# Analysis of variance revisited

### Section 1

## Analysis of variance

# Analysis of variance

- Analysis of variance used with:
  - counted/measured response
  - categorical explanatory variable(s)
  - that is, data divided into groups, and see if response significantly different among groups
  - or, see whether knowing group membership helps to predict response.
- Typically two stages:
  - F-test to detect any differences among/due to groups
  - if *F*-test significant, do *multiple comparisons* to see which groups significantly different from which.
- Need special multiple comparisons method because just doing (say) two-sample t-tests on each pair of groups gives too big a chance of finding "significant" differences by accident.

## **Packages**

#### These:

```
library(tidyverse)
library(broom)
library(car) # for Levene's text
```

## Example: Pain threshold and hair colour

- Do people with different hair colour have different abilities to deal with pain?
- Men and women of various ages divided into 4 groups by hair colour: light and dark blond, light and dark brown.
- Each subject given a pain sensitivity test resulting in pain threshold score: higher score is higher pain tolerance.
- 19 subjects altogether.

## The data

### In hairpain.txt:

hair pain	darkblond 43
lightblond 62	lightbrown 42
lightblond 60	lightbrown 50
lightblond 71	lightbrown 41
lightblond 55	lightbrown 37
lightblond 48	darkbrown 32
darkblond 63	darkbrown 39
darkblond 57	darkbrown 51
darkblond 52	darkbrown 30
darkblond 41	darkbrown 35

# Summarizing the groups

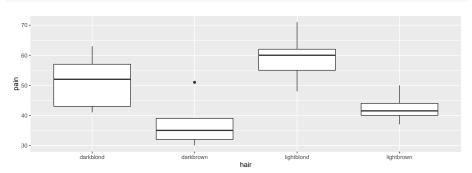
```
my_url <- "http://www.utsc.utoronto.ca/~butler/d29/hairpain.txt"
hairpain <- read_delim(my_url, " ")
hairpain %>%
  group_by(hair) %>%
  summarize(
    n = n(),
    xbar = mean(pain),
    s = sd(pain)
)
```

hair	n	xbar	s
darkblond	5	51.2	9.284396
darkbrown	5	37.4	8.324662
lightblond	5	59.2	8.526430
lightbrown	4	42.5	5.446712

Brown-haired people seem to have lower pain tolerance.

## **Boxplot**

## ggplot(hairpain, aes(x = hair, y = pain)) + geom\_boxplot()



## **Assumptions**

- Data should be:
  - normally distributed within each group
  - same spread for each group
- darkbrown group has upper outlier (suggests not normal)
- darkblond group has smaller IQR than other groups.
- But, groups small.
- Shrug shoulders and continue for moment.

# Testing equality of SDs

• via Levene's test in package car:

```
leveneTest(pain ~ hair, data = hairpain)
```

```
## Warning in leveneTest.default(y = y, group =
## group, ...): group coerced to factor.
```

	Df	F value	Pr(>F)
group	3	0.3927432	0.7600161
	15	NA	NA

- No evidence (at all) of difference among group SDs.
- Possibly because groups small.

# Analysis of variance

```
hairpain.1 <- aov(pain ~ hair, data = hairpain)
summary(hairpain.1)</pre>
```

```
## Df Sum Sq Mean Sq F value Pr(>F)
## hair 3 1361 453.6 6.791 0.00411 **
## Residuals 15 1002 66.8
## ---
## Signif. codes:
## 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

- P-value small: the mean pain tolerances for the four groups are not all the same.
- Which groups differ from which, and how?

# Multiple comparisons

- Which groups differ from which? Multiple comparisons method. Lots.
- ullet Problem: by comparing all the groups with each other, doing many tests, have large chance to (possibly incorrectly) reject  $H_0$ : groups have equal means.
- 4 groups: 6 comparisons (1 vs 2, 1 vs 3, ..., 3 vs 4). 5 groups: 10 comparisons. Thus 6 (or 10) chances to make mistake.
- Get "familywise error rate" of 0.05 (whatever), no matter how many comparisons you're doing.
- My favourite: Tukey, or "honestly significant differences': how far apart might largest, smallest group means be (if actually no differences). Group means more different: significantly different.

## **Tukey**

• TukeyHSD:

```
TukeyHSD(hairpain.1)
```

```
##
     Tukey multiple comparisons of means
##
       95% family-wise confidence level
##
  Fit: aov(formula = pain ~ hair, data = hairpain)
##
  $hair
##
##
                          diff
                                      lwr
                                                 upr
  darkbrown-darkblond
                         -13.8 -28.696741
                                           1.0967407
                        8.0 -6.896741 22.8967407
  lightblond-darkblond
  lightbrown-darkblond -8.7 -24.500380 7.1003795
  lightblond-darkbrown 21.8
                                 6.903259 36.6967407
## lightbrown-darkbrown
                        5.1 -10.700380 20.9003795
  lightbrown-lightblond -16.7 -32.500380 -0.8996205
##
                             p adj
  darkbrown-darkblond
                         0.0740679
  lightblond-darkblond
                         0.4355768
                            Analysis of variance revisited
```

# The old-fashioned way

- List group means in order
- Draw lines connecting groups that are *not* significantly different:

```
darkbrown lightbrown darkblond lightblond 37.4 42.5 51.2 59.2
```

- lightblond significantly higher than everything except darkblond (at  $\alpha=0.05$ ).
- darkblond in middle ground: not significantly less than lightblond, not significantly greater than darkbrown and lightbrown.
- More data might resolve this.
- Looks as if blond-haired people do have higher pain tolerance, but not completely clear.

# Some other multiple-comparison methods

- Work any time you do k tests at once (not just ANOVA).
  - **Bonferroni**: multiply all P-values by k.
  - **Holm**: multiply smallest P-value by k, next-smallest by k-1, etc.
  - False discovery rate: multiply smallest P-value by k/1, 2nd-smallest by k/2, ..., i-th smallest by k/i.
- Stop after non-rejection.

## Example

- $\bullet$  P-values 0.005, 0.015, 0.03, 0.06 (4 tests all done at once) Use  $\alpha=0.05.$
- Bonferroni:
  - Multiply all P-values by 4 (4 tests).
  - Reject only 1st null.
- Holm:
  - Times smallest P-value by 4: 0.005 \* 4 = 0.020 < 0.05, reject.
  - Times next smallest by 3: 0.015 \* 3 = 0.045 < 0.05, reject.
  - Times next smallest by 2: 0.03 \* 2 = 0.06 > 0.05, do not reject. Stop.

#### ...Continued

- With P-values 0.005, 0.015, 0.03, 0.06:
- False discovery rate:
  - Times smallest P-value by 4: 0.005 \* 4 = 0.02 < 0.05: reject.
  - Times second smallest by 4/2: 0.015 \* 4/2 = 0.03 < 0.05, reject.
  - Times third smallest by 4/3: 0.03 \* 4/3 = 0.04 < 0.05, reject.
  - $\bullet$  Times fourth smallest by  $4/4\colon 0.06*4/4=0.06>0.05,$  do not reject. Stop.

### pairwise.t.test

## darkbrown 0.0699 -## lightblond 0.4001 0.0045 ## lightbrown 0.4001 0.4001

## P value adjustment method: holm

##

darkblond darkbrown lightblond

0.0408

```
with(hairpain, pairwise,t,test(pain, hair, p,adi = "none"))
##
   Pairwise comparisons using t tests with pooled SD
##
## data: pain and hair
              darkblond darkbrown lightblond
## darkbrown 0.01748
## lightblond 0.14251 0.00075
## lightbrown 0.13337 0.36695
                                0.00817
##
## P value adjustment method: none
with(hairpain, pairwise,t,test(pain, hair, p,adi = "holm"))
##
   Pairwise comparisons using t tests with pooled SD
##
## data: pain and hair
##
```

## pairwise.t.test part 2

1.0000

## P value adjustment method: bonferroni

0.0490

## darkbrown 0.1049 ## lightblond 0.8550 0.0045 ## lightbrown 0.8002

##

```
with(hairpain, pairwise.t.test(pain, hair, p.adi = "fdr"))
##
   Pairwise comparisons using t tests with pooled SD
##
## data: pain and hair
             darkblond darkbrown lightblond
## darkbrown 0.0350
## lightblond 0.1710 0.0045
## lightbrown 0.1710 0.3670
                               0.0245
##
## P value adjustment method: fdr
with(hairpain, pairwise.t.test(pain, hair, p.adj = "bon"))
##
   Pairwise comparisons using t tests with pooled SD
##
## data: pain and hair
##
             darkblond darkbrown lightblond
```

#### Comments

- P-values all adjusted upwards from "none".
- Required because 6 tests at once.
- Highest P-values for Bonferroni: most "conservative".
- Prefer Tukey or FDR or Holm.
- Tukey only applies to ANOVA, not to other cases of multiple testing.

### Rats and vitamin B

- What is the effect of dietary vitamin B on the kidney?
- A number of rats were randomized to receive either a B-supplemented diet or a regular diet.
- Desired to control for initial size of rats, so classified into size classes lean and obese.
- After 20 weeks, rats' kidneys weighed.
- Variables:
  - Response: kidneyweight (grams).
  - Explanatory: diet, ratsize.
- Read in data:

```
my_url <- "http://www.utsc.utoronto.ca/~butler/d29/vitaminb.txt"
vitaminb <- read_delim(my_url, " ")</pre>
```

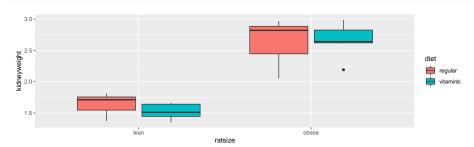
## The data

#### vitaminb

ratsize	diet	${\it kidneyweight}$
lean	regular	1.62
lean	regular	1.80
lean	regular	1.71
lean	regular	1.81
lean	regular	1.47
lean	regular	1.37
lean	regular	1.71
lean	vitaminb	1.51
lean	vitaminb	1.65
lean	vitaminb	1.45
lean	vitaminb	1.44
lean	vitaminb	1.63
lean	vitaminb	1.35

## Grouped boxplot

```
ggplot(vitaminb, aes(
   x = ratsize, y = kidneyweight,
   fill = diet
)) + geom_boxplot()
```



## What's going on?

Calculate group means:

```
summary <- vitaminb %>%
group_by(ratsize, diet) %>%
summarize(mean = mean(kidneyweight))
```

## `summarise()` regrouping output by 'ratsize' (override with `.groups` ar summary

diet	mean
regular	1.641429
vitaminb	1.527143
regular	2.642857
vitaminb	2.672857
	regular vitaminb regular

- Rat size: a large and consistent effect.
- Diet: small/no effect (compare same rat size, different diet).

### ANOVA with interaction

```
vitaminb.1 <- aov(kidneyweight ~ ratsize * diet,
   data = vitaminb
)
summary(vitaminb.1)</pre>
```

- Significance/nonsignificance as we expected.
- Note no significant interaction (can be removed).

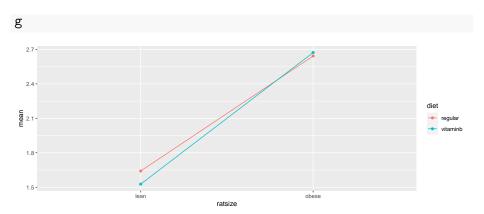
## Interaction plot

 Plot mean of response variable against one of the explanatory, using other one as groups. Start from summary:

```
g <- ggplot(summary, aes(
  x = ratsize, y = mean,
  colour = diet, group = diet
)) +
  geom_point() + geom_line()</pre>
```

• For this, have to give both group and colour.

## The interaction plot



Lines basically parallel, indicating no interaction.

### Take out interaction

```
vitaminb.2 <- update(vitaminb.1, . ~ . - ratsize:diet)
summary(vitaminb.2)</pre>
```

- No Tukey for diet: not significant.
- No Tukey for ratsize: only two sizes, and already know that obese rats have larger kidneys than lean ones.
- Bottom line: diet has no effect on kidney size once you control for size of rat.

# Assessing assumptions: residuals

- In two-way ANOVA, not many observations per treatment group.
- Difficult to check for normality / equal spreads.
- But, any regular ANOVA also a regression.
- Use regression residual ideas.
- In ANOVA, one fitted value per treatment group (based on means).
- Residual: observation minus fitted value.

## Previous ANOVA as regression

```
vitaminb.3 <- lm(kidneyweight ~ ratsize + diet, data = vitaminb)</pre>
summary(vitaminb.3)
##
## Call:
## lm(formula = kidneyweight ~ ratsize + diet, data = vitaminb)
##
## Residuals:
##
      Min 10 Median 30 Max
## -0.62893 -0.12625 0.04071 0.14607 0.35321
##
## Coefficients:
##
             Estimate Std. Error t value Pr(>|t|)
## (Intercept) 1.60536 0.07768 20.67 < 2e-16
## ratsizeobese 1.07357 0.08970 11.97 7.59e-12
##
## (Intercept) ***
## ratsizeobese ***
## dietvitaminb
## ---
## Signif. codes:
## 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## Reproduce ANOVA

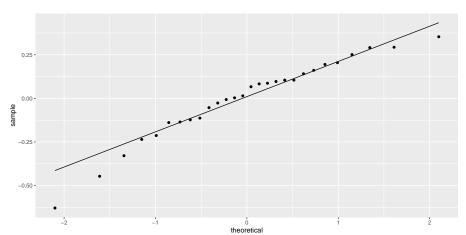
#### tidy(drop1(vitaminb.3, test = "F"))

term	df	sumsq	rss	AIC	statistic	p.value
	NA	NA	1.407946	-77.72202	NA	NA
ratsize	1	8.0678893	9.475836	-26.33687	143.2563257	0.0000000
diet	1	0.0124321	1.420379	-79.47587	0.2207496	0.6425419

- ANOVA and regression drop1 output always the same.
- this time, ANOVA and regression summary output have same
   P-values, but only because categorical variables both have two levels.

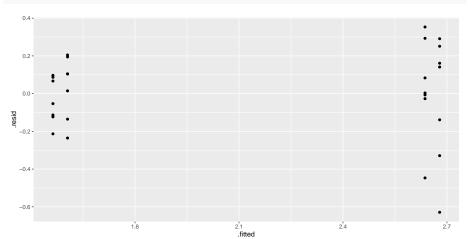
### Are the residuals normal?

```
ggplot(vitaminb.3, aes(sample=.resid)) +
stat_qq() + stat_qq_line()
```



## Residuals against fitted

ggplot(vitaminb.3, aes(x=.fitted, y=.resid)) + geom\_point()



#### Comments

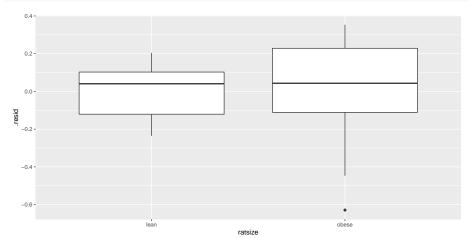
- ullet 2 rat sizes, 2 diets: only  $2 \times 2 = 4$  different fitted values
- larger fitted values have greater spread (fan-out, transformation?)
- add residuals to data to plot residuals against size, diet (augment from broom):

```
vitaminb.3 %>% augment(vitaminb) -> vitaminb.3a
```

 explanatory ratsize, diet categorical, so plot resid vs. them with boxplots.

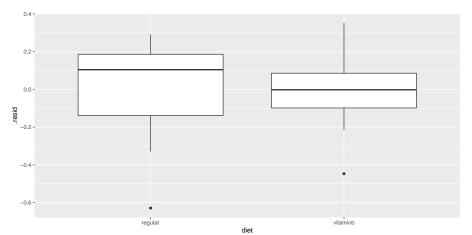
### Residuals vs rat size

```
ggplot(vitaminb.3a, aes(x = ratsize, y = .resid)) +
  geom_boxplot()
```



### Residuals vs diet

```
ggplot(vitaminb.3a, aes(x = diet, y = .resid)) +
  geom_boxplot()
```



#### Comments

- there are low outliers on the plot against diet
- residuals for obese rats seem more spread out than for lean rats
- case for transformation of rat weights
- however, story from our analysis very clear:
  - rat size strongly significant
  - diet nowhere near significant
- and so expect transformation to make no difference to conclusions.

### The auto noise data

In 1973, the President of Texaco cited an automobile filter developed by Associated Octel Company as effective in reducing pollution. However, questions had been raised about the effects of filter silencing. He referred to the data included in the report (and below) as evidence that the silencing properties of the Octel filter were at least equal to those of standard silencers.

```
u <- "http://www.utsc.utoronto.ca/~butler/d29/autonoise.txt"
autonoise <- read_table(u)
##</pre>
```

```
## -- Column specification -----
## cols(
## noise = col_double(),
## size = col_character(),
## type = col_character(),
## side = col_character()
```

### The data

#### autonoise

noise	size	type	side
840	М	Std	R
770	L	Octel	L
820	M	Octel	R
775	L	Octel	R
825	M	Octel	L
840	M	Std	R
845	M	Std	L
825	М	Octel	L
815	М	Octel	L
845	М	Std	R
765	L	Octel	L
835	S	Std	L
775	L	Octel	R

### Making boxplot

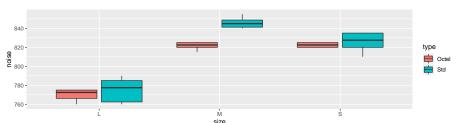
- Make a boxplot, but have combinations of filter type and engine size.
- Use grouped boxplot again, thus:

```
g <- autonoise %>%
ggplot(aes(x = size, y = noise, fill = type)) +
geom_boxplot()
```

### The boxplot

- See difference in engine noise between Octel and standard is larger for medium engine size than for large or small.
- Some evidence of differences in spreads (ignore for now):

g



#### **ANOVA**

##

```
autonoise.1 <- aov(noise ~ size * type, data = autonoise)</pre>
summary(autonoise.1)
```

```
Df Sum Sq Mean Sq F value Pr(>F)
                 26051 13026 199.119 < 2e-16 ***
## size
              1 1056 1056 16.146 0.000363 ***
## type
## size:type 2 804 402 6.146 0.005792 **
## Residuals 30 1962 65
## ---
## Signif. codes:
## 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

- The interaction is significant, as we suspected from the boxplots.
- The within-group spreads don't look very equal, but only based on 6 obs each.

### Tukey: ouch!

```
autonoise.2 <- TukeyHSD(autonoise.1)
autonoise.2$`size:type`</pre>
```

```
diff
##
                                      lwr
                                                 upr
## M:Octel-L:Octel
                    51.6666667
                                37.463511
                                           65.869823
## S:Octel-L:Octel
                    52.5000000
                                38.296844
                                           66.703156
## L:Std-L:Octel
                     5.0000000
                                -9.203156
                                           19,203156
## M:Std-L:Octel 75.8333333
                                61.630177
                                           90.036489
## S:Std-L:Octel
                    55.8333333
                                41.630177
                                           70.036489
## S:Octel-M:Octel
                     0.8333333 -13.369823
                                           15.036489
## I.:Std-M:Octel
                   -46.6666667 -60.869823 -32.463511
## M:Std-M:Octel
                    24.1666667
                                 9.963511
                                           38.369823
## S:Std-M:Octel
                     4.1666667 -10.036489
                                           18.369823
## L:Std-S:Octel
                   -47.5000000 -61.703156 -33.296844
## M:Std-S:Octel
                    23.3333333
                                 9.130177
                                           37.536489
## S:Std-S:Octel
                     3.3333333 -10.869823
                                           17.536489
## M:Std-L:Std
                    70.8333333
                                56.630177
                                           85.036489
## S:Std-L:Std
                    50.8333333
                                36.630177
                                           65.036489
## S:Std-M:Std
                  -20.0000000 -34.203156
                                           -5.796844
##
                          p adj
## M:Octel-L:Octel 6.033496e-11
## S:Octel-L:Octel 4.089762e-11
## L:Std-L:Octel 8.890358e-01
```

### Interaction plot

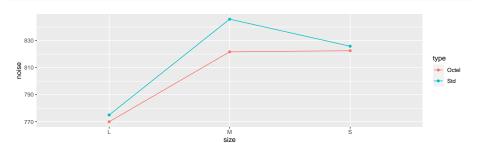
- This time, don't have summary of mean noise for each size-type combination.
- One way is to compute summaries (means) first, and feed into ggplot as in vitamin B example.
- Or, have ggplot compute them for us, thus:

```
g <- ggplot(autonoise, aes(
    x = size, y = noise,
    colour = type, group = type
)) +
    stat_summary(fun = mean, geom = "point") +
    stat_summary(fun = mean, geom = "line")</pre>
```

### Interaction plot

The lines are definitely *not* parallel, showing that the effect of type is different for medium-sized engines than for others:

g

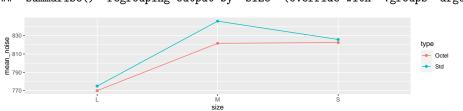


### If you don't like that...

...then compute the means first, in a pipeline:

```
autonoise %>%
 group_by(size, type) %>%
  summarize(mean noise = mean(noise)) %>%
  ggplot(aes(
   x = size, y = mean_noise, group = type,
   colour = type
  )) + geom_point() + geom_line()
```

## `summarise()` regrouping output by 'size' (override with `.groups` argum



# Simple effects for auto noise example

- In auto noise example, weren't interested in all comparisons between car size and filter type combinations.
- Wanted to demonstrate (lack of) difference between filter types for each car type.
- These are called simple effects of one variable (filter type) conditional on other variable (car type).
- To do this, pull out just the data for small cars, compare noise for the two filter types. Then repeat for medium and large cars. (Three one-way ANOVAs.)

### Do it using dplyr tools

Small cars:

```
autonoise %>%
  filter(size == "S") %>%
  aov(noise ~ type, data = .) %>%
  summary()
```

```
## Df Sum Sq Mean Sq F value Pr(>F)
## type 1 33.3 33.33 0.548 0.476
## Residuals 10 608.3 60.83
```

- No filter difference for small cars.
- For Medium, change S to M and repeat.

# Simple effect of filter type for medium cars

```
autonoise %>%
  filter(size == "M") %>%
  aov(noise ~ type, data = .) %>%
  summary()
```

 There is an effect of filter type for medium cars. Look at means to investigate (over).

### Mean noise for each filter type

...for medium engine size:

```
autonoise %>%
  filter(size == "M") %>%
  group_by(type) %>%
  summarize(m = mean(noise))
```

## `summarise()` ungrouping output (override with `.groups` as

type	m
Octel	821.6667
Std	845.8333

Octel filters produce less noise for medium cars.

### Large cars

Large cars:

```
autonoise %>%
  filter(size == "L") %>%
  aov(noise ~ type, data = .) %>%
  summary()
```

```
## Df Sum Sq Mean Sq F value Pr(>F)
## type 1 75 75 0.682 0.428
## Residuals 10 1100 110
```

No significant difference again.

# All at once, using split/apply/combine

The "split" part:

```
autonoise %>%
  group_by(size) %>%
  nest()
```

#### size data

```
M 840, 820, 825, 840, 845, 825, 815, 845, 850, 855, 820, 825, Std, Octel, Octel, Std, Octel, Octel, Std, Octel, Octel, R. R. L. R. L. L. R. L. R. R. R. R.
```

- L 770, 775, 765, 775, 760, 760, 785, 770, 760, 790, 775, 785, Octel, Octel, Octel, Octel, Octel, Std, Std, Std, Std, Std, Octel, Std, L, R, L, R, L, L, R, L, L, R, R, R, R
- S 835, 835, 820, 835, 825, 820, 825, 820, 820, 820, 810, 825, Std, Std, Octel, Std, Octel, Std, Octel, Std, Octel, Std, Octel, L, L, R, L, R, L, R, R, R, R, R, R, R

## **Apply**

 Write function to do aov on a data frame with columns noise and type, returning P-value:

```
aov_pval <- function(x) {</pre>
  noise.1 \leftarrow aov(noise \sim type, data = x)
  gg <- tidy(noise.1)</pre>
  gg$p.value[1]
  Test it:
autonoise %>%
  filter(size == "L") %>%
  aov pval()
```

## [1] 0.428221

Check.

### Combine

 Apply this function to each of the nested data frames (one per engine size):

```
autonoise %>%
  group_by(size) %>%
```

```
nest() %>%
```

<pre>mutate(p_val = map_dbl(data,</pre>	<pre>~ aov_pval(.)))</pre>	
size data		- _val

M 840, 820, 825, 840, 845, 825, 815, 845, 850, 855, 820, 

770 . 775 . 765 . 775 . 760 . 760 . 785 . 770 . 760 . 790 . 775 . 785, Octel, Octel, Octel, Octel, Octel, Std, Std, Std, Std,

0.000825, Std, Octel, Octel, Std, Std, Octel, Octel, Std, Std, Std

0.428 Std, Octel, Std, L, R, L, R, L, R, L, R, R, R, R 835, 835, 820, 835, 825, 820, 825, 820, 820, 820, 810, 0.476

Analysis of variance revisited

### Tidy up

 The data column was stepping-stone to getting answer. Don't need it any more:

```
simple_effects <- autonoise %>%
  group_by(size) %>%
  nest() %>%
  mutate(p_val = map_dbl(data, ~ aov_pval(.))) %>%
  select(-data)
simple_effects
```

size	p_val
М	0.0000085
L	0.4282210
S	0.4761786

### Simultaneous tests

When testing simple effects, doing several tests at once. (In this case,
3.) Have to adjust P-values for this. Eg. Holm:

```
simple_effects %>% ungroup() %>% arrange(p_val) %>%
mutate(multiplier = 4 - row_number()) %>%
mutate(p_val_adj = p_val * multiplier)
```

size	p_val	multiplier	p_val_ad
М	0.0000085	3	0.0000255
L	0.4282210	2	0.8564419
S	0.4761786	1	0.4761786

- No change in rejection decisions.
- Octel filters sig. better in terms of noise for medium cars, and not sig. different for other sizes.
- Octel filters never significantly worse than standard ones.

### Confidence intervals

- Perhaps better way of assessing simple effects: look at confidence intervals rather than tests.
- Gives us sense of accuracy of estimation, and thus whether non-significance might be lack of power: "absence of evidence is not evidence of absence".
- Works here because two filter types, using t.test for each engine type.
- Want to show that the Octel filter is equivalent to or better than the standard filter, in terms of engine noise.

## Equivalence and noninferiority

- Known as "equivalence testing" in medical world. A good read: link. Basic idea: decide on size of difference  $\delta$  that would be considered "equivalent", and if CI entirely inside  $\pm \delta$ , have evidence in favour of equivalence.
- We really want to show that the Octel filters are "no worse" than the standard one: that is, equivalent *or better* than standard filters.
- Such a "noninferiority test" done by checking that upper limit of CI, new minus old, is *less* than  $\delta$ . (This requires careful thinking about (i) which way around the difference is and (ii) whether a higher or lower value is better.)

### CI for small cars

Same idea as for simple effect test:

```
autonoise %>%
  filter(size == "S") %>%
  t.test(noise ~ type, data = .) %>%
  pluck("conf.int")
```

```
## [1] -14.517462 7.850795
## attr(,"conf.level")
## [1] 0.95
```

### CI for medium cars

```
autonoise %>%
  filter(size == "M") %>%
  t.test(noise ~ type, data = .) %>%
  pluck("conf.int")
```

```
## [1] -30.75784 -17.57549
## attr(,"conf.level")
## [1] 0.95
```

### CI for large cars

```
autonoise %>%
  filter(size == "L") %>%
  t.test(noise ~ type, data = .) %>%
  pluck("conf.int")
```

```
## [1] -19.270673 9.270673
## attr(,"conf.level")
## [1] 0.95
```

# Or, all at once: split/apply/combine

```
ci_func <- function(x) {
  tt <- t.test(noise ~ type, data = x)
  tt$conf.int
}
autonoise %>% group_by(size) %>%
  nest() %>%
  mutate(ci = map(data, ~ ci_func(.))) %>%
  unnest(ci) -> cis
```

840,820,825,840,845,825,815,845,850,855,820,

825, Std, Octel, Octel, Std, Std, Octel, Octel, Std, Std, Std

770 . 775 . 765 . 775 . 760 . 760 . 785 . 770 . 760 . 790 . 775 .

### Results

rcsui	
cis	

M

size data

M 840 , 820 , 825 , 840 , 845 , 825 , 815 , 845 , 850 , 855 , 820 , 825 . Std . Octel, Octel, Std . Std . Octel, Octel, Std . Std . Std 

785, Octel, Octel, Octel, Octel, Std, Std, Std, Std, 19.27  $\mathsf{Std}$ ,  $\mathsf{Octel}$ ,  $\mathsf{Std}$ ,  $\mathsf{L}$ ,  $\mathsf{R}$ ,  $\mathsf{L}$ ,  $\mathsf{R}$ ,  $\mathsf{L}$ ,  $\mathsf{R}$ ,  $\mathsf{R}$ ,  $\mathsf{R}$ 770, 775, 765, 775, 760, 760, 785, 770, 760, 790, 775, 9.270 785, Octel, Octel, Octel, Octel, Std, Std, Std, Std,

Std, Octel, Std, L, R, L, R, L, R, L, L, R, R, R, R

63/80

30.75

17.57

835 . 835 . 820 , 835 , 825 , 820 , 825 , 820 , 820 , 820 , 810 , Analysis of variance revisited

#### **Procedure**

- Function to get CI of difference in noise means for types of filter on input data frame
- Group by size, nest (mini-df per size)
- Calculate CI for each thing in data (ie. each size). map: CI is two numbers long
- unnest ci column to see two numbers in each CI.

### Cls and noninferiority test

- Suppose we decide that a 20 dB difference would be considered equivalent. (I have no idea whether that is reasonable.)
- Intervals:

```
hilos=c("lower", "upper")
cis %>%
  mutate(hilo = hilos) %>%
```

<pre>pivot_wider(names_from=hilo, values_from=ci)</pre>	
size data	up- lower per
M 840 , 820 , 825 , 840 , 845 , 825 , 815 , 845 , 850 , 855 , 820 , 825 , Std , Octel, Octel, Std , Std , Octel, Octel, Std , Std	 30.75 <b>78</b> 457

Analysis of variance revisited

770 . 775 . 765 . 775 . 760 . 760 . 785 . 770 . 760 . 790 . 775 . 785, Octel, Octel, Octel, Octel, Octel, Std., Std., Std., Std., Std. 

9.270619.27067

#### Comments

- In all cases, upper limit of CI is less than 20 dB. The Octel filters are "noninferior" to the standard ones.
- Caution: we did 3 procedures at once again. The true confidence level is not 95%. (Won't worry about that here.)

### Contrasts in ANOVA

- Sometimes, don't want to compare all groups, only some of them.
- Might be able to specify these comparisons ahead of time; other comparisons of no interest.
- Wasteful to do ANOVA and Tukey.

## Example: chainsaw kickback

- From link.
- Forest manager concerned about safety of chainsaws issued to field crew. 4 models of chainsaws, measure "kickback" (degrees of deflection) for 5 of each:

So far, standard 1-way ANOVA: what differences are there among models?

# chainsaw kickback (2)

- But: models A and D are designed to be used at home, while models
   B and C are industrial models.
- Suggests these comparisons of interest:
- home vs. industrial
- the two home models A vs. D
- the two industrial models B vs. C.
- Don't need to compare all the pairs of models.

### What is a contrast?

- Contrast is a linear combination of group means.
- Notation:  $\mu_A$  for (population) mean of group A, and so on.
- In example, compare two home models:  $H_0: \mu_A \mu_D = 0$ .
- Compare two industrial models:  $H_0: \mu_B \mu_C = 0$ .
- Compare average of two home models vs. average of two industrial models:  $H_0: \frac{1}{2}(\mu_A+\mu_D)-\frac{1}{2}(\mu_B+\mu_C)=0$  or  $H_0: 0.5\mu_A-0.5\mu_B-0.5\mu_C+0.5\mu_D=0$ .
- Note that coefficients of contrasts add to 0, and right-hand side is 0.

#### Contrasts in R

• Comparing two home models A and D ( $\mu_A - \mu_D = 0$ ):

$$c.home <- c(1, 0, 0, -1)$$

• Comparing two industrial models B and C ( $\mu_B - \mu_C = 0$ ):

c.industrial 
$$\leftarrow c(0, 1, -1, 0)$$

• Comparing home average vs. industrial average  $(0.5\mu_A - 0.5\mu_B - 0.5\mu_C + 0.5\mu_D = 0)$ :

c.home.ind 
$$\leftarrow c(0.5, -0.5, -0.5, 0.5)$$

### Orthogonal contrasts

• What happens if we multiply the contrast coefficients one by one?

```
c.home * c.industrial
```

```
## [1] 0 0 0 0
```

c.home \* c.home.ind

c.industrial \* c.home.ind

 in each case, the results add up to zero. Such contrasts are called orthogonal.

# Orthogonal contrasts (2)

Compare these:

```
c1 <- c(1, -1, 0)
c2 <- c(0, 1, -1)
sum(c1 * c2)
```

```
## [1] -1
```

Not zero, so c1 and c2 are not orthogonal.

- Orthogonal contrasts are much easier to deal with.
- Can use non-orthogonal contrasts, but more trouble (beyond us).

#### Read in data

```
url <- "http://www.utsc.utoronto.ca/~butler/d29/chainsaw.txt"
chain.wide <- read_table(url)
chain.wide</pre>
```

Α	В	С	D
42	28	57	29
17	50	45	29
24	44	48	22
39	32	41	34
43	61	54	30

# **Tidying**

Need all the kickbacks in one column:

# Starting the analysis (2)

### The proper data frame (tiny):

chain

model	kickback
A	42
В	28
C	57
D	29
Α	17
В	50
C	45
D	29
Α	24
В	44
C	48
D	22
Α	39
В	32
C	41
D	34
Α	43
В	61
C	54
D	30

### Setting up contrasts

# ANOVA as if regression

```
chain.1 <- lm(kickback ~ model, data = chain)</pre>
summary(chain.1)
##
## Call:
## lm(formula = kickback ~ model, data = chain)
##
## Residuals:
##
     Min
            10 Median 30 Max
## -16.00 -7.10 0.60 6.25 18.00
##
## Coefficients:
##
                   Estimate Std. Error t value
                     38.450 2.179 17.649
## (Intercept)
## modelc.home
                 2.100 3.081 0.682
## modelc.industrial -3.000 3.081 -0.974
## modelc.home.ind -15.100 4.357 -3.466
                  Pr(>|t|)
##
## (Intercept) 6.52e-12 ***
## modelc.home
               0.50524
## modelc.industrial 0.34469
## modelc.home.ind 0.00319 **
## ---
```

#### Conclusions

```
tidy(chain.1) %>% select(term, p.value)
```

term	p.value
(Intercept)	0.0000000
modelc.home	0.5052396
modelc.industrial	0.3446913
modelc.home.ind	0.0031872

- Two home models not sig. diff. (P-value 0.51)
- Two industrial models not sig. diff. (P-value 0.34)
- Home, industrial models are sig. diff. (P-value 0.0032).

## Means by model

The means:

```
chain %>%
  group_by(model) %>%
  summarize(mean.kick = mean(kickback)) %>%
  arrange(desc(mean.kick))
```

## `summarise()` ungrouping output (override with `.groups` argument)

model	mean.kick
С	49.0
В	43.0
A	33.0
D	28.8

- Home models A & D have less kickback than industrial ones B & C.
- Makes sense because industrial users should get training to cope with additional kickback