# Algorithm for Construction of Phylogenetic Tree Using BioPython

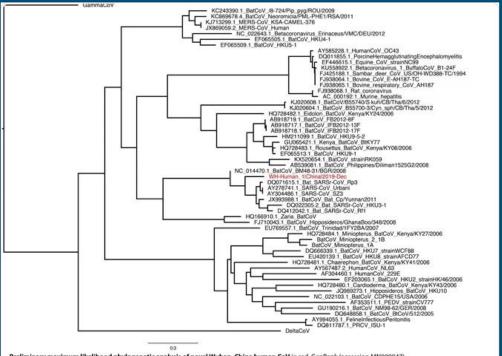
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11 December 2020

# Introduction

#### Phylogenetic Trees

- Spatially represent evolutionary relationships
- Built from sequences (protein, DNA, etc.)
- Long sequences can be unwieldy
- How can we make this process as convenient as possible?



Preliminary maximum likelihood phylogenetic analysis of novel Wuhan, China human CoV in red, GenBank (accession MN908947)

Novel CoV seg data from: http://virological.org/t/initial-genome-release-of-novel-coronavirus/319. The Shanghai Public Health Clinical Center & School of Public Health, in collaboration with the Central Hospital of Wuhan, Huazhong University of Science and Technology, the Wuhan Center for Disease Control and Prevention. the National Institute for Communicable Disease Control and Prevention, Chinese Center for Disease Control, and the University of Sydney, Sydney, Australia.

PhyML tree based on partial RdRp gene sequence (410bp), aligned with representative human and animal CoV sequences from Genbank compiled by Alice Latinne; tree by Kevin Olival Analysis by EcoHealth Alliance - 11 Jan 2020 (12:30pm EST)



Phylogenetic analysis of COVID-19, in red.

From https://www.news-medical.net/health/The-Phylogenetic-Tree-of-the-SARS-CoV-2-Virus.aspx

# Methodology

### BioPython Library

- A variety of computational biology functions
- Most useful modules for us:
  - ClustalW and AlignIO for MSA
  - Phylo for working with phylogenetic trees, uses common modules like Matplotlib to draw phylogenetic trees for visualization
  - o Pandas for data manipulation

```
from Bio import AlignIO
from Bio import Phylo
from Bio.Align import substitution_matrices
from Bio.Align.Applications import ClustalwCommandline
from io import StringIO
from numpy import NaN
import pandas as pd
import re
```

### Sequence Alignment

```
# create a multiple sequence alignment from provided file
def msa(filePath):
    ClustalwCommandline("clustalw2", infile=filePath, clustering="UPGMA")()
    return AlignIO.read(filePath.split(".")[0] + ".aln", "clustal")
```

- ClustalW will accept a variety of file types, including FASTA
- msa(filePath) returns a multiple sequence alignment in a special Sequence Alignment type that you can iterate through (similar to a list).
  - This type stores characteristics of each sequence, such as name, description, etc.

#### FASTA Input:

>Diplocarpon\_rosae

MPPKSAPKAKAASSNTTPGPSYQDMIIHAITTLKDRKGSSRIVLKKFVKSNNDIKAGD SMFTSLFNKAIAVGVEKGVFEQPKGASGTVKLAQKAPKPAAAKAAAKPKVEKKTTEK KPAVKKAPVAKKTTATKVVKPKADPTPKASPKAKAAAKPKAASKAKVAQPKVAAKP KKASAAKAVPAVVEKPSVLNKTKSGRVTKTTSATPKKAAAKKATPKKKATPKKA >Oncorhynchus\_mykiss

MAEVAPAPAAAAPAKAPKKKAAAKPKKAGPSVGELIVKAVSASKERSGVSLAALKKS LAAGGYDVEKNNSRVKIAVKSLVTKGTLVQTKGTGASGSFKLNKKAVEAKKPAKKAA APKAKKVAAKKPAAAKKPKKVAAKKAVAAKKSPKKAKKPATPKKAAKSPKKVKKPA AAAKKAAKSPKKATKAAKPKAAKPKAAKAKKAAPKKK

. . .

### Converting BLOSUM62 to Distance

```
def getBLOSUMDistanceMatrix(alignment):
    blosumMatrix = substitution_matrices.load("BLOSUM62")

    df = pd.DataFrame(columns=list(r.id for r in alignment), index=list(r.id for r in alignment))

    for record1 in alignment:
        for record2 in alignment:
            score = 0
            for i in range(len(record1.seq)):
                aa1 = record1[i] if record1[i] != '-' else '*'
                aa2 = record2[i] if record2[i] != '-' else '*'
                score -= blosumMatrix[aa1][aa2] #subtracts so closer nodes have lower values
            df[record1.id][record2.id] = score

    return df.apply(pd.to_numeric)
```

- Align subpackage substitution\_matrices includes most useful matrices (BLOSUM variations, PAM variations, etc)
- Give alignment generated from msa(), returns a distance matrix ready for UPGMA
- Uses negative BLOSUM scores because lower BLOSUM scores are "worse" (farther)

#### **About Distance Matrices**

	а	b	С	d	е
а	0	<mark>17</mark>	21	31	23
b	<mark>17</mark>	0	30	34	21
С	21	30	0	28	39
d	31	34	28	0	43
е	23	21	39	43	0

	(a,b)	С	d	е
(a,b)	0	25.5	32.5	<mark>22</mark>
С	25.5	0	28	39
d	32.5	28	0	43
е	<mark>22</mark>	39	43	0

"Cluster" - Column/row. Represents a species or strain

Cluster distance - Value at intersection

Merging - Average distance to all sub-clusters

#### **UPGMA**

#### Given a distance matrix:

l. While the distance matrix has more than 1 cluster:

Ci, Cj <- closest clusters

Cnew <- Merge Ci and Cj

Create node representing Cnew (Ci and Cj are children of new node)

Set the age of the new node to distance between closest clusters

Add Cnew to distance matrix

Remove Ci and Cj from the distance matrix

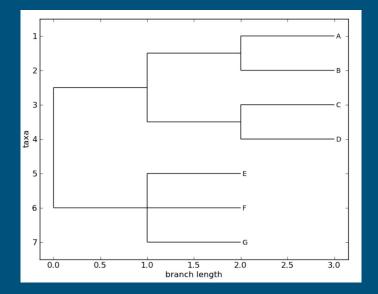
2. Traverse graph starting at the remaining cluster in the distance matrix

Distance between any two nodes is the difference in their age

## Visualizing the Tree

**Newick format** - representing trees using parentheses and commas, recognized by BioPython

Looks like:



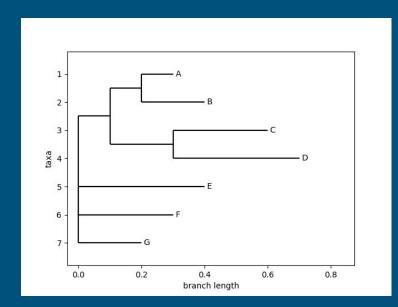
Branch lengths are assumed to be 1.0 when none are specified

Parentheses denote clusters.

### Visualizing the Tree

To denote distance of each node from parent:

```
(((A:0.1,B:0.2):0.1,(C:0.3,D:0.4):0.2):0.1,(E:0.4,F:0.3,G:0.2));
```



\*Note: there are some more robust formats (e.g. phyloxml), but Newick is simple and easy to work with in strings.

Phylo in biopython assumes 0 distance if unspecified when distances are specified elsewhere

So, we need to build a tree using the **unweighted pair group method with arithmetic mean (UPGMA)** method, then draw it.

### Visualizing the Tree

```
s = nodes[distances.columns[0]].toString()
tree = Phylo.read(StringIO(s), "newick")
Phylo.draw(tree)
```

- Phylo uses common libraries (like Matplotlib) to visualize trees
  - \*Note: you can also visualize the tree in ASCII in command line

#### **Implementation**

```
def UPGMA(distances):
    #initialization
    labels = distances.columns

    for cluster in distances.columns:
        distances[cluster][cluster] = NaN

    nodes = {}

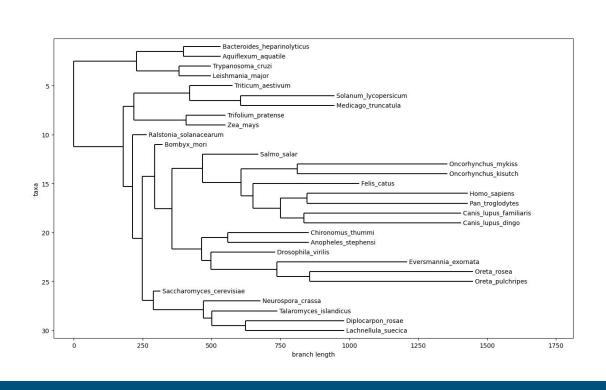
    for name in labels:
        nodes[name] = UPGMANode(name)

    Cnew = len(distances)
    #end initialization
```

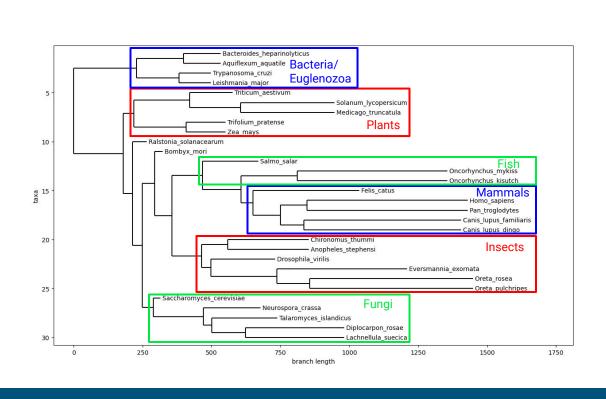
```
while distances.size > 1:
   Ci, Cj = distances.stack().idxmin()
    distances[Cnew] = mergeClusters(Ci, Cj)
   distances.loc[Cnew] = distances[Cnew]
   nodes[Cnew] = UPGMANode(Cnew)
   nodes[Cnew].children.add(nodes[Ci], nodes[Cj])
   nodes[Ci].parent = nodes[Cnew]
   nodes[Cj].parent = nodes[Cnew]
   nodes[Cnew].age = distances[Ci][Cj]
    distances.drop(Ci, Cj)
   Cnew += 1
s = nodes[distances.columns[0]].toString()
tree = Phylo.read(StringIO(s), "newick")
Phylo.draw(tree)
```

# Results

## Histone H1 Tree using BLOSUM Distances

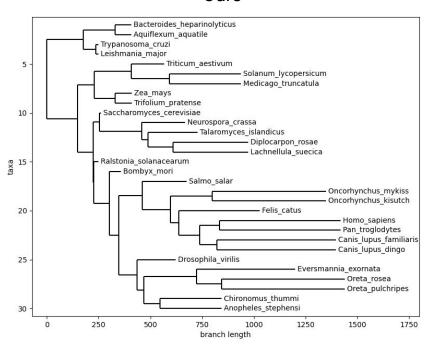


## Histone H1 Tree using BLOSUM Distances

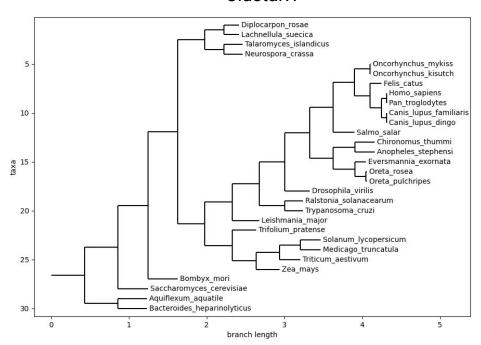


### Compared to ClustalW UPGMA

#### Ours



#### ClustalW



# Conclusions

#### Conclusions

- BioPython methods and tools greatly simplify computational biology applications
  - AlignIO for MSA
  - Phylo for building and drawing trees
  - UPGMA is NOT in BioPython good example of BioPython's strengths
- UPGMA is theoretically simple but is tedious
  - A working Python program greatly reduces this tedium

#### References

https://www.bioinformaticsalgorithms.org/

https://biopython.org/wiki/Documentation

https://github.com/biopython/biopython

https://biopython.org/docs/1.74/api/

https://en.wikipedia.org/wiki/UPGMA

http://rosalind.info/problems/ba7d/

https://www.news-medical.net/health/The-Phylogenetic-Tree-of-the-SARS-CoV-2-Virus.aspx

Our code available at: <a href="https://github.com/nyoma-diamond/BB1003\_Group8-Project">https://github.com/nyoma-diamond/BB1003\_Group8-Project</a>