riojaPlot: User Guide (Version 0.1-1)

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1. Introduction

riojaPlot plots a set of variables in a stratigraphic diagram. Diagrams can be plotted with lines, silhouettes, bars or symbols of any combination of these element. riojaPlot extends the older function strat.plot in several ways: variables can be grouped and displayed with different colours, a cumulative summary plot can be added automatically based on the grouping, a cluster dendrogram and resulting zones can be added automatically, a secondary y-axis can be added to plot both depth and age axes, and the figures margins are determined automatically depending on the size and length of labels. Arguments to riojaPlot that control the appearance of the plot are know as styles. Styles can specified on a plot-by-plot basis or can be saved and applied to multiple figures.

Please send questions, bug reports and suggestions for improvements and additions to Stephen.Juggins@ncl. ac.uk.

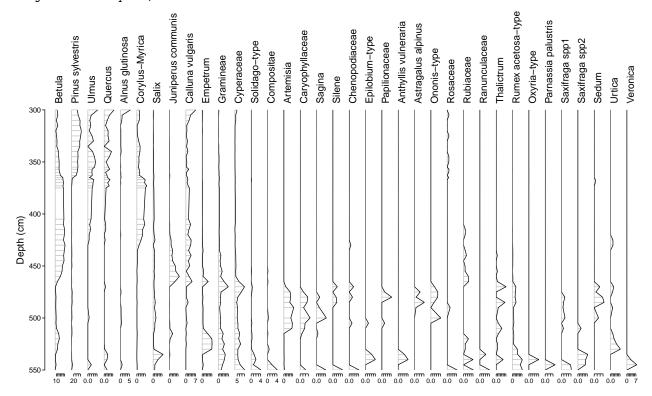
2. Basic diagram

To create a basic stratigraphic diagram just pass riojaPlot a data frame of variables to plot and and a data frame with at least one column containing the y-axis variable (usually sample depth or age). If the y-axis variable name is not specified riojaPlot will use the first column and label the y-axis with the column name. The default plot is shown with lines and bars for non-percentage data and as filled silhouettes (filled curves) and bars for percentage data (style scale.percent=TRUE. Turn silhouettes off and create plots with lines, bars or symbols by setting styles plot.poly, plot.line, plot.bars or plot.symb TRUE or FALSE (all may be overlain).

library(riojaPlot)

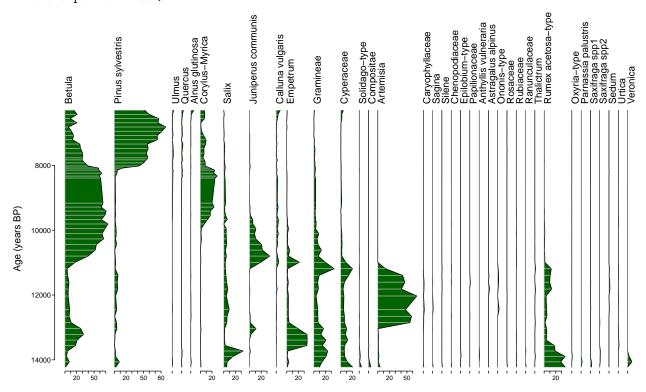
```
# use built-in data from Abernethy Forest
# see ?rioja::aber for citation
data(aber)
# extract pollen percentages
aber.poll <- aber$spec
# replace species codes with full taxon names
colnames(aber.poll) <- aber$names$Name
aber.chron <- aber$ages</pre>
```

plot on depth scale (depth is the first column in chron)
riojaPlot(aber.poll, aber.chron)



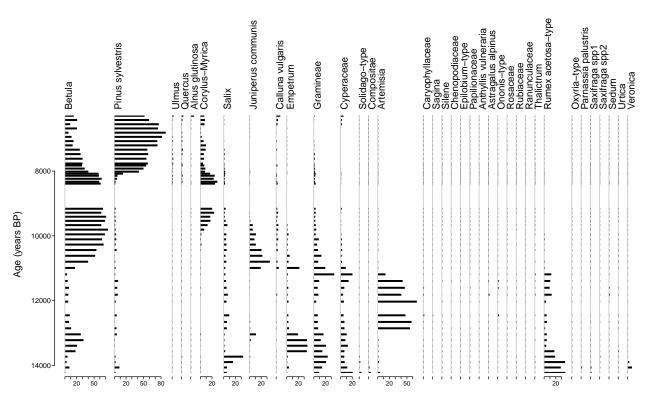
These are percentage data so we scale the diagram for percentages and plot on an age scale by passing the name of the appropriate column to style yvar.name. riojaPlot will take the y-axis from the first column of aber.chron by default.

scale for percentage data
riojaPlot(aber.poll, aber.chron,
 yvar.name="Age (years BP)",
 scale.percent=TRUE)



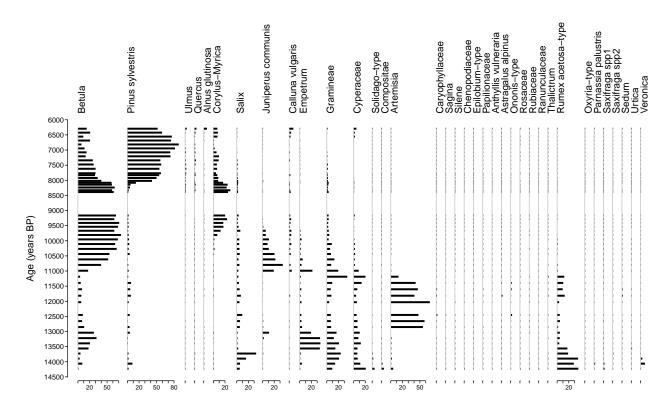
Or plot as bars.

```
# turn of silhouettes and lines, increase bar thickness
riojaPlot(aber.poll, aber.chron,
   yvar.name="Age (years BP)",
   scale.percent=TRUE,
   plot.poly=FALSE,
   plot.line=FALSE,
   lwd.bar=3,
   col.bar="black")
```



The y-axis doesn't cover the whole range so we set the upper and lower limits manually.

```
# Specify y-axis limits
sapply(aber.chron, range)
#>
        Depth (cm) Age (14C years BP) Age (years BP)
#> [1,]
               300
                                  5515
                                                  6295
#> [2,]
               550
                                 12147
                                                14229
riojaPlot(aber.poll, aber.chron,
   yvar.name="Age (years BP)",
   ymin=6000, ymax=14500, yinterval=500,
   scale.percent=TRUE,
   plot.poly=FALSE,
  plot.line=FALSE,
   lwd.bar=3,
   col.bar="black")
```

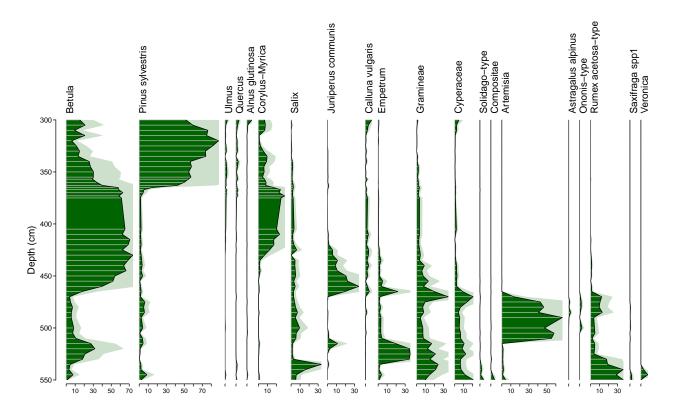


3. Selecting variables to plot

By default riojaPlot will plot all columns in the data frame. In this example there are many rare taxa: we could either remove these from the data frame prior to plotting or pass a character vector of column names to plot to the argument selVars. Here we only plot taxa with max abundance greater than 5%. We also add exaggeration curves. These are shown as silhouettes with a multiplication factor of 2 by default by can be changed to lines with different exaggeration using styles exag.mult, col.exag, and col.line.exag. col.exag="auto" will use a transparent colour the same as the main curve.Useexag.alpha' to change transparency.

```
# reduce number of taxa and add exaggerations
# make a vector of taxon names with max abundance greater that 2 percent
mx <- apply(aber.poll, 2, max)
aber.sel <- names(mx[mx > 2])

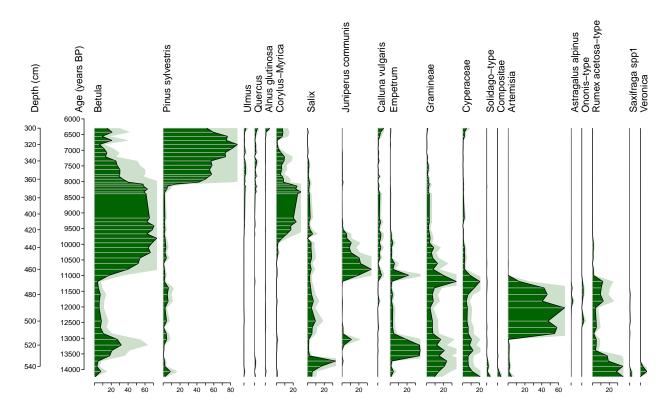
riojaPlot(aber.poll, aber.chron, aber.sel,
    scale.percent=TRUE,
    exag=TRUE)
```



4. Plotting a secondary y-axis

To show a secondary y-axis simply pass the column name of the secondary y-axis variable to style sec.yvar.name. In this example we use age (years BP) for the primary axis and depth for the secondary axis and set plot.sec.axis to TRUE. We also use ymin, ymax and yinterval to fine-tune the start, end, and increment of the y-axis.

```
riojaPlot(aber.poll, aber.chron, aber.sel,
   yvar.name="Age (years BP)",
   sec.yvar.name="Depth (cm)",
   plot.sec.axis=TRUE,
   ymin=6000,
   ymax=14300,
   yinterval=500,
   scale.percent=TRUE,
   exag=TRUE)
```



5. Showing groups

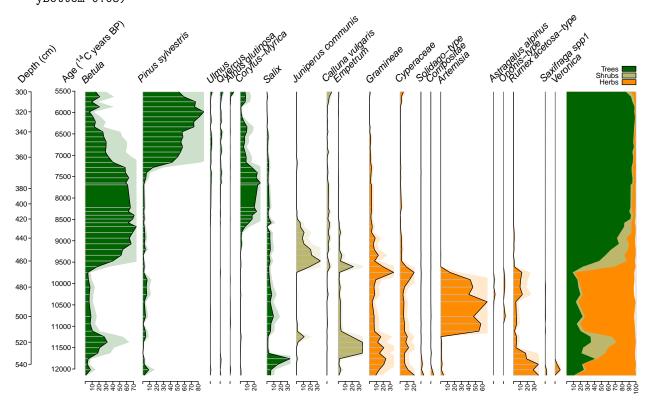
Individual can be assigned to a group and each group assigned a different colour. Cumulative plots can also be added showing the sum of each group.

To assign variables to groups pass a data frame of two columns, the first with the name of each variable (spelt exactly the same as in the data) and a column of group memberships. This should be a factor with the levels determining the order they appear in the legend. The names of the groups will be used in the cumulative plot legend so choose them wisely.

We can also italicise the names using the style names.italicise, rotate them with srt.xlabel, and specify a label for the age variable to override the column name. Here we plot a 14C age scale and use an expression to create a label with superscriped 14C.

```
# group taxa by type and add cumulative graph
# extract types
aber.types <- aber$names[, -1]</pre>
# convert pollen types to a factor
aber.types$Group <- factor(aber.types$Group, levels=c("Trees", "Shrubs", "Herbs"))
ylab <- expression(Age~"("*""^{14}*C~years~BP*")")</pre>
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types,
   yvar.name="Age (14C years BP)",
   sec.yvar.name="Depth (cm)",
   ylabel=ylab,
   plot.sec.axis=TRUE,
   ymin=5500,
   ymax=12200,
   yinterval=500,
   scale.percent=TRUE,
   plot.groups=TRUE,
```

```
plot.cumul=TRUE,
exag=TRUE,
names.italicise = TRUE,
srt.xlabel=45,
las.axis=2,
yBottom=0.05)
```

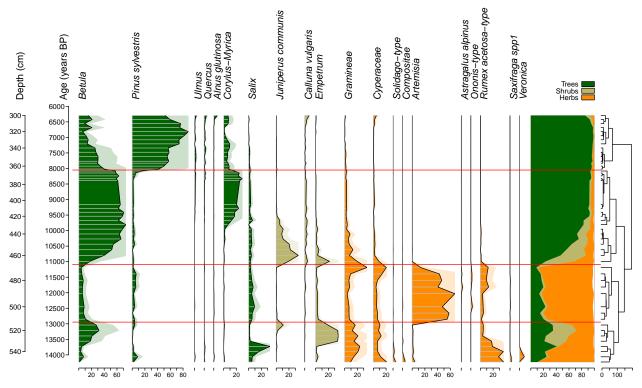


6. Add a zonation

We can add a zonation to the plot in 3 ways. The first is to set the styles do.clust and plot.clust and / or plot.zones to "auto" or the number of zones to show. This will perform a constrained clustering using chclust and either add a dendrogram and/or zone lines to the diagram. The number of zones can be estimated using a broken stick model (plot.zones="auto" see ?rioja::chclust for details) or specified (e.g. plot.zones=3). Colour of the zone lines can be changed with col.zones. The zonation can be based on all columns in the data (style clust.use.selected=FALSE) or only the displayed columns if only a subset are selected using selVars. Data can optionally be transformed (clust.data.trans="sqrt"), scaled (clust.data.trans="scale") prior to clustering or used as is (clust.data.trans="none", the default). Change the width of the dendrogram using style clust.width=N where N is the required width as a fraction of the whole diagram width.

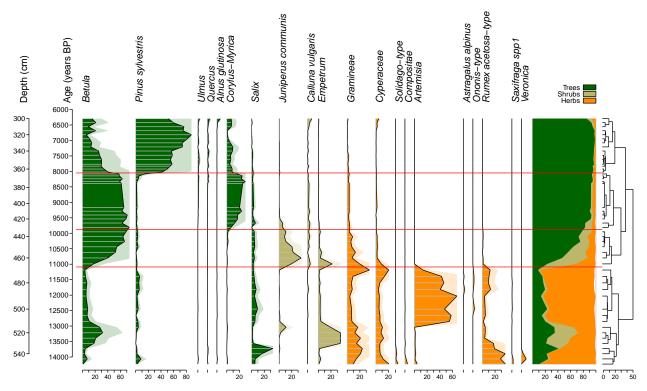
```
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types,
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500,
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
   plot.groups=TRUE,
   plot.cumul=TRUE,
```

```
exag=TRUE,
do.clust=TRUE,
clust.data.trans="sqrt",
plot.clust=TRUE,
plot.zones="auto",
names.italicise = TRUE)
```



We can also generate a constrained cluster object and pass this to riojaPlot for plotting. Here we log10-transform the data prior to clustering.

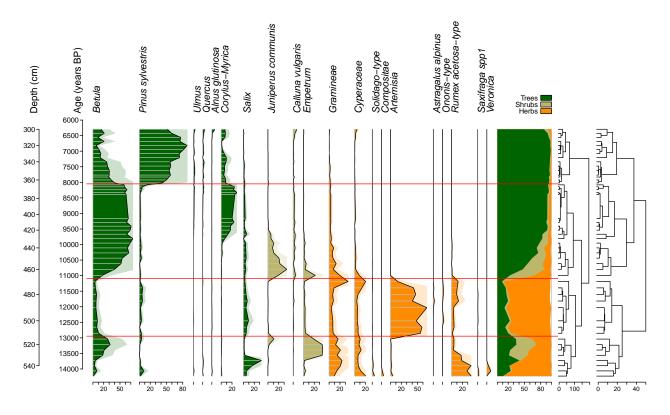
```
aber.clust <- chclust(dist(log10(aber.poll+1)))</pre>
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types, clust=aber.clust,
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax = 14300,
   yinterval=500,
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
   plot.groups=TRUE,
   plot.cumul=TRUE,
   exag=TRUE,
   clust.data.trans="sqrt",
   plot.clust=TRUE,
   plot.zones="auto",
   names.italicise = TRUE)
```



Finally, we can add a zonation at a different position in the diagram. Here we add the zonation based on log10-transformed data to the original diagram with a sqrt-transformed zonation for comparison. In this case we set the right-hand limit of the original diagram with xRight and add the additional zonation to this space. This is a bit of a contrived example but adding the zonation separately to the diagram can be useful if we are plotting multiple datasets and want the zonation on the far-right (see example in section 8).

```
aber.clust <- chclust(dist(log10(aber.poll+1)))</pre>
rp <- riojaPlot(aber.poll, aber.chron, aber.sel, aber.types,</pre>
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500,
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
   plot.groups=TRUE,
   plot.cumul=TRUE,
   exag=TRUE,
   clust.data.trans="sqrt",
   do.clust=TRUE,
   plot.clust=TRUE,
   plot.zones="auto",
   names.italicise = TRUE,
   xRight = 0.9)
```

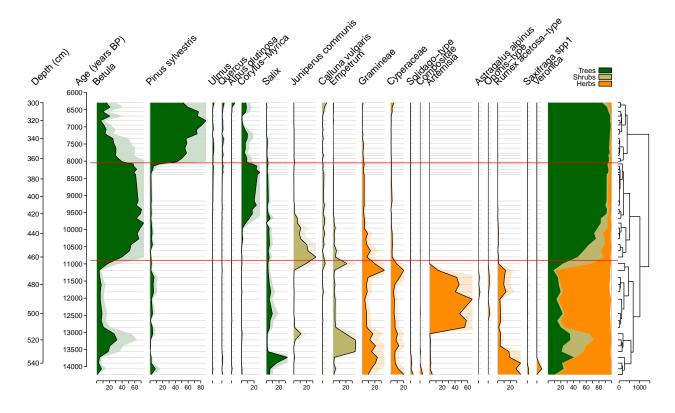
addRPClust(rp, aber.clust, xLeft=0.91, xRight=0.99)



7. Reusing styles

Save settings as a style and apply to figure. Here we also full-width plot bars behind the curves.

```
mystyle <- makeStyles(</pre>
   plot.sec.axis=TRUE,
   scale.percent=TRUE,
   plot.groups=TRUE,
   plot.cumul=TRUE,
   exag=TRUE,
   plot.poly=TRUE,
   plot.bar="full",
   bar.back=TRUE,
   lwd.bar=0.5,
   col.bar="lightgrey",
   do.clust=TRUE,
   plot.clust=TRUE,
   plot.zones="auto",
   srt.xlabel=45)
riojaPlot(aber.poll, aber.chron, aber.sel, aber.types, mystyle,
   sec.yvar.name="Depth (cm)",
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax=14300,
   yinterval=500)
```



8. Combining different datasets in a single diagram

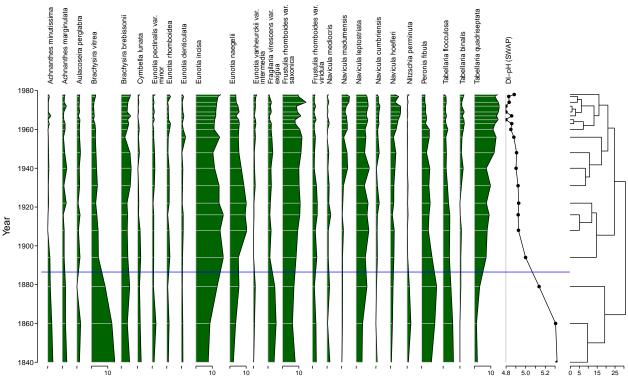
We may want to plot different datasets in the same figure. For example, we may want to plot one set of variables with percentage scaling and the other with normal scaling where each variable is the same width. Or we may have two or more datasets measured at different depths of ages.

To combine different datasets we plot the first with function riojaPlot and the others with function riojaPlot2. We also need to specify the right-hand position of each plot in fractions of the page width.

```
# Diatom data the Round Loch of Glenhead, Galloway, SW Scotland
# Shows recent acidification and small recovery
data(RLGH)
RLGH.names <- RLGH$names
RLGH.diat <- RLGH$spec
# plot only common taxa
mx <- apply(RLGH.diat, 2, max)</pre>
RLGH.sel <- colnames(RLGH.diat)[mx > 2]
RLGH.chron <- RLGH$depths
# data has 210Pb age based on years before coring (1980)
# Add new column with Years (CE)
RLGH.chron$Year <- 1980 - RLGH.chron$Age
# do a pH reconstruction using SWAP modern dataset (see ?rioja::SWAP)
data(SWAP)
# generate a WA model
SWAP.wa <- WA(SWAP$spec, SWAP$pH)
RLGH.pH <- predict(SWAP.wa, RLGH$spec)</pre>
# convert to data frame with single column
RLGH.pH <- data.frame(`DI-pH (SWAP)`=RLGH.pH$fit[, 1], check.names=FALSE)
rp <- riojaPlot(RLGH.diat, RLGH.chron,</pre>
```

```
yvar.name="Year",
                          scale.percent=TRUE,
                         y.rev=FALSE,
                         selVars=RLGH.sel,
                          ymax=1980,
                         x.names=RLGH$names$TaxonName,
                          cex.xlabel=0.7,
                         names.break.n=25,
                          xRight=0.9)
riojaPlot(RLGH.pH, RLGH.chron[, "Year", drop=FALSE],
                            riojaPlot=rp,
                            scale.minmax=FALSE,
                            plot.bar=FALSE,
                            plot.symb=TRUE,
                            symb.cex=0.6)
                 Achnanthes minutissima
                                                                                                                                 Frustulia rhomboides var.
viridula
                       Achnanthes marginulata
                                                                                                                                                                                                      Tabellaria quadriseptata
                                                                                                        Eunotia vanheurckii var.
intermedia
                                                                                                               -ragilaria virescens var
                             Aulacoseira perglabra
                                                Brachysira brebissonii
                                                                                                                                              Vavicula madumensis
                                                                                                                                                            Navicula cumbriensis
                                                                                                                                                                                        Fabellaria flocculosa
                                                                                                                                                    Vavicula leptostriata
                                                                   Eunotia rhomboidea
                                                                                                                                                                         Vitzschia perminuta
                                                                                                                                        Vavicula mediocris
                                                                         Eunotia denticulata
                                                             Eunotia pectinalis v
minor
                                                                                                                                                                                                Tabellaria binalis
                                   Brachysira vitrea
                                                       Cymbella lunata
                                                                                               Eunotia naegelii
                                                                                                                                                                  Vavicula hoefleri
                                                                                                                                                                                                                    DI-pH (SWAP)
                                                                               Eunotia incisa
                                                                                                                                                                               Peronia fibula
      1980
      1960
      1940
      1920
Year
      1900
      1880
      1860
```

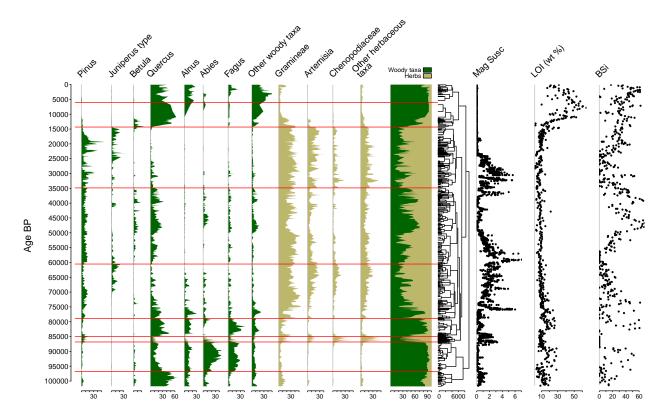
Here we add a zonation to the right of the above figure. There are only two significant zones in these data.



In the next example we combine pollen, magnetic susceptibility, LOI and biogenic silica data recorded in a core from Lago Grande di Monticchio and published by Allen et al. (1999). The core spans the last c. 100 kyr and the four datasets are measured at different depth intervals and resolutions. The data are downloaded from the NOAA paleoclimatology data archive (https://www.ncei.noaa.gov/products/paleoclimatology).

```
library(readxl)
library(dplyr)
# Import the data, assign pollen types to groups
fpath <- system.file("extdata/allen1999.xlsx", package="riojaPlot")
pollen <- read_excel(fpath, sheet="Pollen data", skip=2)
pollen.chron <- pollen %>% select(1)
pollen <- pollen %>% select(Pinus:`Other herbaceous taxa`)
types <- data.frame(Name=colnames(pollen), Group="Woody taxa")
types$Group[9:12] <- "Herbs"
types$Group <- factor(types$Group, levels=c("Woody taxa", "Herbs"))
mag <- read_excel(fpath, sheet="Magnetic susceptibility", skip=2)
mag.chron <- mag %>% select(1)
mag <- mag %>% select(`Mag Susc`) %>% mutate(`Mag Susc` = `Mag Susc` / 1000)
loi <- read_excel(fpath, sheet="Loss on ignition", skip=2)</pre>
```

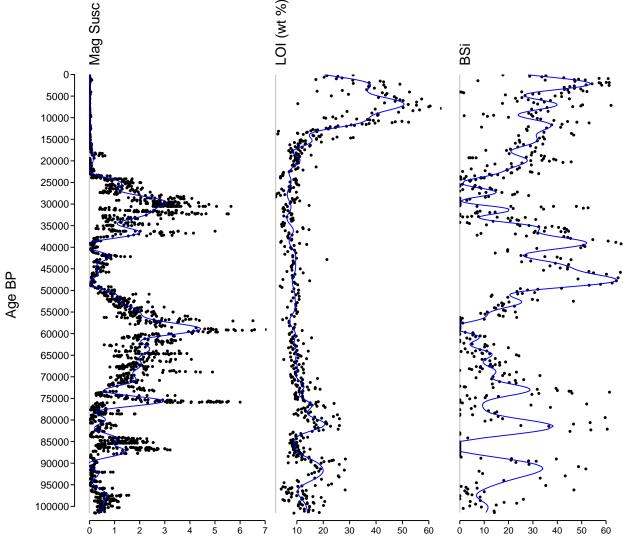
```
loi.chron <- loi %>% select(1)
loi <- loi %>% select("LOI (wt %)")
BSi <- read_excel(fpath, sheet="Biogenic silica", skip=2)
BSi.chron <- BSi %>% select(1)
BSi <- BSi %>% select("BSi")
rp1 <- riojaPlot(pollen, pollen.chron, groups=types,</pre>
          yinterval = 5000,
          ymin = 0,
          ymax=102000,
          yvar.name = "Age BP",
          scale.percent=TRUE,
          plot.groups=TRUE,
          do.clust = TRUE,
          plot.zones = "auto",
          plot.clust=TRUE,
          plot.cumul=TRUE,
          cex.cumul=0.6,
          srt.xlabel=45,
          xSpace = 0.01,
          plot.bar=FALSE,
          tcl=-0.1,
          cex.yaxis=0.7,
          cex.xlabel=0.8,
          xRight = 0.7,
          plot.line=FALSE
rp2 <- riojaPlot(mag, mag.chron[, "Age BP", drop=FALSE],</pre>
           riojaPlot=rp1, xGap = 0.01,
           xRight=0.8, scale.minmax=FALSE,
           plot.bar=FALSE, plot.line=F,
           plot.symb=TRUE, symb.cex=0.3)
rp3 <- riojaPlot(loi, loi.chron[, "Age BP", drop=FALSE],</pre>
           riojaPlot=rp2,
           xRight=0.9,
           scale.minmax=FALSE, plot.bar=F,
          plot.line=F, plot.symb=TRUE, symb.cex=0.3)
riojaPlot(BSi, BSi.chron[, "Age BP", drop=FALSE],
           riojaPlot=rp3,
           xRight=0.99,
           scale.minmax=FALSE, plot.bar=FALSE,
           plot.line=FALSE, plot.symb=TRUE, symb.cex=0.3)
```



9. Adding custom plotting functions

In the above example the physical / chemical variables are measured on a large number of samples and we plot with symbols because these display the variability in the data more clearly than lines. It would be useful to fit a smooth through these data to highlight the trends. Here we plot the three phys/chem variables and fit a gam to each curve using a custom function to add the smooth to each plot.

```
fun.gam <- function(x, y, i, nm) {</pre>
  tmp <- data.frame(x=y, y=x)</pre>
  gam \leftarrow mgcv::gam(y \sim s(x, k=50), data=tmp)
  x2 <- predict(gam, type="response")</pre>
  lines(x2, y, col="blue", lwd=1)
}
rp <- riojaPlot(mag, mag.chron[, "Age BP", drop=FALSE],</pre>
           yinterval = 5000,
           ymin = 0,
           ymax=102000,
           xRight=0.3,
           scale.minmax=FALSE,
           plot.bar=FALSE,
           plot.line=F,
           plot.symb=TRUE,
           plot.poly=FALSE,
           symb.cex=0.3,
           fun.xfront=fun.gam)
rp1 <- riojaPlot(loi, loi.chron[, "Age BP", drop=FALSE],</pre>
            riojaPlot=rp,
            xRight=0.5,
```



10. Frequently asked questions

1. How do I change the order of the variables in the diagram?

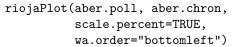
There are several ways to do this. Variables are plotted in the order they appear in the data, so rearrange the order of variables in the original data. dplyr::select offers a convenient way to do this:

```
data.reordered <- data %>% select(var3, var1, var5) # and so on...
```

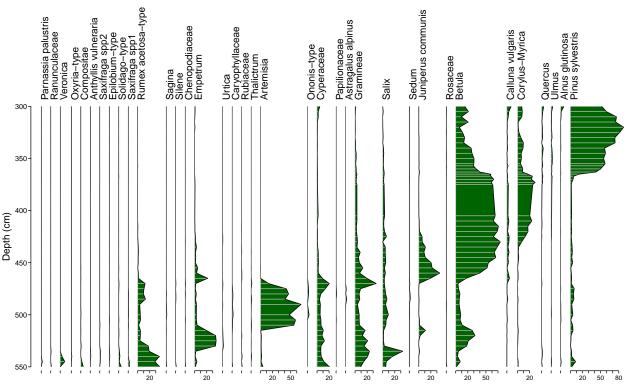
Another way is to create a character vector wi the variables in the required order and pass this to selVars:

```
myorder <- c("var3", "var1", "var5") # and so on...
riojaPlot(spec, chron, selVArs=myorder)</pre>
```

A quick way to sort variables accorging to their position in the diagram is to set the style wa.order="bottomleft" (or use bottomright for the opposite). This will sort variables from those common at the base of the core on the left to those common at the top on the right.

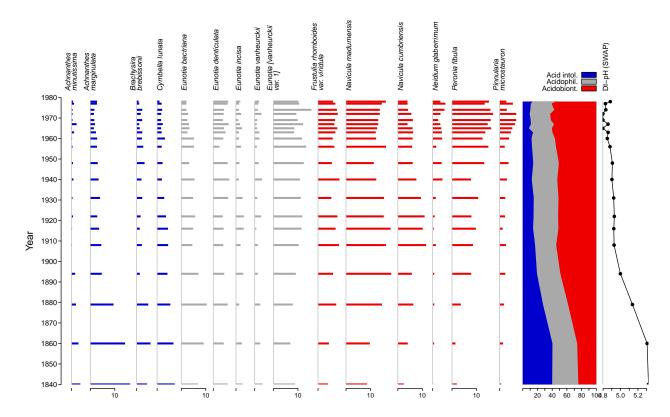


select(CODE, Group)



To have complete control you probably need to use option 1 - change the order in the original data. Here is an extended example in which we sort the taxa in the RLGH diatom data according to their pH optima. We also group the taxa into three pH classes based on their pH optima and display the groups and cumulative plot.

```
RLGH.diat <- RLGH.diat %>% select(optima2$CODE)
mx <- apply(RLGH.diat, 2, max)</pre>
RLGH.sel <- colnames(RLGH.diat)[mx > 3]
rp <- riojaPlot(RLGH.diat, RLGH.chron, groups=optima2,</pre>
          yvar.name="Year",
          scale.percent=TRUE,
          y.rev=FALSE,
          selVars=RLGH.sel,
          ymax=1980,
          yinterval=10,
          x.names=RLGH$names$TaxonName,
          names.italicise=TRUE,
          cex.xlabel=0.7,
          plot.poly=FALSE,
          plot.line=FALSE,
          lwd.bar=3,
          xRight=0.9,
          plot.groups=TRUE,
          plot.cumul=TRUE,
          col.group=c("mediumblue", "darkgrey", "red2"),
          cumul.mult=0.3
          )
riojaPlot(RLGH.pH, RLGH.chron[, "Year", drop=FALSE],
           riojaPlot=rp,
           scale.minmax=FALSE,
           plot.bar=FALSE,
           plot.symb=TRUE,
           symb.cex=0.6)
```



2. How do I change the font sizes?

Use the following styles (note, font sizes are specified relative to the default size of 1): - cex.axis = font size of x-axis values (default 0.6) - cex.yaxis = font size of y-axis values (default 0.7) - cex.xlabel = font size of x-axis labels (default 0.9) - cex.ylabel = font size of y-axis labels (default 0.9) - cex.cumul = font size of group names in cumulative plot (default 0.7)

3. How do I change the limits and labels of the of the y-axes?

Use the style y.rev to reverse the scale of y-axis (default is TRUE, ie. values increase from top to bottom). Use the styles ymin, ymax and yinterval to change the start, end and interval values. The corresponding styles sec.ymin, sec.ymax and sec.yinterval control the secondary y-axis scale.

4. My variable names are too long and take up too much space at the top of the diagram.

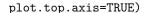
Long names can be split by setting style names.break.long=TRUE (the default). Names will be split so no part if greater than names.break.n characters (default 20).

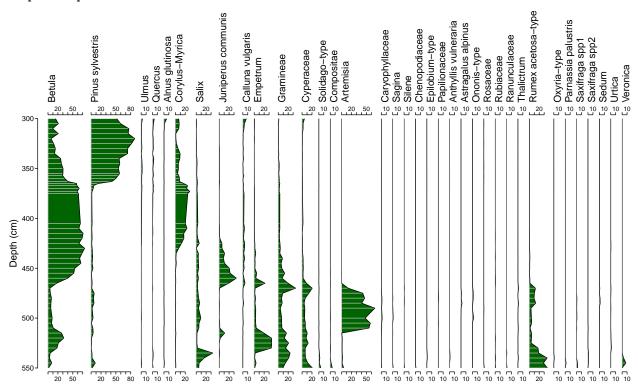
5. How do I add a second x-axis at the top of the plots?

Styles plot.top.axis and plot.bottom.axis allow plotting of the top and bottom x-axes. If there is not enough space between the top axis and the taxon names use style ylabPos to fine-tune the gap (default 0.1).

```
data(aber)
aber.poll <- aber$spec
colnames(aber.poll) <- aber$names$Name
aber.chron <- aber$ages

riojaPlot(aber.poll, aber.chron,
    scale.percent=TRUE,
    min.width.pc=10,</pre>
```



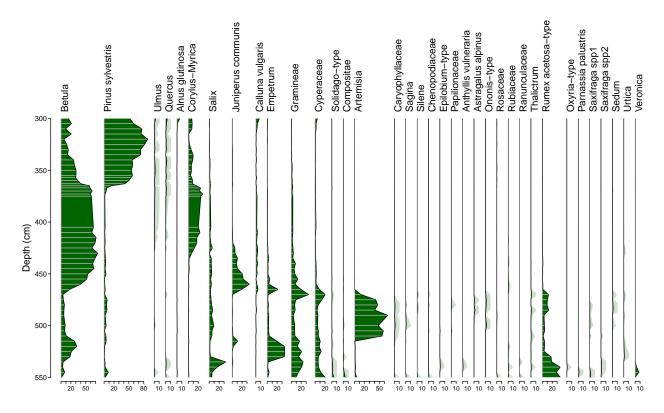


6. How do I add exaggeration curves to selected taxa only?

Style exag takes a single value (TRUE or FALSE) to add exaggerations to all curves but can also take a logical vector of length equal to the number of columns in the x-variables data frame to control exaggerations for each variable. The following example shows how to add exaggerations for pollen types with maximum relative abundance < 5 percent. We also change the exaggeration multiplier to 10 and set the min widths of curves to 10% (default is 5%).

```
aber.poll <- aber$spec
colnames(aber.poll) <- aber$names$Name
aber.chron <- aber$ages
mx <- apply(aber.poll, 2, max)
exag.sel <- mx < 5

riojaPlot(aber.poll, aber.chron,
    scale.percent=TRUE,
    exag=exag.sel,
    exag.mult=10,
    min.width.pc=10)</pre>
```



7. How I plot different styles for different variables?

In this example we use geochemical data from Laguna del Maule published by Frugone-Álvarez et al. (2020) and downloaded from the NOAA Paleoclimate data archive (https://www.ncei.noaa.gov/products/paleo climatology). The data we plot here contain 9 geochemical variables. Some (TS, TC, TIC and TOC) are measured at every interval but others only at selected depths. We plot magnetic susceptibility with symbols, TS, TC, TIC, TOC and BioSi with lines, and d15N and d13C with lines and symbols.

```
fpath <- system.file("extdata/maule2020geochem.txt", package="riojaPlot")</pre>
maule <- readr::read delim(fpath, skip=158, show col types = FALSE)
maule.data <- maule %>% select(-(1:4))
maule.chron <- maule %>% select(1:4)
# How many measurements do we have for each variable:
unlist(lapply(maule.data, function(x) sum(!is.na(x))))
#>
      MS
            TS
                        TIC
                               TOC BioSi
                                            TN
                                                d15N
                                                       d13C
           267
#>
     146
                  267
                        267
                               267
                                     253
                                           108
                                                   57
                                                         57
selsymb <- rep(FALSE, 9)
selsymb[c(1, 8:9)] \leftarrow TRUE
selbar <- rep(FALSE, 9)
selline <- rep(TRUE, 9)
selline[1] <- FALSE
riojaPlot(maule.data, maule.chron,
          yvar.name="age_BP",
          ymin=-100,
          ymax=13300,
          yinterval=500,
```

```
plot.bar=FALSE,
             plot.poly=FALSE,
             plot.line=selline,
             plot.symb=selsymb,
             plot.groups=TRUE,
             ylabPos=2,
             symb.cex=0.5,
   400
   900
  1400
  1900
  2400
  2900
  3400
  3900
  4400
  4900
  5400
ದ್ದ <sup>5900</sup>
6900
6900
  7400
  7900
  8400
  8900
  9400
  9900
 10400
 10900
```

To also show the variables in different colours we need to group them.

0.0

```
groups <- data.frame(names=colnames(maule.data), groups=c(1, 2, 2, 2, 2, 2, 2, 3, 3)) riojaPlot(maule.data, maule.chron, groups=groups,
```

16 20 24

0.2

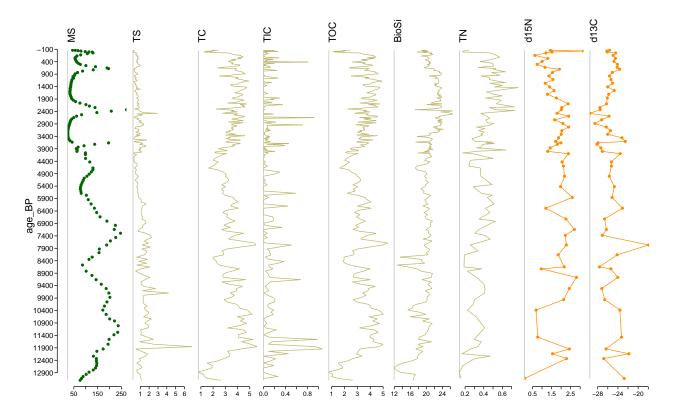
0.4 0.6

2.5

```
yvar.name="age_BP",
ymin=-100,
ymax=13300,
yinterval=500,
plot.bar=selbar,
plot.poly=FALSE,
plot.line=selline,
plot.symb=selsymb,
plot.groups=TRUE,
ylabPos=2,
symb.cex=0.5,
)
```

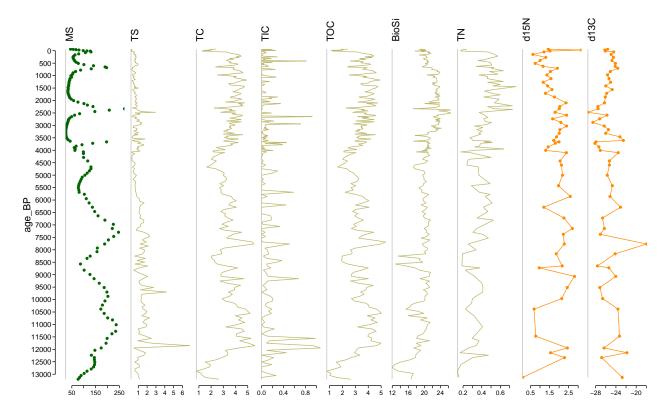
1 2 3 4 5 6

11400 · 11900 · 12400 · 12900 · 1



8. How do I control the tick values on the y-axis?

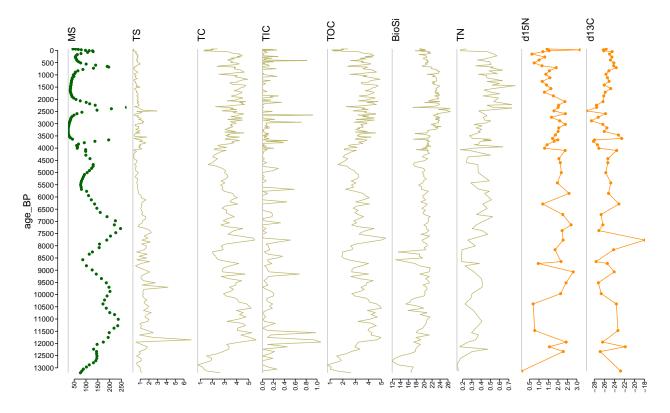
riojaPlot will try to add a y-axis with a sensible upper and lower limit and interval. It doesn't always suceed. If the default doesn't work, try specifying values for ymin, ymax and yinterval (and corresponding sec.ymin etc. value for the seconday axis). If this still doesn;t work you can pass a numeric vector of "tick values" to styles ytks1 and ytks2 for the primary and secondary axes respectively. In the plot above the y-axis values start at -100 and increment in 500 which gives unusal values. To fix this we create a vector of tick values from 0 to 13000 and use this.



9. How do I rotate the tick values on the x-axes?

 $\begin{tabular}{ll} USe the style {\tt las=2}. You may also need to create a larger space at the bottom of the figure using yBottom. \\ \end{tabular}$

```
riojaPlot(maule.data, maule.chron, groups=groups,
          yvar.name="age_BP",
          ymin=-100,
          ymax = 13300,
          yinterval=500,
          plot.bar=FALSE,
          plot.poly=FALSE,
          plot.line=selline,
          plot.symb=selsymb,
          plot.groups=TRUE,
          ylabPos=2.2,
          symb.cex=0.5,
          ytks1=myticks,
          las.axis=2,
          yBottom=0.06
          )
```

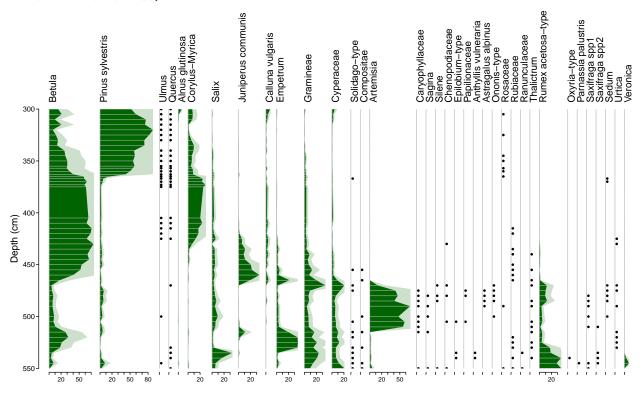


10. How do I plot symbols for rare types instead of bars or curves?

To do this we supply plot.poly etc. with a logical vector to control the plotting of each curve then set the values of these styles to FALSE for the rare types. Then we define a function to plot symbols for the rare types and create a list of functions to plot symbols only for the rare types.

```
aber.poll <- aber$spec
colnames(aber.poll) <- aber$names$Name</pre>
aber.chron <- aber$ages
# calculate of max of each column
mx <- apply(aber.poll, 2, max)</pre>
#create a logical vector which is TRUE for taxa with max < 5</pre>
sel \leftarrow mx < 5
# define a custom function to plot symbols
symb.fun <- function(x, y, i, nm) {</pre>
   sel <- x > 0
   if (sum(sel) > 0) {
      points(rep(3, sum(sel)), y[sel], cex=0.4, pch=19)
}
# create a list of functions of length equal to the number of columns in the data
funlist <- lapply(1:ncol(aber.poll), function(x) symb.fun)</pre>
# now set the elements of the list where we don;t want to plot symbols to NULL
funlist[!sel] <- list(NULL)</pre>
# plot silhouettes and lines for taxa > 5% and apply our function to the others
riojaPlot(aber.poll, aber.chron,
   scale.percent=TRUE,
   plot.poly=!sel,
```

plot.bar = !sel,
plot.line=FALSE,
lwd.bar=0.6,
exag=TRUE,
fun.xfront=funlist)



11. How do I change the widths of individual graphs?

In the above example we plotted rare types as symbols. In the example we can expand the x-axis for rare types.

For percentage diagrams the width of each graph is scaled uniformly so the percentage values for each graph are the same. We can change these using the style graph.widths which takes a vector of relative widths. We can also change the x-axis labels for each graph by supplying a numeric vector to style x.inc.pc to set the interval for x-values. Here we hide columns with maximum relative abundance < 2%, and show those taxa with max < 5% as curves with 5 * normal width, and set the x-interval for these to 1% (rather than the default of 10%). We could, of course, chose the columns manually to have complete control over what is hidden and shown with expanded axes.

```
mx <- apply(aber.poll, 2, max) # calculate max % of each column
widths <- rep(1, ncol(aber.poll)) # generate numeric vector of widths, set to 1
inc <- rep(10, ncol(aber.poll)) # generate numeric vector of x-intervals, set to 10
selTaxa <- names(mx[mx > 2]) # generate character vector of column names to include in the figure
sel <- which(mx < 5) # generate a logical vector of columns to expand
widths[sel] <- 5 # set widths to 5 times normal width
inc[sel] <- 1 # set x-interval to 1%
riojaPlot(aber.poll, aber.chron, selVars=selTaxa,</pre>
```

scale.percent=TRUE,
graph.widths=widths,
min.width.pc=5,

```
x.pc.inc=inc,
                                     cex.axis=0.5,
                                     las.axis=2)
                                                                                                                                                                                                                                                                                                                 Rumex acetosa-type
                                                                                                                                                         Juniperus communis
                                                                                                                                                                                                                                                                                       Astragalus alpinus
                                                                                                                                                                           Calluna vulgaris
Empetrum
                                                     Pinus sylvestris
                                                                                                                     Alnus glutinosa
Corylus-Myrica
                                                                                                                                                                                                                                                                                                                                  Saxifraga spp1
                                                                                                                                                                                                                               Solidago-type
                                                                                                                                                                                                                                                                                                    Ononis-type
                                                                                                                                                                                                                   Cyperaceae
                                                                                                                                                                                                                                            Compositae
                                                                                                                                                                                                  Gramineae
                                                                                                                                                                                                                                                         Artemisia
                                                                                                                                                                                                                                                                                                                                                Veronica
                                                                                                       Quercus
                                                                                          Ulmus
                     Betula
       350
Depth (cm)
       450
```

We can also change the widths for non-percent data. Here we return to the Laguna del Maule data in FAQ 7 and change the width of d15N and d13C columns to twice the standard width.

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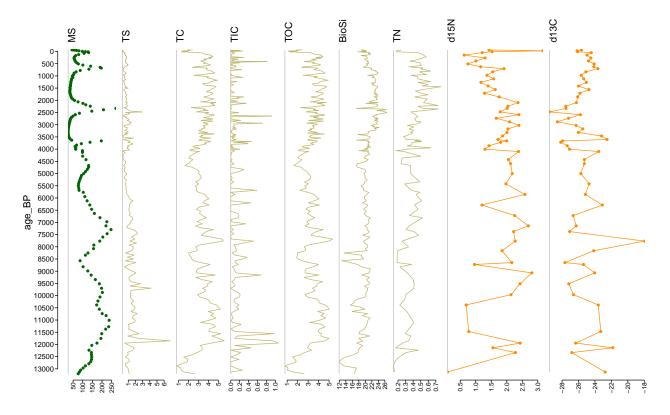
525

```
widths <- rep(1, ncol(maule.data))</pre>
#sel <- names(maule.data) %in% c("d15N", "d13C")</pre>
#widths[sel] <- 2</pre>
widths[8:9] <- 2
riojaPlot(maule.data, maule.chron, groups=groups,
          yvar.name="age_BP",
          ymin=-100,
          ymax = 13300,
          yinterval=500,
          plot.bar=FALSE,
          plot.poly=FALSE,
          plot.line=selline,
          plot.symb=selsymb,
          plot.groups=TRUE,
          ylabPos=2.2,
          symb.cex=0.5,
          ytks1=myticks,
          las.axis=2,
          graph.widths=widths,
          yBottom=0.06
```

\(\frac{1}{4}\) \(\frac{1}{4}

500

550-



12. How do I plot surface pollen or diatom data?

riojaPlot can plot surface sediment data, you just need to think how to arrange samples on the y-axis. The example below plots surface sediment diatom data from small ponds and pools in SE England collected by Bennion (1994) (see ?rioja::Ponds) for details. Here we arrange the samples along the pH gradient and plot the data against Lake name with TP as a secondary y-axis.

```
data(Ponds)
# reorder the rows in decreasing TP
o <- order(Ponds$env$TP, decreasing=TRUE)</pre>
ponds.diat <- Ponds$spec[o, ]</pre>
ponds.TP <- data.frame(TP=round(Ponds$env$TP[o]), LakeName=Ponds$env$Name[o])</pre>
# replace taxon codes with names
colnames(ponds.diat) <- Ponds$names$Name</pre>
# remove rare species
mx <- apply(ponds.diat, 2, max)</pre>
ponds.sel <- colnames(ponds.diat)[mx > 10]
# With a large number of samples we would use points but with only 30
# in this data we use bars. Although the data are arranged in order of
# increasing TP they are not a temporally or spatially contiguous so
# lines or silhouettes are not appropriate.
riojaPlot(ponds.diat, ponds.TP, selVars=ponds.sel,
          yvar.name="LakeName",
          sec.yvar.name="TP",
          sec.yinterval = 20,
          plot.sec.axis=TRUE,
          scale.percent=TRUE,
          plot.poly=FALSE,
```

```
plot.line=FALSE,
                 plot.bar=TRUE,
                 col.bar="black",
                 lwd.bar = 2,
                 symb.cex=0.4,
                 cex.xlabel=0.6,
                 cex.yaxis=0.6,
                 wa.order="topleft",
                 names.italicise=TRUE,
                 las.axis=2,
                  cex.axis=0.5)
 ᅀ
                 Wiremill Lak
                  Abbey Lake
585
485
            Marsworth Reservoir
425
345
325
                  Fleet Pond
               Albury Mill Pond
              Bonnington Lake
305
285
265
             Lily Lake, Matching
              Sissinghurst Lake
225
205
           Lower St. Clere Lake
               Cinder Hill Lake
185
                Hartwell Pond
          Grendon Quarter Pond
165
           Hemingford Park Lake
145
                 Debden Lake
125
                 Blue Lagoon
           Mitchams Pit. Burwell
                Hammer Pond
105
            Beachborough Lake
              Childerditch Pond
                Farthings Lake
```

13. How do I plot different levels or samples with a different colour?

क्षें के के किया के क्षें क्षें क्षें क्षें के के किया के किया के

Eleven Acre Lake Frensham Little Pond

Fowlmere Spring Shortheath Pond Branches Park Lake

Boldermere North Pool, Marsh Works Wintry Wood Lake Decoy Lake, Arlington

65

45

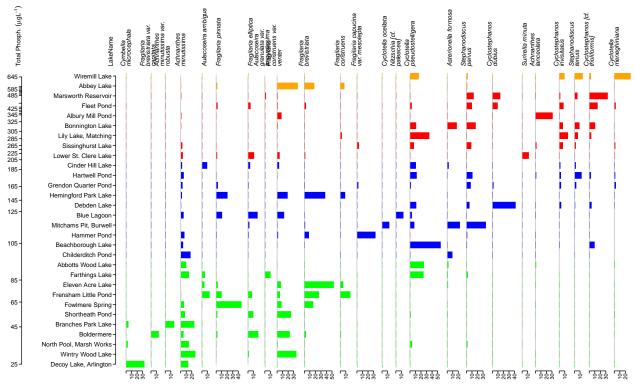
It is currently only possible to plot different samples with different coloured bars. Here we extend the example for FAQ 12 above and classify the samples into different groups based on their TP, then plot these different samples with different colours. We do this by create a character vector of colour names, that corresponds to the samples, then pass this to style col.sep.bar and set sep.bar to true. We also create an expression to label the TP axis in the correct units.

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```
sec.vinterval = 20,
plot.sec.axis=TRUE,
scale.percent=TRUE,
plot.poly=FALSE,
plot.line=FALSE,
plot.bar=TRUE,
col.bar="black",
sep.bar=TRUE,
col.sep.bar=ponds.TP$bar.cols,
lwd.bar = 10,
symb.cex=0.4,
cex.xlabel=0.6,
cex.yaxis=0.6,
wa.order="bottomleft",
names.italicise=TRUE,
las.axis=2,
cex.axis=0.5)
```

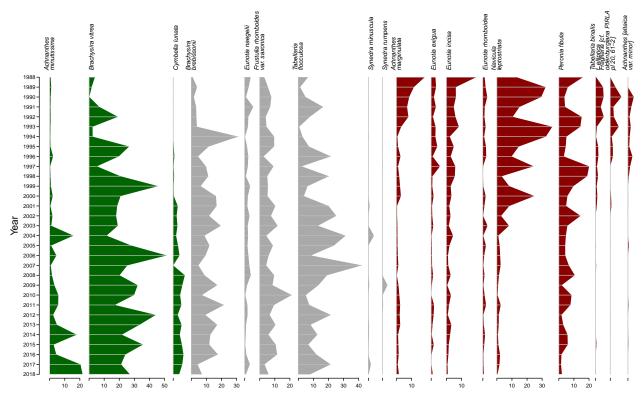


14. How do I plot ecological time series?

riojaPlot can also be used to plot multi-species ecological time series (or spatial transect) data. Here we use diatom data from the UK Upland Waters Monitoring Network (https://uwmn.uk/). The dat at hand are epilithic diatoms samples collected annually from 1988-2018 from Loch Chon (Trossachs, Scotland). Loch Chon has been previously acidified as a result of acid precipitation, these data were collected to monitor the potential recovery of sites from this previous acidification. Also with the diatom counts (expressed as relative abundance) a trend test based on a multivariate generalised linear model using the manyglm function in package mvabund has been conducted to identify taxa that have a significant increasing or decreasing trend.

We first re-order the taxa according to their trend over time and plot the data with silhouettes, coloured to show taxa with significant increasing, decreasing or no trend.

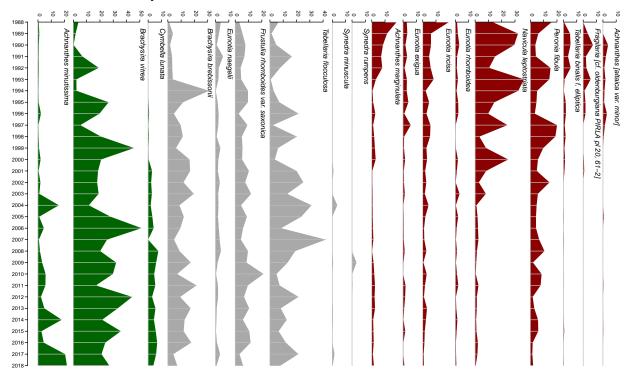
```
fpath <- system.file("extdata/LochChon.xlsx", package="riojaPlot")</pre>
chon <- readxl::read_excel(fpath, sheet="Diatoms")</pre>
sig_test <- readxl::read_excel(fpath, sheet="Sig_test")</pre>
chon.diat <- chon %>% select(-Year)
chon.year <- chon %>% select(Year)
# data has many rare species, remove these first
mx <- apply(chon.diat, 2, max)</pre>
chon.diat <- chon.diat[, mx > 3]
# Create a grouping variable based on significance of trend (increasing, decreasing or not sig.)
chon.groups <- data.frame(TaxonName=colnames(chon.diat)) %>%
  left_join(sig_test, by="TaxonName") %>%
  mutate(group = case_when(p_unadj < 0.1 & slope>0 ~ "Increasing",
                           p_unadj < 0.1 & slope<0 ~ "Decreasing",</pre>
                            TRUE ~ "Not sig"),
         group=factor(group, levels=c("Increasing", "Not sig", "Decreasing"))) %>%
  select(TaxonName, group)
# Sort variables according to significance
diat.order <- chon.groups %>% arrange(group) %>% pull(TaxonName)
chon.ordered <- chon.diat %>% select(!!diat.order)
riojaPlot(chon.ordered, chon.year, groups=chon.groups,
          scale.percent=TRUE,
          yinterval=1,
          plot.poly=TRUE,
          plot.line=FALSE,
          plot.groups=TRUE,
          cex.xlabel=0.6,
          cex.yaxis=0.5,
          names.italicise=TRUE,
          cex.axis=0.5,
          col.group=c("darkgreen", "darkgrey", "darkred"))
```



This looks OK but we can "virtually" rotate this figure so it can be viewed as a stacked diagram. To do this we have to modify the axis labels and orientation of taxon names. We can't easily do this using riojaPlot styles but we can suppress the plotting of names and x-axis and create a custom function to plot these in any style or position we wish.

```
# create list of blank names to suppress taxon names
names <- rep("", ncol(chon.ordered))</pre>
myfun <- function(x, y, i, nm) {</pre>
  usr <- par("usr") # extract the x and y data limits of the plot
  name <- bquote(italic(.(names(nm)))) # extract name from nm and italicise it
  text(usr[2]-2, usr[4]+0.5, name, adj=c(0, 0), xpd=NA, srt=-90, cex=0.7)
  xval <- seq(0, usr[2], by=10) # create a vector of labels for the axis
  xlab <- rep("", length(xval))</pre>
  axis(side=3, at=xval, labels=xlab, tcl=-0.2, cex.axis=0.5, mgp=c(3, 0.1, 0))
  text(xval, usr[4]-0.5, xval, cex=0.5, srt=-90, adj=c(1, 0), xpd=NA) # ad the x-axis vales
}
riojaPlot(chon.ordered, chon.year, groups=chon.groups,
          scale.percent=TRUE,
          vinterval=1,
          x.names=names, # replace taxon names with a vector of blank names
          ylabel = " ", # suppress y-axis label
          plot.poly=TRUE,
          plot.line=FALSE,
          plot.groups=TRUE,
          cex.xlabel=0.6,
          cex.yaxis=0.5,
          names.italicise=TRUE,
          cex.axis=0.5,
          col.group=c("darkgreen", "darkgrey", "darkred"),
```

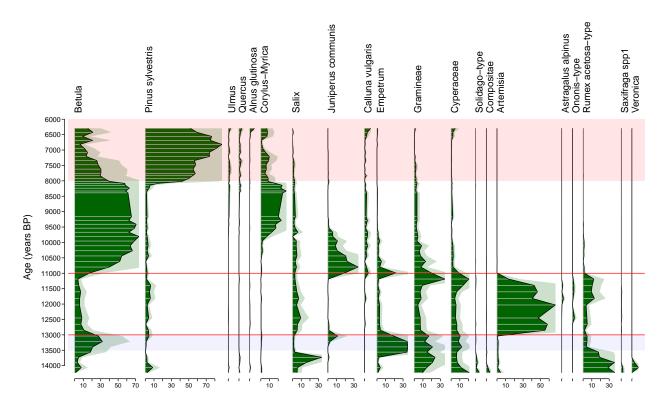
plot.bottom.axis=FALSE,
min.width.pc=10, # set minimum size of x-axes
fun.xfront=myfun)



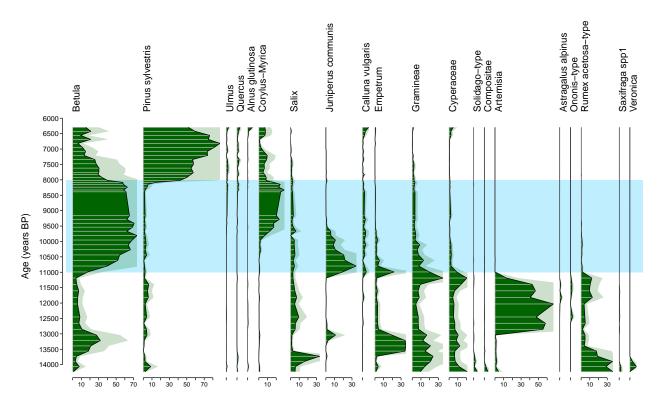
15. How do I add my own zones to a diagram?

Arbitrary zones can be added using function addRPZone, as either single lines, or as shaded rectangles. Simply save the riojaPlot object and pass it to addRPZone.

```
mx <- apply(aber.poll, 2, max)</pre>
aber.sel <- names(mx[mx > 2])
rp <- riojaPlot(aber.poll, aber.chron, aber.sel,</pre>
   yvar.name="Age (years BP)",
   ymin=6000,
   ymax = 14300,
   yinterval=500,
   scale.percent=TRUE,
   exag=TRUE)
# define zone lines at 11000 and 13000 years BP
myzones <- c(11000, 13000)
addRPZone(rp, myzones, col="red")
# add shaded zone from 6000 to 8000 years BP
addRPZone(rp, 6000, 8000)
# change colour and shading
addRPZone(rp, 13000, 13500, col="blue", alpha = 0.05)
```



```
# The zones above are plotted over the existing diagram.
# It is possible to plot zones or other annotatiosn behind
# the diagram using a cuctom function supplied to style fun.plotback
# this takes 2 arguments, usr = vector of 4 numbers giving the
\# coordinates of the plotting area in data units (x is 0-1)
# and fig, coordinates of the plotting area as fractions of the page size
fun.back <- function(usr, fig) {</pre>
 print(usr)
 rect(0, 8000, 1, 11000, col="lightblue1", border=NA)
rp <- riojaPlot(aber.poll, aber.chron, aber.sel,</pre>
  yvar.name="Age (years BP)",
  ymin=6000,
  ymax = 14300,
  yinterval=500,
   scale.percent=TRUE,
   exag=TRUE,
   fun.plotback=fun.back
#> [1]
                 1 14300 6000
```



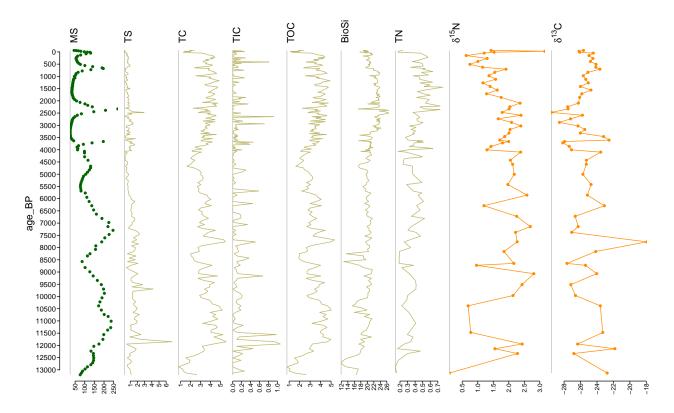
16. How do I include special symbols in the variable names

We can pass a character vector of custom names for each column using style x.names and use expression to create names with superscripts, subscripts or greek letters.

```
nms <- colnames(maule.data)
nms[8] <- expression(delta^15*N)</pre>
```

We can also use function label_geochem in package tidypaleo to automatically add annotations for a slected range of common geochemical variables.

```
library(tidypaleo)
nms <- colnames(maule.data)</pre>
nms2 <- as.expression(sapply(nms, function(x) unlist(label_geochem(x)[[1]])))</pre>
riojaPlot(maule.data, maule.chron, groups=groups,
          yvar.name="age BP",
          x.names=nms2,
          ymin=-100,
          ymax=13300,
          yinterval=500,
          plot.bar=FALSE,
          plot.poly=FALSE,
          plot.line=selline,
          plot.symb=selsymb,
          plot.groups=TRUE,
          ylabPos=2.2,
          symb.cex=0.5,
          ytks1=myticks,
          las.axis=2,
          graph.widths=widths,
          yBottom=0.06
          )
```



11. List of modifiable styles and default values

Style	Value
yvar.name	
sec.yvar.name	
ylabel	
sec.ylabel	
plot.sec.axis	FALSE
scale.percent	FALSE
scale.minmax	FALSE
y.rev	TRUE
ymin	
ymax	
yinterval	
sec.ymin	
sec.ymax	
sec.yinterval	
wa.order	none
plot.bar	TRUE
plot.line	TRUE

Style	Value
plot.poly	FALSE
plot.symb	FALSE
do.clust	FALSE
plot.clust	FALSE
plot.zones	0
lwd.bar	1
lwd.line	1
lwd.poly.line	0.6
lwd.cumul.line	0.6
col.bar	grey
bar.back	FALSE
col.symb	black
col.line	black
col.poly	darkgreen
col.poly.line	
col.cumul.line	
col.zones	red
symb.pch	19
symb.cex	1
cex.axis	0.6
cex.yaxis	0.7
cex.ylabel	0.9
cex.xlabel	0.9
srt.xlabel	90
tcl	-0.2
cex.cumul	0.7
clust.data.trans	none
clust.use.selected	FALSE
clust.width	0.05
graph.widths	1
exag	FALSE
col.exag	auto
col.exag.line	
lwd.exag.line	0.6
exag.mult	2

Style	Value
exag.alpha	0.2
names.break.long	TRUE
names.break.n	20
names.italicise	FALSE
plot.groups	FALSE
plot.cumul	FALSE
cumul.mult	1
col.group1	darkgreen
col.group2	darkkhaki
col.group3	darkorange
col.group4	darkred
col.group5	deepskyblue
col.group6	sienna3
col.group7	${\rm darkgoldenrod} 3$
col.group8	darkseagreen
col.group9	yellow3
col.group10	darkgrey
xRight	0.99
xLeft	
yBottom	0.05
уТор	
fun.xback	
fun.xfront	
fun.plotback	
fun.yaxis	
ylabPos	
xlabPos	0.1
xSpace	0.01
x.pc.omit0	TRUE
lwd.axis	1
col.axis	grey
min.width.pc	5
las.axis	1
las.yaxis	1
ytks1	

Style	Value
ytks2	
omitMissing	TRUE
col.sep.bar	black
sep.bar	FALSE
plot.bottom.axis	TRUE
plot.top.axis	FALSE
cumulSpace	
plot.yaxis	TRUE
start.new.plot	TRUE
xGap	0.01
x.pc.inc	10

12. References

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