Sparse Canononical Correlation Analysis for Neuroimaging (SCCAN): A Tutorial

Introduction

Overview

Sparse canonical correlation analysis for neuroimaging (SCCAN) is a general purpose tool for "two-sided" multiple regression. It is an extension of Hotelling's seminal canonical correlation analysis [1, 2] which itself is a multi-modal extension of principal component analysis (PCA). This technique allows one to symmetrically compare one matrix of data to another and find linear relationships between them in a low-dimensional space. SCCAN derives from classic canonical correlation analysis and also relates to singular value decomposition. To handle data with p >> n (common in medical imaging scenarios) SCCAN uses high-dimensional regularization methods common in ℓ_1 regression and spatial regularization to help ensure the biological plausibility of statistical maps in medical imaging (often referred to as eigenanatomy). This problem is a difficult optimization (NP-hard) and, to improve solution interpretability and stability, SCCAN allows one to to use prior knowledge to constrain the solution space.

Another view (perhaps simpler and broader) is that eigenanatomy is a general framework for reducing the dimensionality of imaging data into interpretable pieces. Eigenanatomy is motivated by two ideas:

Voxels that change together should hang together.

and

Clustering before hypothesis testing, not hypothesis testing then clustering.

These two strategies conserve statistical power in a controllable manner (contrasting with mass univariate techniques) by reducing the number of statistical tests performed.

Comparison with other techniques

Common mass univariate techniques (e.g., voxel-based morphometry [3]) require adjustment for multiple comparisons and subsequent ad hoc clustering of significant voxels into potentially anatomically meaningful clusters [4]. Decomposition techniques, such as SCCAN, invert this statistical directionality by first "decomposing", or "clustering", voxels in an anatomically constrained, yet data-driven, manner followed by significance testing. Repeating from the previous section, this prioritizing of the dimensionality reduction step mitigates the multiple comparison issue.

Traditional decomposition methods, such as the orthogonality-constrained PCA and independent components analysis (ICA), have found widespread utility in neuroimaging (e.g., PCA: [5, 6] and ICA: [7–9]). However, without additional constraints, such solutions are not "sparse" in the sense that the solution space is non-zero over the entire problem domain (e.g., PCA- and ICA-derived eigenvectors) and can produce negative weights which limits biological interpretability [10].

One potentially problematic issue with sparse matrix decompositions, such as SCCAN, is the potential collinearity of the "eigenvectors" (or, more accurately, the "pseudo-eigenvectors" since they do not necessarily satisfy orthogonality) resulting from optimization of the eigenvalue problem modified by constraints (e.g., positivity and sparsity) [10, 11].

Applications

SCCAN-related methods have been applied in the following papers:

- Dementia induces correlated reductions in white matter integrity and cortical thickness: a multivariate neuroimaging study with sparse canonical correlation analysis [12].
- Methodological considerations in longitudinal morphometry of traumatic brain injury [13].
- Sparse canonical correlation analysis relates network-level atrophy to multivariate cognitive measures in a neurodegenerative population [11]. Repeatable cortical structural networks associated with specific pyschometric testing are determined from SCCAN by determining maximal correlations between cognitive measurements from the Philadelphia Brief Assessment of Cognition (e.g., apathy, agitation, and social comportment) and gray matter density determined from T1-weighted MRI. The tutorial in the next section is based on this work.
- Eigenanatomy: Sparse dimensionality reduction for multi-modal medical image analysis [10]. A comparison is performed using decomposition techniques (ICA, PCA, and SCCAN) on multi-modality neuroimaging data (cortical thickness from T1, cerebral blood flow from ASL, and fractional anisotropy from diffusion-weighted MRI) from a publicly available cohort. SCCAN (i.e., eigenanatomy) outperforms the other methods in predicting subject age. Data and analysis scripts for this work are publicly available.
- Subject-specific functional connectivity parcellation via Prior Based Eigenanatomy [14]. Sparse decomposition is performed on individual subject fMRI data (the corresponding image data matrix is $t \times p$ where t is the number of time points and p is the number of voxels in the user-specified mask. A spatial (i.e., anatomical) prior is accommodated by the eigenanatomy framework and is used to describe the default mode network—hippocampus functional connectivity. These connectivity patterns are then used to classify mild cognitive impairment subjects from cognitively normal subjects.
- White matter imaging helps dissociate tau from TDP-43 in frontotemporal lobar degeneration [15].
- The power of neuroimaging biomarkers for screening frontotemporal dementia [16].
- Genetic and neuroanatomic associations in sporadic frontotemporal lobar degeneration [17].

ANTsR implementation

The various SCCAN-related methods and associated functionality are all available in the ANTsR package 1 . The two main R functions are:

- sparseDecom Decomposes a matrix into sparse eigenevectors to maximize explained variance.
- sparseDecom2 Decomposes two matrices into paired sparse eigenevectors to maximize canonical correlation.

Auxiliary functions include:

- initializeEigenanatomy
- eigSegs
- joinEigenanatomy

¹https://github.com/stnava/ANTsR

Tutorial

Below is a series of tutorials based on the work presented in [11] described above where structural networks are determined based on gray matter density computed from T1-weighted MRI and neurocognitive testing using the Philadelphia Brief Assessment of Cognition (PBAC).

Initialization

```
# We include all the necessary R package dependencies. We assume that the user
# is running this script (stitchTutorialDocument.R) in the repo directory.
library( knitr )
library( visreg )
library( pheatmap )
library( pander )
library( png )
library( misc3d )
library( rgl )
library( pixmap )
library( randomForest )
library( ggplot2 )
library( stargazer )
invisible( suppressMessages( library( ANTsR ) ) )
rootDirectory <- "./"</pre>
knitr::opts_knit$set( root.dir = rootDirectory )
knitr::opts_chunk$set( comment = "" )
figuresDirectory <- pasteO( rootDirectory, "Figures/" )</pre>
if( ! dir.exists( figuresDirectory ) )
  dir.create( pasteO( rootDirectory, "Figures/" ) )
dataDirectory <- pasteO( rootDirectory, "Data/" );</pre>
```

Read input data

```
# Load the AAL (Automated Anatomical Labeling) data table from ANTsR as well as
# the AAL label image.

data( aal, package = 'ANTsR' )
aalLabelTable <- aal
aalFileName <- pasteO( dataDirectory, "aal.nii.gz" )
aalImage <- antsImageRead( filename = aalFileName, dimension = 3 )

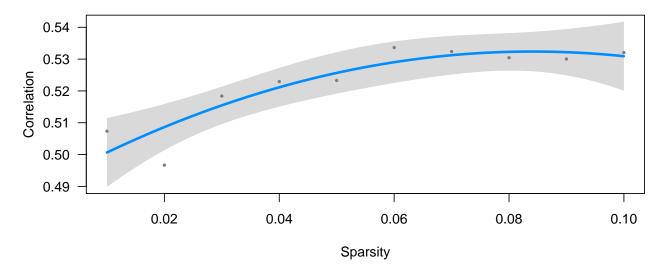
# For convenience, the data set for test/train subjects are stored in 2-D images where
# the rows correspond to different subjects and the columns are the voxels within
# the 3-D template gray matter mask.

trainingFile <- pasteO( dataDirectory, "pbac_train_img.mha" )</pre>
```

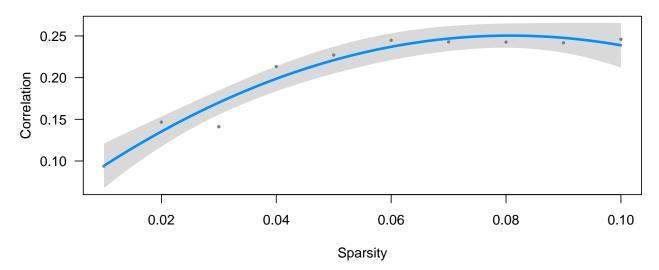
```
trainingImageData <- as.matrix( antsImageRead( filename = trainingFile, dimension = 2 ) )
testingFile <- pasteO( dataDirectory, "pbac_test_img.mha" )
testingImageData <- as.matrix( antsImageRead( filename = testingFile, dimension = 2 ) )
grayMatterMaskFile <- pasteO( dataDirectory, "mask.nii.gz" )
grayMatterMask <- antsImageRead( filename = grayMatterMaskFile, dimension = 3 )
# Read in the corresponding cognitive data for the test/train subjects.
trainingCognitiveData <- read.csv( pasteO( dataDirectory, "pbac_train_cog.csv" ) )
testingCognitiveData <- read.csv( pasteO( dataDirectory, "pbac_test_cog.csv" ) )</pre>
```

SCCAN for sparse regression

```
# Here we use SCCAN to find brain regions relating to age. In this case, SCCAN
# is used as a sparse regression utility. We impose a "cluster threshold"
# regularization to prevent isolated voxels from appearing in the solution.
# Evaluation as a function of sparseness is performed using the testing data.
# This type of approach can be useful in parameter selection i.e., choosing the
# optimization criterion based on the training data.
trainingAgeMatrix <- matrix( trainingCognitiveData$age, ncol = 1 )</pre>
sparsityValues <- seq( from = 0.01, to = 0.1, length = 10 )
trainingAgeCorrelations <- rep( 0, length( sparsityValues ) )</pre>
testingAgeCorrelations <- rep( 0, length( sparsityValues ) )</pre>
for( i in 1:length( sparsityValues ) )
  ageSccanResults <- sparseDecom2(</pre>
   inmatrix = list( scale( trainingImageData ), scale( trainingAgeMatrix ) ),
    sparseness = c( sparsityValues[i], 0.9 ), inmask = c( grayMatterMask, NA ),
   nvecs = 2, mycoption = 0, cthresh = c(1000, 0), ell1 = 10, smooth = 0.5,
   verbose = 0)
  # ageSccanResults$eig1 contain the eigenvectors for "view 1" (in CCA terminology)
  # whereas ageSccanResults$eig2 contain the eigenvectors for "view 2". In this
  # scenario only the first view (imaging data) is relevant. One can view the
  # eigenvectors as images using the ANTsR::matrixToImages() function, e.g.,
      sccanFirstEigenImage <- matrixToImages( t( ageSccanResults$eig1 ),</pre>
             qrayMatterMask )[[1]]
  # determine correlation with training data
  trainingAgePredictors <- trainingImageData %*% ageSccanResults$eig1
  trainingAgeCorrelations[i] <- cor( trainingAgePredictors, trainingCognitiveData$age )
  # validate using testing data
  testingAgePredictors <- testingImageData %*% ageSccanResults$eig1</pre>
  testingAgeCorrelations[i] <- cor( testingAgePredictors, testingCognitiveData$age )</pre>
```



```
testingLm <- lm( testingAgeCorrelations ~ sparsityValues + I( sparsityValues^2 ) )
visreg( testingLm, xlab = "Sparsity", ylab = "Correlation" )</pre>
```



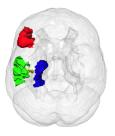
SCCAN with prior initialization

```
# Here we use the same data as in the previous example but use SCCAN to find brain
# regions relating to the battery of tests that measure language-related cognitive
# function. Due to the language association, we initialize SCCAN with left
# hemispheric regions. In this case, the initialization controls the sparseness
# parameters for each eigenvector. Thus, the sparsity parameter will be overridden
# by the priors, thereby enabling a "per eigenvector" sparsity value.
languageBatteryTypes <- c( "Speech", "Writing", "Semantic", "Reading", "Naming" )</pre>
trainingCognitiveMatrix <- cbind( trainingCognitiveData$speech_adj,</pre>
                                   trainingCognitiveData$writing_adj,
                                   trainingCognitiveData$semantic_adj,
                                   trainingCognitiveData$reading_adj,
                                   trainingCognitiveData$naming_adj )
colnames( trainingCognitiveMatrix ) <- languageBatteryTypes</pre>
testingCognitiveMatrix <- cbind( testingCognitiveData$speech_adj,</pre>
                                  testingCognitiveData$writing adj,
                                  testingCognitiveData$semantic_adj,
                                  testingCognitiveData$reading_adj,
                                  testingCognitiveData$naming adj )
colnames( testingCognitiveMatrix ) <- languageBatteryTypes</pre>
# Initialize the left hemispheric AAL regional priors associated with the language
# centers. More information re. AAL can be found at various neuroimaging sites, e.g.,
     http://neuro.imm.dtu.dk/wiki/Automated\_Anatomical\_Labeling
# The specific regions of interest for the testing below are:
      Label
                                Region
#
        13
                        Left area triangularis
#
        81
                        Left superior temporal gyrus
        39
                         Left parahippocampal gyrus
        79
                         Left transverse temporal gyri
aalLanguageRegionalLabels <- c( 13, 81, 39, 79 )
\# Create a label matrix ( size = number Of Cognitive Labels x number Of Voxels In Mask )
# to be used as a spatial prior
numberOfCognitiveLabels <- length( aalLanguageRegionalLabels )</pre>
numberOfVoxelsInMask <- sum( grayMatterMask > 0.5 )
aalLanguageRegionalLabelMatrix <- matrix(</pre>
  rep( 0, numberOfVoxelsInMask * numberOfCognitiveLabels ),
 nrow = numberOfCognitiveLabels )
for( i in 1:numberOfCognitiveLabels )
  perLabelVector <- aalImage[grayMatterMask == 1] == aalLanguageRegionalLabels[i]</pre>
  aalLanguageRegionalLabelMatrix[i,] <- as.numeric( perLabelVector )</pre>
eigenInitialization <- initializeEigenanatomy( aalLanguageRegionalLabelMatrix,
```

```
grayMatterMask )
priorWeights <- c(0.9, 0.5, 0.05)
# Create 3-D brain volumetric rendering of the initial AAL language labels
# and create some variables for rendering the results.
languagePriorWeightsSccanPlotFiles <- rep( '', length( priorWeights ) )</pre>
languagePriorWeightsSccanPlot3DFiles <- rep( '', length( priorWeights ) )</pre>
cognitiveSccanColors <- c( "red", "green", "blue", "yellow" )</pre>
brain <- renderSurfaceFunction( surfimg = list( aalImage ), alphasurf = 0.1,</pre>
    funcimg = eigenInitialization$initlist, smoothsval = 1.5, smoothfval = 0,
    mycol = cognitiveSccanColors )
id <- par3d( "userMatrix" )</pre>
rid <- rotate3d( id, -pi / 2, 1, 0, 0 )
rid2 <- rotate3d( id, pi / 2, 0, 0, 1 )
par3d( userMatrix = id )
languageInitializationSccanPlot3DFile <- paste0( figuresDirectory,</pre>
  "cognitiveSccanEigenImages3D_Initialization" )
dd <- make3ViewPNG( rid, id, rid2, paste0( figuresDirectory,</pre>
  "cognitiveSccanEigenImages3D_Initialization" ) )
languageInitializationSccanPlot3DFile <- pasteO( figuresDirectory,</pre>
  "cognitiveSccanEigenImages3D Initialization", ".png" )
par3d( userMatrix = id )
for( i in 1:length( priorWeights ) )
  cognitiveSccanResult <- sparseDecom2(</pre>
       inmatrix = list( scale( trainingImageData ), scale( trainingCognitiveMatrix ) ),
       its = 20, mycoption = 0, sparseness = c(0, -0.5), nvecs = numberOfCognitiveLabels,
       inmask = c(grayMatterMask, NA), cthresh = c(1000, 0), verbose = 0, ell1 = 10,
       initializationList = eigenInitialization$initlist, priorWeight = priorWeights[i],
       smooth = 0, perms = 0)
  # calculate the predictors for both (imaging and cognitive testing) views
  trainingImagePredictors <- trainingImageData %*% cognitiveSccanResult$eig1
  colnames( trainingImagePredictors ) <- paste0( "GM", c( 1:numberOfCognitiveLabels ) )</pre>
  trainingCognitivePredictors <- trainingCognitiveMatrix %*% cognitiveSccanResult$eig2
  bestPredictor <- which.max(</pre>
    abs( diag( cor( trainingImagePredictors, trainingCognitivePredictors ) ) ) )
  trainingDataFrame <- data.frame( trainingImagePredictors, trainingCognitivePredictors )</pre>
  # Set up a linear model with the training data to calculate cognitive predictors
  # with the testing data using the best predictor.
  lmFormula <- as.formula( paste0( "Variate00" ,bestPredictor-1 , "~ GM1+GM2+GM3+GM4" ) )</pre>
  trainingLm <- lm( lmFormula, data = trainingDataFrame )</pre>
  testingImagePredictors <- testingImageData %*% cognitiveSccanResult$eig1
```

```
colnames( testingImagePredictors ) <- pasteO( "GM", c( 1:numberOfCognitiveLabels ) )</pre>
  testingCognitivePredictors <- testingCognitiveMatrix %*% cognitiveSccanResult$eig2
  testingDataFrame <- data.frame( testingImagePredictors, testingCognitivePredictors )</pre>
  # How well does the gray matter image view predict language-cognitive ability?
  # We don't print this test but report the correlation and p-values directly in
  # caption for the corresponding 3-D brain renderings produced below.
  testingCorrelationTest <- cor.test( testingDataFrame[, bestPredictor],</pre>
          predict( trainingLm, newdata = testingDataFrame ) )
  cognitiveSccanImages <- matrixToImages( t( cognitiveSccanResult$eig1 ), grayMatterMask )</pre>
  for( image in cognitiveSccanImages )
    {
    image[grayMatterMask == 1] <- abs( image[grayMatterMask == 1] )</pre>
    image[grayMatterMask == 1] <- image[grayMatterMask == 1] /</pre>
       max( image[grayMatterMask == 1] )
    }
  # Create 2-D slice mosaics in saqittal view. We don't render them in the
  # pdf document but write them to disk for perusal and to illustrate use
  # case.
  languagePriorWeightsSccanPlotFiles[i] <- pasteO( figuresDirectory,</pre>
    "cognitiveSccanEigenImages_PriorWeight", priorWeights[i], ".jpg")
  plot( grayMatterMask, cognitiveSccanImages, color.overlay = cognitiveSccanColors,
        axis = 1, nslices = 20, outname = languagePriorWeightsSccanPlotFiles[i] )
  # Create 3-D brain volumetric rendering. These are rendered in Figures 1-4.
  brain <- renderSurfaceFunction( surfimg = list( aalImage ), alphasurf = 0.1,</pre>
      funcimg = cognitiveSccanImages, smoothsval = 1.5, smoothfval = 0,
      mycol = cognitiveSccanColors )
  id <- par3d( "userMatrix" )</pre>
  rid <- rotate3d( id, -pi / 2, 1, 0, 0 )
  rid2 <- rotate3d( id, pi / 2, 0, 0, 1 )
  par3d( userMatrix = id )
  languagePriorWeightsSccanPlot3DFiles[i] <- paste0( figuresDirectory,</pre>
    "cognitiveSccanEigenImages3D_PriorWeight", priorWeights[i] )
  dd <- make3ViewPNG( rid, id, rid2, languagePriorWeightsSccanPlot3DFiles[i] )</pre>
  languagePriorWeightsSccanPlot3DFiles[i] <- paste0( figuresDirectory,</pre>
    "cognitiveSccanEigenImages3D PriorWeight", priorWeights[i], ".png" )
  par3d( userMatrix = id )
# Figures 1 through 4 show that the best results initialized by the prior can
# drift away from that initialization. A fundamental question is --- Where in the
# brain do the solution vectors end up? We write a quick function to answer this
# question for the weak prior case (priorWeighting = 0.1).
reportAnatomy <- function( eigenImageList, mask, weight = 0.3 )</pre>
```

```
sccanAalLabels <- c()
for( eigenImage in eigenImageList )
   {
   nonZeroIndices<- abs( eigenImage[mask == 1] ) > 0
   sccanAalLabels <- append( sccanAalLabels, aalImage[mask == 1][nonZeroIndices] )
   }
   anatomicalCount <- hist( sccanAalLabels, breaks = 0:100, plot = FALSE )$count
   anatomicalCount[anatomicalCount < weight * max( anatomicalCount )] <- 0
   aalIndices <- which( anatomicalCount > 0 )
   return( aalLabelTable$label_name[aalIndices] )
}
reportedAnatomy <- reportAnatomy( cognitiveSccanImages, grayMatterMask )</pre>
```





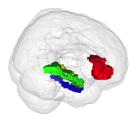
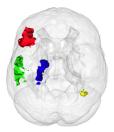
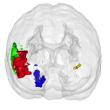


Figure 1: 3-D renderings of the initial language regions from the AAL image. Regional labels are left area triangularis (red), left superior temporal gyrus (green), left parahippocampal gyrus (yellow), left transverse temporal gyri (blue).





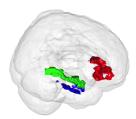
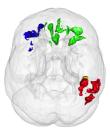
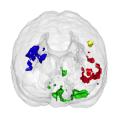


Figure 2: 3-D renderings of the eigenvectors for the imaging (gray matter) view constructed from a strong prior (= 0.9). Pearson's product-moment correlation results in a correlation value of 0.4 (p-value = 0.0002)





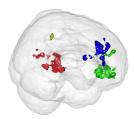
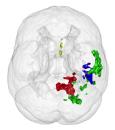
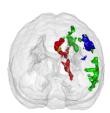


Figure 3: 3-D renderings of the eigenvectors for the imaging (gray matter) view constructed from a medium prior (=0.5). Pearson's product-moment correlation results in a correlation value of 0.65 (p-value =2e-11)





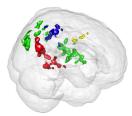


Figure 4: 3-D renderings of the eigenvectors for the imaging (gray matter) view constructed from a weak prior (= 0.1). Pearson's product-moment correlation results in a correlation value of 0.4 (p-value = 0.0001)

```
reportedAnatomyDataFrame <- data.frame( newRegions = reportedAnatomy )
colnames( reportedAnatomyDataFrame ) <- c( 'Predictor regions' )
pander( reportedAnatomyDataFrame, style = "rmarkdown", caption = "Using a weak prior
    (= 0.1), the solution migrates from the initialized regions. The new regions
    associated with the best \"language\" predictor are provided below." )</pre>
```

Table 1: Using a weak prior (=0.1), the solution migrates from the initialized regions. The new regions associated with the best "language" predictor are provided below.

```
Predictor regions

Calcarine_R
Occipital_Sup_R
Occipital_Mid_R
Parietal_Inf_R
SupraMarginal_R
Angular_R
Precuneus_R
Temporal_Sup_R
Temporal_Mid_R
```

How good were our original hypothetical regions as predictors? -- Good question.

A closer look at the SCCAN eigenvectors

```
# Recalling that CCA maximizes PearsonCorrelation$(XW^T$,ZY^T)$, where $X$ and $Z$ are
# data matrices, we can study the eigenvector matrix $Y$ (or $W$) which contrasts or
# combines columns of the associated data matrix. In this example, we look at $Y$
# (prior weighting = 0.1) which operates on the language-related cognition/design matrix.
# Technical note: In order to break this tutorial into reasonably sized chunks and take
# advantage of caching, we need to re-run SCCAN with the final parameters (prior weighting
# = 0.1) from the previous chunk. ANTSR stores an external pointer to the image which is
# not preserved between document compilations ("stitching").
eigenInitialization <- initializeEigenanatomy( aalLanguageRegionalLabelMatrix,</pre>
                        grayMatterMask )
cognitiveSccanResult <- sparseDecom2(</pre>
     inmatrix = list( scale( trainingImageData ), scale( trainingCognitiveMatrix ) ),
     its = 20, mycoption = 0, sparseness = c(0, -0.5), nvecs = numberOfCognitiveLabels,
     inmask = c(grayMatterMask, NA), cthresh = c(1000, 0), verbose = 0, ell1 = 10,
     initializationList = eigenInitialization$initlist, priorWeight = 0.1,
     smooth = 0, perms = 0)
cognitiveSccanHeatMapFile <- pasteO( figuresDirectory,</pre>
  "cognitiveSccanHeatMap_PriorWeight0.1.png" )
rownames( cognitiveSccanResult$eig2 ) <- languageBatteryTypes</pre>
```

```
pheatmap( cognitiveSccanResult$eig2, cluster_rows = TRUE, cluster_cols = TRUE,
    filename = cognitiveSccanHeatMapFile )
```

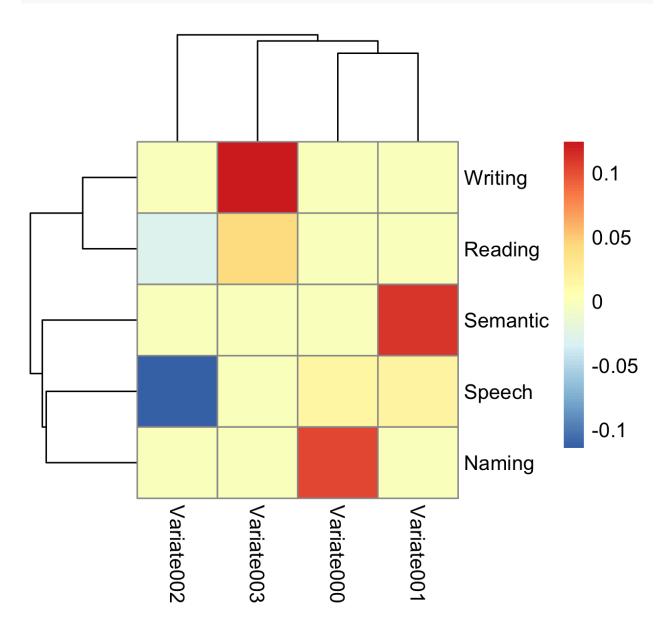


Figure 5: Heat map of the eigenvectors for the language/cognitive view constructed from a weak prior (= 0.1).

SCCAN regression with nuisance variables

```
# One often wants to control for the presence of nuisance variables in conjunction
# with SCCAN results. There are several options including:
# (1) control after you do dimensionality reduction,
# (2) orthogonalize the predictors before decomposition, and
```

```
# (3) use alternative SCCAN formulations (e.g. set ``mycoption`` to 0 or 2).
# We first try the options (1) and (2) as they are more traditional.
# Option (1)---control for nuisance variables (e.g., age and mmse) after dimensionality
# reduction.
eigenInitialization <- initializeEigenanatomy( aalLanguageRegionalLabelMatrix,</pre>
                        grayMatterMask )
cognitiveSccanResult <- sparseDecom2(</pre>
     inmatrix = list( scale( trainingImageData ), scale( trainingCognitiveMatrix ) ),
     its = 20, mycoption = 0, sparseness = c(0, -0.9), nvecs = numberOfCognitiveLabels,
     inmask = c(grayMatterMask, NA), cthresh = c(1000, 0), verbose = 0, ell1 = 10,
     initializationList = eigenInitialization$initlist, priorWeight = 0.1,
     smooth = 0, perms = 0)
trainingImagePredictors <- trainingImageData %*% cognitiveSccanResult$eig1
colnames( trainingImagePredictors ) <- paste0( "GM", c( 1:numberOfCognitiveLabels ) )</pre>
trainingCognitivePredictors <- trainingCognitiveMatrix %*% cognitiveSccanResult$eig2
bestPredictor <- which.max(</pre>
  abs( diag( cor( trainingImagePredictors, trainingCognitivePredictors ) ) ) )
# Set up a linear model with the training data to get the cognitive predictors
# with the testing data using the best predictor (as before) but this time, add
# mmse and age as covariates.
trainingDataFrame <- data.frame( trainingImagePredictors, trainingCognitivePredictors,
  MMSE = trainingCognitiveData$mmse, Age = trainingCognitiveData$age )
lmFormula <- as.formula( paste0( "Variate00" ,bestPredictor-1 ,</pre>
   "~ GM1+GM2+GM3+GM4+Age+MMSE" ) )
trainingLm <- lm( lmFormula, data = trainingDataFrame )</pre>
testingImagePredictors <- testingImageData %*% cognitiveSccanResult$eig1
colnames( testingImagePredictors ) <- pasteO( "GM", c( 1:numberOfCognitiveLabels ) )</pre>
testingCognitivePredictors <- testingCognitiveMatrix %*% cognitiveSccanResult$eig2
testingDataFrame <- data.frame( testingImagePredictors, testingCognitivePredictors,
 MMSE = testingCognitiveData$mmse, Age = testingCognitiveData$age )
testingLm <- lm( lmFormula, data = testingDataFrame )</pre>
# We output the two linear models in Table 2 using the stargazer package.
stargazer( trainingLm, testingLm, model.names = FALSE, header = FALSE,
 no.space = TRUE, ci = TRUE, ci.level = 0.90, single.row = TRUE, type = 'latex',
  table.placement = 'h', dep.var.caption = "", dep.var.labels.include = FALSE,
  column.labels = c( "{\\bf Training}", "{\\bf Testing}" ), model.numbers = FALSE,
 title = "Summary of training and testing linear models with nuisance variables
  included after decomposition (option 1). 90\ confidence intervals for the
 covariates are given in parentheses." )
```

```
# Now we try option (2)---orthogonalize the predictors against MMSE and age. In # other words, we regress out the effects of MMSE on both the imaging and # language/cognitive views prior to decomposition.
```

Table 2: Summary of training and testing linear models with nuisance variables included after decomposition (option 1). 90% confidence intervals for the covariates are given in parentheses.

	Training	Testing	
GM1	22.123^{***} (16.405, 27.842)	20.017*** (14.187, 25.846)	
GM2	$3.099^{**} (0.855, 5.342)$ $6.237^{***} (2.866, 9.609)$		
GM3	-0.609 (-3.324, 2.105)	5) 0.812 (-2.173, 3.797)	
GM4	5.636^{***} (3.064, 8.207) -0.999 (-4.104, 2.107)		
Age	$-0.001 \; (-0.003, 0.001)$ $0.0003 \; (-0.002, 0.002)$		
MMSE	-0.009^{***} $(-0.012, -0.006)$ -0.008^{***} $(-0.012, -0.004)$		
Constant	$0.757^{***} (0.337, 1.177)$	$0.889^{***} (0.452, 1.326)$	
Observations	89	83	
\mathbb{R}^2	0.683	0.532	
Adjusted \mathbb{R}^2	0.660	0.495	
Residual Std. Error	0.093 (df = 82)	0.097 (df = 76)	
F Statistic	$29.440^{***} (df = 6; 82)$	$14.393^{***} (df = 6; 76)$	
Note:		*n<0.1: **n<0.05: ***n<0.01	

Note:p<0.1; **p<0.05; ***p<0.01

```
trainingCognitiveResiduals <- residuals( lm(</pre>
  trainingCognitiveMatrix ~ trainingCognitiveData$mmse + trainingCognitiveData$age ) )
trainingImageResiduals <- residuals( lm(</pre>
  trainingImageData ~ trainingCognitiveData$mmse + trainingCognitiveData$age ) )
eigenInitialization <- initializeEigenanatomy( aalLanguageRegionalLabelMatrix,
                        grayMatterMask )
cognitiveSccanResult <- sparseDecom2(</pre>
     inmatrix = list( scale( trainingImageResiduals ), scale( trainingCognitiveResiduals ) ),
     its = 20, mycoption = 0, sparseness = c(0, -0.9), nvecs = numberOfCognitiveLabels,
     inmask = c(grayMatterMask, NA), cthresh = c(1000, 0), verbose = 0, ell1 = 10,
     initializationList = eigenInitialization$initlist, priorWeight = 0.1,
     smooth = 0, perms = 0)
# Decomposition of the residualized data produces eigenvectors which we can apply
# to the original data. We then construct the linear models with the original
# testing and training data but including the nuisance variables in the model formula.
trainingImagePredictors <- trainingImageData %*% cognitiveSccanResult$eig1
colnames( trainingImagePredictors ) <- paste0( "GM", c( 1:numberOfCognitiveLabels ) )</pre>
trainingCognitivePredictors <- trainingCognitiveMatrix %*% cognitiveSccanResult$eig2
bestPredictor <- which.max(</pre>
  abs( diag( cor( trainingImagePredictors, trainingCognitivePredictors ) ) ) )
trainingDataFrame <- data.frame( trainingImagePredictors, trainingCognitivePredictors,
  MMSE = trainingCognitiveData$mmse, Age = trainingCognitiveData$age )
lmFormula <- as.formula( paste0( "Variate00" ,bestPredictor-1 ,</pre>
   "~ GM1+GM2+GM3+GM4+Age+MMSE" ) )
trainingLm <- lm( lmFormula, data = trainingDataFrame )</pre>
testingImagePredictors <- testingImageData %*% cognitiveSccanResult$eig1
colnames( testingImagePredictors ) <- paste0( "GM", c( 1:numberOfCognitiveLabels ) )</pre>
```

```
# We output the two linear models in Table 3 using the stargazer package.

stargazer( trainingLm, testingLm, model.names = FALSE, header = FALSE,
  no.space = TRUE, ci = TRUE, ci.level = 0.90, single.row = TRUE, type = 'latex',
  table.placement = 'h', dep.var.caption = "", dep.var.labels.include = FALSE,
  column.labels = c( "{\\bf Training}", "{\\bf Testing}" ), model.numbers = FALSE,
  title = "Summary of training and testing linear models with nuisance variables
  included after decomposition on the residualized data matrices (option 2).
  90\\% confidence intervals for the covariates are given in parentheses." )
```

Table 3: Summary of training and testing linear models with nuisance variables included after decomposition on the residualized data matrices (option 2). 90% confidence intervals for the covariates are given in parentheses.

	Training	Testing
GM1	-5.166^{***} (-8.318, -2.014)	-3.388* (-6.658, -0.118)
GM2	18.949^{***} (14.521, 23.377)	17.008^{***} (12.492, 21.525)
GM3	-0.561 (-4.049, 2.928)	-1.589 (-5.498, 2.319)
GM4	2.007 (-1.462, 5.477)	3.489 (-0.361, 7.339)
Age	$0.001 \ (-0.002, \ 0.003)$	$-0.001 \ (-0.003, \ 0.001)$
MMSE	$0.007^{***} (0.004, 0.010)$	$0.006^{**} (0.001, 0.010)$
Constant	$-1.472^{***} (-2.028, -0.915)$	$-1.272^{***} (-1.853, -0.691)$
Observations	89	83
\mathbb{R}^2	0.607	0.502
Adjusted R ²	0.578	0.463
Residual Std. Error	0.111 (df = 82)	0.118 (df = 76)
F Statistic	$21.104^{***} (df = 6; 82)$	$12.779^{***} (df = 6; 76)$
Note:		*p<0.1; **p<0.05; ***p<0.01

Predicting the full cognitive battery from the neuroimaging data

```
# Try to predict all the demographic variability from the imaging data. We use
# `mycoption O` to try to reduce correlation in low-dimensional space. This
# enforces a new SCCAN constraint (not previously reported). (Nick: We've been
# using mycoption = O this whole time.)

trainingCognitiveMatrix <- as.matrix( trainingCognitiveData )

trainingCognitiveResiduals <- residuals( lm(
    trainingCognitiveMatrix ~ trainingCognitiveData$mmse + trainingCognitiveData$age ) )

trainingImageResiduals <- residuals( lm( trainingImageData ~ trainingCognitiveData$mmse ) )

cognitiveSccanResult <- sparseDecom2(
    inmatrix = list( scale( trainingImageResiduals ),</pre>
```

```
scale( trainingCognitiveResiduals ) ),
     its = 20, mycoption = 0, sparseness = c(0.02, -0.05), nvecs = 11,
     inmask = c(grayMatterMask, NA), cthresh = c(1000, 0), verbose = TRUE,
     smooth = 0.5)
trainingImagePredictors <- trainingImageData %*% cognitiveSccanResult$eig1
colnames( trainingImagePredictors ) <-</pre>
  paste0( "GM", c( 1:ncol( trainingImagePredictors ) ) )
trainingCognitivePredictors <- trainingCognitiveResiduals %*% cognitiveSccanResult$eig2
testingImagePredictors <- testingImageData %*% cognitiveSccanResult$eig1
colnames( testingImagePredictors ) <-</pre>
  pasteO( "GM", c( 1:ncol( testingImagePredictors ) ) )
testingCognitiveMatrix <- as.matrix( testingCognitiveData )</pre>
\texttt{testingCognitivePredictors} \ \texttt{\leftarrow} \ \texttt{testingCognitiveMatrix} \ \%*\% \ \texttt{cognitiveSccanResult\$eig2}
eigenImageList <- matrixToImages( t( cognitiveSccanResult$eig1 ), grayMatterMask )</pre>
cognitivePredictorNames <- rep('NA', ncol( cognitiveSccanResult$eig2 ) )</pre>
weights <- rep( 'NA', ncol( cognitiveSccanResult$eig2 ) )</pre>
correlations <- rep( 'NA', ncol( cognitiveSccanResult$eig2 ) )</pre>
predictorRegions <- rep( 'NA', ncol( cognitiveSccanResult$eig2 ) )</pre>
predictorRegionPlot3DFiles <- rep( 'NA', ncol( cognitiveSccanResult$eig2 ) )</pre>
for( i in 1:ncol( cognitiveSccanResult$eig2 ) )
  trainingCognitiveDataFrame <- data.frame( Cognitive = trainingCognitivePredictors[, i],</pre>
    trainingImagePredictors, Age = trainingCognitiveData$age,
    MMSE = trainingCognitiveData$mmse )
  lmFormula <- formula( paste( "Cognitive ~ Age + MMSE + ",</pre>
    paste0( "GM", c( 1:ncol( trainingImagePredictors ) ), collapse = '+' ),
    collapse = '+' ) )
  trainingLm <- lm( lmFormula, data = trainingCognitiveDataFrame )</pre>
  trainingLmStats <- bigLMStats( trainingLm )</pre>
  testingCognitiveDataFrame <- data.frame( Cognitive = testingCognitivePredictors[, i],</pre>
    testingImagePredictors, Age = testingCognitiveData$age,
    MMSE = testingCognitiveData$mmse )
  testingCorrelation <- cor( testingCognitivePredictors[, i], predict( trainingLm,</pre>
    newdata = testingCognitiveDataFrame ) )
  correlations[i] <- format( testingCorrelation, digits = 3 )</pre>
  nonZeroIndices <- which( abs( cognitiveSccanResult$eig2[, i] ) > 0 )
  nonZeroNames <- colnames( trainingCognitiveData )[nonZeroIndices]</pre>
  nonZeroWeights <- abs( cognitiveSccanResult$eig2[nonZeroIndices, i] )</pre>
  cognitivePredictorNames[i] <- paste( nonZeroNames, collapse = ', ' )</pre>
  weights[i] <- paste( format( nonZeroWeights, digits = 3 ), collapse = ', ' )</pre>
  regions <- reportAnatomy( list( eigenImageList[[i]] ), grayMatterMask, 0.5 )</pre>
  predictorRegions[i] <- paste( regions, collapse = ", " )</pre>
```

```
significantIndices \leftarrow which( p.adjust( trainingLmStats$beta.pval ) < 0.1 ) - 2
  visualizationImages <- list()</pre>
  for( j in 1:length( significantIndices ) )
    visualizationImages[[j]] <-</pre>
      abs( antsImageClone( eigenImageList[[significantIndices[j]]] ) )
    }
  brain <- renderSurfaceFunction( surfimg = list( aalImage ), alphasurf = 0.1,</pre>
      funcimg = visualizationImages, smoothsval = 1.5, smoothfval = 0,
      mycol = rainbow( length( visualizationImages ) ) )
  id <- par3d( "userMatrix" )</pre>
  rid <- rotate3d( id, -pi / 2, 1, 0, 0 )
  rid2 <- rotate3d( id, pi / 2, 0, 0, 1 )
  par3d( userMatrix = id )
  predictorRegionPlot3DFiles[i] <- paste0( figuresDirectory,</pre>
    "predictorSccanEigenImages3D_Vector", i )
  dd <- make3ViewPNG( rid, id, rid2, predictorRegionPlot3DFiles[i] )</pre>
  predictorRegionPlot3DFiles[i] <- paste0( figuresDirectory,</pre>
    "predictorSccanEigenImages3D_Vector", i, ".png" )
  par3d( userMatrix = id )
  }
cognitiveSccanDataFrame <- data.frame( 1:ncol( cognitiveSccanResult$eig2 ),</pre>
  cognitivePredictorNames, weights, correlations )
colnames( cognitiveSccanDataFrame ) <- c( 'Vector', 'Cognitive predictors',</pre>
  'Eigenvector weights', 'Correlation')
pander( cognitiveSccanDataFrame, style = "rmarkdown", caption = "The set of
  cognitive predictors (with non-zero weights) for each eigenvector." )
```

Table 4: The set of cognitive predictors (with non-zero weights) for each eigenvector.

Vector	Cognitive predictors	Eigenvector weights	Correlation
1	socialcomportment	0.107	0.108
2	naming_adj, JOLO_adj	1.28e-01, 6.52e-07	0.41
3	fluency_adj	0.123	0.228
4	apathy	0.122	0.463
5	fluency_adj, recog_adj	3.20e-07, 1.11e-01	0.000591
6	$speech_adj$	0.109	-0.108
7	reading_adj	0.108	0.185
8	$semantic_adj$	0.111	-0.0848
9	age, $JOLO_adj$	1.07e-01, 7.83e-08	-0.319
10	$\operatorname{dig_bwd_adj}$	0.107	0.198
11	agitation	0.136	0.176

```
cognitiveSccanDataFrame <- data.frame( 1:ncol( cognitiveSccanResult$eig2 ),
   predictorRegions )
colnames( cognitiveSccanDataFrame ) <- c( 'Eigenvector', 'Predictor regions' )</pre>
```

pander(cognitiveSccanDataFrame, style = "rmarkdown", caption = "The set of
 predictor regions for each eigenvector.")

Table 5: The set of predictor regions for each eigenvector. (continued below)

Eigenvector
1
2
3
4
5
6
7
8
9
10
11

Predictor regions

Frontal_Inf_Oper_R, Frontal_Inf_Tri_R, Frontal_Inf_Orb_R, Insula_R
Temporal_Mid_R, Temporal_Inf_R
Precuneus_R

Frontal_Sup_Medial_L, Frontal_Med_Orb_L, Cingulum_Ant_L
Temporal_Mid_L, Temporal_Inf_L
Fusiform_L, Temporal_Inf_L
ParaHippocampal_R, Fusiform_R
SupraMarginal_R, Temporal_Sup_R
Frontal_Sup_R
SupraMarginal_L, Temporal_Sup_L, Temporal_Mid_L
Frontal_Mid_L







Figure 6: Eigenvector 1 of the gray matter view (cf Table 5).





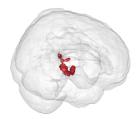


Figure 7: Eigenvector 2 of the gray matter view (cf Table 5).







Figure 8: Eigenvector 3 of the gray matter view (cf Table 5).





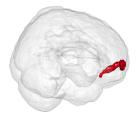


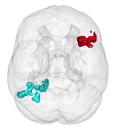
Figure 9: Eigenvector 4 of the gray matter view (cf Table 5).

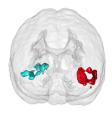






Figure 10: Eigenvector 5 of the gray matter view (cf Table 5).





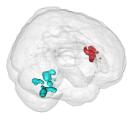


Figure 11: Eigenvector 6 of the gray matter view (cf Table 5).

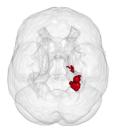






Figure 12: Eigenvector 7 of the gray matter view (cf Table 5).





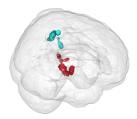


Figure 13: Eigenvector 8 of the gray matter view (cf Table 5).







Figure 14: Eigenvector 9 of the gray matter view (cf Table 5).





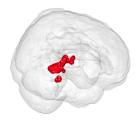


Figure 15: Eigenvector 10 of the gray matter view (cf Table 5).





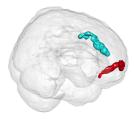


Figure 16: Eigenvector 11 of the gray matter view (cf Table 5).

Questions for Brian:

• Are the pseudo-eigenvectors ordered in terms of the variance explained? It would seem not. In the R code chunk priorinitialization, I can do the following

```
> i <- 1
> cognitiveSccanResult <- sparseDecom2(</pre>
inmatrix = list( scale( trainingImageData ), scale( trainingCognitiveMatrix ) ),
its = 20, mycoption = 0, sparseness = c(0, -0.5), nvecs = numberOfCognitiveLabels,
inmask = c(grayMatterMask, NA), cthresh = c(1000, 0), verbose = 0, ell1 = 10,
initializationList = eigenInitialization$initlist, priorWeight = priorWeightings[i]
smooth = 0, perms = 0)
> norm( as.matrix( cognitiveSccanResult$eig1[,1] ), type = "f" )
[1] 0.003823065
> norm( as.matrix( cognitiveSccanResult$eig1[,2] ), type = "f" )
[1] 0.005176968
> norm( as.matrix( cognitiveSccanResult$eig1[,3] ), type = "f" )
[1] 0.008712231
> norm( as.matrix( cognitiveSccanResult$eig1[,4] ), type = "f" )
[1] 0.01931158
> norm( as.matrix( cognitiveSccanResult$eig2[,1] ), type = "f" )
[1] 0.1086147
> norm( as.matrix( cognitiveSccanResult$eig2[,2] ), type = "f" )
[1] 0.08315364
> norm( as.matrix( cognitiveSccanResult$eig2[,3] ), type = "f" )
[1] 0.1380107
> norm( as.matrix( cognitiveSccanResult$eig2[,4] ), type = "f" )
[1] 0.1814885
```

It would seem that they're coordinated between the two views. Also, it would seem that this accounts for the colors not seeming to coordinate across Figures 1, 2, and 3.

- What does a negative sparseness value imply?
- (priorinitialization, line 350) Is the abs() call necessary? I thought the elements of the eigenvector are constrained to be positive.

- In the tutorial, you set copt <- 0 and write # 0 for most applications, 1 for priors. However, in the rcode chunk priorinitialization you use mycoption = 0. Was there a reason for this? It seems like 0 represents a compromise between spatial orthogonality and low-dimensional orthogonality as these might be at odds, i.e., the notion of eigenvector vs. pseudo-eigenvector.
- For the various options of dealing with nuisance variables, could one residualize out the effects of the nuisance variables and then just work on the "clean" data?

References

- 1. Hotelling, H. "Canonical Correlation Analysis (CCA)" J. Educ. Psychol. (1935):
- 2. Hotelling, H. "Relations Between Two Sets of Variants" Biometrika (1936): 321–377.
- 3. Maguire, E. A., Gadian, D. G., Johnsrude, I. S., Good, C. D., Ashburner, J., Frackowiak, R. S., and Frith, C. D. "Navigation-Related Structural Change in the Hippocampi of Taxi Drivers" *Proc Natl Acad Sci U S A* 97, no. 8 (2000): 4398–403. doi:10.1073/pnas.070039597
- 4. Nichols, T. and Hayasaka, S. "Controlling the Familywise Error Rate in Functional Neuroimaging: A Comparative Review" Stat Methods Med Res 12, no. 5 (2003): 419–46. doi:10.1191/0962280203sm341ra
- 5. Habeck, C., Stern, Y., and Alzheimer's Disease Neuroimaging Initiative. "Multivariate Data Analysis for Neuroimaging Data: Overview and Application to Alzheimer's Disease" Cell Biochem Biophys 58, no. 2 (2010): 53–67. doi:10.1007/s12013-010-9093-0
- 6. Shamy, J. L., Habeck, C., Hof, P. R., Amaral, D. G., Fong, S. G., Buonocore, M. H., Stern, Y., Barnes, C. A., and Rapp, P. R. "Volumetric Correlates of Spatiotemporal Working and Recognition Memory Impairment in Aged Rhesus Monkeys" Cereb Cortex 21, no. 7 (2011): 1559–73. doi:10.1093/cercor/bhq210
- 7. McKeown, M. J., Makeig, S., Brown, G. G., Jung, T. P., Kindermann, S. S., Bell, A. J., and Sejnowski, T. J. "Analysis of FMRI Data by Blind Separation into Independent Spatial Components" *Hum Brain Mapp* 6, no. 3 (1998): 160–88.
- 8. Calhoun, V. D., Adali, T., Pearlson, G. D., and Pekar, J. J. "A Method for Making Group Inferences from Functional MRI Data Using Independent Component Analysis" *Hum Brain Mapp* 14, no. 3 (2001): 140–51.
- 9. Calhoun, V. D., Liu, J., and Adali, T. "A Review of Group ICA for FMRI Data and ICA for Joint Inference of Imaging, Genetic, and ERP Data" Neuroimage 45, no. 1 Suppl (2009): S163–72. doi:10.1016/j.neuroimage.2008.10.057
- 10. Kandel, B. M., Wang, D. J. J., Gee, J. C., and Avants, B. B. "Eigenanatomy: Sparse Dimensionality Reduction for Multi-Modal Medical Image Analysis" *Methods* 73, (2015): 43–53. doi:10.1016/j.ymeth.2014.10.016
- 11. Avants, B. B., Libon, D. J., Rascovsky, K., Boller, A., McMillan, C. T., Massimo, L., Coslett, H. B., Chatterjee, A., Gross, R. G., and Grossman, M. "Sparse Canonical Correlation Analysis Relates Network-Level Atrophy to Multivariate Cognitive Measures in a Neurodegenerative Population." Neuroimage 84, (2014): 698–711. doi:10.1016/j.neuroimage.2013.09.048, Available at http://dx.doi.org/10.1016/j.neuroimage.2013.09.048
- 12. Avants, B. B., Cook, P. A., Ungar, L., Gee, J. C., and Grossman, M. "Dementia Induces Correlated Reductions in White Matter Integrity and Cortical Thickness: A Multivariate Neuroimaging Study with Sparse Canonical Correlation Analysis" *Neuroimage* 50, no. 3 (2010): 1004–16. doi:10.1016/j.neuroimage.2010.01.041
- 13. Kim, J., Avants, B., Whyte, J., and Gee, J. C. "Methodological Considerations in Longitudinal Morphometry of Traumatic Brain Injury" Front Hum Neurosci 7, (2013): 52. doi:10.3389/fnhum.2013.00052
- 14. Dhillon, P. S., Wolk, D. A., Das, S. R., Ungar, L. H., Gee, J. C., and Avants, B. B. "Subject-Specific Functional Parcellation via Prior Based Eigenanatomy." Neuroimage (2014): doi:10.1016/j.neuroimage.2014.05.026, Available at http://dx.doi.org/10.1016/j.neuroimage.2014.05.026
- 15. McMillan, C. T., Irwin, D. J., Avants, B. B., Powers, J., Cook, P. A., Toledo, J. B., McCarty Wood, E., Van Deerlin, V. M., Lee, V. M.-Y., Trojanowski, J. Q., and Grossman, M. "White Matter Imaging Helps Dissociate Tau from TDP-43 in Frontotemporal Lobar Degeneration." *J Neurol Neurosurg*

 $Psychiatry~84,~\text{no.}~9~(2013):~949-955.~\text{doi:}10.1136/\text{jnnp-}2012-304418,~\text{Available at http://dx.doi.org/}10.1136/\text{jnnp-}2012-304418}$

- 16. McMillan, C. T., Avants, B. B., Cook, P., Ungar, L., Trojanowski, J. Q., and Grossman, M. "The Power of Neuroimaging Biomarkers for Screening Frontotemporal Dementia." *Hum Brain Mapp* (2014): doi:10.1002/hbm.22515, Available at http://dx.doi.org/10.1002/hbm.22515
- 17. McMillan, C. T., Toledo, J. B., Avants, B. B., Cook, P. A., Wood, E. M., Suh, E., Irwin, D. J., Powers, J., Olm, C., Elman, L., McCluskey, L., Schellenberg, G. D., Lee, V. M.-Y., Trojanowski, J. Q., Van Deerlin, V. M., and Grossman, M. "Genetic and Neuroanatomic Associations in Sporadic Frontotemporal Lobar Degeneration." Neurobiol Aging 35, no. 6 (2014): 1473–1482. doi:10.1016/j.neurobiolaging.2013.11.029, Available at http://dx.doi.org/10.1016/j.neurobiolaging.2013.11.029