)$ and $D_{mRGCRF}(\epsilon)$ being the local spatial densities of the cone mosaic
1900 and of the mRGC RF centers, respectively, at the eccentricity, ϵ , of the target mRGC.
1901 We draw from the nearest cones that have not yet been connected and whose distance
1902 to the mRGC RF center does not exceed a fraction of the local mRGC RF center
1903 spacing. This fraction is a parameter of the model and for the work presented here
1904 was set to 0.6.

1905 Continuing with these assignments of cones to mRGC RF centers, we move outward
1906 to more peripheral locations in the mRGC mosaic, connecting cones to each mRGC
1907 RF center. Any L- and M-cones that remain unconnected at the end of this sub-stage
1908 are then connected to their nearest mRGC RF center, so that all cones are connected
1909 to one mRGC RF center.

1910 This sub-stage can result in local inhomogeneities in both the number of cones and
1911 the type of cones pooled within neighboring mRGC RF centers. These inhomogeneities
1912 are smoothed out as part of the next sub-stage.

1913
1914 **A.2 Optimizing cone connections to mRGC RF centers
1915 (stage 2B)**

1916 In the second sub-stage of the cone to mRGC RF center connectivity, convergent
1917 connections from multiple cones to single mRGC RF centers are optimized according
1918 to a desired balance between spatial homogeneity and spectral purity. This is achieved
1919 by reassigning cones between nearby mRGC RF centers, which itself occurs in two
1920 steps.

1921 In the first step, we allow cone reassessments to a target mRGC from neighboring
1922 mRGCs that have a higher input cone numerosity in their RF centers. In the second
1923 step, we allow cone swaps between a target mRGC and its neighbors, independently
1924 of their input cone numerosities.

1925 The heuristics followed in the first step are as follows. We begin by targeting
1926 mRGCs with a single input cone and continue to target mRGCs with progressively
1927 higher input cone numerosity. Within each set of targeted input cone numerosity,
1928 mRGCs are sorted based on ascending retinal eccentricity. For each targeted mRGC
1929 we determine up to 6 neighboring mRGCs which have input numerosity that exceeds
1930 that of the target mRGC by at least 2 cones.

Cone reassessments from the candidate donor mRGCs to the target mRGC are	1933
executed in multiple passes. Starting with the neighboring mRGC of the highest input	1934
numerosity, we determine the best transfer of a single cone. If there are no eligible	1935
donor nearby mRGCs, we move to the next targeted mRGC. If there is a single candi-	1936
date, we accept it and execute the cone transfer. If there are more than one candidates,	1937
for each candidate donor mRGC we compute a cost function, C, for reassigning each of	1938
its cones to the target mRGC, and pick the transfer that minimizes C across all cones	1939
and all candidate donor mRGCs. The cost function is described in more detail below.	1940
Once the optimal cone transfers for each mRGC of the targeted input cone	1941
numerosity are executed, we move to the next pass, examining possible transfers from	1942
neighboring mRGCs of lower input cone numerosity than before, but still higher than	1943
the input cone numerosity of the targeted mRGCs. Once all passes are executed, this	1944
process is repeated, now targeting mRGCs with increasing input cone numerosity,	1945
until all input cone numerosities have been targeted.	1946
In the second step, we only allow for cone swaps between an mRGC RF center and	1947
one of its neighbors. For each mRGC of the targeted input cone numerosity, we deter-	1948
mine its 6 closest neighbors, but now without regard to their input cone numerosity.	1949
For each of these neighboring mRGCs, we evaluate the cost function, C, for all pos-	1950
sible combinations of cones from the target mRGC and cones from the neighboring	1951
mRGC and pick the combination that minimizes C. The selected cone swap is exe-	1952
cuted only if the post-swap value of C is lower than its pre-swap value. Multiple passes	1953
through the entire mRGC mosaic, are executed, with each pass targeting mRGCs with	1954
progressively higher input cone numerosity.	1955
The cost function, C, employed to determine the optimal transfer/swapping is based on	1956
the position and types of the cones pooled by the target mRGC, t , and the examined	1957
neighboring mRGC, t_i . For each examined pair of mRGCs, (t, t_i) , $C^{(t, t_i)}$ is defined as:	1958
$C^{(t, t_i)} = \phi \cdot C_{\chi}^{(t, t_i)} + (1 - \phi) \cdot C_{\lambda}^{(t, t_i)} \quad (\text{A2})$	1959
where $C_{\chi}^{(t, t_i)}$ quantifies the degree of spatial incompactness, $C_{\lambda}^{(t, t_i)}$ quantifies the degree	1960
of spectral impurity. The ϕ parameter controls the desired trade-off between spatial	1961
incompactness and spectral impurity of the RF centers. When $\phi = 1$, cone reassig-	1962
nements/swaps are selected so as to minimize the spatial incompactness score, when	1963
$\phi = 0$, cone reassessments are chosen so as to minimize the spectral impurity score,	1964
and for intermediate values of ϕ , cone reassessments are chosen so as to minimize a	1965
ratio of the two scores.	1966
The spatial incompactness score, $C_{\chi}^{(t, t_i)}$, in Eq. A2 is defined as:	1967
$C_{\chi}^{(t, t_i)} = C_{\chi_N}^{(t, t_i)} + C_{\chi_o}^{(t, t_i)} \quad (\text{A3})$	1968
The $C_{\chi_N}^{(t, t_i)}$ term quantifies the differential input cone numerosity between the	1969
examined pair of mRGCs, and is defined as:	1970
$C_{\chi_N}^{(t, t_i)} = (N_L^t + N_M^t) - (N_L^{t_i} + N_M^{t_i}) \quad (\text{A4})$	1971

1979 with N_L^t and N_M^t are the numbers of L– and M–cones pooled by the RF center of
 1980 mRGC t , respectively. The $C_{\chi_o}^{(t,t_i)}$ term is a measure of the spatial overlap of the two
 1981 sets of cones pooled by the two mRGCs, and is defined as the inverse of the distance
 1982 between the centroids, (P^t, P^{t_i}) , of the sets of pooled cones normalized by the sum of
 1983 their respective spatial standard deviations, (σ^t, σ^{t_i}) :

1984

$$1985 \quad C_{\chi_o}^{(t,t_i)} = 1 / \left(\frac{\|P^t - P^{t_i}\|}{\sigma^t + \sigma^{t_i}} \right) \quad (\text{A5})$$

1987

1988 A low value of $C_{\chi_o}^{(t,t_i)}$ indicates low overlap between the sets of cones pooled by the
 1989 examined pair of mRGCs and conversely, a high value indicates a large overlap.

1990 The spectral impurity score, C_λ^{t,t_i} , in Eq. A2, is defined as the sum of spectral
 1991 impurities of the RF centers of the pair of analyzed mRGCs:

1992

$$1993 \quad C_\lambda^{t,t_i} = C_\lambda^t + C_\lambda^{t_i} \quad (\text{A6})$$

1994

1995 with C_λ^t , quantifying the degree of non-specificity, with regard to the type of cone, in
 1996 the pooling within the RF center of an mRGC, defined as:

1997

$$1998 \quad C_\lambda^t = \frac{\min([N_L^t, N_M^t])}{N_L^t + N_M^t} \quad (\text{A7})$$

2000

2001 Values of C_λ^t near zero indicate a low amount of mixture of L– and M–cones, and
 2002 therefore a RF with a high degree of spectral purity, and conversely, values of C_λ^t , near
 2003 0.5, indicate an equal mixture of L– and M–cones, and therefore a RF center with a
 2004 low degree of spectral purity.

2005

2006 A.3 Divergent cone connections to multiple mRGC RF 2007 centers (stage 2C)

2009 In the final sub-stage of establishing the RF center connectivity, the exclusivity of
 2010 connections is relaxed, and cone connections are allowed to diverge to more than one
 2011 mRGC RF center. This divergence is guided by *in-vitro* measurements of mRGC RF
 2012 center overlap in the macaque [39].

2013 According to these observations, neighboring mRGC RF centers abut at approxi-
 2014 mately one standard deviation of their Gaussian RF profile. One caveat of using these
 2015 *in-vitro* measurements to establish cone divergence in the model, is that these measure-
 2016 ments are only available in the far periphery (30–40 degrees), with no data available
 2017 for more central locations. Anatomical studies suggest, however, that, in the central
 2018 retina, there must be little to no divergence of cone signals to mRGCs RF centers, so
 2019 we chose to implement an eccentricity-varying divergence in our model.

2020 To achieve this, we begin by fitting an ellipsoid to the spatial pooling map of cones
 2021 that are exclusively connected to the RF center of an mRGC, and extract the rotation,
 2022 α , and the major/minor axes, σ_x, σ_y of the fitted ellipsoid. Next, a supra-Gaussian
 2023

2024

ellipsoid function, $G(x, y, n)$, defined as:

$$G(x, y, n) = \exp \left[-0.5 \times \left(\sqrt{(y'^2 + y'^2)} \right)^n \right] \quad (A8)$$

with:

$$\begin{bmatrix} x' & y' \end{bmatrix} = \begin{bmatrix} x & y \end{bmatrix} \cdot \begin{bmatrix} \cos(\alpha) & -\sin(\alpha) \\ \sin(\alpha) & \cos(\alpha) \end{bmatrix} \cdot \begin{bmatrix} 1/\sigma_x & 0 \\ 0 & 1/\sigma_y \end{bmatrix} \quad (A9)$$

is computed by scaling the values of σ_x, σ_y by a common factor, so that the value of $G(x, y, n)$, evaluated at the most remote exclusively-connected cone(s) is $k \times e^{-1/2}$. The value of k is determined empirically so that RF maps of nearby mRGCs computed under diffraction-limited optics abut when their sensitivities drop to $e^{-1/2}$ (per [39]).

By varying the exponent of the supra-Gaussian, n , we model varying degrees of cone divergence. When $n = 10$, we obtain a flat-top Gaussian with very sharp fall-offs, modeling minimal cone divergence. When $n = 2$, we get a standard Gaussian modeling cone divergence that is consistent with the *in-vitro* measurements of RF center overlap at peripheral locations.

By allowing n to vary with eccentricity using a sigmoidal function we obtain a gradual transition in cone divergence with eccentricity. The slope and mid-point of the sigmoidal variation of n are currently chosen arbitrarily, with the only restrictions that above 15° , n is stable at 2.0, and below 7° , n is stable at 10.0. The weights of divergent cone-mRGC RF center connections are computed by evaluating the supra-Gaussian ellipsoid at the positions of all cones in the vicinity of the examined mRGC.

Appendix B Deriving cone weights to the mRGC RF surrounds

B.1 Choosing physiology-based constraints for deriving surround cone weights in stage 3B

The optimization of the parameters of the surround cone pooling functions at each iteration is driven by the residual between the visual STF that is computed based on the surround pooling weights at the previous iteration and the Difference of Gaussians model fit to it, $DoG(\omega)$, which is given by:

$$DoG(\omega) = K_c \cdot R_c^2 \cdot \exp[-\pi \cdot R_c \cdot \omega]^2 - K_s \cdot R_s^2 \cdot \exp[-\pi \cdot R_s \cdot \omega]^2 \quad (B10)$$

This aspect of the optimization captures the observation that the DoG model provides a reasonable fit to the *in-vivo* measured STFs of macaque mRGCs. To ensure adherence to the *in-vivo* data of Croner & Kaplan, the DoG model fit is constrained so that the ratio of surround to center radii, R_s/R_c , and the ratio of surround to center integrated sensitivities, $K_s/K_c \times (R_s/R_c)^2$, both remain within a specified tolerance range from the corresponding macaque data.

Specifically, for the model's R_s/R_c ratio, we enforce

$$\frac{R_s^m}{R_{c_*}^m} \times (1 - \tau) \leq \frac{R_s}{R_c} \leq (1 + \tau) \times \frac{R_s^m}{R_{c_*}^m} \quad (\text{B11})$$

2076 where R_c^m and R_s^m are the mean values of center and surround radii across the Croner
 2077 & Kaplan population of macaque mRGCs at the eccentricity of the synthesized mRGC.
 2078 The model's $K_s/K_c \times (R_s/R_c)^2$ ratio is constrained in the same way.

2079 The residual between the visual STF and the Difference of Gaussians model fit to
 2080 it, drives the optimization of the surround pooling function. This function is a double
 2081 exponent (following the H1 horizontal cell spatial RF in the macaque [43]):

$$W_s(r) = K_{\text{wide}} \times \exp[-r/R_{\text{wide}}] + K_{\text{narrow}} \times \exp[-r/R_{\text{narrow}}] \quad (\text{B12})$$

To ensure that the surround pooling function remains consistent with parameter values observed in macaque H1 cell [43], the optimization of $W_s(r)$ is also constrained so that ratio of radii, $R_{\text{narrow}}/R_{\text{wide}}$, and the ratio of volumes, $V_{\text{narrow}}/V_{\text{wide}} = K_{\text{narrow}}/K_{\text{wide}} \times (R_{\text{narrow}}/R_{\text{wide}})^2$, of the two exponentials both remain within a specified tolerance range of the macaque data.

In the present work, the tolerance range for $R_{\text{narrow}}/R_{\text{wide}}$ was set to [0.07, 0.35] for all mosaics, whereas the tolerance range for $V_{\text{narrow}}/V_{\text{wide}}$ was set to [0.01, 0.6] for mosaics at eccentricities $\leq 15^\circ$, to [0.3, 0.9] for eccentricities in $15^\circ \dots 25^\circ$, and to [0.6, 1.3], for eccentricities $\geq 25^\circ$.

The joint manipulation of the tolerance values applied to the parameters of the DoG model fit to the vSTF, and to the parameters of the double exponential surround pooling model, $W_s(r)$, allows for different options for deriving spatial pooling functions in synthetic mRGC surrounds.

One option is to set very strict tolerances on the parameters of DoG model fit while allowing for a large tolerance in the parameters of $W_s(r)$. Results of this choice are depicted in the left-most column of Figure B1. A second option would be to allow medium tolerance levels in both the DoG model fit and the $W_s(r)$. Results of this choice are depicted in the middle column of Figure B1. A third option would be to enforce strict tolerances in $W_s(r)$, for example matching parameters of individual H1 horizontal cells, while allowing for a loose tolerance in the DoG model fit. Results of this choice are depicted in the right column of Figure B1. In the present work, we chose the second option.

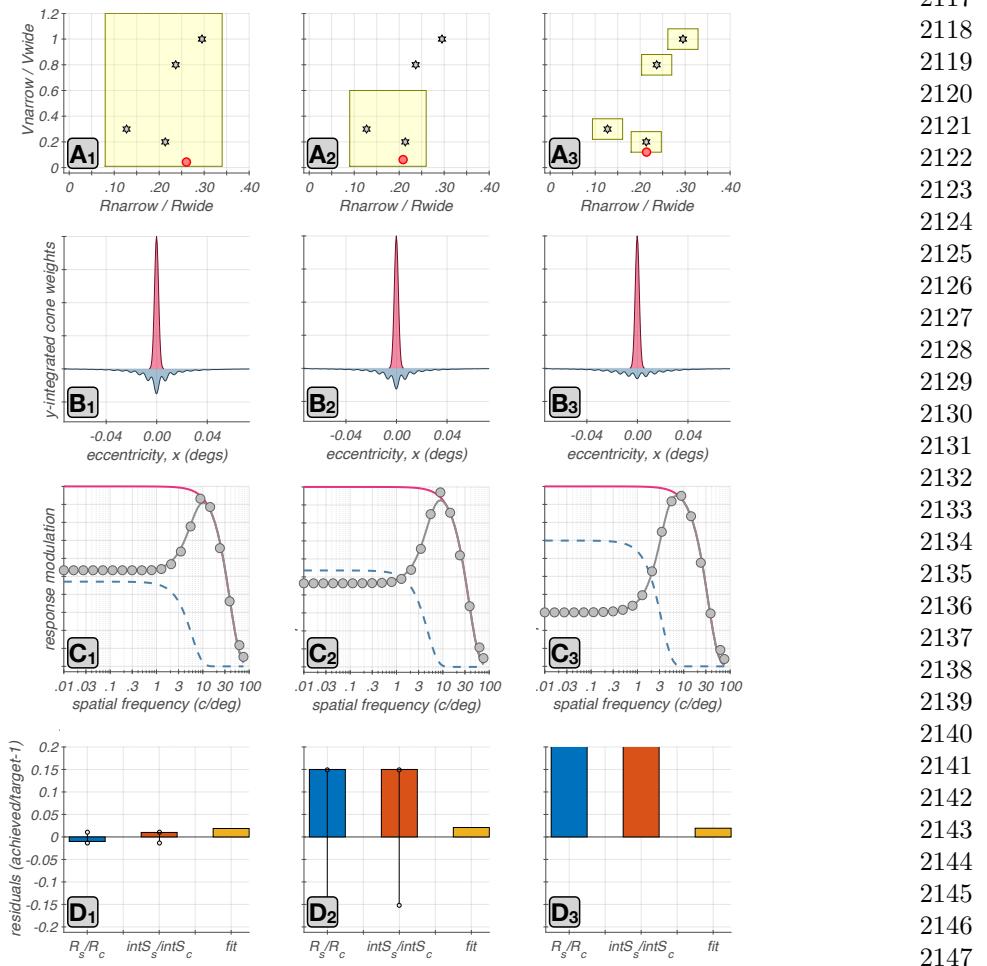


Fig. B1 Effect of constraints on surround cone pooling. Results from three options for constraining the surround optimization. Left column: tight tolerance in the parameters of the DoG model fit to the vSTF and loose tolerance in the parameters of the double exponential surround pooling model, $W_s(r)$. Middle column: medium tolerance in both sets of parameters. Right column: loose tolerance in the DoG parameters and tight tolerance in the $W_s(r)$ parameters. **A1-A3:** The yellow rectangles indicate the tolerance range in the joint space of the two surround cone pooling related parameters, $V_{\text{narrow}}/V_{\text{wide}}$ and $R_{\text{narrow}}/R_{\text{wide}}$. Stars depict the corresponding parameter values in four macaque H1 horizontal cells from the study of Packer & Dacey. The red disk depicts the achieved parameter values under each strategy for an example foveal synthetic mRGC. **B1-B3:** Line weighting functions of the retinal space referred center and surround cone pooling weights under the three examined strategies. **C1-C3:** The vSTF computed under the three strategies (gray disks) and corresponding DOG model fits (gray lines). The red and blue lines depict the center and surround components of the fitted DOG model. **D1-D3:** Blue and orange bars depict the residuals for the ratios of visual space-referred R_s/R_c and $K_s/K_c \times (R_s/R_c)^2$ ratios. Black circles connected by a black line depict the enforced tolerance range in these ratios. The enforced tolerance value in D3 was $\tau = 0.5$, and is not visualized. The orange bars depict the $\|\text{vSTF}(\omega) - \text{DOG}(\omega)\|$ residual.

2163 **References**

- 2164 [1] Wandell, B.A., Brainard, D.H., Cottaris, N.P.: Visual encoding: Principles and
2165 software. *Prog Brain Res* **273**(1), 199–229 (2022) <https://doi.org/10.1016/bs.pbr.2022.04.006>
- 2166 [2] Cottaris, N.P., Jiang, H., Ding, X., Wandell, B.A., Brainard, D.H.: A
2167 computational-observer model of spatial contrast sensitivity: Effects of wave-
2168 front-based optics, cone-mosaic structure, and inference engine. *J Vis* **19**(4), 8
2169 (2019) <https://doi.org/10.1167/19.4.8>
- 2170 [3] Cottaris, N.P., Wandell, B.A., Rieke, F., Brainard, D.H.: A computational
2171 observer model of spatial contrast sensitivity: Effects of photocurrent encod-
2172 ing, fixational eye movements, and inference engine. *J Vis* **20**(7), 17 (2020)
2173 <https://doi.org/10.1167/jov.20.7.17>
- 2174 [4] Lian, T., MacKenzie, K.J., Brainard, D.H., Cottaris, N.P., Wandell, B.A.: Ray
2175 tracing 3d spectral scenes through human optics models. *J Vis* **19**(12), 23 (2019)
2176 <https://doi.org/10.1167/19.12.23>
- 2177 [5] Nankivil, D., Cottaris, N.P., Brainard, D.H.: Theoretical impact of chromatic
2178 aberration correction on visual acuity. *Biomed Opt Express* **15**(5), 3265–3284
2179 (2024) <https://doi.org/10.1364/BOE.516049>
- 2180 [6] Zhang, L.Q., Cottaris, N.P., Brainard, D.H.: An image reconstruction framework
2181 for characterizing initial visual encoding. *eLife* **11** (2022) <https://doi.org/10.7554/eLife.71132>
- 2182 [7] Kupers, E.R., Carrasco, M., Winawer, J.: Modeling visual performance differences
2183 'around' the visual field: A computational observer approach. *PLoS Comput Biol*
2184 **15**(5), 1007063 (2019) <https://doi.org/10.1371/journal.pcbi.1007063>
- 2185 [8] Singh, V., Cottaris, N.P., Heasly, B.S., Brainard, D.H., Burge, J.: Computational
2186 luminance constancy from naturalistic images. *J Vis* **18**(13), 19 (2018) <https://doi.org/10.1167/18.13.19>
- 2187 [9] Ding, X., Radonjic, A., Cottaris, N.P., Jiang, H., Wandell, B.A., Brainard, D.H.:
2188 Computational-observer analysis of illumination discrimination. *J Vis* **19**(7), 11
2189 (2019)
- 2190 [10] Godat, T., Cottaris, N.P., Patterson, S., Kohout, K., Parkins, K., Yang, Q.,
2191 Strazzeri, J.M., McGregor, J.E., Brainard, D.H., Merigan, W.H., Williams, D.R.:
2192 In vivo chromatic and spatial tuning of foveolar retinal ganglion cells in macaca
2193 fascicularis. *PLoS One* **17**(11), 0278261 (2022) <https://doi.org/10.1371/journal.pone.0278261>
- 2194 [11] Osterberg, G., Andersen, H.: Topography of the Layer of Rods and Cones in
2195 2196 2197 2198 2199 2200 2201 2202 2203 2204 2205 2206 2207 2208

- the Human Retina / by G. Østerberg. A. Busck, Copenhagen (1935). Includes bibliographical references (pages 94-96). 2209
2210
2211
- [12] Polyak, S.L.: The Retina; the Anatomy and the Histology of the Retina in Man, Ape, and Monkey, Including the Consideration of Visual Functions, the History of Physiological Optics, and the Histological Laboratory Technique, p. 607. The University of Chicago press, Chicago, Ill., (1941) 2212
2213
2214
2215
2216
- [13] Dacey, D.M.: The mosaic of midget ganglion cells in the human retina. *J Neurosci* **13**(12), 5334–55 (1993) <https://doi.org/10.1523/JNEUROSCI.13-12-05334.1993> 2217
2218
- [14] Patterson, S.S., Neitz, M., Neitz, J.: Reconciling color vision models with midget ganglion cell receptive fields. *Front Neurosci* **13**, 865 (2019) <https://doi.org/10.3389/fnins.2019.00865> 2219
2220
2221
2222
- [15] Field, G.D., Gauthier, J.L., Sher, A., Greschner, M., Machado, T.A., Jepson, L.H., Shlens, J., Gunning, D.E., Mathieson, K., Dabrowski, W., Paninski, L., Litke, A.M., Chichilnisky, E.J.: Functional connectivity in the retina at the resolution of photoreceptors. *Nature* **467**(7316), 673–7 (2010) <https://doi.org/10.1038/nature09424> 2223
2224
2225
2226
2227
2228
- [16] Wool, L.E., Crook, J.D., Troy, J.B., Packer, O.S., Zaidi, Q., Dacey, D.M.: Non-selective wiring accounts for red-green opponency in midget ganglion cells of the primate retina. *J Neurosci* **38**(6), 1520–1540 (2018) <https://doi.org/10.1523/JNEUROSCI.1688-17.2017> 2229
2230
2231
2232
2233
- [17] Croner, L.J., Kaplan, E.: Receptive fields of p and m ganglion cells across the primate retina. *Vision Res* **35**(1), 7–24 (1995) [https://doi.org/10.1016/0042-6989\(94\)e0066-t](https://doi.org/10.1016/0042-6989(94)e0066-t) 2234
2235
2236
2237
- [18] Lee, B.B., Shapley, R.M., Hawken, M.J., Sun, H.: Spatial distributions of cone inputs to cells of the parvocellular pathway investigated with cone-isolating gratings. *J Opt Soc Am A Opt Image Sci Vis* **29**(2), 223–32 (2012) <https://doi.org/10.1364/JOSAA.29.00A223> 2238
2239
2240
2241
2242
- [19] Reid, R.C., Shapley, R.M.: Spatial structure of cone inputs to receptive fields in primate lateral geniculate nucleus. *Nature* **356**(6371), 716–8 (1992) <https://doi.org/10.1038/356716a0> 2243
2244
2245
2246
- [20] Chen, Q., Ingram, N.T., Baudin, J., Angueyra, J.M., Sinha, R., Rieke, F.: Predictably manipulating photoreceptor light responses to reveal their role in downstream visual responses. *eLife* **13** (2024) <https://doi.org/10.7554/eLife.93795> 2247
2248
2249
2250
- [21] Freeman, J., Field, G.D., Li, P.H., Greschner, M., Gunning, D.E., Mathieson, K., Sher, A., Litke, A.M., Paninski, L., Simoncelli, E.P., Chichilnisky, E.J.: Mapping nonlinear receptive field structure in primate retina at single cone resolution. *eLife* 2251
2252
2253
2254

- 2255 4 (2015) <https://doi.org/10.7554/eLife.05241>
- 2256
- 2257 [22] Hong, A., Rieke, F.: On-off asymmetries of nonlinear spatial integration impact
2258 the encoding of natural stimuli in primate retina. *Investigative Ophthalmology
2259 Visual Science* **65**(7), 4346–4346 (2024)
- 2260
- 2261 [23] Raghavan, R.T., Kelly, J.G., Hasse, J.M., Levy, P.G., Hawken, M.J., Movshon,
2262 J.A.: Contrast and luminance gain control in the macaque's lateral geniculate
2263 nucleus. *eNeuro* **10**(3) (2023) <https://doi.org/10.1523/ENEURO.0515-22.2023>
- 2264 [24] Turner, M.H., Rieke, F.: Synaptic rectification controls nonlinear spatial integra-
2265 tion of natural visual inputs. *Neuron* **90**(6), 1257–1271 (2016) <https://doi.org/10.1016/j.neuron.2016.05.006>
- 2266
- 2267 [25] Benardete, E.A., Kaplan, E.: The receptive field of the primate p retinal ganglion
2268 cell, i: Linear dynamics. *Vis Neurosci* **14**(1), 169–85 (1997) <https://doi.org/10.1017/s0952523800008853>
- 2269
- 2270 [26] Croner, L.J., Purpura, K., Kaplan, E.: Response variability in retinal ganglion
2271 cells of primates. *Proc Natl Acad Sci U S A* **90**(17), 8128–30 (1993) <https://doi.org/10.1073/pnas.90.17.8128>
- 2272
- 2273 [27] Kling, A., Cooler, S., Manookin, M.B., Rhoades, C., Brackbill, N., Field, G.,
2274 Rieke, F., Sher, A., Litke, A., Chichilnisky, E.J.: Functional diversity in the output
2275 of the primate retina. *bioRxiv* (2024) <https://doi.org/10.1101/2024.10.31.621339>
- 2276
- 2277 [28] Wohrer, A., Kornprobst, P.: Virtual retina: a biological retina model and sim-
2278 ulator, with contrast gain control. *J Comput Neurosci* **26**(2), 219–49 (2009)
2279 <https://doi.org/10.1007/s10827-008-0108-4>
- 2280
- 2281 [29] Ly, K., Italiano, M.L., Shivdasani, M.N., Tsai, D., Zhang, J.Y., Jiang, C., Lovell,
2282 N.H., Dokos, S., Guo, T.: Virtual human retina: Simulating neural signalling,
2283 degeneration, and responses to electrical stimulation. *Brain Stimul* **18**(1), 144–163
2284 (2025) <https://doi.org/10.1016/j.brs.2025.01.013>
- 2285
- 2286 [30] Somaratna, M.A., Freeman, A.W.: The receptive field construction of midget
2287 ganglion cells in primate retina. *J Neurophysiol* **133**(1), 268–285 (2025) <https://doi.org/10.1152/jn.00302.2024>
- 2288
- 2289 [31] Cottaris, N., Wandell, B., Brainard, D.: Midget retinal ganglion cell surrounds
2290 in macaque: cone-selective or not? *Journal of Vision* **24**(10), 1090–1090 (2024)
2291 <https://doi.org/10.1167/jov.24.10.1090>
- 2292
- 2293 [32] Hennig, M.H., Funke, K., Worgotter, F.: The influence of different retinal sub-
2294 circuits on the nonlinearity of ganglion cell behavior. *J Neurosci* **22**(19), 8726–38
2295 (2002) <https://doi.org/10.1523/JNEUROSCI.22-19-08726.2002>
- 2296
- 2297
- 2298
- 2299
- 2300

- [33] Pillow, J.W., Paninski, L., Uzzell, V.J., Simoncelli, E.P., Chichilnisky, E.J.: Prediction and decoding of retinal ganglion cell responses with a probabilistic spiking model. *J Neurosci* **25**(47), 11003–13 (2005) <https://doi.org/10.1523/JNEUROSCI.3305-05.2005> 2301
2302
2303
2304
2305
- [34] Watson, A.B.: A formula for human retinal ganglion cell receptive field density as a function of visual field location. *J Vis* **14**(7) (2014) <https://doi.org/10.1167/14.7.15> 2306
2307
2308
2309
- [35] Curcio, C.A., Allen, K.A.: Topography of ganglion cells in human retina. *J Comp Neurol* **300**(1), 5–25 (1990) <https://doi.org/10.1002/cne.903000103> 2310
2311
- [36] Curcio, C.A., Sloan, K.R., Kalina, R.E., Hendrickson, A.E.: Human photoreceptor topography. *J Comp Neurol* **292**(4), 497–523 (1990) <https://doi.org/10.1002/cne.902920402> 2312
2313
2314
2315
- [37] Klug, K., Herr, S., Ngo, I.T., Sterling, P., Schein, S.: Macaque retina contains an s-cone off midget pathway. *J Neurosci* **23**(30), 9881–7 (2003) <https://doi.org/10.1523/JNEUROSCI.23-30-09881.2003> 2316
2317
2318
2319
- [38] Patterson, S.S., Kuchenbecker, J.A., Anderson, J.R., Bordt, A.S., Marshak, D.W., Neitz, M., Neitz, J.: An s-cone circuit for edge detection in the primate retina. *Sci Rep* **9**(1), 11913 (2019) <https://doi.org/10.1038/s41598-019-48042-2> 2320
2321
2322
2323
- [39] Gauthier, J.L., Field, G.D., Sher, A., Shlens, J., Greschner, M., Litke, A.M., Chichilnisky, E.J.: Uniform signal redundancy of parasol and midget ganglion cells in primate retina. *J Neurosci* **29**(14), 4675–80 (2009) <https://doi.org/10.1523/JNEUROSCI.5294-08.2009> 2324
2325
2326
2327
2328
- [40] Kling, A., Gogliettino, A.R., Shah, N.P., Wu, E.G., Brackbill, N., Sher, A., Litke, A.M., Silva, R.A., Chichilnisky, E.J.: Functional organization of midget and parasol ganglion cells in the human retina (2020) <https://doi.org/10.1101/2020.08.07.240762> 2329
2330
2331
2332
2333
- [41] Kolb, H., Marshak, D.: The midget pathways of the primate retina. *Doc Ophthalmol* **106**(1), 67–81 (2003) <https://doi.org/10.1023/a:1022469002511> 2334
2335
- [42] McMahon, M.J., Lankheet, M.J., Lennie, P., Williams, D.R.: Fine structure of parvocellular receptive fields in the primate fovea revealed by laser interferometry. *J Neurosci* **20**(5), 2043–53 (2000) <https://doi.org/10.1523/JNEUROSCI.20-05-02043.2000> 2336
2337
2338
2339
2340
- [43] Packer, O.S., Dacey, D.M.: Receptive field structure of h1 horizontal cells in macaque monkey retina. *J Vis* **2**(4), 272–92 (2002) <https://doi.org/10.1167/2.4.1> 2341
2342
2343
- [44] Smith, V.C., Lee, B.B., Pokorny, J., Martin, P.R., Valberg, A.: Responses 2344
2345
2346

- 2347 of macaque ganglion cells to the relative phase of heterochromatically modu-
2348 lated lights. *J Physiol* **458**, 191–221 (1992) <https://doi.org/10.1113/jphysiol.1992.sp019413>
- 2349
2350
- 2351 [45] Polans, J., Jaeken, B., McNabb, R.P., Artal, P., Izatt, J.A.: Wide-field optical
2352 model of the human eye with asymmetrically tilted and decentered lens that
2353 reproduces measured ocular aberrations. *Optica* **2**(2), 124–134 (2015) <https://doi.org/10.1364/OPTICA.2.000124>
- 2354
2355
- 2356 [46] Tuten, W.S., Cooper, R.F., Tiruveedhula, P., Dubra, A., Roorda, A., Cottaris,
2357 N.P., Brainard, D.H., Morgan, J.I.W.: Spatial summation in the human fovea:
2358 Do normal optical aberrations and fixational eye movements have an effect? *J Vis*
2359 **18**(8), 6 (2018) <https://doi.org/10.1167/18.8.6>
- 2360
2361
- 2362 [47] Schein, S.J.: Anatomy of macaque fovea and spatial densities of neurons in foveal
2363 representation. *J Comp Neurol* **269**(4), 479–505 (1988) <https://doi.org/10.1002/cne.902690403>
- 2364
2365
- 2366 [48] Packer, O., Hendrickson, A.E., Curcio, C.A.: Photoreceptor topography of the
2367 retina in the adult pigtail macaque (macaca nemestrina). *J Comp Neurol* **288**(1),
2368 165–83 (1989) <https://doi.org/10.1002/cne.902880113>
- 2369
2370
- 2371 [49] Gogliettino, A.R., Madugula, S.S., Grosberg, L.E., Vilkhu, R.S., Brown, J.,
2372 Nguyen, H., Kling, A., Hottowy, P., Dabrowski, W., Sher, A., Litke, A.M.,
2373 Chichilnisky, E.J.: High-fidelity reproduction of visual signals by electrical stim-
2374 ulation in the central primate retina. *J Neurosci* **43**(25), 4625–4641 (2023)
2375 <https://doi.org/10.1523/JNEUROSCI.1091-22.2023>
- 2376
2377
- 2378 [50] Grunert, U., Martin, P.R.: Cell types and cell circuits in human and non-human
2379 primate retina. *Prog Retin Eye Res*, 100844 (2020) <https://doi.org/10.1016/j.preteyeres.2020.100844>
- 2380
2381
- 2382 [51] Banks, M.S., Geisler, W.S., Bennett, P.J.: The physical limits of grating visi-
2383 bility. *Vision Res* **27**(11), 1915–24 (1987) [https://doi.org/10.1016/0042-6989\(87\)90057-5](https://doi.org/10.1016/0042-6989(87)90057-5)
- 2384
2385
- 2386 [52] Geisler, W.S.: Sequential ideal-observer analysis of visual discriminations. *Psychol
2387 Rev* **96**(2), 267–314 (1989) <https://doi.org/10.1037/0033-295x.96.2.267>
- 2388
2389
- 2390 [53] Sekiguchi, N., Williams, D.R., Brainard, D.H.: Efficiency in detection of isolumi-
2391 nant and isochromatic interference fringes. *J Opt Soc Am A Opt Image Sci Vis*
2392 **10**(10), 2118–33 (1993) <https://doi.org/10.1364/josaa.10.002118>
- 2393
2394
- 2395 [54] Casile, A., Victor, J.D., Rucci, M.: Contrast sensitivity reveals an oculomotor
2396 strategy for temporally encoding space. *Elife* **8** (2019) <https://doi.org/10.7554/eLife.40924>
- 2397
2398
- 2399
2400

- [55] Houston, A.J.H., Brainard, D.H., Smithson, H.E., Read, D.J.: Information, movement and adaptation in human vision. bioRxiv, 2025–0905674315 (2025) <https://doi.org/10.1101/2025.09.05.674315> 2393
2394
2395
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2397
2398
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2425
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2428
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2431
2432
2433
2434
2435
2436
2437
2438
- [56] Mullen, K.T.: The contrast sensitivity of human colour vision to red-green and blue-yellow chromatic gratings. *J Physiol* **359**, 381–400 (1985) <https://doi.org/10.1113/jphysiol.1985.sp015591> 2393
2394
2395
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2397
2398
2399
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2425
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2428
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2430
2431
2432
2433
2434
2435
2436
2437
2438
- [57] Chaparro, A., Stromeier, r. C. F., Huang, E.P., Kronauer, R.E., Eskew, J. R. T.: Colour is what the eye sees best. *Nature* **361**(6410), 348–50 (1993) <https://doi.org/10.1038/361348a0> 2400
2401
2402
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2405
2406
2407
2408
2409
2410
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2425
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2427
2428
2429
2430
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2433
2434
2435
2436
2437
2438
- [58] Sekiguchi, N., Williams, D.R., Brainard, D.H.: Aberration-free measurements of the visibility of isoluminant gratings. *J Opt Soc Am A Opt Image Sci Vis* **10**(10), 2105–17 (1993) <https://doi.org/10.1364/josaa.10.002105> 2400
2401
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2404
2405
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2409
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2425
2426
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2428
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2435
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2437
2438
- [59] Hong, F., Cottaris, N.P., Brainard, D.H.: Limits on human contrast sensitivity imposed by the initial visual encoding. *Journal of Vision* **24**(10), 658–658 (2024) <https://doi.org/10.1167/jov.24.10.658> 2400
2401
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2425
2426
2427
2428
2429
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2437
2438
- [60] Cruz, Y.T., He, J., Eskew, J. Rhea T.: Using s-cone signals to detect the effects of longitudinal chromatic aberration. *Journal of Vision* **19**(15), 6–6 (2019) <https://doi.org/10.1167/19.15.6> 2400
2401
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2438
- [61] Oh, S., Hong, F., Nankivil, D., Haggerty, E.B., Stauble, R., Cottaris, N.P., Buch, J., Aguirre, G.K., Brainard, D.H.: Variations in human optics explain idiosyncratic patterns in the red-green spatial contrast sensitivity function. *Journal of Vision* **25**(9), 2440–2440 (2025) <https://doi.org/10.1167/jov.25.9.2440> 2400
2401
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2428
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2438
- [62] Hansen, T., Pracejus, L., Gegenfurtner, K.R.: Color perception in the intermediate periphery of the visual field. *J Vis* **9**(4), 26–112 (2009) <https://doi.org/10.1167/9.4.26> 2400
2401
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2438
- [63] Stockman, A., Brainard, D.H.: In: Bass, M., DeCusatis, C., Enoch, J., Lakshminarayanan, V., Li, G., Macdonald, C., Mahajan, V., Stryland, E. (eds.) *Color vision mechanisms*, pp. 11–111104. McGraw Hill, New York (2010) 2400
2401
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2410
2411
2412
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2425
2426
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2428
2429
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2431
2432
2433
2434
2435
2436
2437
2438
- [64] Lennie, P., Haake, P.W., Williams, D.R.: In: Landy, M.S., Movshon, J.A. (eds.) *The design of chromatically opponent receptive fields*, pp. 71–82. MIT Press, Cambridge, Mass. (1991). TY - JOUR 2400
2401
2402
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2434
2435
2436
2437
2438
- [65] Mullen, K.T., Kingdom, F.A.: Losses in peripheral colour sensitivity predicted from "hit and miss" post-receptoral cone connections. *Vision Res* **36**(13), 1995–2000 (1996) [https://doi.org/10.1016/0042-6989\(95\)00261-8](https://doi.org/10.1016/0042-6989(95)00261-8) 2400
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- [66] Wandell, B.A.: Foundations of Vision. Sinauer, Sunderland, MA (1995) 2400
2401
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2436
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- 2439 [67] Kim, Y.J., Packer, O., Pollreisz, A., Martin, P.R., Grunert, U., Dacey, D.M.:
2440 Comparative connectomics reveals noncanonical wiring for color vision in human
2441 foveal retina. *Proc Natl Acad Sci U S A* **120**(18), 2300545120 (2023) <https://doi.org/10.1073/pnas.2300545120>
- 2442
- 2443 [68] Sun, H., Ruttiger, L., Lee, B.B.: The spatiotemporal precision of ganglion cell sig-
2444 nals: a comparison of physiological and psychophysical performance with moving
2445 gratings. *Vision Res* **44**(1), 19–33 (2004) <https://doi.org/10.1016/j.visres.2003.08.017>
- 2446
- 2447 [69] Angueyra, J.M., Rieke, F.: Origin and effect of phototransduction noise in primate
2448 cone photoreceptors. *Nat Neurosci* **16**(11), 1692–700 (2013) <https://doi.org/10.1038/nn.3534>
- 2449
- 2450 [70] Roy, S., Jun, N.Y., Davis, E.L., Pearson, J., Field, G.D.: Inter-mosaic coordination
2451 of retinal receptive fields. *Nature* **592**(7854), 409–413 (2021) <https://doi.org/10.1038/s41586-021-03317-5>
- 2452
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