Class18

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Read

```
library(tximport)
library(rhdf5)

# setup the folder and filenames to read
folders <- dir(pattern="SRR21568*")
samples <- sub("_quant", "", folders)
files <- file.path( folders, "abundance.h5" )
names(files) <- samples

txi.kallisto <- tximport(files, type = "kallisto", txOut = TRUE)</pre>
```

1 2 3 4

See how many transcripts we have for each sample and how many are detected are in at least one sample.

```
head(txi.kallisto$counts)
```

	SRR2156848	SRR2156849	SRR2156850	SRR2156851
ENST00000539570	0	0	0.00000	0
ENST00000576455	0	0	2.62037	0
ENST00000510508	0	0	0.00000	0
ENST00000474471	0	1	1.00000	0

```
ENST00000381700 0 0.00000 0
ENST00000445946 0 0.00000 0
```

```
colSums(txi.kallisto$counts)
```

SRR2156848 SRR2156849 SRR2156850 SRR2156851 2563611 2600800 2372309 2111474

```
sum(rowSums(txi.kallisto$counts)>0)
```

[1] 94561

filter out those annotated transcripts with no reads:

```
to.keep <- rowSums(txi.kallisto$counts) > 0
kset.nonzero <- txi.kallisto$counts[to.keep,]</pre>
```

no change

```
keep2 <- apply(kset.nonzero,1,sd)>0
x <- kset.nonzero[keep2,]</pre>
```

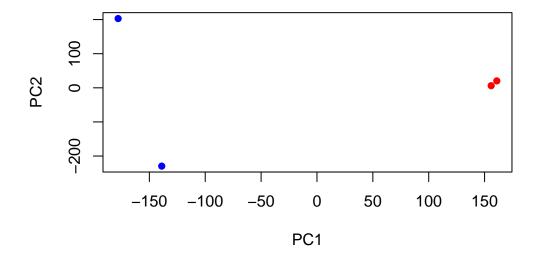
PCA

```
pca <- prcomp(t(x), scale=TRUE)
summary(pca)</pre>
```

Importance of components:

```
PC1 PC2 PC3 PC4
Standard deviation 183.6379 177.3605 171.3020 1e+00
Proportion of Variance 0.3568 0.3328 0.3104 1e-05
Cumulative Proportion 0.3568 0.6895 1.0000 1e+00
```

visualizing the summarized transcriptomic profiles of each sample:



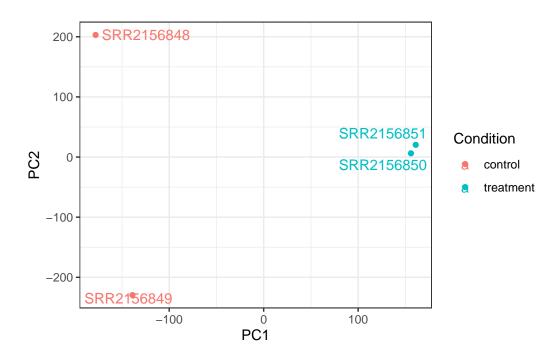
Q. Use ggplot to make a similar figure of PC1 vs PC2 and a seperate figure PC1 vs PC3 and PC2 vs PC3.

```
library(ggrepel)

# Make metadata object for the samples
colData <- data.frame(condition = factor(rep(c("control", "treatment"), each = 2)))
rownames(colData) <- colnames(txi.kallisto$counts)

# Make the data.frame for ggplot
y <- as.data.frame(pca$x)
y$Condition <- as.factor(colData$condition)</pre>
ggplot(y) +
```

```
aes(PC1, PC2, col=Condition) +
geom_point() +
geom_text_repel(label=rownames(y)) +
theme_bw()
```



Differential-Expression Analysis

library(DESeq2)

Loading required package: S4Vectors

Loading required package: stats4

Loading required package: BiocGenerics

Attaching package: 'BiocGenerics'

The following objects are masked from 'package:stats':

IQR, mad, sd, var, xtabs

The following objects are masked from 'package:base':

anyDuplicated, aperm, append, as.data.frame, basename, cbind, colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget, order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank, rbind, Reduce, rownames, sapply, saveRDS, setdiff, table, tapply, union, unique, unsplit, which.max, which.min

Attaching package: 'S4Vectors'

The following object is masked from 'package:utils':

findMatches

The following objects are masked from 'package:base':

expand.grid, I, unname

Loading required package: IRanges

Attaching package: 'IRanges'

The following object is masked from 'package:grDevices':

windows

Loading required package: GenomicRanges

Loading required package: GenomeInfoDb

Loading required package: SummarizedExperiment

Loading required package: MatrixGenerics

Loading required package: matrixStats

Warning: package 'matrixStats' was built under R version 4.4.2

Attaching package: 'MatrixGenerics'

The following objects are masked from 'package:matrixStats':

colAlls, colAnyNAs, colAnys, colAvgsPerRowSet, colCollapse, colCounts, colCummaxs, colCummins, colCumprods, colCumsums, colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs, colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats, colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds, colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads, colWeightedMeans, colWeightedMedians, colWeightedSds, colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgsPerColSet, rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods, rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps, rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins, rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks, rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars, rowWeightedMads, rowWeightedMeans, rowWeightedMedians, rowWeightedSds, rowWeightedVars

Loading required package: Biobase

Welcome to Bioconductor

Vignettes contain introductory material; view with 'browseVignettes()'. To cite Bioconductor, see 'citation("Biobase")', and for packages 'citation("pkgname")'.

Attaching package: 'Biobase'

The following object is masked from 'package:MatrixGenerics':

rowMedians

using counts and average transcript lengths from tximport

DESeq

```
estimating size factors

using 'avgTxLength' from assays(dds), correcting for library size

estimating dispersions

gene-wise dispersion estimates

mean-dispersion relationship

-- note: fitType='parametric', but the dispersion trend was not well captured by the function: y = a/x + b, and a local regression fit was automatically substituted. specify fitType='local' or 'mean' to avoid this message next time.

final dispersion estimates

fitting model and testing
```

res <- results(dds) head(res)</pre>

log2 fold change (MLE): condition treatment vs control

Wald test p-value: condition treatment vs control

DataFrame with 6 rows and 6 columns

Datariame with 0 10%s and		O COLUMNS				
		${\tt baseMean}$	${\tt log2FoldChange}$	lfcSE	stat	pvalue
		<numeric></numeric>	<numeric></numeric>	<numeric></numeric>	<numeric></numeric>	<numeric></numeric>
	ENST00000539570	0.000000	NA	NA	NA	NA
	ENST00000576455	0.761453	3.155061	4.86052	0.6491203	0.516261
	ENST00000510508	0.000000	NA	NA	NA	NA
	ENST00000474471	0.484938	0.181923	4.24871	0.0428185	0.965846
	ENST00000381700	0.000000	NA	NA	NA	NA
	ENST00000445946	0.000000	NA	NA	NA	NA
		padj				
		<numeric></numeric>				
	ENST00000539570	NA				
	ENST00000576455	NA				
	ENST00000510508	NA				
	ENST00000474471	NA				
	ENST00000381700	NA				
	ENST00000445946	NA				