

The game of 15 sticks in *E. coli*

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1 Introduction

1.1 The game of 15 sticks¹

We design genetic circuits in *E. coli* that play the following game:

Two players start with 15 sticks and alternately take 1, 2 or 3 sticks.
The last player to take a stick loses.

1.2 General design

We encode in binary the number of sticks left as

$$\text{number of sticks left} = 8s_3 + 4s_2 + 2s_1 + s_0. \quad (1)$$

By the 4-periodicity of the winning strategy (see §1.4), only the two lowest bits s_1/s_0 matter to the player. The response of a player is a number 1, 2, or 3, which we encode in binary as

$$\text{number of sticks taken by a player} = 2r_1 + r_0. \quad (2)$$

The bits in (1)–(2) are transmitted between compute units by intercellular signaling molecules, cf. §1.5. Although our players are essentially identical, we model them separately with one difference: Player A (B) becomes active when the extracellular signal w_A (w_B) is supplied.

The *Subtractor* module keeps track of the number of sticks left (1). To appreciate the key design issue, suppose we manually transmit the response $r_1 r_0 = 01$ of a player into the medium of the subtractor. It should compute the new number of the sticks, yet, it has to do so once and not keep subtracting the value $r_1 r_0$ that is still present. One solution is single-use subtractors but then the result of each computation has to be imparted to an unused one. A feasible design seems to require some separation of reading/storing the current state, computing the new state and communicating it – temporally or spatially. We propose a

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subtractor composed of two parts A and B, and a workflow as shown in Table 1. Thus, in Phase A characterized by the supply of the master signal w_A , Subtractor A emits the current state s , Player A responds with r , while Subtractor B reads s/r and computes the new state silently and remembers the result in order to emit it in the subsequent Phase B. We use the variables $d_3/d_2/d_1/d_0$ for this internal state. The medium is cleared, before Phase B is initiated by supplying the master signal w_B . Briefly, we require a gated memory module and a delayed response, cf. §1.4.

	Phase A	Interphase I	Phase B
Experimenter	supply w_A	clear medium	supply w_B
Player A	in: s , out: r	–	–
Subtractor A	out: new s	–	in: s/r ; compute
Subtractor B	in: s/r ; compute	–	out: new s
Player B	–	–	in: s , out: r

Table 1: Workflow: Phases I/A/I/B alternate until the game is over.

1.3 Conventions

As shown in Table 4, we encode w_A by L-arabinose, w_B by IPTG, etc. Our convention is $w_A = 1$ if the corresponding messenger molecule L-arabinose is present in the medium/cell in sufficiently high quantity, and $w_A = 0$ otherwise. We write $\#w_A$ for the *number* of L-arabinose molecules in the numerical simulations. Where suitable, we indicate the semantics of an element thus: $\langle \text{signal} \rangle$, (protein) – often a transcription factor, and $[\text{promoter}]$; this is consistent with the symbols that we employ in the SimBiology schemes. By $\langle w_A \rangle$ we mean “the signal molecule that encodes w_A ”, i.e. L-arabinose. Ditto for the other signals.

Abbreviations: RBS = ribosome binding site, QS = quorum sensing, AHL = acyl homoserine lactone, 3OC6 = 3OC6-HSL (etc.), SM = supplementary material, DNF = disjunctive normal form, txn = transcription, TF = transcription factor.

1.4 Logic circuits

Player. Both players follow the same deterministic strategy, which is a winning strategy for the first-mover. They are identical except that player A responds when w_A is present and player B responds upon w_B . The logic is shown in Table 2.

Subtractor. In Phase A, Subtractor B senses the current number of sticks (1), emitted by Subtractor A, and the response (2) from Player A, and computes the difference d . It has four compute units Bit 0 to Bit 3 that are also spatially separated (cf. §1.5). The subtraction proceeds from Bit 0 to Bit 3. Bit 0 senses the current s_0/r_0 and computes d_0 (silently) and the carry flag c_1 (emitted immediately). The carry flag is passed on to Bit 1. Bit 1 subtracts

r_1 and c_1 from s_1 . The resulting carry flag c_2 is passed on to Bit 2, which subtracts c_2 from s_2 . Then Bit 3 subtracts the resulting carry flag c_3 from s_3 . Each Bit 0/2/3 is in fact a “half-subtractor” with 2 inputs, whilst Bit 1 is a “full subtractor” with 3 inputs. See Table 3 for their truth table. Fig. 6 shows a reference logic circuit for Bit 0 and Bit 1 that illustrates the principle of delayed response. Since Bits 0/2/3 are a subfunctionality of Bit 1, we will not discuss them in detail.

Winning strategy:					w_A	s_1	s_0	r_1	r_0
					0	0	0	0	0
					0	0	1	0	0
					0	1	0	0	0
					0	1	1	0	0
					1	0	0	1	1
					1	0	1	0	1
					1	1	0	0	1
					1	1	1	1	0

Sticks left				Take
15	11	7	3	2
14	10	6	2	1
13	9	5	1	1
12	8	4		3

Table 2: Both players follow the same strategy. Truth table and logic circuit for Player A. The input is a wake-up signal w_A and the bits s_1/s_0 of the current number of sticks (1). The output is the number of sticks taken (2). The output is cut off unless w_A is present. The circuit for Player B is identical except that the output is controlled by w_B .

Half-subtractor:				s_1	r_1	c_1	d_1	c_2
				0	0	0	0	0
				0	0	1	1	1
				0	1	0	1	1
				0	1	1	0	1
				1	0	0	1	0
				1	0	1	0	0
				1	1	0	0	0
				1	1	1	1	1

s_3	c_3	d_3	—
s_2	c_2	d_2	c_3
s_0	r_0	d_0	c_1
0	0	0	0
0	1	1	1
1	0	1	0
1	1	0	0

$$\begin{aligned}
 & 8s_3 + 4s_2 + 2s_1 + s_0 \\
 & - 2r_1 + r_0 \\
 & = 8d_3 + 4d_2 + 2d_1 + d_0
 \end{aligned}$$

Table 3: The subtractor computes $d = s - r \pmod{16}$. As r only has these two bits, a half-subtractor is sufficient for the bits $d_0/d_2/d_3$ and the carry flags c_1/c_3 , and only d_1/c_2 require the full subtractor. We have $d_0 = (s_0 \text{ XOR } r_0)$; $c_1 = \text{not}(s_0 \geq r_0)$; $d_1 = ((s_1 + r_1 + c_1) \bmod 2)$; $c_2 = \text{not}(s_1 \geq r_1 \geq c_1)$. See Fig. 6 for a logic circuit.

1.5 Experimental design

We distinguish *extracellular* chemical signals that are supplied by the experimenter (the master signals w_A/w_B) and *intercellular* signals that are produced and sensed by cells. For an experiment in a single test tube we need orthogonal intercellular channels: *three* for (1) (s_3 has no downstream), *two* for (2), and *three* for the carry flags (see §1.4). Thus we need

eight orthogonal intercellular channels. One channel could be spared at the expense of a more involved computation of the last useful carry flag c_3 . Du et al. (2020) designed ten de novo intercellular signaling pathways and optimized four known QS pathways. However, only *eight* worked well orthogonally on the sensor level and seven on the promoter level (Du et al., 2020, Fig. 3c/g). We have assigned in Table 4 the signals to minimize crosstalk, in particular avoiding LuxR and RpaR in one cell, heeding the advice of Du et al. (2020, p.6).

	$\langle \text{signal} \rangle$, (txn factor), [promoter]	Primary reference	Details
w_A	$\langle \text{Ara} \rangle \rightarrow (\text{AraC}^*) \rightarrow [\text{BAD}]$	Nielsen et al. 2016, SM:VII.M	§2.5/p.6
w_B	$\langle \text{IPTG} \rangle \rightarrow (\text{LacI}) \rightarrow [\text{Tac}]$	Nielsen et al. 2016, SM:VII.M	§2.5/p.6
r_0	$\langle \text{3OC6-HSL} \rangle \rightarrow (\text{LuxR}) \rightarrow [\text{Lux}^*]$	Du et al. 2020, SM:p.3	§2.6/p.6
r_1	$\langle \text{IV-HSL} \rangle \rightarrow (\text{BjaR}) \rightarrow [\text{Bja}^*]$	Du et al. 2020, SM:p.2	§2.6/p.7
s_0	$\langle \text{DAPG} \rangle \rightarrow (\text{PhlF}) \rightarrow [\text{PhlF}^*]$	Du et al. 2020, SM:p.3	§2.6/p.7
c_1	$\langle \text{Sal} \rangle \rightarrow (\text{NahR}) \rightarrow [\text{Sal}]$	Du et al. 2020, SM:p.3	§2.6/p.7
s_1	$\langle \text{pC-HSL} \rangle \rightarrow (\text{RpaR}) \rightarrow [\text{Rpa}^*]$	Du et al. 2020, SM:p.2	§2.6/p.7
c_2	$\langle \text{MMF} \rangle \rightarrow (\text{MmfR}) \rightarrow [\text{Mmf}^*]$	Du et al. 2020, SM:p.2	§2.6/p.7
s_2	$\langle \text{3OC12-HSL} \rangle \rightarrow (\text{LasR}) \rightarrow [\text{Las}^*]$	Du et al. 2020, SM:p.3	§2.6/p.8
c_3	$\langle \text{NG} \rangle \rightarrow (\text{FdeR}) \rightarrow [\text{FdeA}]$	Du et al. 2020, SM:p.3	§2.6/p.8

Table 4: Extra-/intercellular $\langle \text{signals} \rangle$ with downstream (factors) and [promoters].

To facilitate debugging we partition the workflow into six spatial compartments (for example, 10 mL centrifuge tubes) that require manual transmission of the medium: two for the players (A and B) and four for the subtractor (Bits 0–3). Unfortunately, this does not automatically reduce the number of intercellular channels needed. Each Bit compartment contains two subpopulations of cells as part of Subtractor A or B, referred to as Bit 0A, Bit 0B, etc.

To initialize the experiment, add $\langle w_B \rangle$ with $\langle s_i \rangle$ to each Bit i , causing Subtractor A to compute and remember $d_i = 1$. Then wash and replace the medium. In Phase A, do the following steps, adding $\langle w_A \rangle$ before each step.

1. Having added $\langle w_A \rangle$ to Bit 0 and Bit 1, wait for emission of s_1 and s_0 .
2. Admix the media from Bits 0/1 containing s_0/s_1 to Player A, wait for response. Note, the medium may or may not contain the actual signal molecules $\langle s_0 \rangle / \langle s_1 \rangle$.
3. Admix the medium from Player A containing $r_0/r_1/s_0/s_1$ to Bit 0. Wash Player A, replenish with fresh medium. Wait for: Bit 0B to compute the difference d_0 (silent); Bit 0A to compute the carry flag c_1 .
4. Admix the medium from Bit 0 containing c_1 to Bit 1. Wash and replenish Bit 0. Wait for: Bit 1B to compute the difference d_1 (silent); Bit 1A to compute the carry flag c_2 .
5. Admix the medium from Bit 1 containing c_2 to Bit 2. Wash and replenish Bit 1. Wait for: Bit 2B to compute the difference d_2 (silent); Bit 2A to compute the carry flag c_3 .

6. Admix the medium from Bit 2 containing c_3 to Bit 3. Wash and replenish Bit 2. Wait for Bit 3B to compute the difference d_3 (silent). Wash and replenish Bit 3.

This concludes Phase A. In the interphase it may be necessary to clear the media again and generally wait for the steady state. Phase B proceeds similarly. The game finishes when the internal states d are zero. We can detect this, for example, by coupling expression of reporter proteins to the memory units (details omitted).

Refinements could include a) dealing with a player who attempts to take more sticks than (1); b) dedicated sensor cells that report on signaling molecules in order to monitor the communication channels; c) cells that emit $\langle s_3 \rangle / \langle s_2 \rangle / \langle s_1 \rangle / \langle s_0 \rangle$ upon one signal, e.g. tetracycline, in order to reboot the game.

2 Genetic toolbox

2.1 Constitutive parts

Constitutive promoter. The iGEM part BBa_J23100 is a strong constitutive *E. coli* σ^{70} promoter. We refer to it as P_c .

cI. The optimized and orthogonal variants $cl^a := cl_{opt}$ and $cl^b := cl_{5G6G,P}$ of the transcription factor λ cI from (Brödel et al., 2016, Fig. 4c/d) require no cofactors and show about 10-fold activation of their “forward” promoter ($[cl^a_+]$ and $[cl^b_+]$) and a similar-fold repression of their “backward” promoter ($[cl^a_-]$ and $[cl^b_-]$). Arkin et al. (1998) performed a detailed stochastic computational analysis of the phage λ lysis-lysogeny decision circuit.

2.2 3-input gates by Cello

Nielsen et al. (2016) reported among the good performers: $r_0 = 0x0E(s_0, s_1, w_A)$; $r_1 = \text{not } 0xF6(s_0, s_1, w_A)$; $c_2 = \text{not } 0x8E(c_1, r_1, s_1) = 0x4D(c_1, s_1, r_1)$, which is symmetric in r_1/c_1 ; but we couldn’t find the 3-XOR, see §2.9.

We used their 0x0E for r_0 , replacing the sensors and replacing the repressor PhlF (which clashes with the s_0 sensor) by HlyIIR (also from (Nielsen et al., 2016)). The last operation is a “poor man’s OR gate”; we let it control (cl^a) (§2.1) that triggers the synthesis of $\langle r_0 \rangle$. We observed that r_1 can reuse most of this circuitry and implemented the remainder using ribo-computing (details omitted). For c_2 , we took not 0x8E, replacing the sensors and replacing the final “not(not(\cdot) OR not(\cdot))” by a ribo-AND gate with cross-inhibition (see §2.8/p.8).

2.3 Intracellular pathways (transducers)

AmtR. The TetR-like AmtR regulates the nitrogen starvation response in the Gram-positive *C. glutamicum* (Jakoby et al., 2000). This repressor is released by the trimeric

adenylylated signaling protein GlnK (Beckers et al., 2005; Sevvana et al., 2017), which is present upon nitrogen starvation. The AmtR box and the regulon is characterized in (Beckers et al., 2005) and several variants are compared in (Muhl et al., 2009). It seems clear that AmtR sits on the DNA as a dimer (Sevvana et al., 2017), (Schwab, 2019, §3.4.2).

Further, we use the transcription repressors BetI and HlyIIR, see (Nielsen et al., 2016).

2.4 Insulation of genetic context and RBS

To reduce “part-junction interference” and maintain relative promoter strengths, Lou et al. (2012) (and Nielsen et al. (2016)) inserted a ribozyme just upstream of the RBS, which cuts off the 5'-UTR leader sequence and the scar, if present. This remedies early saturation of transcription (Lou et al., 2012, SM:Fig. 5) and thus increases the choice of RBS for regulating expression on the translation level (Lou et al., 2012, SM:Fig. 4), (Salis, 2011). An additional hairpin helps expose the RBS (Lou et al., 2012; Carrier and Keasling, 1997). For our purposes, standard RBS sequences such as BBa_J61100 can be used.

2.5 Extracellular signaling

Recall that the two master signals w_A/w_B are encoded by L-arabinose/IPTG. Following Nielsen et al. (2016, SM, VII.M) we posit a truncated (AraC*) that has reduced crosstalk with IPTG. The sequences can be found in (Nielsen et al., 2016, SM, Table S8) for (Ara*) and (LacI), and in (Nielsen et al., 2016, SM, Table S9) for [BAD] and [Tac].

2.6 Intercellular pathways

Here we collect key aspects of the intercellular receivers. 3OC6, IV and 3OC12 belong to the AHL QS family, whereas pC has a p-coumaric acid group rather than a fatty acyl group.

According to Hirakawa et al. (2011, p.2598), LuxR homologs (which function as homoserine lactones receivers) are homodimeric. Du et al. (2020) used the LuxR and LuxR homologs LasR (Fuqua et al., 1996, Table 1), RpaR (Hirakawa et al., 2011, p.2598), BjaR (Lindemann et al., 2011). The TetR family are transcriptional repressors that sit on the operons and block the corresponding promoters (Ramos et al., 2005). They are released by specific inducers (often antibiotics) via a conformational change. It is suggested (Ramos et al., 2005; Cuthbertson and Nodwell, 2013) that the TetR-type proteins are usually homodimers with two slots for the inducer. PhlF and MmFR are TetR homologs.

Of note, Du et al. (2020, SM:p.8) reported that sender cells enter the log-phase with delay compared to control samples (Sal, DAPG: $\approx 12h$; 3OC6, 3OC12, pC, MMF: $\approx 3h$; IV, NG: no delay). See (Du et al., 2020, SM) for sequences, biosynthesis and induction curves.

3OC6. The transcription activator LuxR occurs in Gram-negatives such as *V. fischeri*, that is permeable to the (auto)inducer 3-oxo-C6-homoserine lactone (3OC6-HSL). It binds

to the N-terminal of (LuxR), which otherwise inhibits its functional C-terminal (Stevens et al., 1994). The purified C-terminal binds upstream of the *lux* box (which is centered at -42.5bp (Egland and Greenberg, 1999)); however, together with the RNA Pol, it protects the *lux* box and the *lux* operon promoter (Stevens et al., 1994).

IV. Isovaleryl-HSL (IV) is a branched-chain fatty acyl-HSL from soybean symbiont *B. japonicum* that regulates (BjaR). In the presence of IV, dimerized BjaR activates [Bja] (Lindemann et al., 2011).

DAPG. 2,4-diacetylphloroglucinol (DAPG) is a broad-spectrum antibiotic phenol produced by some Gram-negative *P. fluorescens*. Bangera and Thomashow (1999) identified the *phlABCD* operon (in strain Q2-87) required for the synthesis of 2,4-DAPG and its precursor MAPG. This operon is flanked by genes *phlE* (efflux protein) and *phlF* (repressor protein, (Bangera and Thomashow, 1999, p.3162)), which are separately transcribed. The helix-turn-helix motif of *phlF* resembles that of the TetR-family (Bangera and Thomashow, 1999, p.3161). Schnider-Keel et al. (2000) investigated auto-induction of DAPG synthesis (in strain CHA0), and reported, in particular, strong repression by extracellular $\langle \text{Sal} \rangle$ (indeed, *P. fluorescens* has a NahR-homolog HQ912.21075); but the *phlF* mutant was unaffected by it. CHA0 also contains the gene *phlG* to specifically degrade DAPG to MAPG (Bottiglieri and Keel, 2006), and another TetR-analog *PhlH* already found by Schnider-Keel et al. (2000, p.1219) was shown by Yan et al. (2017) to repress *phlG*.

Sal. The transcription activator (NahR) binds to the recognition site of [Sal] at -83 to -45 without the inducer (Huang and Schell, 1991, p.10837), (Schell and Wender, 1986). Schell et al. (1990) suggested that the active configuration of (NahR) is a tetramer, while Park et al. (2005) reported that there could be three different complexes (NahR):[Sal]. Possibly (Peking iGEM 2013 team, 2013), $4 \times (\text{NahR})$ bind to the DNA, and transcription starts once a $\langle \text{Sal} \rangle$ binds to each (NahR).

pC. The transcription factor RpaR protein senses P-coumaroyl-HSL with p-coumaric acid rather than fatty acids. Hence, it shows good orthogonality to many AHL-HSL receiver proteins (Schaefer et al., 2008). The *R. palustris* transcriptional regulator RpaR, when purified, binds an inverted repeat element centered at -48.5bp of its promoter (Hirakawa et al., 2011). The complex $\langle \text{pC} \rangle : (\text{RpaR})$ bound to the promoter activates transcription (Hirakawa et al., 2011, Discussion).

MMF. Methylenomycin furans (Mmf) are precursors of methylenomycin, an antibiotic produced by *S. coelicolor* (Zhou et al., 2020). As a TetR-family transcriptional regulator, *MmfR* represses [Mmf], but accumulation of $\langle \text{Mmf} \rangle$ releases it (O'Rourke et al., 2009). The TetR homolog *MmfR* forms a homodimer (Zhou et al., 2020), which contains two Mmf recognition sites. Different R-groups of Mmfs may alter the binding affinity (Zhou et al., 2020, p.2-3). Du et al. (2020, p.2) use an MMF with a branched side chain.

3OC12. Two 3-oxo-dodecanoyl-homoserine lactone (3OC12-HSL) molecules bind irreversibly to the dimerized (LasR) from *P. aeruginosa*. The activated LasR activates several promoters with high affinity with no or positive cooperativity (Schuster et al., 2004; Grant et al., 2016). Du et al. (2020, p.2) use a custom promoter [Las*].

NG. Naringenin (NG) is the main flavone from grapefruits (Siedler et al., 2014; Hein et al., 2014). It binds the homodimeric protein FdeR from *H. seropedicae*. There are two slots for NG, whose binding induces a conformational change of the dimer. The activated FdeR promotes the “Flavanone degradation” operon [FdeA].

2.7 Memory module

As explained in §1.2 (cf. Table 1/3 and Fig. 6), we need a memory module to record and recall the result of the subtraction. To that end we use a “reversible” recombinase. Specifically, the Bxb1 gp35/gp47 serine-integrase/excisionase (Int/Xis) direct the infection cycle of Bxb1 in *M. smegmatis*. Stoichiometry, start codons, degradation rates (proteolysis tags) and DNA substrate copy number were optimized in *E. coli* in (Bonnet et al., 2012) to assemble a memory bit that is stable and responsive over several generations. A promoter of choice is flanked by Int recognition sites and its direction is *set* by Int and *reset* by Int+Xis. Switching times of ~ 4 h were demonstrated but the authors expected 30min to be realistic. Their final construct (Bonnet et al., 2012, Fig. 4A) consists of an Int gene and a Xis+Int operon induced by two separate signals, which facilitates stoichiometry optimization. However, in our design, an external pulse (say, $\langle w_B \rangle$) controls Int, and an internal compute unit (say, $s_0 \text{ XOR } r_0$) controls Xis. The *reset* state corresponds to positive output, i.e. the memory bit is *on*. The flanked promoter is activated by another external signal (say, $\langle w_A \rangle$).

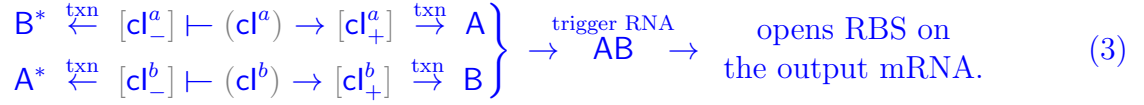
2.8 Enhanced AND gate

HrpR/HrpS. Wang et al. (2011, Fig. 1) proposed an AND gate in *E. coli* using the heterodimeric transcription factor HrpR:HrpS from *P. syringae*. The dimer binds upstream of the promoter hrpL and opens the σ^{54} -RNAP-hrpL transcription complex. Increasing the copy number of this weak promoter improves the dynamic range (Wong et al., 2015, p.4).

Ribocomputing. Green et al. (2017, Fig. 2e) combined short RNAs A and B into a “trigger” RNA which opens an RBS-occluding stemloop on the output mRNA (cf. Fig. 1c).

Cross-inhibition. Even in our idealized simulations such AND gates produce undesired leakage within larger circuits. We propose that *low* input *a* inhibit the downstream of *b*, and vice versa: we let $a \rightarrow cl^a$ and $b \rightarrow cl^b$ produce two orthogonal bidirectional transcription

factors as in §2.1. They, in turn, activate $[cl_+^a]/[cl_+^b]$ and repress $[cl_-^a]/[cl_-^b]$:



Herein, A and B form a trigger RNA AB as in (Green et al., 2017, Fig. 2e), including toeholds for preferential binding of the complementary RNAs A* and B*. The cross-inhibition is more effective if $[cl_-^a] > [cl_+^b]$ and $[cl_-^b] > [cl_+^a]$ in terms of the output of the promoters.

2.9 3-input XOR

The subtractor result bit (cf. Table 3 and §2.2)

$$d_1 = 0x69(s_1, r_1, c_1) = \text{not } 0x96(s_1, r_1, c_1) = 3\text{-XOR}(s_1, r_1, c_1) = ((s_1 + r_1 + c_1) \bmod 2) \quad (4a)$$

$$= (s_1 \wedge \bar{r}_1 \wedge \bar{c}_1) \vee (\bar{s}_1 \wedge r_1 \wedge \bar{c}_1) \vee (\bar{s}_1 \wedge \bar{r}_1 \wedge c_1) \vee (s_1 \wedge r_1 \wedge c_1) \quad (4b)$$

is particularly difficult because the result flips whenever any input is flipped. It is a concatenation of two XOR gates (cf. Fig. 6). We considered:

- Cascading two XOR gates built on tandem promoters as in (Nielsen et al., 2016, Fig. 3).
- Cascading two XOR gates, each as in (Wong et al., 2015, Fig. 5) but with two orthogonal heterodimeric transcription activators (cf. §2.8).
- RNA AND-gates as in Fig. 1b instead of the protein heterodimer.
- **RNA-only gate** inspired by (Green et al., 2017, Fig. 2e). Each input expresses its own mRNA, whose 5' tail opens its RBS stemloop. Each input also expresses complementary RNAs that sequester other tails thus closing up other RBS. Upon all three inputs, all complementary RNAs form a ring, so that the 5' tails reopen their RBS.
- Three orthogonal resettable recombinases as in (Chiu and Jiang, 2017, Fig. 2g).
- Cas13/ribozymes to cut mRNA instead of zipping up the RBS as in the final design.
- The recombinase-based (Weinberg et al., 2017) and the multi-cell (Ausländer et al., 2017) one-bit adders, mentioned here for completeness.

Eventually, we implemented the DNF (4b) using RBS switches. Each input signal produces 1) an output mRNA; 2) two short RNAs that inhibit the output mRNA of the other two inputs using the “three-way junction” mechanism of (Kim et al., 2019, Fig. 1c); 3) a co-trigger RNA: upon all 3 signals, the co-trigger RNAs form a trimer that opens the RBS of a fourth (constitutively transcribed) output mRNA as in (Green et al., 2017, Fig. 2e). Fig. 2 shows the corresponding genetic circuits:

- The first circuit constitutively provides the transcription factors to sense the signals s_1 (pC-HSL), r_1 (IV-HSL) and c_1 (Sal).

- The second circuit is activated by pC-HSL through four independent promoters [Rpa*] (the copy number and strength of each may have to be tuned): two kinds of inhibitory trigger RNAs (bd and cd), one “co-trigger” RNA mk and an output mRNA. The trigger RNA bd / cd inhibits the output from the IV-HSL / Sal responsive circuit as in Fig. 1a.
- IV-HSL and Sal responsive circuits work analogously. Thus, the output from the responsive circuits is translated if exactly one of the inducers is present.
- Another copy of output mRNA is constitutively transcribed but is silenced by its RBS stemloop by default. Only when the 3 co-trigger RNAs form a trimer, the exposed sequence mn pries open the RBS, see Fig. 1c. Therefore, the output is expressed when all 3 signals are present.

2.10 Shelving

The 3-XOR gate from §2.9 activates or represses translation of an output mRNA. We take this output to be the transcription factor cl^a that activates the expression of a trigger RNA Y with toehold, which in turn triggers the translation of Xis . Thus, the presence of Y stands for $d_1 = 1$. Now, suppose $w_B = 1$ and $s_1 r_1 c_1 = 101$, in which case $d_1 = 0$. Then Y is low, but around the timepoint when w_B is turned off and the inputs begin to change, Y can exhibit a transient bump (which we observed in some simulations). This can cause production of Xis and an unintended *reset*. One way to mitigate this is to condition the expression of Xis on w_B as well, which requires w_B to go down early and also tuning of degradation rates. We implemented another, complementary way, which is a “shelf” (a buffer) for Y that i) has high affinity for Y ; ii) sequesters about $30 \times EC50_{Y \rightarrow Xis}$ copies of Y ; iii) is degraded when w_A is on. In this way, a transient increase in Y is shelved but a sustained signal (while w_B is up) will overflow the shelves and allow translation of Xis , cf. Fig. 5. As a concrete mechanism we propose transcription of a repeated complement of Y , viz. $\langle w_A \rangle \rightarrow (AraC^*) \rightarrow [BAD] \rightarrow (cl^b) \dashv [cl_-^b] \rightarrow (Y^*)^n \dashv Y \rightarrow (Xis)$.

3 Simulations

3.1 General considerations

Recall from §2.3 that the dimerized (AmtR) represses [AmtR], which we can express as

$$2(AmtR) + [AmtR]_{active} \rightleftharpoons (AmtR)_2 : [AmtR] = \text{repressed state.} \quad (5)$$

In steady state, we have the Hill equation

$$[AmtR]_{active} = (1 + ((AmtR)/EC50)^n)^{-1} [AmtR]_{total}. \quad (6)$$

In case of an activator we use, for example,

$$[Sal]_{active} = (1 + (EC50/\langle Sal \rangle)^n)^{-1} [Sal]_{total}. \quad (7)$$

All species are in units of *molecules*. For simplicity, we use the above Hill equations (which, notably, have no baseline term) for all species with $n = 2$ and mostly $EC50 = 1$ molecules (cf. §4). We simulate production and degradation such that typically about 10 molecules of output from a fully active promoter correspond to the steady state, for example $\frac{d}{dt}(\text{Output}) = k_+[\text{Sal}]_{\text{active}} - k_-(\text{Output})$ with $k_+ = 1\text{s}^{-1}$ and $k_- = 0.1\text{s}^{-1}$ models $[\text{Sal}]_{\text{active}} \rightarrow (\text{Output})$. Generally, we take 0.1 and 10 molecules to mean *off* and *on*, respectively. In the figures we refer to “a.u.” instead of seconds, because these choices lead to timescales that are too fast to be realistic (perhaps by a factor of 10).

3.2 Player A

We implemented the response r_0 and r_1 of Player A as sketched out in §2.2. The steady-state response is shown in Fig. 3.

3.3 Bit 1

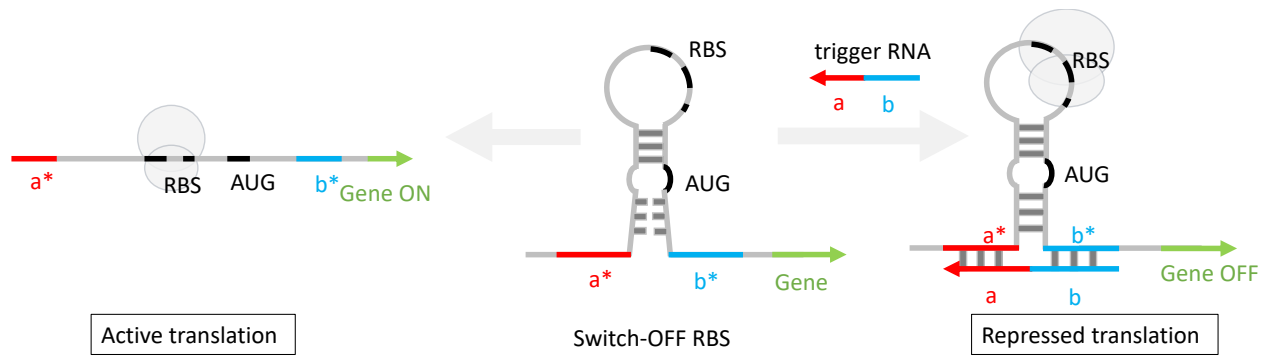
Carry flag c_2 . We implemented c_2 as indicated in §2.2. Recall that c_2 is emitted immediately, bypassing the memory unit. The steady-state response is shown in Fig. 4.

Difference d_1 . We implemented d_1 as in §2.9 with the buffer/shelving mechanism from §2.10 and the recombinase memory unit from §2.7. Fig. 4 shows the write-then-emit response of this stack of circuits. Fig. 5 shows the time-course of key components over four phases (compute-read-compute-read) with a particular choice of inputs and illustrates the effect of shelving for isolating the memory unit from spurious output of the compute unit.

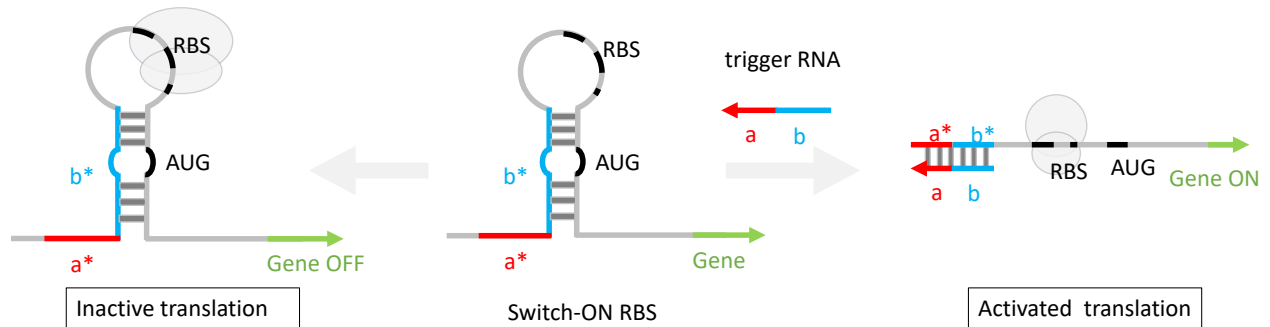
4 Discussion

The focus of this note is the design of a multi-cell genetic count-down computer which is *reusable*. Our solution relies on computing, remembering and emitting the new status in alternating phases. In particular, we propose an RNA-based 3-XOR gate. The memory unit is based on a resettable recombinase. We also propose to insulate undesirable fluctuations in the output of the compute unit by means of a buffer.

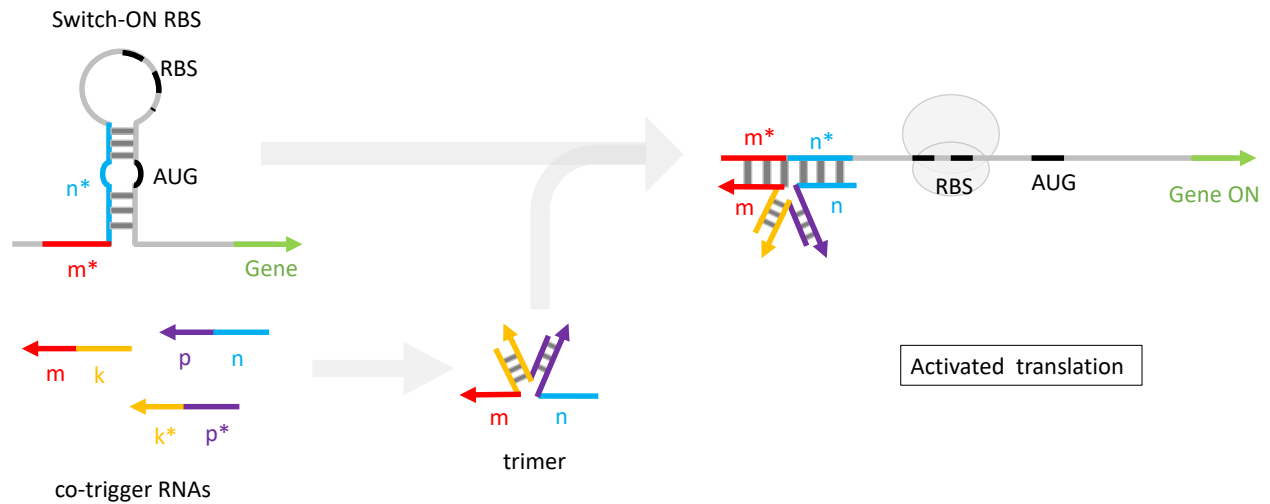
Our simulations have helped identify some issues but are highly stylized. For example, in reality, the EC50 of the signals vary over several orders of magnitude (Du et al., 2020, SM:Fig.8) and the Hill equation parameters depend on the context (Nielsen et al., 2016, SM:p.10); we rely heavily on orthogonality of the signaling channels and ignore potential toxicity (Du et al., 2020, SM:p.8); we did not simulate diffusion of signaling molecules in and out of the cell and have not systematically investigated robustness against mistiming of signaling.



(a) The unstable hairpin is sealed by the trigger RNA ab , repressing translation. Adapted from (Kim et al., 2019, Fig. 1c).



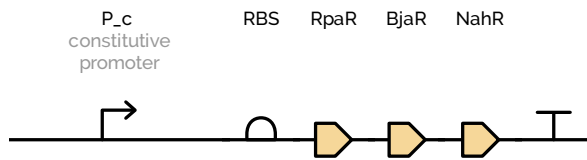
(b) The stem of the stable hairpin is opened by the trigger RNA ab , exposing the RBS for translation. Adapted from (Green et al., 2014, Fig. 1b).



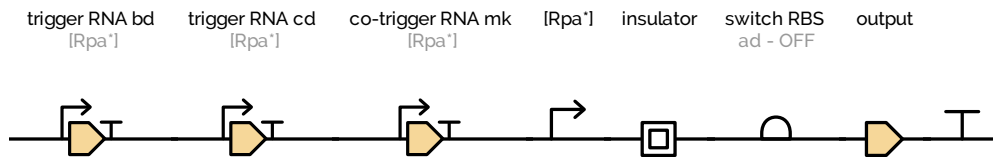
(c) The co-trigger RNAs form a trimer that switches ON the RBS. Adapted from (Green et al., 2017, Fig. 3a).

Figure 1: Switch-OFF (1a) and Switch-ON (1b) RBS; RNAs 3-AND gate (1c). Cf. §2.9/§2.8.

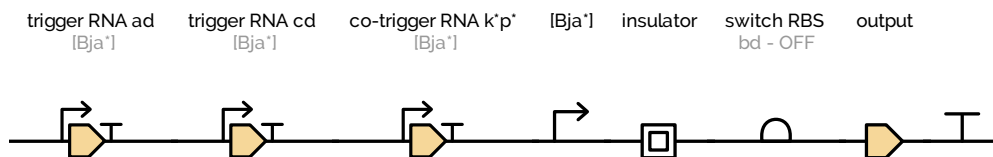
Transcription factors



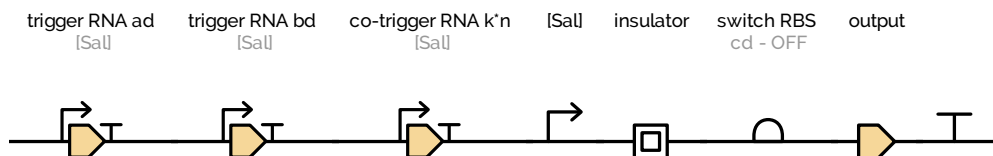
pC-HSL responsive circuit



IV-HSL responsive circuit



Sal responsive circuit



co-trigger RNA AND gate

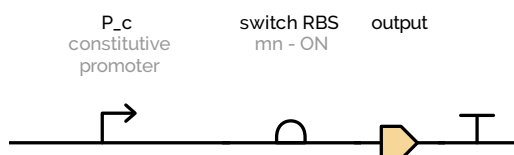


Figure 2: Genetic circuits for the 3-input XOR gate. See §2.9.

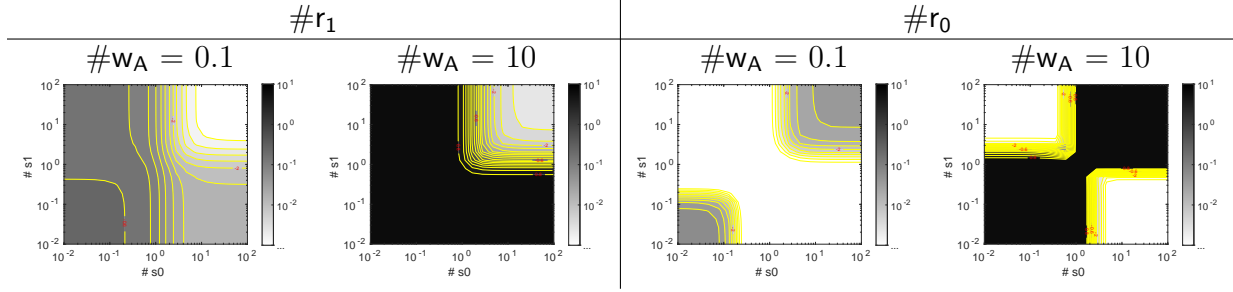


Figure 3: Response surfaces of Player A for varying $\#s_1(\uparrow)$ and $\#s_0(\rightarrow)$. See §3.2. The log-colorbar goes from 10^{-3} -or-less (white) to 10^1 (black). Yellow contours at 10^t , $t = 0.8, 0.6, 0.4, \dots$

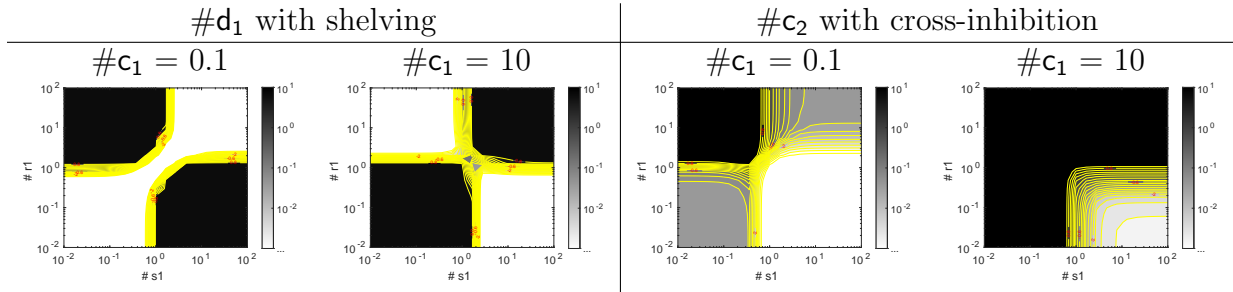


Figure 4: Response of Subtractor A for varying $\#r_1(\uparrow)$ and $\#s_1(\rightarrow)$. See §3.3. The output $\#d_1$ is measured after a compute-, inter- and readout-phase, cf. Fig. 5. The log-colorbar goes from 10^{-3} -or-less (white) to 10^1 (black). Yellow contours at 10^t , $t = 0.8, 0.6, 0.4, \dots$

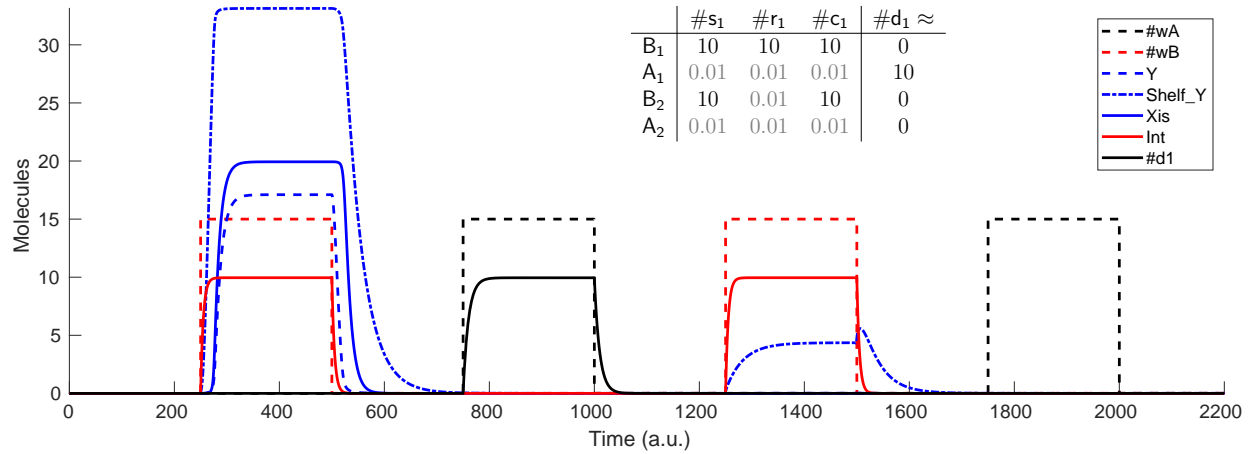


Figure 5: Subtractor A, Bit 1, output $\#d_1$ across phases B₁-A₁-B₂-A₂ (with interphases). The table shows the inputs for each phase. We have $\#d_1 \approx 10$ in the recall phase A₁ and $\#d_1 \approx 0$ in A₂. Observe the slight delay in the rise of Y at the beginning of B₁ while it is being “shelved”; conversely, the leakage of Y is “shelved” during B₂ preventing unwanted expression of Xis. At the end of B₂, a small deliberate mistiming in signal decay leads to a bump in Y that is absorbed by the shelf. See also §3.3/p.11.

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Appendix

4.1 Subtractor module

Fig. 6 shows the principles of the Subtractor module (bit 0 and bit 1), but our implementation differs in the details. In particular, we use a reversible recombinase for the memory module, see §2.7/p.8.

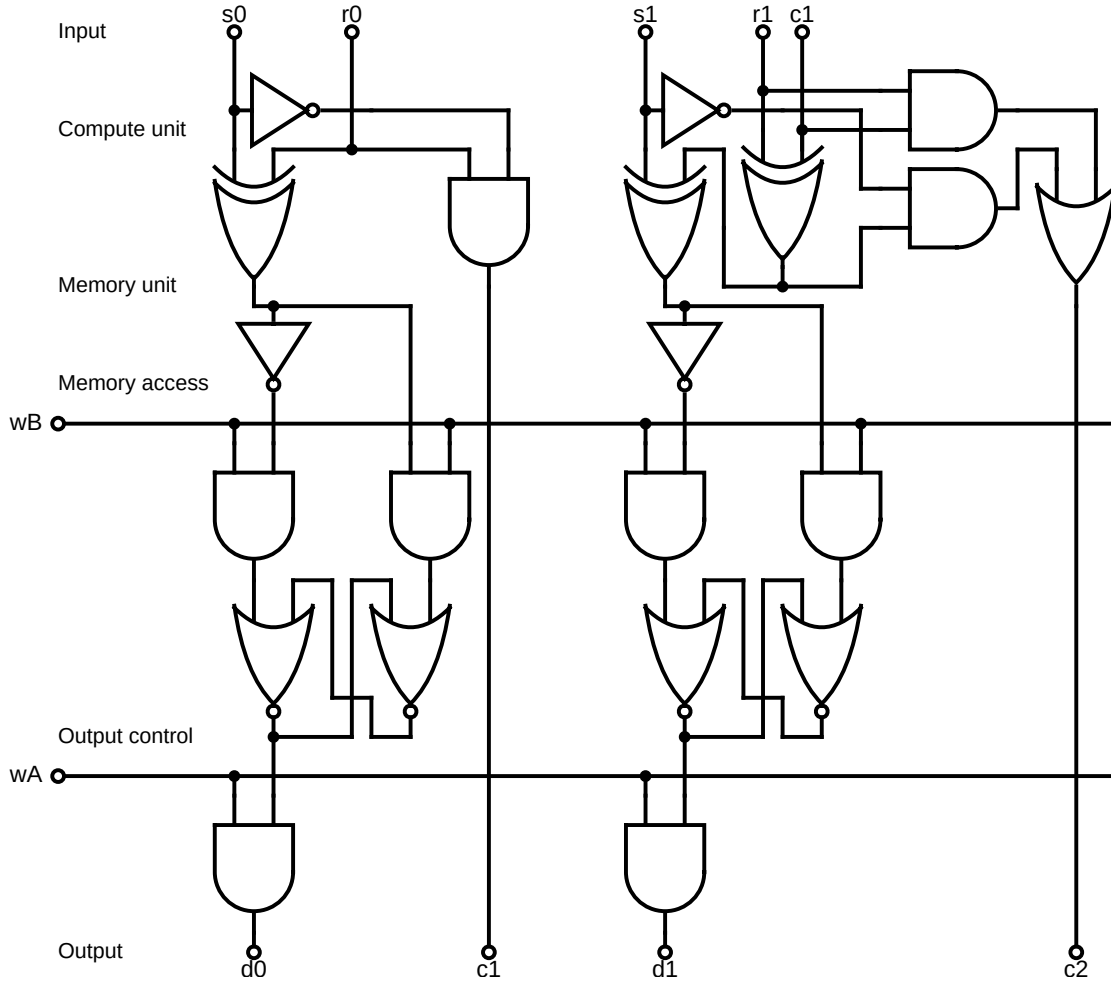


Figure 6: Bit 0 and Bit 1 of Subtractor A. The result of the compute unit is written to the memory unit (e.g. a bistable switch) when the signal w_B is present. The value of the memory unit is forwarded to the output when the signal w_A is present.

TODOs:

Also:

TODO(1): spacing of act and rep

TODO(2): check that Table 4 is correct

TODO(3): stars

TODO(4): acknowledgements

TODO(5): review the references section