riojaPlot: User Guide (Version 0.1-20)

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Contents

1. Introduction	1
2. Basic diagram	2
3. Selecting variables to plot	6
4. Plotting a secondary y-axis	7
5. Showing groups	8
6. Add a zonation	9
	12
	13
0 1 0	17
10. Adding a column showing lithology	18
11. Frequently asked questions	19
	19
	22
3. How do I change the limits and labels of the of the y-axes?	22
	22
1 1	22
v v	23
1	24
\checkmark	26
	27
1 0	28
0 1	29
1	31
	32
14. How do I plot ecological time series?	33
v e	36
16. How do I include special symbols in the variable names	38
\mathbf{y}	39
18. Can I use a pipe with riojaPlot	41
	41
	42
12. References	46

1. Introduction

riojaPlot plots a set of variables in a stratigraphic diagram. Diagrams can be plotted with lines, silhouettes, bars or symbols of any combination of these element. riojaPlot extends the older function strat.plot in several ways: variables can be grouped and displayed with different colours, a cumulative summary plot can be added automatically based on the grouping, a cluster dendrogram and resulting zones can be added automatically, a secondary y-axis can be added to plot both depth and age axes, and the figures margins are

determined automatically depending on the size and length of labels. Arguments to riojaPlot that control the appearance of the plot are know as **styles**. Styles can specified on a plot-by-plot basis or can be saved and applied to multiple figures.

Please send questions, bug reports and suggestions for improvements and additions to Stephen.Juggins@ncl. ac.uk.

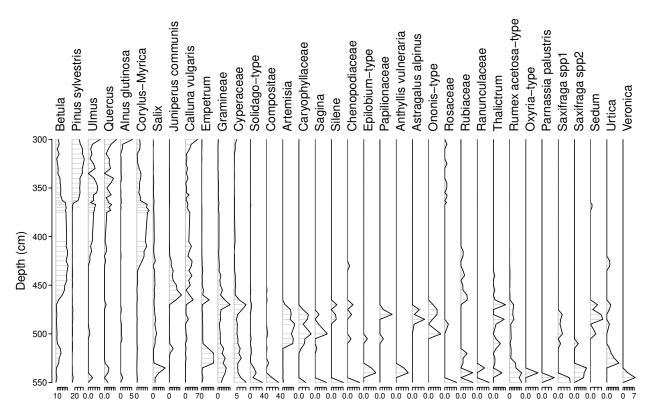
2. Basic diagram

To create a basic stratigraphic diagram just pass riojaPlot a data frame of variables to plot and and a data frame with at least one column containing the y-axis variable (usually sample depth or age). If the y-axis variable name is not specified riojaPlot will use the first column and label the y-axis with the column name. The default plot is shown with lines and bars for non-percentage data and as filled silhouettes (filled curves) and bars for percentage data (style scale.percent=TRUE. Turn silhouettes off and create plots with lines, bars or symbols by setting styles plot.poly, plot.line, plot.bars or plot.symb TRUE or FALSE (all may be overlain).

```
library(riojaPlot)
library(rioja)

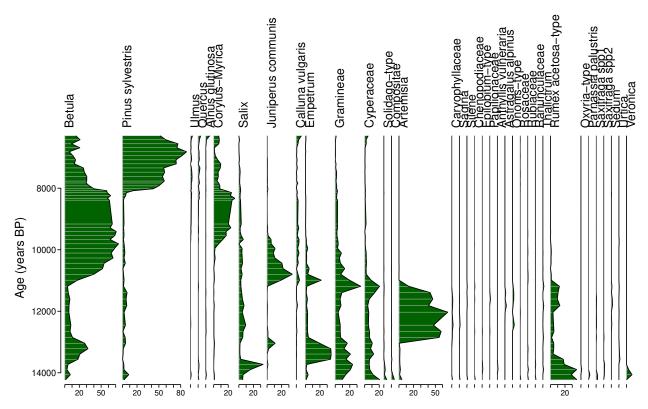
# use built-in data from Abernethy Forest
# see ?rioja::aber for citation
data(aber)
# extract pollen percentages
aber.poll <- aber$spec
# replace species codes with full taxon names
colnames(aber.poll) <- aber$names$Name
aber.chron <- aber$ages

# plot on depth scale (depth is the first column in chron)
riojaPlot(aber.poll, aber.chron)</pre>
```



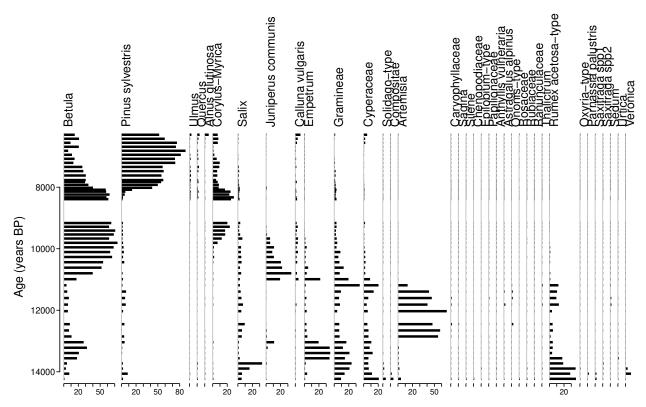
These are percentage data so we scale the diagram for percentages and plot on an age scale by passing the name of the appropriate column to style yvar.name. riojaPlot will take the y-axis from the first column of aber.chron by default.

```
# scale for percentage data
riojaPlot(aber.poll, aber.chron,
    yvar.name="Age (years BP)",
    scale.percent=TRUE)
```



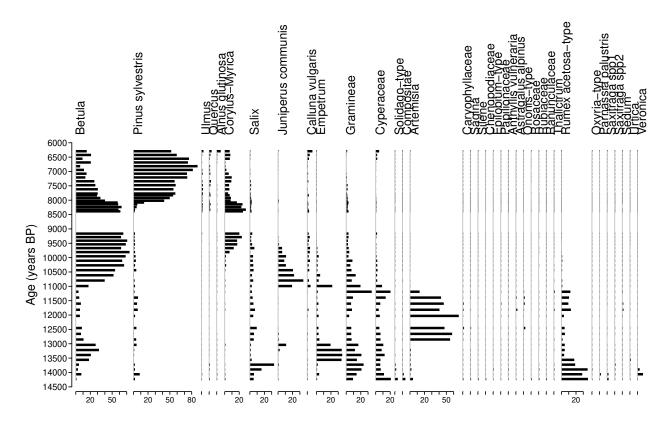
Or plot as bars.

```
# turn of silhouettes and lines, increase bar thickness
riojaPlot(aber.poll, aber.chron,
    yvar.name="Age (years BP)",
    scale.percent=TRUE,
    plot.poly=FALSE,
    plot.line=FALSE,
    lwd.bar=3,
    col.bar="black")
```



The y-axis doesn't cover the whole range so we set the upper and lower limits manually.

```
# Specify y-axis limits
sapply(aber.chron, range)
        Depth (cm) Age (14C years BP) Age (years BP)
#>
#> [1,]
               300
                                  5515
                                                 6295
#> [2,]
               550
                                 12147
                                                14229
riojaPlot(aber.poll, aber.chron,
   yvar.name="Age (years BP)",
   ymin=6000, ymax=14500, yinterval=500,
   scale.percent=TRUE,
   plot.poly=FALSE,
  plot.line=FALSE,
   lwd.bar=3,
   col.bar="black")
```

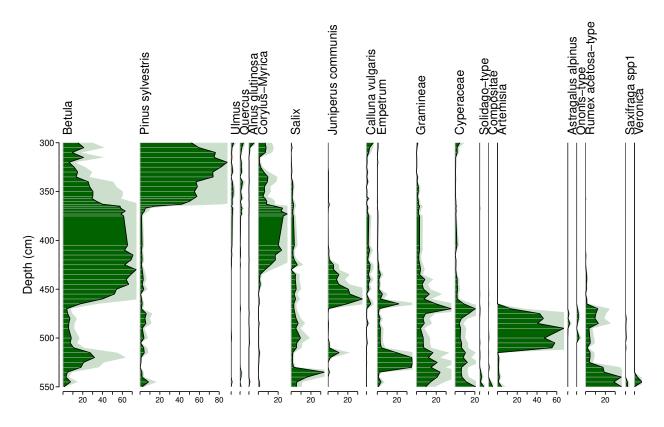


3. Selecting variables to plot

By default riojaPlot will plot all columns in the data frame. In this example there are many rare taxa: we could either remove these from the data frane prior to plotting or pass a character vector of column names to plot to the argument selVars. Here we only plot taxa with max abundance greater than 5%. We also add exaggeration curves. These are shown as silhouettes with a multiplication factor of 2 by default by can be changed to lines with different exaggeration using styles exag.mult, col.exag, and col.line.exag. col.exag="auto" will use a transparent colour the same as the main curve.Useexag.alpha' to change transparency.

```
# reduce number of taxa and add exaggerations
# make a vector of taxon names with max abundance greater that 2 percent
mx <- apply(aber.poll, 2, max)
aber.sel <- names(mx[mx > 2])

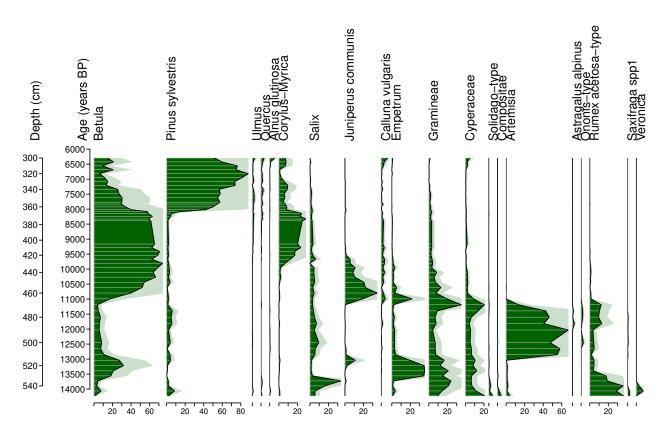
riojaPlot(aber.poll, aber.chron, aber.sel,
    scale.percent=TRUE,
    plot.exag=TRUE)
```



4. Plotting a secondary y-axis

To show a secondary y-axis simply pass the column name of the secondary y-axis variable to style sec.yvar.name. In this example we use age (years BP) for the primary axis and depth for the secondary axis and set plot.sec.axis to TRUE. We also use ymin, ymax and yinterval to fine-tune the start, end, and increment of the y-axis.

```
riojaPlot(aber.poll, aber.chron, aber.sel,
   yvar.name="Age (years BP)",
   sec.yvar.name="Depth (cm)",
   plot.sec.axis=TRUE,
   ymin=6000,
   ymax=14300,
   yinterval=500,
   scale.percent=TRUE,
   plot.exag=TRUE)
```



5. Showing groups

Individual can be assigned to a group and each group assigned a different colour. Cumulative plots can also be added showing the sum of each group.

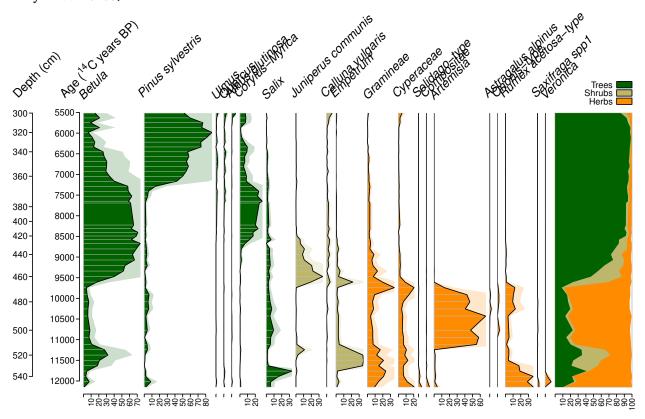
To assign variables to groups pass a data frame of two columns, the first with the name of each variable (spelt exactly the same as in the data) and a column of group memberships. This should be a factor with the levels determining the order they appear in the legend. The names of the groups will be used in the cumulative plot legend so choose them wisely.

We can also italicise the names using the style labels.italicise, rotate them with srt.xlabel, and specify a label for the age variable to override the column name. Here we plot a 14C age scale and use an expression to create a label with superscriped 14C.

```
# group taxa by type and add cumulative graph
# extract types
aber.types <- aber$names[, -1]
# convert pollen types to a factor
aber.types$Group <- factor(aber.types$Group, levels=c("Trees", "Shrubs", "Herbs"))
ylab <- expression(Age~"("*""^{14}*C~years~BP*")")

riojaPlot(aber.poll, aber.chron, aber.sel, aber.types,
   yvar.name="Age (14C years BP)",
   sec.yvar.name="Depth (cm)",
   ylabel=ylab,
   plot.sec.axis=TRUE,
   ymin=5500,
   ymax=12200,
   yinterval=500,
   scale.percent=TRUE,</pre>
```

```
plot.groups=TRUE,
plot.cumul=TRUE,
plot.exag=TRUE,
labels.italicise = TRUE,
srt.xlabel=45,
las.xaxis=2,
yBottom=0.05)
```



6. Add a zonation

We can add a zonation to the plot in 3 ways. The first is to set the styles do.clust and plot.clust and / or plot.zones to "auto" or the number of zones to show. This will perform a constrained clustering using chclust and either add a dendrogram and/or zone lines to the diagram. The number of zones can be estimated using a broken stick model (plot.zones="auto" see ?rioja::chclust for details) or specified (e.g. plot.zones=3). Colour of the zone lines can be changed with col.zones. The zonation can be based on all columns in the data (style clust.use.selected=FALSE) or only the displayed columns if only a subset are selected using selVars. Data can optionally be transformed (clust.data.trans="sqrt"), scaled (clust.data.trans="scale") prior to clustering or used as is (clust.data.trans="none", the default). Change the width of the dendrogram using style clust.width=N where N is the required width as a fraction of the whole diagram width.

```
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types,
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500,
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
```

```
plot.groups=TRUE,
    plot.cumul=TRUE,
    plot.exag=TRUE,
    do.clust=TRUE,
    clust.data.trans="sqrt",
    plot.clust=TRUE,
    plot.zones="auto",
    labels.italicise = TRUE)
                                                           Juniperus communis
                            Pinus sylvestris
           Age (years BP)
                                                                                Cyperaceae
                                                                          Gramineae
 Depth (cm)
                                                                                                                       Trees Shrubs Herbs
         6000-
300-
         6500-
320
         7000
340
         7500
360
         8000
380
         8500-
         9000-
400
         9500
420
        10000-
440
        10500-
460
        11000
        11500
480
        12000
500-
        12500
        13000
520
        13500
540-
```

We can also generate a constrained cluster object and pass this to riojaPlot for plotting. Here we log10transform the data prior to clustering.

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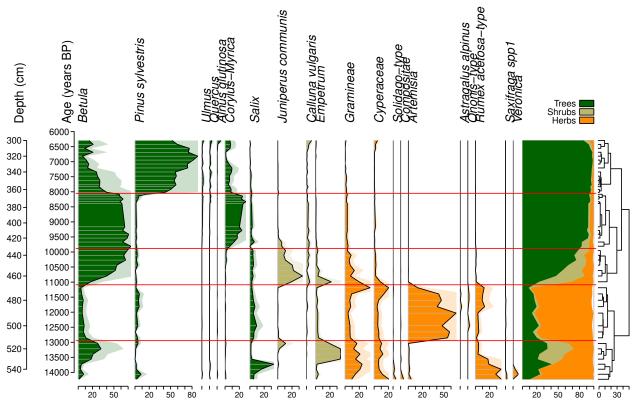
```
aber.clust <- chclust(dist(log10(aber.poll+1)))</pre>
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types, clust=aber.clust,
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
  ymax=14300,
  yinterval=500,
  plot.sec.axis=TRUE,
   scale.percent=TRUE,
  plot.groups=TRUE,
  plot.cumul=TRUE,
  plot.exag=TRUE,
   clust.data.trans="sqrt",
  plot.clust=TRUE,
  plot.zones="auto",
   labels.italicise = TRUE)
```

20

20

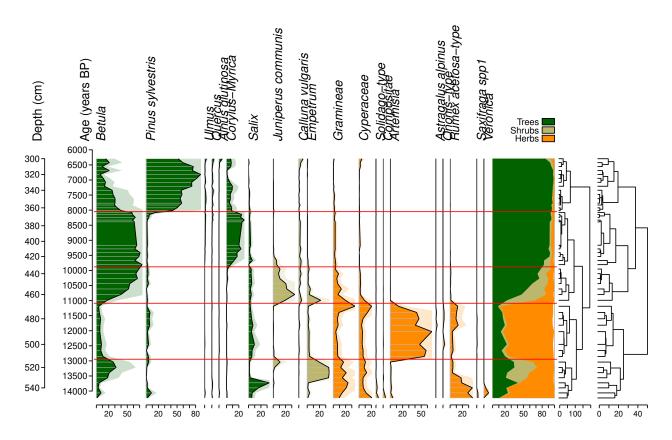
20

14000



Finally, we can add a zonation at a different position in the diagram. Here we add the zonation based on log10-transformed data to the original diagram with a sqrt-transformed zonation for comparison. In this case we set the right-hand limit of the original diagram with xRight and add the additional zonation to this space. This is a bit of a contrived example but adding the zonation separately to the diagram can be useful if we are plotting multiple datasets and want the zonation on the far-right (see example in section 8).

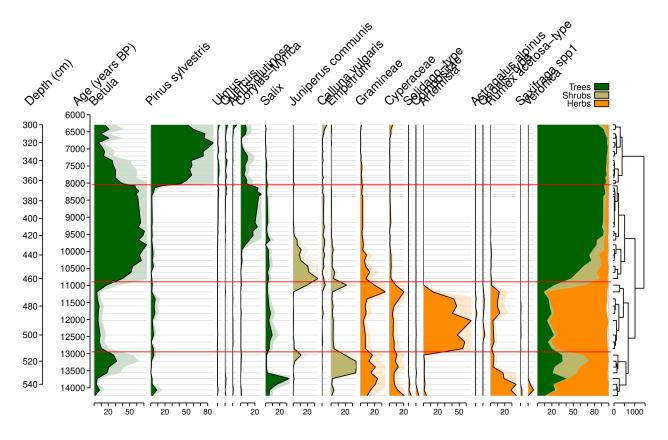
```
aber.clust <- chclust(dist(log10(aber.poll+1)))</pre>
rp <- riojaPlot(aber.poll, aber.chron, aber.sel, aber.types,</pre>
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500,
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
   plot.groups=TRUE,
   plot.cumul=TRUE,
   plot.exag=TRUE,
   clust.data.trans="sqrt",
   do.clust=TRUE,
   plot.clust=TRUE,
   plot.zones="auto",
   labels.italicise = TRUE,
   xRight = 0.9)
addRPClust(rp, aber.clust, xLeft=0.91, xRight=0.99)
```



7. Reusing styles

Save settings as a style and apply to figure. Here we also full-width plot bars behind the curves.

```
mystyle <- makeStyles(</pre>
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
   plot.groups=TRUE,
   plot.cumul=TRUE,
   plot.exag=TRUE,
   plot.poly=TRUE,
   plot.bar="full",
   bar.back=TRUE,
   lwd.bar=0.5,
   col.bar="lightgrey",
   do.clust=TRUE,
   plot.clust=TRUE,
   plot.zones="auto",
   srt.xlabel=45)
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types, mystyle,
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500)
```



8. Combining different datasets in a single diagram

We may want to plot different datasets in the same figure. For example, we may want to plot one set of variables with percentage scaling and the other with normal scaling where each variable is the same width. Or we may have two or more datasets measured at different depths of ages.

To combine different datasets we plot the first with function riojaPlot and the others with function riojaPlot2. We also need to specify the right-hand position of each plot in fractions of the page width.

```
# Diatom data the Round Loch of Glenhead, Galloway, SW Scotland
# Shows recent acidification and small recovery
data(RLGH)
RLGH.names <- RLGH$names
RLGH.diat <- RLGH$spec
# plot only common taxa
mx <- apply(RLGH.diat, 2, max)</pre>
RLGH.sel <- colnames(RLGH.diat)[mx > 2]
RLGH.chron <- RLGH$depths
# data has 210Pb age based on years before coring (1980)
# Add new column with Years (CE)
RLGH.chron$Year <- 1980 - RLGH.chron$Age
# do a pH reconstruction using SWAP modern dataset (see ?rioja::SWAP)
data(SWAP)
# generate a WA model
SWAP.wa <- WA(SWAP$spec, SWAP$pH)
RLGH.pH <- predict(SWAP.wa, RLGH$spec)</pre>
```

convert to data frame with single column

```
RLGH.pH <- data.frame(`DI-pH (SWAP)`=RLGH.pH$fit[, 1], check.names=FALSE)
rp <- riojaPlot(RLGH.diat, RLGH.chron,</pre>
                     yvar.name="Year",
                      scale.percent=TRUE,
                      y.rev=FALSE,
                      selVars=RLGH.sel,
                      ymax=1980,
                     xlabels=RLGH$names$TaxonName,
                      cex.xlabel=0.7,
                      labels.break.n=25,
                      xRight=0.9)
riojaPlot(RLGH.pH, RLGH.chron[, "Year", drop=FALSE],
                       riojaPlot=rp,
                       scale.minmax=FALSE,
                       plot.bar=FALSE,
                       plot.symb=TRUE,
                        symb.cex=0.6)
                                                                                                           Frustulia rhomboides var.
viridula
              Achnanthes minutissima
                                                                                                jlia rhomboides var
                   Achnanthes marginulata
                                                                                                                                                                  Tabellaria quadriseptata
                                                                                      Eunotia vanheurckii var.
                                                                                           ria virescens var.
                       Aulacoseira perglabra
                                                                                                                     Navicula madumensis
                                        Brachysira brebissoni
                                                  Eunotia pectinalis var
                                                                                                                                 Navicula cumbriensis
                                                                                                                          Navicula leptostriata
                                                                                                                                                        Tabellaria flocculosa
                                                     minor
Eunotia rhomboidea
                                                                                                              viridula
Navicula mediocris
                                                                                                                                           Nitzschia perminuta
                                                            Eunotia denticulata
                                                                                                                                                             Tabellaria binalis
                            Brachysira vitrea
                                             Cymbella lunata
                                                                                                                                      Navicula hoefleri
                                                                              Eunotia naegelii
                                                                                                                                                                                DI-pH (SWAP)
                                                                Eunotia incisa
                                                                                                                                                Peronia fibula
     1980
     1960
     1940
     1920
Year
     1900
     1880
     1860
```

Here we add a zonation to the right of the above figure. There are only two significant zones in these data.

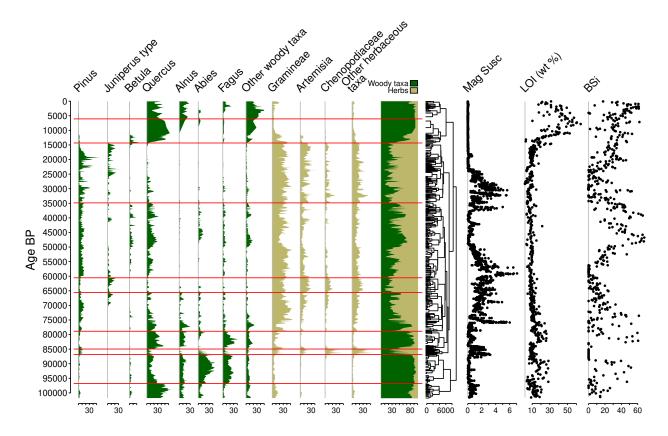
```
labels.break.n=25,
                         xRight=0.8)
rp2 <- riojaPlot(RLGH.pH, RLGH.chron[, "Year", drop=FALSE],</pre>
                           riojaPlot=rp,
                           xRight=0.9,
                           scale.minmax=FALSE,
                           plot.bar=FALSE,
                           plot.symb=TRUE,
                           symb.cex=0.6)
rp3 <- addRPClustZone(rp2, RLGH.clust, col="blue")</pre>
addRPClust(rp3, RLGH.clust)
                                                                                                   gua
Istulia rhomboides var
                                                                                                               ustulia rhomboides var
                Achnanthes minutissima
                     Achnanthes marginulata
                                                                                                                                                                       Tabellaria quadriseptata
                                                                                          Eunotia vanheurckii var.
                          Aulacoseira perglabra
                                           Brachysira brebissoni
                                                                                                                         Vavicula madumensis
                                                     Eunotia pectinalis var
                                                                                                                                      Vavicula cumbriensis
                                                                                                                              Vavicula leptostriata
                                                                                                                                                             Tabellaria flocculosa
                                                                                                                                                Vitzschia perminuta
                                                               Eunotia denticulata
                                                                                                                                                                   Tabellaria binalis
                               Brachysira vitrea
                                                                                                                                          Vavicula hoefleri
                                                Symbella lunata
                                                                                 Eunotia naegelii
                                                                                                                                                                                     DI-pH (SWAP)
                                                                    Eunotia incisa
                                                                                                                                                     Peronia fibula
      1980
       1960
      1940
      1920
Year
      1900
      1880
      1860
```

In the next example we combine pollen, magnetic susceptibility, LOI and biogenic silica data recorded in a core from Lago Grande di Monticchio and published by Allen et al. (1999). The core spans the last c. 100 kyr and the four datasets are measured at different depth intervals and resolutions. The data are downloaded from the NOAA paleoclimatology data archive (https://www.ncei.noaa.gov/products/paleoclimatology).

5.0

```
library(readxl)
library(dplyr)
# Import the data, assign pollen types to groups
fpath <- system.file("extdata/allen1999.xlsx", package="riojaPlot")
pollen <- read_excel(fpath, sheet="Pollen data", skip=2)
pollen.chron <- pollen %>% select(1)
pollen <- pollen %>% select(Pinus: Other herbaceous taxa`)
types <- data.frame(Name=colnames(pollen), Group="Woody taxa")
types$Group[9:12] <- "Herbs"
types$Group <- factor(types$Group, levels=c("Woody taxa", "Herbs"))
mag <- read_excel(fpath, sheet="Magnetic susceptibility", skip=2)</pre>
```

```
mag.chron <- mag %>% select(1)
mag <- mag %% select(`Mag Susc`) %>% mutate(`Mag Susc` = `Mag Susc` / 1000)
loi <- read_excel(fpath, sheet="Loss on ignition", skip=2)</pre>
loi.chron <- loi %>% select(1)
loi <- loi %>% select("LOI (wt %)")
BSi <- read_excel(fpath, sheet="Biogenic silica", skip=2)
BSi.chron <- BSi %>% select(1)
BSi <- BSi %>% select("BSi")
rp1 <- riojaPlot(pollen, pollen.chron, groups=types,</pre>
          yinterval = 5000,
          ymin = 0,
          ymax=102000,
          yvar.name = "Age BP",
          scale.percent=TRUE,
          plot.groups=TRUE,
          do.clust = TRUE,
          plot.zones = "auto",
          plot.clust=TRUE,
          plot.cumul=TRUE,
          cex.cumul=0.6,
          srt.xlabel=45,
          xSpace = 0.01,
          plot.bar=FALSE,
          tcl=-0.1,
          cex.yaxis=0.7,
          cex.xlabel=0.8,
          xRight = 0.7,
          plot.line=FALSE
rp2 <- riojaPlot(mag, mag.chron[, "Age BP", drop=FALSE],</pre>
           riojaPlot=rp1, xGap = 0.01,
           xRight=0.8, scale.minmax=FALSE,
           plot.bar=FALSE, plot.line=F,
           plot.symb=TRUE, symb.cex=0.3)
rp3 <- riojaPlot(loi, loi.chron[, "Age BP", drop=FALSE],</pre>
           riojaPlot=rp2,
           xRight=0.9,
           scale.minmax=FALSE, plot.bar=F,
          plot.line=F, plot.symb=TRUE, symb.cex=0.3)
riojaPlot(BSi, BSi.chron[, "Age BP", drop=FALSE],
           riojaPlot=rp3,
           xRight=0.99,
           scale.minmax=FALSE, plot.bar=FALSE,
           plot.line=FALSE, plot.symb=TRUE, symb.cex=0.3)
```



9. Adding custom plotting functions

In the above example the physical / chemical variables are measured on a large number of samples and we plot with symbols because these display the variability in the data more clearly than lines. It would be useful to fit a smooth through these data to highlight the trends. Here we plot the three phys/chem variables and fit a gam to each curve using a custom function to add the smooth to each plot.

```
fun.gam <- function(x, y, i, nm, style) {</pre>
  tmp <- data.frame(x=y, y=x)</pre>
  gam \leftarrow mgcv::gam(y \sim s(x, k=50), data=tmp)
  x2 <- predict(gam, type="response")</pre>
  lines(x2, y, col="blue", lwd=1)
rp <- riojaPlot(mag, mag.chron[, "Age BP", drop=FALSE],</pre>
           yinterval = 5000,
           ymin = 0,
           ymax=102000,
           xRight=0.3,
           scale.minmax=FALSE,
           plot.bar=FALSE,
           plot.line=F,
           plot.symb=TRUE,
           plot.poly=FALSE,
           symb.cex=0.3,
           fun.xfront=fun.gam)
rp1 <- riojaPlot(loi, loi.chron[, "Age BP", drop=FALSE],</pre>
            riojaPlot=rp,
```

```
xRight=0.5,
            scale.minmax=FALSE, plot.bar=F, plot.line=F, plot.symb=TRUE,
            symb.cex=0.3, fun.xfront=fun.gam)
riojaPlot(BSi, BSi.chron[, "Age BP", drop=FALSE],
            riojaPlot=rp1,
            xRight=0.7,
            scale.minmax=FALSE, plot.bar=FALSE, plot.line=FALSE, plot.symb=TRUE,
            symb.cex=0.3, fun.xfront=fun.gam)
           Mag Susc
                                                              BSi
     5000
    10000
    15000
    20000
    25000
    30000
    35000
    40000
Age BP
    45000
    50000
    55000
    60000
    65000
    70000
    75000
    80000
    85000
    90000
    95000
    100000
                     3
                                           20
                                              30
                                                  40
                                                      50
                                                         60
                                                                10
                                                                   20
```

10. Adding a column showing lithology

Adding a column showing lithogy is experimental at the moment but it is possible to show basic lithology with different colours and / or pattern fills. To do this create a data frame with at least 3 columns - the top and bottom of each lithological unit, and the fill colour for that unit. You also have a define a custom function that takes this data frame and draws the lithology. Here we create a function that uses apply to loop through each row in the lithology dataframe and plot a filled rectangle in the lithology column.

```
myfun2 <- function(x, style) {</pre>
  x %>% pwalk(~rect(0, ..1, 1, ..2, col=..4))
mx <- apply(aber.poll, 2, max)</pre>
mx5 <- mx[mx>10]
rp <- riojaPlot(aber.poll, aber.chron, selVars=names(mx5), lithology=lithology,</pre>
              sec.yvar.name="Age (years BP)",
              sec.ymin=6000, sec.ymax=14000, sec.yinterval=500,
              yvar.name="Depth (cm)",
              plot.sec.axis = TRUE,
              scale.percent=TRUE,
              fun.lithology=myfun2)
                                                                                                                     Rumex acetosa-type
                                                                       Juniperus communis
                                   Pinus sylvestris
                                                       Corylus-Myrica
   Age (years BP)
                                                                                                Cyperaceae
                                                                                        Gramineae
          Depth (cm)
                                                                               Empetrum
                                                                                                     Artemisia
                  Betula
                                                              Salix
         300-
 6500-
 7000
 7500
         350-
 8000
 8500
 9000
         400-
 9500
10000
10500
         450-
11000
11500
12000
         500
12500
13000
13500
```

11. Frequently asked questions

60

14000

550

1. How do I change the order of the variables in the diagram?

20 40 60 80

There are several ways to do this. Variables are plotted in the order they appear in the data, so rearrange the order of variables in the original data. dplyr::select offers a convenient way to do this:

20

20

20

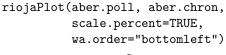
```
data.reordered <- data %>% select(var3, var1, var5) # and so on...
```

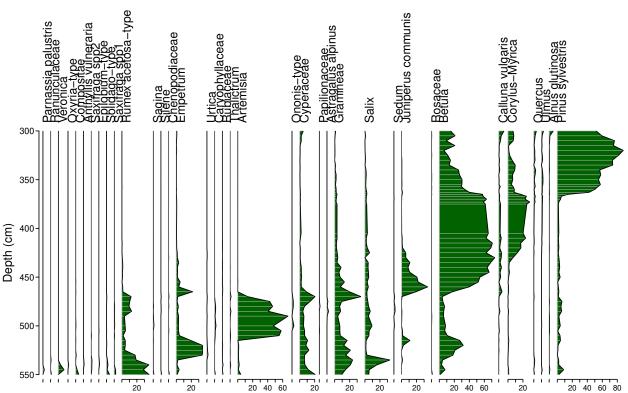
Another way is to create a character vector wi the variables in the required order and pass this to selVars:

```
myorder <- c("var3", "var1", "var5") # and so on...
riojaPlot(spec, chron, selVArs=myorder)</pre>
```

A quick way to sort variables accorging to their position in the diagram is to set the style

wa.order="bottomleft" (or use bottomright for the opposite). This will sort variables from those common at the base of the core on the left to those common at the top on the right.





To have complete control you probably need to use option 1 - change the order in the original data. Here is an extended example in which we sort the taxa in the RLGH diatom data according to their pH optima. We also group the taxa into three pH classes based on their pH optima and display the groups and cumulative plot.

RLGH.diat <- RLGH.diat %>% select(optima2\$CODE)

```
mx <- apply(RLGH.diat, 2, max)</pre>
RLGH.sel <- colnames(RLGH.diat)[mx > 3]
rp <- riojaPlot(RLGH.diat, RLGH.chron, groups=optima2,</pre>
              yvar.name="Year",
              scale.percent=TRUE,
              y.rev=FALSE,
              selVars=RLGH.sel,
              ymax=1980,
              yinterval=10,
              xlabels=RLGH$names$TaxonName,
              labels.italicise=TRUE,
              cex.xlabel=0.7,
              plot.poly=FALSE,
              plot.line=FALSE,
              lwd.bar=3,
              xRight=0.9,
              plot.groups=TRUE,
             plot.cumul=TRUE,
              col.group=c("mediumblue", "darkgrey", "red2"),
              cumul.mult=0.3
riojaPlot(RLGH.pH, RLGH.chron[, "Year", drop=FALSE],
               riojaPlot=rp,
               scale.minmax=FALSE,
               plot.bar=FALSE,
               plot.symb=TRUE,
               symb.cex=0.6)
                                                                              Neidium glaberrimum
                                                             Vavicula madumensis
                                                                        Navicula cumbriensis
                                    Eunotia denticulata
                             Eunotia bactriana
                         Cymbella lunata
                                                                                                                 DI-pH (SWAP)
                                        Eunotia incisa
                                                                                  Peronia fibula
                                                                                          Pinnularia
microstauron
                                                                                                   Acid intol.
Acidophil.
Acidobiont.
   1980
   1970
   1960
   1950
   1940
   1930
   1920
   1910
   1900
   1890
   1880·
   1870
   1860
   1850
   1840
                                                                                                  20 40 60 80
                                                                                                               4.8 5.0 5.2
```

2. How do I change the font sizes?

Use the following styles (note, font sizes are specified relative to the default size of 1): - cex.xaxis = font size of x-axis values (default 0.6) - cex.yaxis = font size of y-axis values (default 0.7) - cex.xlabel = font size of x-axis labels (default 0.9) - cex.ylabel = font size of y-axis labels (default 0.9) - cex.cumul = font size of group names in cumulative plot (default 0.7)

3. How do I change the limits and labels of the of the y-axes?

Use the style y.rev to reverse the scale of y-axis (default is TRUE, ie. values increase from top to bottom). Use the styles ymin, ymax and yinterval to change the start, end and interval values. The corresponding styles sec.ymin, sec.ymax and sec.yinterval control the secondary y-axis scale.

4. My variable names are too long and take up too much space at the top of the diagram.

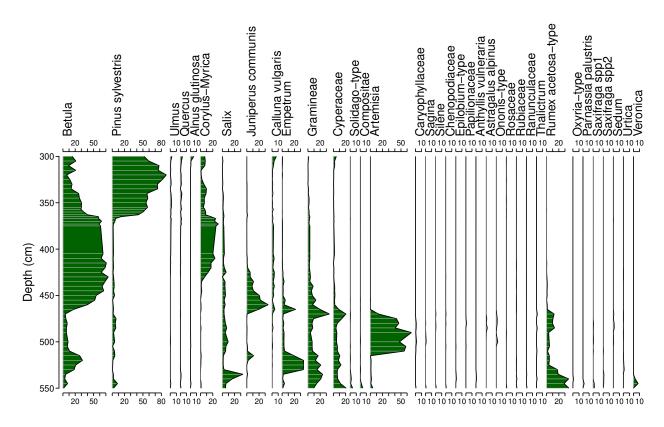
Long names can be split by setting style labels.break.long=TRUE (the default). Names will be split so no part if greater than labels.break.n characters (default 20).

5. How do I add a second x-axis at the top of the plots?

Styles plot.top.axis and plot.bottom.axis allow plotting of the top and bottom x-axes. If there is not enough space between the top axis and the taxon names use style ylabPos to fine-tune the gap (default 0.1).

```
data(aber)
aber.poll <- aber$spec
colnames(aber.poll) <- aber$names$Name
aber.chron <- aber$ages

riojaPlot(aber.poll, aber.chron,
    scale.percent=TRUE,
    min.width.pc=10,
    plot.top.axis=TRUE)</pre>
```

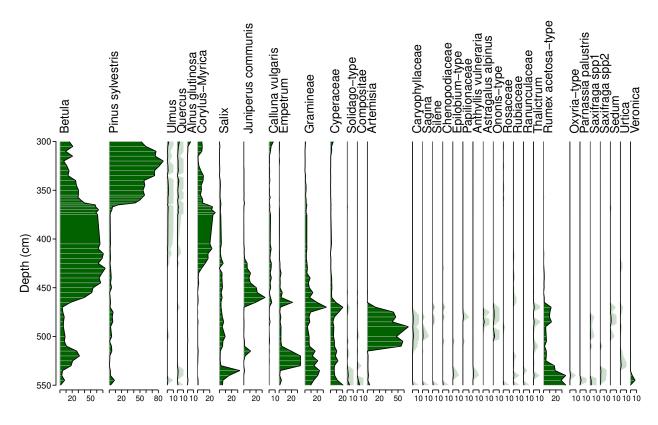


6. How do I add exaggeration curves to selected taxa only?

Style plot.exag takes a single value (TRUE or FALSE) to add exaggerations to all curves but can also take a logical vector of length equal to the number of columns in the x-variables data frame to control exaggerations for each variable. The following example shows how to add exaggerations for pollen types with maximum relative abundance < 5 percent. We also change the exaggeration multiplier to 10 and set the min widths of curves to 10% (default is 5%).

```
aber.poll <- aber$spec
colnames(aber.poll) <- aber$names$Name
aber.chron <- aber$ages
mx <- apply(aber.poll, 2, max)
exag.sel <- mx < 5

riojaPlot(aber.poll, aber.chron,
    scale.percent=TRUE,
    plot.exag=exag.sel,
    exag.mult=10,
    min.width.pc=10)</pre>
```

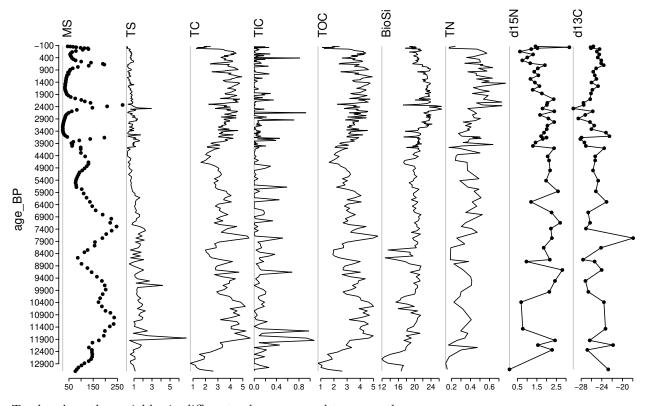


7. How I plot different styles for different variables?

In this example we use geochemical data from Laguna del Maule published by Frugone-Álvarez et al. (2020) and downloaded from the NOAA Paleoclimate data archive (https://www.ncei.noaa.gov/products/paleo climatology). The data we plot here contain 9 geochemical variables. Some (TS, TC, TIC and TOC) are measured at every interval but others only at selected depths. We plot magnetic susceptibility with symbols, TS, TC, TIC, TOC and BioSi with lines, and d15N and d13C with lines and symbols.

```
fpath <- system.file("extdata/maule2020geochem.txt", package="riojaPlot")</pre>
maule <- readr::read_delim(fpath, skip=158, show_col_types = FALSE)
maule.data <- maule %>% select(-(1:4))
maule.chron <- maule %>% select(1:4)
# How many measurements do we have for each variable:
unlist(lapply(maule.data, function(x) sum(!is.na(x))))
      MS
                               TOC BioSi
                                                 d15N
                                                        d13C
             TS
                   TC
                         TIC
                                             TN
     146
            267
                  267
                         267
                               267
                                      253
                                            108
                                                    57
                                                          57
selsymb <- rep(FALSE, 9)
selsymb[c(1, 8:9)] \leftarrow TRUE
selbar <- rep(FALSE, 9)</pre>
selline <- rep(TRUE, 9)</pre>
selline[1] <- FALSE
riojaPlot(maule.data, maule.chron,
          yvar.name="age_BP",
          ymin=-100,
          ymax=13300,
          yinterval=500,
```

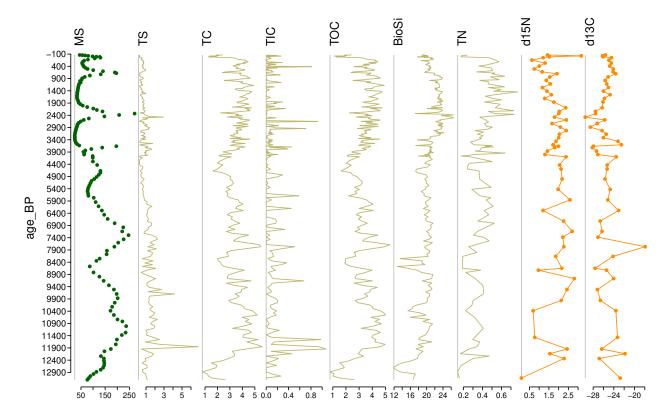
```
plot.bar=FALSE,
plot.poly=FALSE,
plot.line=selline,
plot.symb=selsymb,
symb.cex=0.5,
)
```



To also show the variables in different colours we need to group them.

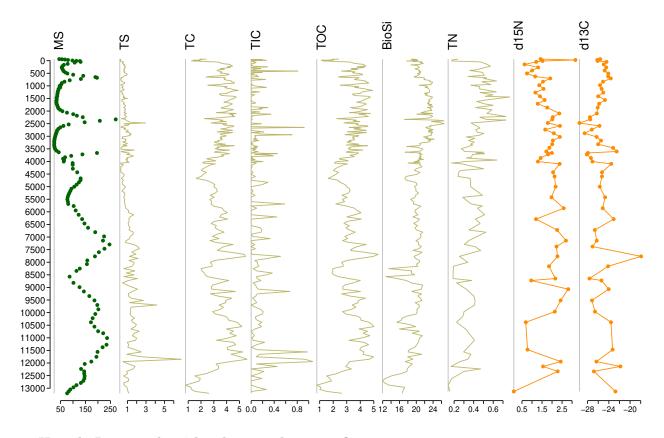
```
groups <- data.frame(names=colnames(maule.data), groups=c(1, 2, 2, 2, 2, 2, 2, 3, 3))
riojaPlot(maule.data, maule.chron, groups=groups,</pre>
```

```
yvar.name="age_BP",
ymin=-100,
ymax=13300,
yinterval=500,
plot.bar=selbar,
plot.poly=FALSE,
plot.line=selline,
plot.symb=selsymb,
plot.groups=TRUE,
symb.cex=0.5
```



8. How do I control the tick values on the y-axis?

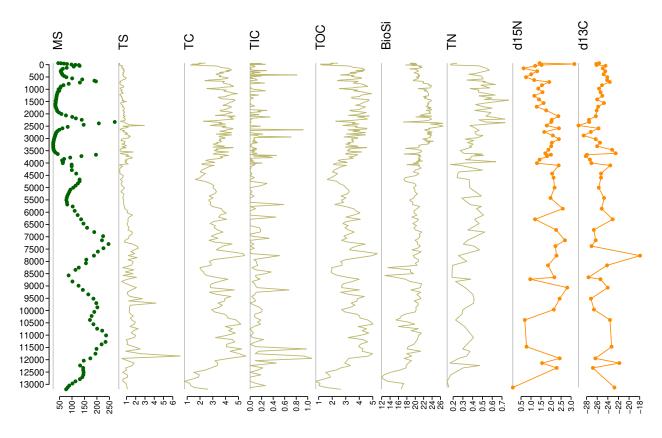
riojaPlot will try to add a y-axis with a sensible upper and lower limit and interval. It doesn't always suceed. If the default doesn't work, try specifying values for ymin, ymax and yinterval (and corresponding sec.ymin etc. value for the seconday axis). If this still doesn;t work you can pass a numeric vector of "tick values" to styles ytks1 and ytks2 for the primary and secondary axes respectively. In the plot above the y-axis values start at -100 and increment in 500 which gives unusal values. To fix this we create a vector of tick values from 0 to 13000 and use this.



9. How do I rotate the tick values on the x-axes?

 $\begin{tabular}{ll} USe the style {\tt las=2}. You may also need to create a larger space at the bottom of the figure using yBottom. \\ \end{tabular}$

```
yvar.name="age_BP"
ymin=-100,
ymax=13300,
yinterval=500,
plot.bar=FALSE,
plot.poly=FALSE,
plot.line=selline,
plot.symb=selsymb,
plot.groups=TRUE,
ylabPos=2.2,
symb.cex=0.5,
ytks1=myticks,
las.xaxis=2,
yBottom=0.06
```

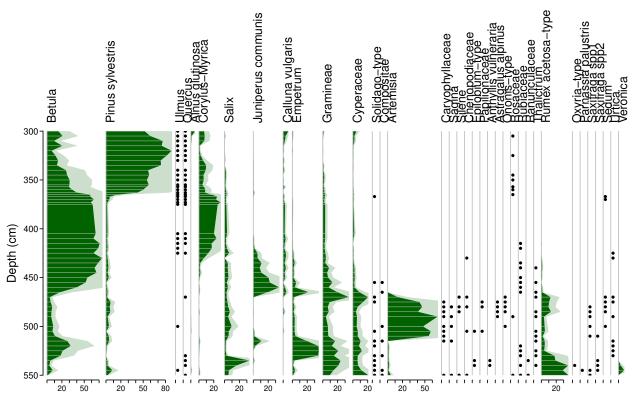


10. How do I plot symbols for rare types instead of bars or curves?

To do this we supply plot.poly etc. with a logical vector to control the plotting of each curve then set the values of these styles to FALSE for the rare types. Then we define a function to plot symbols for the rare types and create a list of functions to plot symbols only for the rare types.

```
aber.poll <- aber$spec</pre>
colnames(aber.poll) <- aber$names$Name</pre>
aber.chron <- aber$ages
# calculate of max of each column
mx <- apply(aber.poll, 2, max)</pre>
\#create a logical vector which is TRUE for taxa with max < 5
sel \leftarrow mx < 5
# define a custom function to plot symbols
symb.fun <- function(x, y, i, nm, style) {</pre>
   sel \leftarrow x > 0
   if (sum(sel) > 0) {
      points(rep(3, sum(sel)), y[sel], cex=0.4, pch=19)
   }
}
# create a list of functions of length equal to the number of columns in the data
funlist <- lapply(1:ncol(aber.poll), function(x) symb.fun)</pre>
# now set the elements of the list where we don;t want to plot symbols to NULL
funlist[!sel] <- list(NULL)</pre>
# plot silhouettes and lines for taxa > 5% and apply our function to the others
riojaPlot(aber.poll, aber.chron,
   scale.percent=TRUE,
```

plot.poly=!sel,
plot.bar = !sel,
plot.line=FALSE,
lwd.bar=0.6,
plot.exag=TRUE,
fun.xfront=funlist)



11. How do I change the widths of individual graphs?

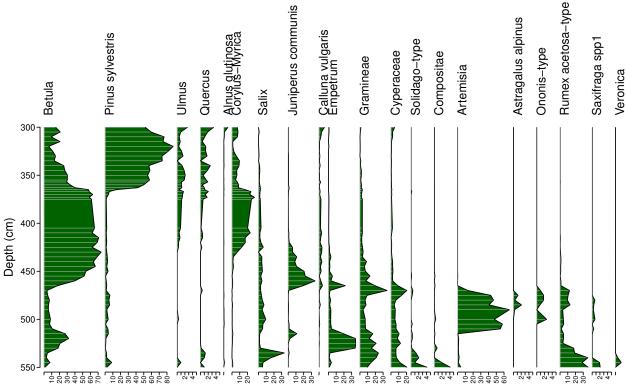
scale.percent=TRUE,
graph.widths=widths,

In the above example we plotted rare types as symbols. In the example we can expand the x-axis for rare types.

For percentage diagrams the width of each graph is scaled uniformly so the percentage values for each graph are the same. We can change these using the style graph.widths which takes a vector of relative widths. We can also change the x-axis labels for each graph by supplying a numeric vector to style x.inc.pc to set the interval for x-values. Here we hide columns with maximum relative abundance < 2%, and show those taxa with max < 5% as curves with 5 * normal width, and set the x-interval for these to 1% (rather than the default of 10%). We could, of course, chose the columns manually to have complete control over what is hidden and shown with expanded axes.

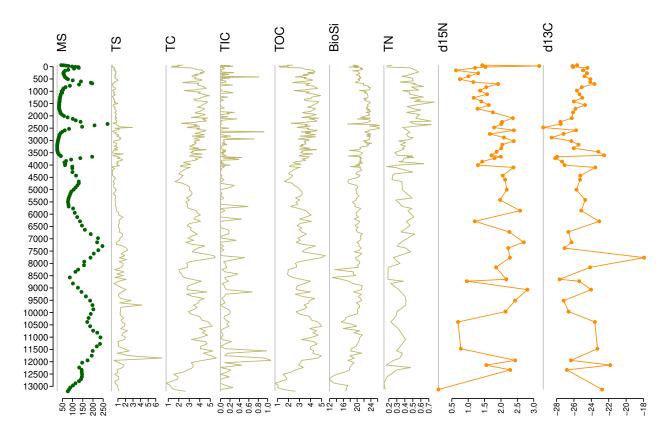
```
mx <- apply(aber.poll, 2, max) # calculate max % of each column
widths <- rep(1, ncol(aber.poll)) # generate numeric vector of widths, set to 1
inc <- rep(10, ncol(aber.poll)) # generate numeric vector of x-intervals, set to 10
selTaxa <- names(mx[mx > 2]) # generate character vector of column names to include in the figure
sel <- which(mx < 5) # generate a logical vector of columns to expand
widths[sel] <- 5 # set widths to 5 times normal width
inc[sel] <- 1 # set x-interval to 1%
riojaPlot(aber.poll, aber.chron, selVars=selTaxa,</pre>
```

```
min.width.pc=5,
x.pc.inc=inc,
cex.xaxis=0.5,
las.xaxis=2)
```



We can also change the widths for non-percent data. Here we return to the Laguna del Maule data in FAQ 7 and change the width of d15N and d13C columns to twice the standard width.

```
widths <- rep(1, ncol(maule.data))</pre>
#sel <- names(maule.data) %in% c("d15N", "d13C")</pre>
#widths[sel] <- 2</pre>
widths[8:9] <- 2
riojaPlot(maule.data, maule.chron, groups=groups,
          yvar.name="age_BP",
          ymin=-100,
          ymax = 13300,
          yinterval=500,
          plot.bar=FALSE,
          plot.poly=FALSE,
          plot.line=selline,
          plot.symb=selsymb,
          plot.groups=TRUE,
          ylabPos=2.2,
          symb.cex=0.5,
          ytks1=myticks,
          las.xaxis=2,
          graph.widths=widths,
          yBottom=0.06
```



12. How do I plot surface pollen or diatom data?

riojaPlot can plot surface sediment data, you just need to think how to arrange samples on the y-axis. The example below plots surface sediment diatom data from small ponds and pools in SE England collected by Bennion (1994) (see ?rioja::Ponds) for details. Here we arrange the samples along the TP gradient and plot the data against Lake name with TP as a secondary y-axis.

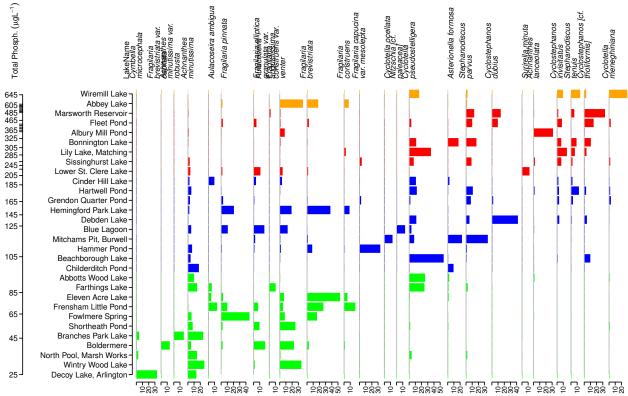
```
data(Ponds)
# reorder the rows in decreasing TP
o <- order(Ponds$env$TP, decreasing=TRUE)</pre>
ponds.diat <- Ponds$spec[o, ]</pre>
ponds.TP <- data.frame(TP=round(Ponds$env$TP[o]), LakeName=Ponds$env$Name[o])</pre>
# replace taxon codes with names
colnames(ponds.diat) <- Ponds$names$Name</pre>
# remove rare species
mx <- apply(ponds.diat, 2, max)</pre>
ponds.sel <- colnames(ponds.diat)[mx > 10]
# With a large number of samples we would use points but with only 30
# in this data we use bars. Although the data are arranged in order of
# increasing TP they are not a temporally or spatially contiguous so
# lines or silhouettes are not appropriate.
riojaPlot(ponds.diat, ponds.TP, selVars=ponds.sel,
          yvar.name="LakeName",
          sec.yvar.name="TP",
          sec.yinterval = 20,
          plot.sec.axis=TRUE,
          scale.percent=TRUE,
```

```
plot.poly=FALSE,
               plot.line=FALSE,
               plot.bar=TRUE,
               col.bar="black",
               lwd.bar = 2,
                symb.cex=0.4,
                cex.xlabel=0.6,
                cex.yaxis=0.6,
                wa.order="topleft",
                labels.italicise=TRUE,
                las.xaxis=2,
                cex.xaxis=0.5)
                                                                    Asterionella formosa
 Р
645-
                 Wiremill Lake
605
485
465
                   Abbey Lake
           Marsworth Reservoir
                   Fleet Pond
365
               Albury Mill Pond
325
              Bonnington Lake
305
285
             Lily Lake, Matching
              Sissinghurst Lake
245
           Lower St. Clere Lake
205
               Cinder Hill Lake
                 Hartwell Pond
          Grendon Quarter Pond
165
          Hemingford Park Lake
145
125-
                 Blue Lagoon
           Mitchams Pit, Burwell
                Hammer Pond
105
            Beachborough Lake
              Childerditch Pond
            Abbotts Wood Lake
                Farthings Lake
 85
              Eleven Acre Lake
           Frensham Little Pond
 65
              Fowlmere Spring
              Shortheath Pond
 45
            Branches Park Lake
                  Boldermere
        North Pool, Marsh Works
             Wintry Wood Lake
 25-
           Decoy Lake, Arlington
```

13. How do I plot different levels or samples with a different colour?

It is currently only possible to plot different samples with different coloured bars. Here we extend the example for FAQ 12 above and classify the samples into different groups based on their TP, then plot these different samples with different colours. We do this by creating a character vector of colour names, that corresponds to the samples, then pass this to style col.sep.bar and set sep.bar to true. We also create an expression to label the TP axis in the correct units.

```
sec.vvar.name="TP",
sec.ylabel=ylab,
sec.yinterval = 20,
plot.sec.axis=TRUE,
scale.percent=TRUE,
plot.poly=FALSE,
plot.line=FALSE,
plot.bar=TRUE,
col.bar="black",
sep.bar=TRUE,
col.sep.bar=ponds.TP$bar.cols,
lwd.bar = 10,
symb.cex=0.4,
cex.xlabel=0.6,
cex.yaxis=0.6,
wa.order="bottomleft",
labels.italicise=TRUE,
las.xaxis=2,
cex.xaxis=0.5)
```

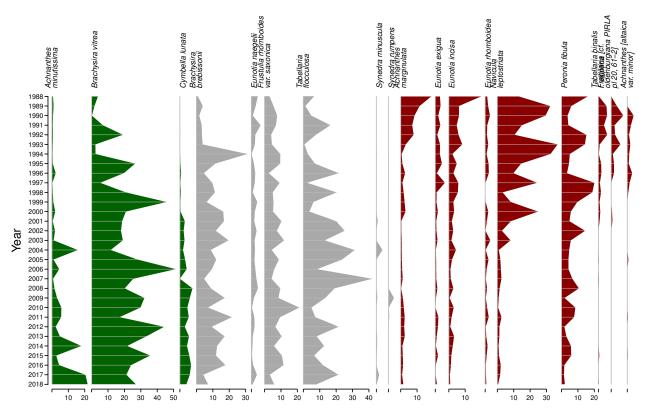


14. How do I plot ecological time series?

riojaPlot can also be used to plot multi-species ecological time series (or spatial transect) data. Here we plot diatom data from the UK Upland Waters Monitoring Network (https://uwmn.uk/). These are epilithic diatom samples collected annually from 1988-2018 from Loch Chon (Trossachs, Scotland). These data were collected to monitor the potential recovery of the loch from previous acidification. The file of diatom counts (expressed as relative abundance) also contains results of a trend test based on a multivariate generalised linear model using the manyglm function in package mvabund used to identify taxa that have a significant increasing or decreasing temporal trend.

We first re-order the taxa according to their trend over time and plot the data with silhouettes, coloured to show taxa with significant increasing, decreasing or no trend.

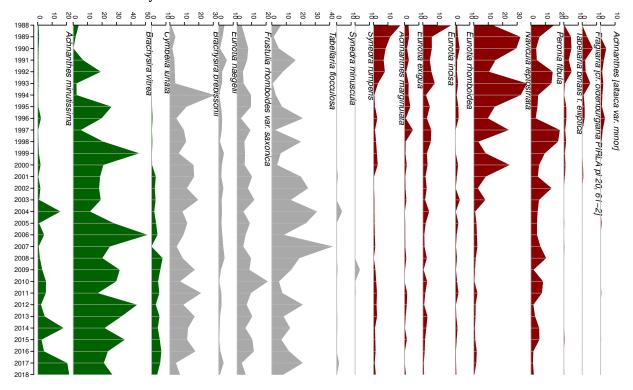
```
fpath <- system.file("extdata/LochChon.xlsx", package="riojaPlot")</pre>
chon <- readxl::read_excel(fpath, sheet="Diatoms")</pre>
sig_test <- readxl::read_excel(fpath, sheet="Sig_test")</pre>
chon.diat <- chon %>% select(-Year)
chon.year <- chon %>% select(Year)
# data has many rare species, remove these first
mx <- apply(chon.diat, 2, max)</pre>
chon.diat <- chon.diat[, mx > 3]
# Create a grouping variable based on significance of trend (increasing, decreasing or not sig.)
chon.groups <- data.frame(TaxonName=colnames(chon.diat)) %>%
  left_join(sig_test, by="TaxonName") %>%
  mutate(group = case_when(p_unadj < 0.1 & slope>0 ~ "Increasing",
                           p_unadj < 0.1 & slope<0 ~ "Decreasing",</pre>
                            TRUE ~ "Not sig"),
         group=factor(group, levels=c("Increasing", "Not sig", "Decreasing"))) %>%
  select(TaxonName, group)
# Sort variables according to significance
diat.order <- chon.groups %>% arrange(group) %>% pull(TaxonName)
chon.ordered <- chon.diat %>% select(!!diat.order)
riojaPlot(chon.ordered, chon.year, groups=chon.groups,
          scale.percent=TRUE,
          yinterval=1,
          plot.poly=TRUE,
          plot.line=FALSE,
          plot.groups=TRUE,
          cex.xlabel=0.6,
          cex.yaxis=0.5,
          labels.italicise=TRUE,
          cex.xaxis=0.5.
          col.group=c("darkgreen", "darkgrey", "darkred"))
```



This looks OK but we can "virtually" rotate this figure so it can be viewed as a stacked diagram. To do this we have to modify the axis labels and orientation of taxon names. We can't easily do this using riojaPlot styles but we can suppress the plotting of names and x-axis and create a custom function to plot these in any style or position we wish.

```
# create list of blank names to suppress taxon names
names <- rep("", ncol(chon.ordered))</pre>
myfun <- function(x, y, i, nm, style) {</pre>
  usr <- par("usr") # extract the x and y data limits of the plot
  name <- bquote(italic(.(names(nm)))) # extract name from nm and italicise it</pre>
  text(usr[2]-2, usr[4]+0.5, name, adj=c(0, 0), xpd=NA, srt=-90, cex=0.7)
  xval <- seq(0, usr[2], by=10) # create a vector of labels for the axis
  xlab <- rep("", length(xval))</pre>
  axis(side=3, at=xval, labels=xlab, tcl=-0.2, cex.axis=0.5, mgp=c(3, 0.1, 0))
  text(xval, usr[4]-0.5, xval, cex=0.5, srt=-90, adj=c(1, 0), xpd=NA) # ad the x-axis vales
}
riojaPlot(chon.ordered, chon.year, groups=chon.groups,
          scale.percent=TRUE,
          yinterval=1,
          xlabels=names, # replace taxon names with a vector of blank names
          ylabel = " ", # suppress y-axis label
          plot.poly=TRUE,
          plot.line=FALSE,
          plot.groups=TRUE,
          cex.xlabel=0.6,
          cex.yaxis=0.5,
          labels.italicise=TRUE,
          cex.xaxis=0.5,
```

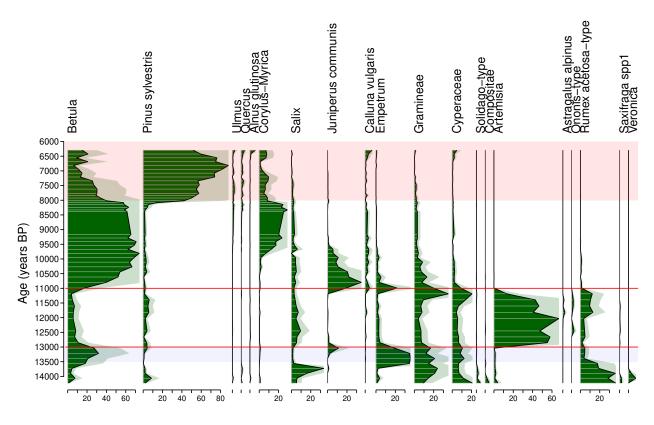
```
col.group=c("darkgreen", "darkgrey", "darkred"),
plot.bottom.axis=FALSE,
min.width.pc=10, # set minimum size of x-axes
fun.xfront=myfun)
```



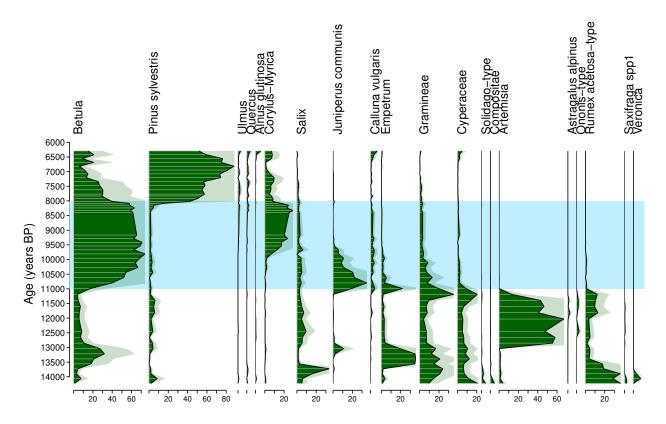
15. How do I add my own zones to a diagram?

Arbitrary zones can be added using function addRPZone, as either single lines, or as shaded rectangles. Simply save the riojaPlot object and pass it to addRPZone.

```
mx <- apply(aber.poll, 2, max)</pre>
aber.sel <- names(mx[mx > 2])
rp <- riojaPlot(aber.poll, aber.chron, aber.sel,</pre>
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500,
   scale.percent=TRUE,
   plot.exag=TRUE)
# define zone lines at 11000 and 13000 years BP
myzones <- c(11000, 13000)
addRPZone(rp, myzones, col="red")
# add shaded zone from 6000 to 8000 years BP
addRPZone(rp, 6000, 8000)
# change colour and shading
addRPZone(rp, 13000, 13500, col="blue", alpha = 0.05)
```



```
# The zones above are plotted over the existing diagram.
# It is possible to plot zones or other annotations behind
# the diagram using a custom function supplied to style fun.plotback
# this takes 2 arguments, usr = vector of 4 numbers giving the
# coordinates of the plotting area in data units (x is 0-1)
# and fig, coordinates of the plotting area as fractions of the page size
fun.back <- function(usr, fig, style) {</pre>
  print(usr)
  rect(0, 8000, 1, 11000, col="lightblue1", border=NA)
rp <- riojaPlot(aber.poll, aber.chron, aber.sel,</pre>
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500,
   scale.percent=TRUE,
   plot.exag=TRUE,
   fun.plotback=fun.back
#> [1]
                 1 14300 6000
```



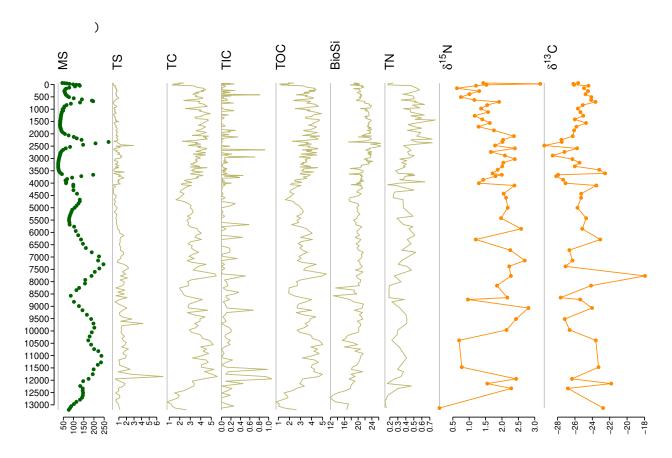
16. How do I include special symbols in the variable names

We can pass a character vector of custom names for each column using style xlabels and use expression to create names with superscripts, subscripts or greek letters.

```
nms <- colnames(maule.data)
nms[8] <- expression(delta^15*N)</pre>
```

We can also use function label_geochem in package tidypaleo to automatically add annotations for a slected range of common geochemical variables.

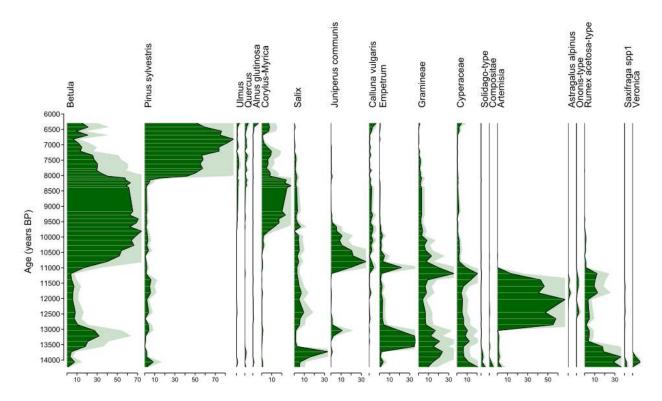
```
library(tidypaleo)
nms <- colnames(maule.data)</pre>
nms2 <- as.expression(sapply(nms, function(x) unlist(label_geochem(x)[[1]])))</pre>
riojaPlot(maule.data, maule.chron, groups=groups,
          yvar.name="age_BP",
          xlabels=nms2,
          ymin=-100,
          ymax=13300,
          yinterval=500,
          plot.bar=FALSE,
          plot.poly=FALSE,
          plot.line=selline,
          plot.symb=selsymb,
          plot.groups=TRUE,
          ylabPos=2.2,
          symb.cex=0.5,
          ytks1=myticks,
          las.xaxis=2,
          graph.widths=widths,
          yBottom=0.06
```



17. What is the best way to save riojaPlot diagrams?

'riojaPlot figures can be saved to file or copied to the clipboard from R or RStudio. This will work but will not always give the best diagram because the plot is scaled to the window it is drawn in, and saving a plot that is already drawn to a different size or aspect ratio may result in unexpected gaps between axes or truncated text. To avoid this, place device calls around the plot to explicitly draw the figure to a graphics file. For a stand-alone figure pdf is appropriate. For a figure to include in a document or presentation svg will usually work best for embedding in recent MS Office documents, or png for earlier documents. Just be careful that if you get an error during the plot to remember to call dev.off() to return plotting to the screen.

```
svg("Myplot1.svg", width=10, height=6) # figure dimensions in inches
# or pdf("Myplot.pdf", width=10, height=6) for output to a pdf file
riojaPlot(aber.poll, aber.chron, aber.sel,
    yvar.name="Age (years BP)",
    ymin=6000,
    ymax=14300,
    yinterval=500,
    scale.percent=TRUE,
    plot.exag=TRUE,
    )
dev.off()
#> pdf
#> 2
To view a saved svg or pdf file use cowplot::draw_image.
cowplot::ggdraw() + cowplot::draw image("Myplot1.svg")
```



Or save the plots to an object and combine using plot_grid:

```
svg("Myplot2.svg", width=10, height=6) # figure dimensions in inches
riojaPlot(aber.poll, aber.chron, aber.sel,
    yvar.name="Age (years BP)",
    ymin=6000,
    ymax=14300,
    yinterval=500,
    scale.percent=TRUE,
    plot.poly=FALSE
    )

dev.off()
#> pdf
#> 2
pl <- cowplot::ggdraw() + cowplot::draw_image("Myplot1.svg")
p2 <- cowplot::ggdraw() + cowplot::draw_image("Myplot2.svg")
cowplot::plot_grid(p1, p2, align="v", axis="tblr")</pre>
```

```
file.remove("Myplot1.svg")
#> [1] TRUE
file.remove("Myplot2.svg")
#> [1] TRUE
```

18. Can I use a pipe with riojaPlot

Yes, for diagrams with multiple parts you can pipe the output from one section into the next. Use function riojaPlot2 to add additional dataset to the plot. This is just a wrapper around riojaPlot() that takes a riojaPlot object as the first argument. Here is the example from section 8 above rewritten to use the pipe.

```
riojaPlot(RLGH.diat, RLGH.chron,
                   yvar.name="Year",
                   scale.percent=TRUE,
                   y.rev=FALSE,
                   selVars=RLGH.sel,
                   ymax=1980,
                   xlabels=RLGH$names$TaxonName,
                   cex.xlabel=0.7,
                   labels.break.n=25,
                   xRight=0.9) |>
    riojaPlot2(RLGH.pH, RLGH.chron[, "Year", drop=FALSE],
                   scale.minmax=FALSE,
                   plot.bar=FALSE,
                   plot.symb=TRUE,
                   symb.cex=0.6)
                                                                                          Frustulia rhomboides var.
viridula
             Achnanthes minutissima
                  Achnanthes marginulata
                                                                            Eunotia [vanheurckii var.
1]
                                                                                                                                                      Pinnularia microstauron
                                 Brachysira brebissonii
                                                                                                    Vavicula madumensis
                                                                                                                                Neidium glaberrimum
                                                                                                                     Navicula cumbriensis
                                                                      Eunotia vanheurckii
                                                          Eunotia denticulata
                                               Eunotia bactriana
                                        Cymbella lunata
                                                                                                                                                              DI-pH (SWAP)
                                                                 Eunotia incisa
                                                                                                                                      Peronia fibula
     1980
     1960
     1940
     1920
Year
     1900
     1880
     1860
                                                                                                                                                                   5.0
```

19. How do I cite riojaPlot in a publication

```
#>
#> To cite package 'riojaPlot' in publications use:
#>
```

```
#>
     Juggins, S. (2022) riojaPlot: Stratigraphic diagrams in R, package
#>
     version (0.1-19). (https://cran.r-project.org/package=riojaPlot).
#>
#> A BibTeX entry for LaTeX users is
#>
#>
     @Manual{,
       title = {riojaPlot: Stratigraphic diagrams in R},
#>
       author = {Steve Juggins},
#>
#>
       year = {2022},
       note = {R package version 0.1-19},
#>
#>
       url = {https://cran.r-project.org/package=riojaPlot},
#>
```

20. I get an error "Too many variables, curves will be too small"

If you try to plot too many variable in a single diagram the width of each will be too small to show in a meaningful way and riojaPlot will issue this error message. To fix this either reduce the number of variables or increase the size of the plotting window. If you want to view onscreen you can open a new plotting window with the windows(width=10, height=5) (on Windows) or quartz(width=10, height=5) (on MacOS), where width and height are in inches. Don't forget to close the windows when finished to return plotting to the R/Rstudio plot window. If you want to save to file just use function pdf() or svg() with appropriate width / height (see FAQ 17 above).

```
fpath <- system.file("extdata/LochChon.xlsx", package="riojaPlot")</pre>
chon <- readxl::read_excel(fpath, sheet="Diatoms")</pre>
chon.diat <- chon %>% select(-Year)
chon.year <- chon %>% select(Year)
# this will give an error
riojaPlot(chon.diat, chon.year,
          scale.percent=TRUE,
          yinterval=1,
          plot.poly=TRUE,
          plot.line=FALSE,
          plot.groups=TRUE,
          cex.xlabel=0.6,
          cex.yaxis=0.5,
          labels.italicise=TRUE,
          cex.xaxis=0.5)
#> Error in .riojaPlot2(d, yvar = yvar, y.rev = style$y.rev, scale.percent = style$scale.percent, : Too
# but this will work
svg("MyPlot.svg", width=20, height=8)
riojaPlot(chon.diat, chon.year,
          scale.percent=TRUE,
          yinterval=1,
          plot.poly=TRUE,
          plot.line=FALSE,
```

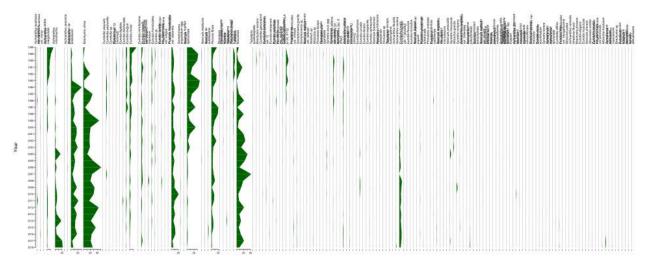
plot.groups=TRUE, cex.xlabel=0.6, cex.yaxis=0.5,

cex.xaxis=0.5)

labels.italicise=TRUE,

dev.off()
#> pdf
#> 2

cowplot::ggdraw() + cowplot::draw_image("Myplot.svg")



file.remove("Myplot.svg")
#> [1] TRUE

"' ## 11. List of modifiable styles and default values

Style	Value
yvar.name	
sec.yvar.name	
ylabel	
sec.ylabel	
plot.sec.axis	FALSE
scale.percent	FALSE
scale.minmax	FALSE
minmax	NA
y.rev	TRUE
ymin	NA
ymax	NA
yinterval	NA
sec.ymin	NA
sec.ymax	NA
sec.yinterval	NA
xlabels	NA
wa.order	none

plot.bar TRUE plot.line TRUE plot.poly FALSE plot.symb FALSE do.clust FALSE plot.clust FALSE plot.zones 0 lwd.bar 0.5 lwd.line 1 lwd.poly.line 0.5 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.cumul.line NA col.zones red col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cst.xlabel 90 srt.xlabel PALSE tcl -0.2 cex.cumul 0.7 clust.data.trans	Style	Value
plot.poly FALSE plot.symb FALSE do.clust FALSE plot.clust FALSE plot.zones 0 lwd.bar 0.5 lwd.line 1 lwd.poly.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	plot.bar	TRUE
plot.symb FALSE do.clust FALSE plot.clust FALSE plot.zones 0 lwd.bar 0.5 lwd.line 1 lwd.poly.line 0.5 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.zones red col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	plot.line	TRUE
do.clust FALSE plot.zones 0 lwd.bar 0.5 lwd.line 1 lwd.poly.line 0.5 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	plot.poly	FALSE
plot.clust FALSE plot.zones 0 lwd.bar 0.5 lwd.line 1 lwd.poly.line 0.5 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zones.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	plot.symb	FALSE
plot.zones 0 lwd.bar 0.5 lwd.line 1 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.zones red col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	do.clust	FALSE
lwd.bar 0.5 lwd.line 1 lwd.poly.line 0.5 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	plot.clust	FALSE
lwd.line 1 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	plot.zones	0
lwd.poly.line 0.5 lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	lwd.bar	0.5
lwd.cumul.line 0.5 col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	lwd.line	1
col.bar grey bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	lwd.poly.line	0.5
bar.back FALSE col.symb black col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	lwd.cumul.line	0.5
col.symb col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.xlabel NA centre.xlabel tcl -0.2 cex.cumul black darkgreen NA darkgreen NA NA col.zone. Darkgreen NA col.zone. NA col.zone. Darkgreen NA col.zone. Darkgreen NA col.zone. Darkgreen Col.zo	col.bar	grey
col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.xlabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	bar.back	FALSE
col.line black col.poly darkgreen col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.symb	black
col.poly.line NA col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.line	black
col.cumul.line NA col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.poly	darkgreen
col.zones red col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.poly.line	NA
col.zone.column grey lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.cumul.line	NA
lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.zones	red
lwd.zones 1 symb.pch 19 symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	col.zone.column	grey
symb.cex 1 cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.xlabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	lwd.zones	
cex.xaxis 0.6 cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	symb.pch	19
cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	symb.cex	1
cex.yaxis 0.7 cex.ylabel 0.9 cex.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	cex.xaxis	0.6
cex.ylabel 0.9 cex.xlabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	cex.yaxis	0.7
cex.xlabel 0.9 srt.xlabel 90 srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	cex.ylabel	0.9
srt.ylabel NA centre.xlabel FALSE tcl -0.2 cex.cumul 0.7		0.9
centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	srt.xlabel	90
centre.xlabel FALSE tcl -0.2 cex.cumul 0.7	srt.ylabel	NA
tcl -0.2 cex.cumul 0.7		FALSE
		-0.2
clust.data.trans none	cex.cumul	0.7
	clust.data.trans	none
clust.use.selected FALSE	clust.use.selected	FALSE
clust.width 0.05	clust.width	0.05

Style	Value
graph.widths	1
plot.exag	FALSE
col.exag	auto
col.exag.line	NA
lwd.exag.line	0.6
exag.mult	2
exag.alpha	0.2
labels.break.long	TRUE
labels.break.n	20
labels.italicise	FALSE
plot.groups	FALSE
plot.cumul	FALSE
cumul.mult	1
col.group	darkgreen, darkkhaki, darkorange, darkred, deepskyblue, sienna3, darkgoldenrod3, darkseagreen, yellow3, darkgrey
xRight	0.99
xLeft	NA
yBottom	0.05
уТор	NA
fun.xback	NA
fun.xfront	NA
fun.plotback	NA
fun.yaxis	NA
ylabPos	NA
xlabPos	0.1
xSpace	0.005
x.pc.omit0	TRUE
lwd.axis	1
col.axis	grey
min.width.pc	5
las.xaxis	1
las.yaxis	1
ytks1	NA
ytks2	NA
omitMissing	TRUE

Style	Value
col.sep.bar	black
sep.bar	FALSE
plot.bottom.axis	TRUE
plot.top.axis	FALSE
cumulSpace	NA
plot.yaxis	TRUE
start.new.plot	TRUE
xGap	0.01
x.pc.inc	10
fun.lithology	NA
lithology.width	0.03
user1	NA
user2	NA
user3	NA
user4	NA

12. References

Allen, J.R.M. $et\ al.\ (1999)$ Rapid environmental changes in southern Europe during the last glacial period. Nature, $400,\ 740-743.$

Bennion, H. (1994) A diatom-phosphorus transfer function for shallow, eutrophic ponds in southeast England. Hydrobiologia, 275/276, 391-410.

Frugone-Álvarez, M. et al. (2020) Volcanism and climate change as drivers in Holocene depositional dynamic of Laguna del Maule (Andes of central Chile - 36S). Climate of the Past, DOI 10.5194/cp-2019-147.