

# Impact of heparin on kinetic parameters of DNA endonuclease Cas9





Target Strand

Non-Target

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### **Motivation**

- Cas9, a DNA cutting protein important in bacterial defense against viruses, has potential as a DNAediting technology in humans.
- Experiments to date use a polymer called Heparin to reduce unspecific binding of Cas9 when studying its binding to target DNA
- We hypothesize Heparin interferes with the Cas9-DNA interaction and leads to inaccurately measured rate constants
- This work uses switchSENSE technology to show Heparin reduces the ability to characterize Cas9

## Background



Figure 1. Cas9 uses a guide RNA to find complementary target DNA for cleavage.

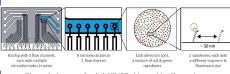


Figure 2. Layout of switchSENSE chip used in all experiments

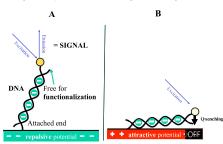


Figure 3. The DNA strand switches between a horizontal (A) and vertical (B) orientation based on the voltage applied to the gold surface.

# Single stranded Double Stranded DNA dsDNA with dsOverhang Figure 4. An overhang that is targeted by the Cas9-gRNA complex is attached to the surface tethered DNA.

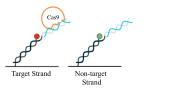


Figure 5. Red and green fluorophore are used to distinguish between specific and unspecific binding of Cas9.

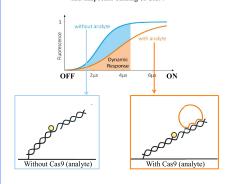


Figure 6: Upon analyte binding, DNA levers move more slowly, decreasing the dynamic response. The reverse occurs during unbinding.



Figure 7. Heparin is used in standard Cas9 studies, but may interfere with the Cas9-DNA interaction.

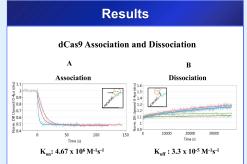
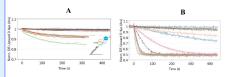


Figure 8. Decrease in dynamic response represents Cas9 association (a) while increase in dynamic response represent dissociation of Cas9 from DNA (b).

### 20 nM dCas9 Association with Heparin



**Figure 9. (a)** Unspecific binding was monitored via the fluorescence of green fluorophore tagged non-target DNA levers. **(b)** Heparin concentrations of 375 ng/mL (black) and higher led to a decrease in K<sub>on</sub> until no binding of Cas9 to target DNA could be observed.

### 200 nM dCas9 Association with Heparin

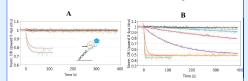
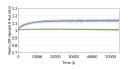
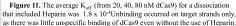


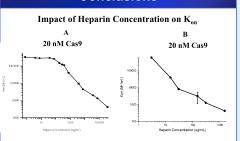
Figure 10. (a) Unspecific binding is observed in control samples (pink and green), but this is removed upon the addition of 3 ug/mL Heparin (b) At Heparin concentration of 3 ug/mL (orange), specific binding of dCas9 decreases, shown by the lower  $K_{on}$  rate constants as the Heparin concentration

### Dissociation of 20 nM, 40 nM, and 80 nM Cas9 with Heparin





### Conclusions



**Figure 12.**  $K_{on}$  values are shown to decrease as higher concentrations of Heparin are used in the association of 20 nM Cas9 (a) and 200 nm Cas9 (b).

### apoCas9 Association and Dissociation

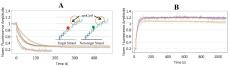


Figure 13. (a) Association of apoCas9 (b).apoCas9 dissociates completely, so lack of complete dissociation in previous experiments is specific to dCas9.

- Heparin reduces the ability for kinetic parameters of Cas9 to be measured
- Unspecific binding should not be completely repressed
- The switchSENSE method is able to measure specific and unspecific binding of Cas9 without the need for Heparin

### References

[Lang13] A. Langer, P.A. Hampel, W. Kaiser, J. Knezevic, T. Welte, V. Villa, M. Maruyama, M. Svejdal, S. Jähner, F. scher, R. Strasser, U. Rant, "Protein Analysis by Time-Resolved Measurements with an Electro-Switchable DNA Chip," attree Communications, 1, (2013).

sature Communications. 1, (2013).
. [Ran13] F.A. Ran, P.D. Hsu, J. Wright, V. Agarwala, D.A. Scott, F. Zhang, "Genome Engineering Using the CRISPI 2as 9 System," Nature Protocols. 8, 2281, (2013).

inding Kinetics of Protein Analytes with a Dynamically Switchable Biosurface, "J. Am. Chem. Soc. 134, 15225-15228, 012)

2012) [Bolul 6] M.F. Bolukbasi, A. Gupta, S.A. Wolfe, "Creating and Evaluating Accurate CRISPR-Cas9 Scalpels for Genom largery," Nature Method. 13, 41, (2016).
[Mall 13] P.M.Ri, K.M. Esvelt, G.M. Church, "Cas9 as a Versatile Tool for Engineering Biology," Nature Methods, 10,

5. [Mali 3]P. Mali, K.M. Esvelt, G.M. Church, "Cas9 as a Fersatile Fool for Engineering Biology," "Nature Methods, It 957, (2013).
6. [Rich16] C.D. Richardson, G.R. Ray, M.A. DeWitt, G.L. Curie, J.E. Corn, "Enhancing Homology Directed Genome Fatimes by Candaviscally Active and Inactive CRISPR-Cas9 Using Asymmetric Donor DNA," Nature Biotechnology, 34

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